

Review

Novel Biomarkers of Renal Dysfunction and Congestion in Heart Failure

Agata Zdanowicz ¹, Szymon Urban ^{1,*}, Barbara Ponikowska ², Gracjan Iwanek ¹, Robert Zymliński ¹, Piotr Ponikowski ¹ and Jan Biegus ¹

¹ Institute of Heart Diseases, Medical University, 50-556 Wrocław, Poland; agatazdanowicz@gmail.com (A.Z.); giwanek95@gmail.com (G.I.); robert.zymliński@umw.edu.pl (R.Z.); piotrponikowski@4wsk.pl (P.P.); janbiegus@gmail.com (J.B.)

² Student Scientific Organization, Institute of Heart Diseases, Medical University, 50-556 Wrocław, Poland; barbara.ponikowska@student.umw.edu.pl

* Correspondence: s.urban@umw.edu.pl; Tel.: +48-71-733-11-12

Abstract: Heart failure is a major public health problem and, despite the constantly emerging, new, effective treatments, it remains a leading cause of morbidity and mortality. Reliable tools for early diagnosis and risk stratification are crucial in the management of HF. This explains a growing interest in the development of new biomarkers related to various pathophysiological mechanisms of HF. In the course of this review, we focused on the markers of congestion and renal dysfunction in terms of their interference with cardiovascular homeostasis. Congestion is a hallmark feature of heart failure, contributing to symptoms, morbidity, and hospitalizations of patients with HF and has, therefore, become a therapeutic target in AHF. On the other hand, impaired renal function by altering the volume status contributes to the development and progression of HF and serves as a marker of an adverse clinical outcome. Early detection of congestion and an adequate assessment of renal status are essential for the prompt administration of patient-tailored therapy. This review provides an insight into recent advances in the field of HF biomarkers that could be potentially implemented in diagnosis and risk stratification of patients with HF.

Keywords: heart failure; congestion; renal dysfunction; biomarker

Citation: Zdanowicz, A.; Urban, S.; Ponikowska, B.; Iwanek, G.; Zymliński, R.; Ponikowski, P.; Biegus, J. Novel Biomarkers of Renal Dysfunction and Congestion in Heart Failure. *J. Pers. Med.* **2022**, *12*, 898. <https://doi.org/10.3390/jpm12060898>

Academic Editors: Josep Comín-Colet, Cristina Enjuanes-Grau and Luis Almenar-Bonet

Received: 14 April 2022

Accepted: 26 May 2022

Published: 29 May 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Heart failure (HF) is a multifaceted medical condition with a complex pathogenesis. Implementation of biomarkers that provide insight into a variety of biological processes involved in HF progression will potentially enable better diagnosis and prognosis of this disease [1]. Although many biomarkers have been identified, only a few have been incorporated into clinical practice, while others have yet to prove their applicability. In the course of this review, we explored the capabilities of the biomarkers related to renal dysfunction and congestion, which constitute the major pathological pathways of heart failure (HF).

Congestion is one of the most important mechanisms of heart failure (HF) development as well as being a therapeutic target during acute heart failure (AHF) episodes [1–3]. The pathophysiological processes underlying congestion in HF are multidirectional and go far beyond “simple” tissue fluid retention [2]. The HF-induced inability to maintain and control water-ion homeostasis leads to several disturbances that result in fluid accumulation and misdistribution [4,5]. Congestion is also the leading cause of hospitalizations for HF and is imminently associated with a poor outcome [6–8]. On the other hand, kidneys play a pivotal role in the management of the homeostasis and, therefore, significantly contribute to the development and reduction of fluid overload in HF.

The incorporation of biomarkers that are involved in the aforementioned mechanisms might potentially increase the complexity of clinical assessment and improve the prognosis of HF patients by personalizing treatment for different phenotypes of the disease. The classification of selected biomarkers is presented in Table 1.

Table 1. Classification of selected biomarkers based on pathophysiological pathways of heart failure.

Pathophysiological Pathway	Biomarkers
Kidney injury and dysfunction	Neutrophil gelatinase-associated lipocalin (NGAL) Kidney injury molecule-1 (KIM-1) Cystatin C (cysC) N-acetyl- β -D-glucosaminidase (NAG) Fibroblast growth factor 23 (FGF-23) Natriuresis
Congestion	Cancer antigen 125 (Ca-125) Adrenomedullin (ADM) NT-proBNP
Neurohumoral activation	Adrenomedullin (ADM) Arginine vasopressin (AVP) Copeptin (CT-proAVP) Chromogranin A (CgA)
Wide spectrum of pathological pathways	microRNA

2. Natriuretic Peptides

Natriuretic peptides, particularly B-type natriuretic peptide (BNP) and its precursor N-terminal pro-B-type natriuretic peptide (NT-proBNP), have an established role as gold-standard biomarkers in HF management [9,10]. This view is supported by ESC guidelines, which advocate their application in HF diagnosis and prognosis [11]. NPs exert a wide range of biological effects with the primary aim to maintain blood pressure–volume homeostasis. These molecules are released in response to an elevated intracardiac pressure, which correlates with HF severity [9]. Furthermore, they contribute to the identification of new HF biomarkers and, since the discovery of benefits from neprilysin inhibition, they have become a novel therapeutic target [12]. Although their significance and clinical application in HF are undeniable, they possess certain limitations. Their lack of specificity has resulted in their tendency to increase in many medical conditions (such as arrhythmias, valvular heart disease, pulmonary hypertension, pulmonary thromboembolism, and sepsis) [13,14]. It has been demonstrated that their concentration might be altered by several confounding variables, including age, kidney function, and body mass index [14]. In some groups of patients, the use of NPs is unsatisfactory. They do not serve their diagnostic purpose in patients with renal disease at any stage. Additionally, their application in guiding decongestion therapy in patients on hemodialysis is not recommended [10]. Concerning their utility as markers of congestion, it should be pointed out that, since they are released in response to stretch and cardiac transmural pressure, they solely reflect intravascular volume overload and do not serve as a proxy for interstitial fluid accumulation. The biomarkers presented in the course of this review could, perhaps, complement NPs in these “gray areas” and provide additive clinical value in the HF setting.

3. Biomarkers of Renal Dysfunction and Injury

The relationship between the heart and renal dysfunction has been widely described and investigated. Mutual cardiac and renal dysfunction worsens the prognosis of patients with the so-called cardiorenal syndrome [15]. Classical serum creatinine level assessment can be misleading. The transient creatinine rise during decongestive therapy may represent the physiological response to fluid removal (hemoconcentration) and does not unequivocally reflect the renal injury or dysfunction [16]. Although many molecules have been postulated as potential biomarkers for the assessment of cardiorenal syndrome, only a few have been thoroughly evaluated. Notable among them are neutrophil gelatinase-associated lipocalin (NGAL), kidney injury molecule-1 (KIM-1), cystatin C (cysC), N-acetyl- β -D-glucosaminidase (NAG), fibroblast growth factor 23 (FGF-23), and natriuresis. The summary of prognostic impact of selected biomarkers in HF is presented in Table 2, the predictive characteristics of the biomarkers are presented in Table 3.

Table 2. Prognostic impact of selected biomarkers in heart failure.

Biomarker	Pathophysiological Mechanism	Clinical Applicability in AHF	Clinical Applicability in CHF
NAG	Tubulointerstitial damage	Worse clinical outcome (death, worsening HF) [17].	Increase in mortality and re-hospitalization [18].
KIM-1	Tubulointerstitial damage	No impact on prognosis [19]. Correlation with worsening of renal function in AHF [20].	Increase in 10-year all-cause mortality [18]. Increase in mortality and re-hospitalization [18].
NGAL	Tubulointerstitial damage	Strong prognostic indicator of 30-day outcome [21]. Correlation with worsening of renal function in AHF [20].	Increase in all-cause mortality and rehospitalization [22].
FGF-23	Renal function mineral metabolism	Increased risk of all-cause mortality and HF hospitalization [23].	Increased risk of mortality in HFrEF [24].
Spot urine sodium	Renal function	Low urinary sodium at hospital admission is independently associated with all-cause mortality [25].	Chronically low urine, high risk of hospitalization for decompensation [26].
CA-125	Congestion	Increase in mortality and readmission [27–31].	Increase in mortality and readmission [32–34].
ADM	Residual congestion neurohumoral activation	Increased risk of all-cause mortality and HF hospitalization [35]. High risk of early hospital readmission [36].	Increased risk of all-cause mortality and HF hospitalization [35].
AVP/CT-proAVP	Neurohumoral activation	Increase in 90-day mortality. High risk of rehospitalization [37]. Increase in all-cause mortality [38].	Increase in all-cause mortality [38].
Chromogranin A	Neurohumoral activation	Increase in mortality [39].	Increase in mortality [17].
MicroRNA	Broad spectrum of mechanisms and correlations with HF prognosis, depending on the specific molecule.		

Table 3. Predictive characteristics of selected biomarkers in heart failure.

Biomarker	Cutoff Value	Specificity	Sensitivity	AUC	Clinical Value
NGAL	84 ng/mL	0.6	0.8	0.72	Mortality in CHF [40].
NAG	4.69	-	-	0.708	AKI prediction in critically ill patients [41].

KIM1	1.62	0.44	0.80	0.757	AKI in ADHF [42].
FGF-23	1180 RU/mL	0.8	0.5	0.686	28-day mortality in cardiogenic shock [43].
Spot urine sodium	50–70 mEq/L	-	-	-	Diuretic response prognosis and evaluation [44].
CA-125	32 U/mL	0.72	0.83	0.784	1-year death in CHF [45].
MR-proADM	4.6 nmol/L	0.810	0.577	0.729	Myocardial injury [46].
	3.5	0.605	0.80	0.730	Mortality at 28 days [46].
CT-proAVP	112.5 pg/mL	87%	86%	0.91	Early diagnosis of acute myocardial infarction [47].
Chromogranin A	158 pmol/L	-	-	0.697	1-year death and hospitalization in AHF [17].

Data presented in the table are considered provisional. These biomarkers are a subject of ongoing studies and official guidelines are yet to be established.

3.1. NGAL

Neutrophil gelatinase-associated lipocalin is a siderophore molecule associated with neutrophils' activation and an injury of epithelial cells, most interestingly, those located in the kidney. NGAL is also described as an iron-traffic regulator [12,48]. The molecule was originally proposed as an early marker of acute kidney injury (AKI), which anticipates the increase of serum creatinine [49].

3.1.1. Clinical Value in Diagnosis and/or Prognosis

The role of NGAL in heart failure (HF) is multidimensional. It has been proven to be independently associated with poor prognosis in terms of mortality and rehospitalizations [20–22,50–53]. An elevation of NGAL has been described as a predictor of cardio-renal syndrome 1 in the population of patients admitted to the hospital due to HF [52,53]. Moreover, increased NGAL was observed in a population of HF patients that did not present symptoms of AKI (defined as elevated serum creatinine level) [22].

It is noteworthy that there is some evidence that NGAL expression is related to heart failure, possibly as a sign of neutrophil activation and inflammation [54]. Further reports on correlations between NGAL and inflammatory mediators, such as tumor necrosis factor alpha or matrix metalloproteinase [55], might provide additional information.

3.1.2. Practical Considerations and Limitations

NGAL's superiority over creatinine resides in its independence from diuretic therapy since it reflects renal injury rather than kidney function [56]. On the other hand, further studies, such as the Acute Kidney Injury N-gal Evaluation of Symptomatic Heart Failure Study (AKINESIS) and Placebo-Controlled Randomized Study of the Selective A(1) Adenosine Receptor Antagonist Rolofylline for Patients Hospitalized with Acute Decompensated Heart Failure and Volume Overload to Assess Treatment Effect on Congestion and Renal Function (PROTECT), have not confirmed its advantage over creatinine in the prediction of an adverse outcome or worsening renal function in acute heart failure patients [57–59].

3.2. KIM-1

Kidney injury molecule-1 is a transmembrane glycoprotein expressed in the proximal tubule [60]. KIM-1 is absent in the healthy tubule and its synthesis can be induced by ischemia, toxic injury, or the process of dedifferentiation of the epithelium [61–63]. It is also related to the conversion of the epithelial cell into the phagocyte [64]. KIM-1 has been thoroughly evaluated as an early marker of AKI.

3.2.1. Clinical Value in Diagnosis and/or Prognosis

A recent meta-analysis of 14 studies and 3300 patients confirmed its value as a diagnostic tool for AKI, which reached the sensitivity of 0.74 and specificity of 0.84 [65]. KIM-1 has also been proposed as a tool for monitoring nephrotoxicity during pharmacotherapy [62,66]. Levels of KIM-1 in the symptomatic HF population have been reported to be higher than in healthy controls, regardless of the glomerular filtration status. Moreover, KIM-1 correlated with the New York Heart Association (NYHA) classification and left ventricular ejection fraction (LVEF) [67]. Different studies have shown its correlation with the n-terminal pro-brain natriuretic peptide (NT-pro-BNP) levels and increased risk of death and HF hospitalizations, independent of the initial glomerular filtration rate (GFR) [68,69]. KIM-1 was also evaluated as a prognostic factor of the all-cause mortality in chronic heart failure patients in a 10-year follow-up, but was not proven to be its independent predictor [18].

3.2.2. Practical Considerations and Limitations

KIM-1, in comparison to the rest of the tubular injury markers (NGAL, NAG, CysC), has been most thoroughly evaluated as a drug-induced nephrotoxicity monitoring tool. Conversely, its role in the mortality prognosis of HF remains unclear and is a subject of many studies.

3.3. CysC

Cystatin C is a protein produced by all nucleated cells. CysC is filtered, reabsorbed, and catabolized in the proximal tubule [70]. CysC is an alternative for the serum creatinine for the GFR calculation. The level of CysC is not affected by muscle mass or diet and depends on age, sex, and race less than creatinine. Conversely, it can be affected by smoking, inflammation, obesity, thyroid dysfunctions, glucocorticosteroids, and malignant processes [71,72]. Such characteristics prompted researchers to evaluate its value in the cardiorenal syndrome assessment.

3.3.1. Clinical Value in Diagnosis and/or Prognosis

The Heart and Soul study revealed that an increased level of CysC predicts all-cause mortality, cardiovascular events, and incidence of HF among ambulatory patients with coronary heart disease [73]. A meta-analysis, which included 10 prospective studies and 3155 patients, showed that an elevated CysC level is associated with an increased risk of all-cause mortality and rehospitalizations in the HF population, independently of creatinine and GFR [74]. CysC is reported to be increased in hypertensive heart failure with preserved ejection fraction (HFpEF) patients and, therefore, associated with left ventricular diastolic dysfunction and collagen alterations [75,76]. CysC is correlated with left ventricular diastolic diameter, left ventricular ejection fraction, and NYHA class [77]. Some studies suggest that CysC can predict in-hospital mortality [78] or 2-year cardiac events' incidence better than NT-proBNP in AHF patients [77].

3.3.2. Practical Considerations and Limitations

Although CysC can constitute a valuable complementary biomarker in the AKI diagnosis and prognosis assessment in heart failure patients, it does not provide much additional data compared to the other renal tubule injury markers. More sophisticated aspects of the cystatin C value in heart failure management require further studies.

3.4. NAG

N-acetyl- β -D-glucosaminidase is the brush border enzyme expressed in proximal tubule cells [79]. As with the aforementioned biomarkers, such as NGAL, KIM-1, and CysC, it reflects the tubular damage and has been used as an early marker of AKI [67].

3.4.1. Clinical Value in Diagnosis and/or Prognosis

An increased level of NAG in CHF patients in comparison to healthy controls has been noticed regardless of GFR. The NAG was also the predictor of death, heart transplantation, cardiovascular event, or HF hospitalization [68]. NAG was reported to be the strongest predictor among the novel renal biomarkers, NGAL and KIM-1, of 10-year all-cause mortality in HF patients [18]. Further studies confirmed the NAG increment in stable HF and revealed its associations with NT-proBNP, NYHA class, and LVEF [67]. NAG, conversely to the serum creatinine level, rose significantly after the withdrawal of diuretics during decongestion HF therapy and returned to the baseline after its re-initiation. Therefore, it can be considered as a renal marker of decongestion exhaustiveness [80]. Connotations between the NAG level and an accelerated progression of chronic kidney disease in HF patients has been described [81,82].

3.4.2. Practical Considerations and Limitations

Among novel tubular injury markers, NAG seems to be the strongest predictor of long-term mortality in CHF patients [18].

NAG has many advantages, including high sensitivity and a simple method of quantification (easily reproducible spectrophotometric enzymatic assays). As for limitations, it should be emphasized that this marker is nonspecific. Increased levels of NAG have been observed in conditions other than AKI, such as hyperthyroidism and rheumatoid diseases. Additionally, endogenous urea and other neurotoxic substances alter NAG's concentration [80,81].

3.5. FGF-23

Fibroblast growth factor-23 is a hormone dominantly secreted by osteocytes and osteoblasts in bones; however, it can also be produced by liver and heart muscle under stress [82,83]. FGF-23 plays a role in phosphate homeostasis by inducing its renal excretion. Further, FGF-23 downregulates the synthesis of vitamin D and parathormone [84]. A number of studies have shown the associations between elevated FGF-23 and left ventricular hypertrophy [83]. FGF-23 has been particularly linked to concentric hypertrophy-compensated cardiac hypertrophy without dilation [85]. This initiated a search for FGF-23 and heart failure associations.

3.5.1. Clinical Value in Diagnosis and/or Prognosis

FGF-23 was reported to be significantly higher in the HF versus healthy controls' groups [86]. Initially, small-cohort studies suggested that FGF-23 was a strong predictor of outcome in the HF group, even stronger than classical predictors such as GFR, age, brain natriuretic peptide, diabetes, left ventricular mass index, or LVEF [87,88]. Then, some controversies about the differences of FGF-23 impact on prognosis between HFpEF and heart failure with reduced ejection fraction (HFrEF) arose. Several studies revealed that FGF-23 promotes HFpEF but predicts events in HFrEF [24,89]. Higher levels of FGF-23 were associated with volume overload, more frequent failure in reaching optimal doses of angiotensin-converting enzyme inhibitors/angiotensin II receptor blockers, and increased all-cause mortality and HF hospitalizations [23]. Conversely, another study showed that FGF-23 did not present an advantage over classical parameters in predicting the outcome in HF patients [90]. The predictive value, distinctions between the role of FGF-23 in HFpEF and HFrEF, and its involvement in cardiorenal cross-talk require further investigation.

3.5.2. Practical Considerations and Limitations

FGF-23 provides novel insight into the previously not evaluated field of the heart-bone axis; however, its predictive value and the differences between its significance in HFpEF and HFrEF have to be elucidated before its implementation into clinical practice.

3.6. Urinary Sodium Excretion

European Society of Cardiology (ESC) guidelines from 2021 recommend navigating decongestive therapy based on the urine sodium excretion [44].

3.6.1. Clinical Value in Diagnosis and/or Prognosis

Indeed, a number of studies have confirmed the prognostic and diagnostic value of spot urinary sodium assessment [91,92]. In the cohort of chronic heart failure patients, a decrease in urinary sodium excretion was an ominous sign of forthcoming decompensation and hospitalization due to acute HF [26]. Furthermore, an initial low urinary sodium level and a lack of its increment during diuretic therapy were associated with poor diuretic response, the elevation of NGAL and KIM-1, and increased 1-year all-cause mortality [25,93]. The prognostic value of the urinary sodium level in acute heart failure seems to be the most powerful during the first days of hospitalization, at the beginning of the decongestive therapy, with probably limited value at discharge [94,95].

3.6.2. Practical Considerations and Limitations

Urine sodium level, at the time of writing this article, is probably the most trustworthy and widely applied biomarker of kidney functions in heart failure, excluding serum creatinine [91].

This review did not entirely cover the topic of cardiorenal cross-talk biomarkers. The role of TIMP2/IGFBP7 ratio assessment is noteworthy, as it is commercially available in the NephroCheck Test. Two multicenter studies confirmed its value in the AKI prediction, with an AUC of 0.8, superior to all existing biomarkers [12,96].

4. Exosomes and Non-Coding RNA

Exosomes are small vesicles excreted by a variety of cells. They are composed of a double layer of the lipid membrane and contain protein, lipid, mRNA, long non-coding RNA, circular RNA, microRNA, DNA, and other molecules [97,98]. Initially, exosomes were perceived as an excretion material from the cell. The turning point of exosomes investigations was the study that showed that exosomes from B-lymphocytes play an antigen-presenting role [99,100]. Currently, exosomes are considered to be a means of information's transmission between the cells. Importantly, exosomes are the carriers of non-coding RNA [101]. Exosomes are structures that protect vulnerable RNA from the harsh environment. They can stably exist in the human body fluids and plasma, which makes it a promising marker for monitoring the pathophysiological processes. The exosome and the non-coding RNA are inseparable concepts, as exosomes are the means of transport for the unstable RNA molecules. The Human Genome Project revealed that only 3% of human DNA is coding proteins. The remaining 97% was considered junk DNA. Further findings rejected this concept, associating the non-coding RNA with mainly regulatory functions [102]. Non-coding RNA can be divided according to the length of nucleic acid and its structure into smaller fractions. The most clinically and scientifically interesting of them are microRNA (miRNA), circular RNA (circRNA), and long non-coding RNA (lncRNA). The role of exosomes and RNA in HF pathophysiology is still underinvestigated. Exosomal miRNA has been proposed as a regulatory molecule in, e.g., cardiomyocyte hypertrophy [103–105], cardiac fibrosis [106], and myocardial angiogenesis [107]. Different subtypes of miRNA have been evaluated in terms of their applicability in the clinical setting. Elevated serum levels of miRNA successfully identified acute and chronic heart failure patients from healthy controls and were associated with natriuretic peptide levels, wide QRS, dilatation of the left ventricle and atrium [108,109], NYHA class [110], mortality [111], and echocardiographic parameters [112]. Little is known about the clinical utility of circRNA and lncRNA. Nevertheless, several studies suggested their important role in the regulation of pathophysiological pathways in HF,

including cardiac hypertrophy, ischemic remodeling, inflammatory process, and the regulation of intracellular Ca^{2+} level [102,113].

5. Multimarker Panels and Clustering

Heart failure is an end stage of cardiovascular diseases and a consequence of a number of pathological pathways. Multimarker evaluation can provide significant information regarding the disease phenotype by including molecules of a different pathophysiological origin [70,114]. Currently, the American College of Cardiology (ACC)/American Heart Association (AHA) heart failure guidelines suggest that assessing troponins, soluble suppression of tumorigenicity-2, and galectin-3 could enhance the natriuretic peptides' predictive value in the risk stratification [115]. One of the most important limitations of multimarker assessment is the arbitrary choice of the biomarkers. Conversely, in a recent sub-analysis of the Treatment of Preserved Cardiac Function Heart Failure with an Aldosterone Antagonist (TOPCAT) study, the authors implemented a machine learning model, which automatically selected significant molecules (out of 49 available) to predict the outcome [116]. Importantly, the authors implemented clustering techniques to analyze the heterogeneity of the biomarkers. Clustering is the unsupervised machine learning technique that divides a set of numerical variables into smaller groups based on their similarity. Clusters are composed of variables that are consistent with each other but not with other clusters [116]. In the recent study, 577 urine peptides from heart failure patients and matched controls were analyzed [117]. Algorithm-based clustering automatically divided participants based on their urinary peptides profiles; 83% of non-HF controls were allocated into cluster 1, while 65% of HF patients were in cluster 2. This experiment revealed the natural, machine-detectable differences between HF and non-HF patients' urinary profiles. Moreover, the study showed clustering techniques' ability to automatically reveal the group of proteins that can present the diagnostic and therapeutic potential.

6. Congestions

6.1. Cancer Antigen 125 (CA-125)

Cancer antigen 125 (CA-125), otherwise known as mucin 16, is a membrane glycoprotein synthesized by mesothelial cells. Traditionally, its use has been restricted to screening and therapeutic monitoring of ovarian cancer [118,119]. However, the CA-125 level has been found to be upregulated in other nonmalignant processes, such as heart failure. A growing body of evidence has implicated CA-125 in the pathophysiological processes underlying HF [120]. In the latter context, the glycoprotein is released in response to extravascular congestion and cytokine-induced inflammation as well as myocardial stress and injury [121]. Taking all this into consideration, CA-125, as a surrogate for fluid overload and inflammation in HF patients, emerges as a potential novel biomarker of this pathology [118,119].

6.1.1. Clinical Value in Diagnosis and/or Prognosis

Several studies have established an association between the CA-125 concentration and clinical manifestation of congestion (including peripheral edema and serosal effusion) in the HF population [27–34,122–129]. The elevation of CA-125 is positively correlated with the severity of an extravascular congestion and is consistent with clinical, hemodynamic, and echocardiographic parameters of fluid overload [27,124].

Patients with more severe congestion had, on average, higher level of CA-125 (mean difference of 54.8 U/mL) than patients without congestion [31]. Among patients presenting to an emergency department with an acute dyspnea of uncertain origin, the highest CA-125 values were recorded in patients with AHF, particularly in those with a worsening heart failure phenotype [125].

Recently, there has been a substantial increase in the number of studies investigating a prognostic value of CA-125 in risk stratification of the HF population. Results of the various trials demonstrated that the elevation of the CA-125 level was positively correlated with an increased risk of mortality and readmission for HF. CA-125 remained a significant prognostic predictor of poor prognosis, even after adjustment for the relevant baseline covariates [28–30,32–34,126–128]. CA-125 had a similar positive predictive value as NT-proBNP in AHF but was superior to NT-proBNP when effusion was present [129]. Key findings from a recent sub-analysis of BIOSTAT-CHF are in line with these findings. It was reported that CA125 was positively correlated with the clinical parameters of congestion (namely, peripheral edema, hepatomegaly, orthopnea, etc.), and a higher level of plasma CA125 was associated with an increased risk of a 1-year all-cause mortality and the composite of death/HF readmission [123]. In the group of AHF patients, those who had a high CA-125 level and low concentration of NT-proBNP at admission had significantly worse prognoses than patients with low CA-125 levels and low NT-proBNP. Likewise, those with high NT-proBNP and high CA125 levels had the least favorable prognoses [30]. Thus, it can be inferred that the implementation of CA125 in conjunction with NT-proBNP is more accurate in risk stratification of patients with AHF than the conventional use of NT-proBNP alone.

6.1.2. Role of CA-125 in Heart Failure Treatment

The substantial number of studies have recently demonstrated that the concentration of CA-125 changes with the degree of fluid overload and, thus, may reflect treatment-induced alterations in a patient's volume status [31,32,130–132]. Considering this finding, CA125 emerges as a potential tool to guide effective decongestion therapy. The authors of the CHANCE-HF trial concluded that CA-125-tailored diuretic therapy was superior to a conventional one in reducing the risk of the composite of 1-year death or AHF readmission [27]. CA-125-guided diuretic therapy was associated with a significant reduction in death and readmission for AHF at 30 days. The CA-125-guided group exhibited a considerably greater improvement in renal function (increase in GFR) in comparison to the standard treatment group [133]. Given the long half-life (over a week) of CA-125, it does not provide any information about acute response to therapy [132]. It might, however, be a reasonable approach to measure CA-125 concentrations at admission for HF as well as at least 7 days after the initial examination.

6.1.3. Practical Considerations and Limitations

CA-125 is associated with congestion, right-sided HF parameters, and an increased risk of adverse clinical events in AHF, beyond standard prognostic factors such as natriuretic peptides [133]. Broad availability, standardized measurement, and low cost make it a compelling candidate for routine use in decompensated HF. Variations in CA-125 concentrations according to the clinical situation make it a potential tool for both monitoring and guiding HF treatment following a decompensated HF event [133]. There are several limitations that currently prevent CA-125 from being used in the HF setting. These include an insufficient understanding of CA-125 biology, a lack of an optimal cut-off value, and indefinite data from large multicenter trials [133]. Hence, future large-scale research is required to validate the role of CA-125 in heart failure.

7. Neurohumoral Activation

Neurohormonal activation (expressed by stimulation of the sympathetic system and RAAS) is believed to be a central mechanism underlying HF pathophysiology. Reduced cardiac output induced by myocardial dysfunction activates the neurohormonal axis to maintain hemodynamic stability. Although activation of this system is presumed to be compensatory, over the long term, it contributes to further deterioration of cardiac systolic function and development of heart failure symptoms [134,135]. While a substantial

body of evidence points to a prognostic role of neurohumoral markers, they are not measured in daily practice. It stems from the fact that the treatment of HF is based on the use of angiotensin-converting enzyme inhibitors and beta-adrenergic receptor blockers, which can significantly modulate plasma concentrations of these hormones, thereby interfering with their interpretation and reducing their predictive value in risk stratification [135]. Therefore, further research is required to identify a stable, yet sensitive, and relatively easy to measure biomarker.

7.1. Adrenomedullin

Adrenomedullin (ADM) is a newly discovered vasoactive and natriuretic peptide hormone involved in cardiorenal regulation. ADM is widely distributed in the cardiovascular system, kidneys, and adrenal glands [135–137]. This hormone is released by endothelial and vascular smooth muscles in response to fluid overload and endothelial barrier disruption. ADM-induced preservation of vascular integrity contributes to the reduction in tissue congestion. Conversely, the inhibition of the ADM results in vascular leakage and systemic/pulmonary edema development [137,138]. Hence, elevated levels of ADM are frequently observed among patients with HF or septic shock. This finding stems from the fact that vascular leakage and organ hypoperfusion are commonly encountered in these two pathologies [138,139]. It stimulates vasodilatation, positive inotropic action, and cardiac hypertrophy inhibition. Thus, it decreases blood pressure and increases blood flow [140–143]. It was observed that even relatively low doses of ADM produced significant vascular dilatation. This finding suggests that, in conditions such as HF, ADM might be in the range that is capable of affecting vascular tone [143,144]. Additionally, by suppressing RAAS, ADM exerts diuretic and natriuretic effects. Its action contributes to an improvement in the glomerular filtration rate [145–147]. Given the cardioprotective role of ADM, it has been suggested that it has a potential to become a therapeutic target in patients with HF [148].

7.1.1. Clinical Value in Diagnosis and/or Prognosis

ADM might be utilized as a marker of congestion in patients with a new onset and worsening HF [35,36,149–153]. It has been observed that patients with a high concentration of this vasoactive peptide exhibited more severe signs of congestion (edema, orthopnea, and elevated jugular venous pressure) [36]. The association between a degree of fluid overload and bio-ADM concentration at baseline remained significant even after adjustment for a multivariable model [35]. A strong association between the ADM level and the degree of hemodynamic instability, impaired cardiac function, and prognosis in cardiogenic shock was demonstrated in a recent cohort study. Results revealed that a high level of ADM (observed in a non-survivor group) was positively correlated with an impaired cardiac index, mean arterial pressure, and central venous pressure [35]. MR-proADM was superior to BNP and troponin for predicting 90-day all-cause mortality in AHF patients with acute dyspnea [149]. Findings of the PROTECT trial indicated that, among patients hospitalized for AHF, elevation of bio-ADM at discharge reflected residual congestion and was related to an increased risk of early rehospitalization [153]. When combined with BNP and NT-proBNP, Mr-proADM provided significant incremental predictive value for 90-day mortality [150].

7.1.2. Practical Considerations and Limitations

The use of MR-proADM in the diagnosis of AHF is limited due to its low specificity. MR-proADM is elevated in various medical conditions (systemic hypertension, myocardial infarction, kidney failure, or sepsis) [151]. It is also crucial to point out that the major disadvantage of MR-proADM is the fact that its level is influenced by a patient's baseline covariates. Higher concentrations of MR-proADM were reported in female elderly patients, kidney disease, and systolic dysfunction. The biomarker's level decreased with a

higher body mass index [152]. Further large, multi-center studies are required to establish the role of this biomarker and determine its value in conjunction with other biomarkers.

7.2. Arginine Vasopressin and Copeptin

Arginine vasopressin (AVP) is an antidiuretic and vasoconstrictive peptide hormone, released in response to hyperosmolality and hypovolemia. It plays a crucial role in hemodynamics and osmoregulation [154].

Key functions of AVP include solute-free water reabsorption in kidney tubules, an increase in peripheral vascular resistance, and a consequential rise in arterial blood pressure [155,156]. Moreover, AVP is a crucial component of the endocrine stress response, triggering ACTH and cortisol release [154]. Due to the poor stability and short half-life (15–20 min) of AVP, copeptin (CT-proAVP, C-terminal segment of pre-provasopressin) was introduced into clinical practice as a reliable surrogate of this hormone [154,155].

7.2.1. Clinical Value in Diagnosis and/or Prognosis

The evidence of AVP elevation in HF patients has been well documented in the literature [157–160]. Among patients with HFpEF, AVP was independently associated with LV hypertrophy and a higher risk of death or HF readmissions. In line with the previous findings, copeptin has been found to be a strong predictor of poor prognosis in the HF population [37]. The BACH study indicated that patients with elevated copeptin levels (especially those with hyponatremia) had significantly higher 90-day mortality and a higher risk of rehospitalization [38]. Furthermore, a meta-analysis, which included 10 prospective cohort studies, revealed that an increased concentration of copeptin was positively correlated with an all-cause mortality in the HF population. The prognostic role of copeptin was found to be equivalent to NT-proBNP for all-cause mortality in patients with HF [38]. Copeptin provided independent prognostic information in severe HF, although its prognostic impact was inferior to NT-proBNP [161].

7.2.2. Practical Considerations and Limitations

Although the biological effect of this biomarker has been widely explored, the optimal use of copeptin in the HF setting remains a question of debate. Copeptin is a sensitive and easily measurable surrogate of AVP, yet it is more stable and has a longer half-life. CT-proAVP exhibits low specificity. The elevation of AVP/CT-proAVP has been associated with a number of diseases, particularly those that are characterized by acute stress [162]. The proper assessment of copeptin's results requires knowledge about confounding factors that interfere with its concentration. Copeptin has been shown to be increased in males and to be associated with a decreased glomerular filtration rate, probably as a result of decreased renal copeptin clearance [154,155]. This marker is noteworthy for its potential value in identifying high-risk patients in a critical condition and, thus, implementing individualized patient care.

7.3. Chromogranin A

Widely recognized as a major marker of neuroendocrine tumor (NET), chromogranin A (CgA) is an acidic protein and a pro-hormone of an active particle, which potentially exerts a biological effect in CHF [163,164]

7.3.1. Clinical Value in Diagnosis and/or Prognosis

A considerable interest in this protein stems from the fact that CgA has been found to be related to the clinical deterioration and higher risk of mortality in patients with AHF and CHF [39,165,166]. The concentration of CgA was measured in a group of 160 patients with CHF to evaluate the association between the CgA level and HF severity (based on the NYHA scale). The results demonstrated that class IV NYHA patients had

the highest level of CgA (median 545.0 ng·mL⁻¹) while class I had the lowest concentration of this protein (median 109.7 ng·mL⁻¹). Furthermore, it was concluded that CgA might be a predictive factor for mortality in HF [17]. CgA was found to have comparable prognostic value to that of NT-proBNP in AHF patients [163]. High concentrations of CgA were independently associated with 1-year death and hospitalization for heart failure [163].

7.3.2. Practical Considerations and Limitations

Data regarding CgA measurement in cardiology are limited. Its role has been investigated only in studies with a small number of patients. In the current state of research, CgA plasma measurement as a biomarker in heart failure is still being explored and cannot be recommended for general use [163]. Furthermore, complex and extensive CgA processing has hampered its use as a routine biomarker due to methodological problems with its measurement [165].

8. Conclusions

With a growing prevalence and incidence of HF, novel and reliable diagnostic tools are thoroughly investigated. Currently, B-type natriuretic peptide (BNP) and N-terminal proBNP are the most recognized biomarkers for the diagnosis and treatment of HF; however, they do have certain limitations. They do not serve their prognostic role in some group of patients such as in those with renal failure and are not specific for HFpEF patients. Therefore, further research to explore new markers that would provide additional prognostic information, especially in areas uncovered by classic biomarkers, is required. Novel biomarkers certainly comprise those related to congestion and renal dysfunction, as, in fact, these pathological pathways are closely related to the development and progression of HF. Although it is important to note that the individual clinical value of the aforementioned biomarkers in diagnosis and prognosis is limited due to their lack of specificity, as such, the future of biomarker application in HF management relies on a multimarker panel and clustering strategies that would include a more specific combination of biomarkers representing different pathophysiological processes underlying HF. Multimarker panels' analysis represents the great potential in prediction, risk stratification, and therapy tailoring in cardiovascular disease. A combination of various biomarkers reflects the cross talk of different pathophysiological pathways, including cardiac remodeling, inflammatory process, renal dysfunction, and neurohormonal activation, all of which play a significant role in heart failure. Machine learning techniques, such as clustering, can constitute helpful tools for elucidating the heterogeneity of the heart failure population and distinguishing important parameters from the immense amounts of data.

Author Contributions: All authors contributed significantly to this article. All authors have seen and approved the final version of the manuscript.

Funding: This research was financially supported by subsidy no. SUB.e190.21.105 and SUBZ.A460.22.055 from the Institute of Heart Diseases, Wroclaw Medical University, Poland.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Miller, W.L. Fluid Volume Overload and Congestion in Heart Failure. *Circulation: Heart Fail.* **2016**, *9*, 90–95. <https://doi.org/10.1161/CIRCHEARTFAILURE.115.002922>.

2. Boorsma, E.M.; ter Maaten, J.M.; Damman, K.; Dinh, W.; Gustafsson, F.; Goldsmith, S.; Burkhoff, D.; Zannad, F.; Udelson, J.E.; Voors, A.A. Congestion in Heart Failure: A Contemporary Look at Physiology, Diagnosis and Treatment. *Nat. Rev. Cardiol.* **2020**, *17*, 641–655. <https://doi.org/10.1038/s41569-020-0379-7>.
3. Sokolska, J.M.; Sokolski, M.; Zymliński, R.; Biegus, J.; Siwołowski, P.; Nawrocka-Millward, S.; Swoboda, K.; Gajewski, P.; Jankowska, E.A.; Banasiak, W.; et al. Distinct Clinical Phenotypes of Congestion in Acute Heart Failure: Characteristics, Treatment Response, and Outcomes. *ESC Heart Fail.* **2020**, *7*, 3830–3840. <https://doi.org/10.1002/ehf2.12973>.
4. Zymliński, R.; Biegus, J.; Ponikowski, P. Not All Fluid Overloads Are the Same: Some Practical Considerations for Better Decongestion. *Eur. J. Heart Fail.* **2021**, *23*, 1106–1109. <https://doi.org/10.1002/ehf.2187>.
5. Fudim, M.; Hernandez, A.F.; Felker, G.M. Role of Volume Redistribution in the Congestion of Heart Failure. *J. Am. Heart Assoc.* **2017**, *6*, e006817. <https://doi.org/10.1161/JAHA.117.006817>.
6. Ambrosy, A.P.; Cerbin, L.P.; Armstrong, P.W.; Butler, J.; Coles, A.; DeVore, A.D.; Dunlap, M.E.; Ezekowitz, J.A.; Felker, G.M.; Fudim, M.; et al. Body Weight Change During and After Hospitalization for Acute Heart Failure: Patient Characteristics, Markers of Congestion, and Outcomes: Findings From the ASCEND-HF Trial. *JACC Heart Fail.* **2017**, *5*, 1–13. <https://doi.org/10.1016/J.JCHF.2016.09.012>.
7. Voors, A.A.; Davison, B.A.; Teerlink, J.R.; Felker, G.M.; Cotter, G.; Filippatos, G.; Greenberg, B.H.; Pang, P.S.; Levin, B.; Hua, T.A.; et al. Diuretic Response in Patients with Acute Decompensated Heart Failure: Characteristics and Clinical Outcome—An Analysis from RELAX-AHF. *Eur. J. Heart Fail.* **2014**, *16*, 1230–1240. <https://doi.org/10.1002/ehf.170>.
8. Gheorghiade, M.; Follath, F.; Ponikowski, P.; Barsuk, J.H.; Blair, J.E.A.; Cleland, J.G.; Dickstein, K.; Drazner, M.H.; Fonarow, G.C.; Jaarsma, T.; et al. Assessing and Grading Congestion in Acute Heart Failure: A Scientific Statement from the Acute Heart Failure Committee of the Heart Failure Association of the European Society of Cardiology and Endorsed by the European Society of Intensive Care Medicine. *Eur. J. Heart Fail.* **2010**, *12*, 423–433. <https://doi.org/10.1093/eurjhf/hfq045>.
9. Januzzi, J.L. Natriuretic Peptides as Biomarkers in Heart Failure. *J. Investig. Med.* **2013**, *61*, 950–955. <https://doi.org/10.2310/JIM.0b013e3182946b69>.
10. Koratala, A.; Kazory, A. Natriuretic Peptides as Biomarkers for Congestive States: The Cardiorenal Divergence. *Dis. Markers* **2017**, *2017*, 1454986. <https://doi.org/10.1155/2017/1454986>.
11. Ponikowski, P.; Voors, A.A.; Anker, S.D.; Bueno, H.; Cleland, J.G.F.; Coats, A.J.S.; Falk, V.; González-Juanatey, J.R.; Harjola, V.-P.; Jankowska, E.A.; et al. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC) Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. *Eur. Heart J.* **2016**, *37*, 2129–2200. <https://doi.org/10.1093/eurheartj/ehw128>.
12. Ibrahim, N.E.; Januzzi, J.L. Established and Emerging Roles of Biomarkers in Heart Failure. *Circ. Res.* **2018**, *123*, 614–629. <https://doi.org/10.1161/CIRCRESAHA.118.312706>.
13. Burke, M.A.; Cotts, W.G. Interpretation of B-type natriuretic peptide in cardiac disease and other comorbid conditions. *Heart Fail. Rev.* **2007**, *12*, 23–36. <https://doi.org/10.1007/s10741-007-9002-9>.
14. Schou, M.; Gustafsson, F.; Kistorp, C.N.; Corell, P.; Kjaer, A.; Hildebrandt, P.R. Effects of Body Mass Index and Age on N-Terminal Pro-Brain Natriuretic Peptide Are Associated with Glomerular Filtration Rate in Chronic Heart Failure Patients. *Clin. Chem.* **2007**, *53*, 1928–1935. <https://doi.org/10.1373/clinchem.2006.084426>.
15. Ronco, C.; McCullough, P.; Anker, S.D.; Anand, I.; Aspromonte, N.; Bagshaw, S.M.; Bellomo, R.; Berl, T.; Bobek, I.; Cruz, D.N.; et al. Cardio-Renal Syndromes: Report from the Consensus Conference of the Acute Dialysis Quality Initiative. *Eur. Heart J.* **2010**, *31*, 703–711. <https://doi.org/10.1093/EURHEARTJ/EHP507>.
16. Costanzo, M.R. The Cardiorenal Syndrome in Heart Failure. *Heart Fail. Clin.* **2020**, *16*, 81–97. <https://doi.org/10.1016/j.hfc.2019.08.010>.
17. Ceconi, C.; Ferrari, R.; Bachetti, T.; Opasich, C.; Volterrani, M.; Colombo, B.; Parrinello, G.; Corti, A. Chromogranin A in Heart Failure; a Novel Neurohumoral Factor and a Predictor for Mortality. *Eur. Heart J.* **2002**, *23*, 967–974. <https://doi.org/10.1053/euhj.2001.2977>.
18. Strack, C.; Bauer, S.; Hubauer, U.; Ücer, E.; Birner, C.; Luchner, A.; Maier, L.; Jungbauer, C. N-Acetyl-β-D-Glucosaminidase Is Predictive of Mortality in Chronic Heart Failure: A 10-Year Follow-Up. *Biomark Med.* **2021**, *15*, 1143–1153. <https://doi.org/10.2217/BMM-2020-0366>.
19. Grodin, J.L.; Perez, A.L.; Wu, Y.; Hernandez, A.F.; Butler, J.; Metra, M.; Felker, G.M.; Voors, A.A.; McMurray, J.J.; Armstrong, P.W.; et al. Circulating Kidney Injury Molecule-1 Levels in Acute Heart Failure. *JACC Heart Fail.* **2015**, *3*, 777–785. <https://doi.org/10.1016/j.jchf.2015.06.006>.
20. Sokolski, M.; Zymliński, R.; Biegus, J.; Siwołowski, P.; Nawrocka-Millward, S.; Todd, J.; Yerramilli, M.R.; Estis, J.; Jankowska, E.A.; Banasiak, W.; et al. Urinary Levels of Novel Kidney Biomarkers and Risk of True Worsening Renal Function and Mortality in Patients with Acute Heart Failure. *Eur. J. Heart Fail.* **2017**, *19*, 760–767. <https://doi.org/10.1002/EJHF.746>.
21. Maisel, A.S.; Mueller, C.; Fitzgerald, R.; Brikhan, R.; Hiestand, B.C.; Iqbal, N.; Clopton, P.; van Veldhuisen, D.J. Prognostic Utility of Plasma Neutrophil Gelatinase-Associated Lipocalin in Patients with Acute Heart Failure: The NGAL Evaluation Along with B-Type Natriuretic Peptide in Acutely Decompensated Heart Failure (GALLANT) Trial. *Eur. J. Heart Fail.* **2011**, *13*, 846–851. <https://doi.org/10.1093/EURJHF/HFR087>.

22. Damman, K.; Masson, S.; Hillege, H.L.; Maggioni, A.P.; Voors, A.A.; Opasich, C.; van Veldhuisen, D.J.; Montagna, L.; Cosmi, F.; Tognoni, G.; et al. Clinical Outcome of Renal Tubular Damage in Chronic Heart Failure. *Eur. Heart J.* **2011**, *32*, 2705–2712. <https://doi.org/10.1093/EURHEARTJ/EHR190>.
23. ter Maaten, J.M.; Voors, A.A.; Damman, K.; van der Meer, P.; Anker, S.D.; Cleland, J.G.; Dickstein, K.; Filippatos, G.; van der Harst, P.; Hillege, H.L.; et al. Fibroblast Growth Factor 23 Is Related to Profiles Indicating Volume Overload, Poor Therapy Optimization and Prognosis in Patients with New-Onset and Worsening Heart Failure. *Int. J. Cardiol.* **2018**, *253*, 84–90. <https://doi.org/10.1016/J.IJCARD.2017.10.010>.
24. Koller, L.; Kleber, M.E.; Brandenburg, V.M.; Goliash, G.; Richter, B.; Sulzgruber, P.; Scharnagl, H.; Silbernagel, G.; Grammer, T.B.; Delgado, G.; et al. Fibroblast Growth Factor 23 Is an Independent and Specific Predictor of Mortality in Patients With Heart Failure and Reduced Ejection Fraction. *Circ. Heart Fail.* **2015**, *8*, 1059–1067. <https://doi.org/10.1161/CIRCHEARTFAILURE.115.002341>.
25. Damman, K.; ter Maaten, J.M.; Coster, J.E.; Krikken, J.A.; van Deursen, V.M.; Krijnen, H.K.; Hofman, M.; Nieuwland, W.; van Veldhuisen, D.J.; Voors, A.A.; et al. Clinical Importance of Urinary Sodium Excretion in Acute Heart Failure. *Eur. J. Heart Fail.* **2020**, *22*, 1438–1447. <https://doi.org/10.1002/EJHF.1753>.
26. Martens, P.; Dupont, M.; Verbrugge, F.H.; Damman, K.; Degryse, N.; Nijst, P.; Reynders, C.; Penders, J.; Tang, W.H.W.; Testani, J.; et al. Urinary Sodium Profiling in Chronic Heart Failure to Detect Development of Acute Decompensated Heart Failure. *JACC Heart Fail* **2019**, *7*, 404–414. <https://doi.org/10.1016/J.JCHF.2019.02.011>.
27. Núñez, J.; Llacer, P.; Bertomeu-González, V.; Bosch, M.J.; Merlos, P.; García-Blas, S.; Montagud, V.; Bodí, V.; Bertomeu-Martínez, V.; Pedrosa, V.; et al. Carbohydrate Antigen-125–Guided Therapy in Acute Heart Failure. *JACC Heart Fail.* **2016**, *4*, 833–843. <https://doi.org/10.1016/j.jchf.2016.06.007>.
28. Núñez, J.; Sanchis, J.; Bodí, V.; Fonarow, G.C.; Núñez, E.; Bertomeu-González, V.; Miñana, G.; Consuegra, L.; Bosch, M.J.; Carratalá, A.; et al. Improvement in Risk Stratification with the Combination of the Tumour Marker Antigen Carbohydrate 125 and Brain Natriuretic Peptide in Patients with Acute Heart Failure. *Eur. Heart J.* **2010**, *31*, 1752–1763. <https://doi.org/10.1093/eurheartj/ehq142>.
29. Mansour, I.N.; Napan, S.; Tarek Alahdab, M.; Stamos, T.D. Carbohydrate Antigen 125 Predicts Long-Term Mortality in African American Patients with Acute Decompensated Heart Failure. *Congest. Heart Fail.* **2010**, *16*, 15–20. <https://doi.org/10.1111/j.1751-7133.2009.00110.x>.
30. Yoon, J.Y.; Yang, D.H.; Cho, H.J.; Kim, N.K.; Kim, C.-Y.; Son, J.; Roh, J.-H.; Jang, S.Y.; Bae, M.H.; Lee, J.H.; et al. Serum Levels of Carbohydrate Antigen 125 in Combination with N-Terminal pro-Brain Natriuretic Peptide in Patients with Acute Decompensated Heart Failure. *Korean J. Intern. Med.* **2019**, *34*, 811–818. <https://doi.org/10.3904/kjim.2017.313>.
31. Li, K.H.C.; Gong, M.; Li, G.; Baranchuk, A.; Liu, T.; Wong, M.C.S.; Jesuthasan, A.; Lai, R.W.C.; Lai, J.C.L.; Lee, A.P.W.; et al. Cancer Antigen-125 and Outcomes in Acute Heart Failure: A Systematic Review and Meta-Analysis. *Heart Asia* **2018**, *10*, e011044. <https://doi.org/10.1136/heartasia-2018-011044>.
32. Nägele, H.; Bahlo, M.; Klapdor, R.; Schaeperkoetter, D.; Rödiger, W. CA 125 and Its Relation to Cardiac Function. *Am. Heart J.* **1999**, *137*, 1044–1049. [https://doi.org/10.1016/S0002-8703\(99\)70360-1](https://doi.org/10.1016/S0002-8703(99)70360-1).
33. D'Aloia, A.; Faggiano, P.; Aurigemma, G.; Bontempi, L.; Ruggeri, G.; Metra, M.; Nodari, S.; Dei Cas, L. Serum Levels of Carbohydrate Antigen 125 in Patients with Chronic Heart Failure. *J. Am. Coll. Cardiol.* **2003**, *41*, 1805–1811. [https://doi.org/10.1016/S0735-1097\(03\)00311-5](https://doi.org/10.1016/S0735-1097(03)00311-5).
34. Becerra-Muñoz, V.M.; Sobrino-Márquez, J.M.; Rangel-Sousa, D.; Fernández-Cisnal, A.; Lage-Gallé, E.; García-Pinilla, J.M.; Martínez-Martínez, Á.; de Teresa-Galván, E. Long-Term Prognostic Role of CA-125 in Noncongestive Patients Undergoing a Cardiac Transplantation. *Biomark Med.* **2017**, *11*, 239–243. <https://doi.org/10.2217/bmm-2016-0247>.
35. Tolppanen, H.; Rivas-Lasarte, M.; Lassus, J.; Sans-Roselló, J.; Hartmann, O.; Lindholm, M.; Arrigo, M.; Tarvasmäki, T.; Köber, L.; Thiele, H.; et al. Adrenomedullin: A Marker of Impaired Hemodynamics, Organ Dysfunction, and Poor Prognosis in Cardiogenic Shock. *Ann. Intensive Care* **2017**, *7*, 6. <https://doi.org/10.1186/s13613-016-0229-2>.
36. ter Maaten, J.M.; Kremer, D.; Demissei, B.G.; Struck, J.; Bergmann, A.; Anker, S.D.; Ng, L.L.; Dickstein, K.; Metra, M.; Samani, N.J.; et al. Bio-Adrenomedullin as a Marker of Congestion in Patients with New-Onset and Worsening Heart Failure. *Eur. J. Heart Fail.* **2019**, *21*, 732–743. <https://doi.org/10.1002/ehf.1437>.
37. Schill, F.; Timpka, S.; Nilsson, P.M.; Melander, O.; Enhörning, S. Copeptin as a Predictive Marker of Incident Heart Failure. *ESC Heart Fail.* **2021**, *8*, 3180–3188. <https://doi.org/10.1002/ehf2.13439>.
38. Maisel, A.; Xue, Y.; Shah, K.; Mueller, C.; Nowak, R.; Peacock, W.F.; Ponikowski, P.; Mockel, M.; Hogan, C.; Wu, A.H.B.; et al. Increased 90-Day Mortality in Patients with Acute Heart Failure with Elevated Copeptin: Secondary Results from the Biomarkers in Acute Heart Failure (BACH) Study. *Circ. Heart Fail* **2011**, *4*, 613–620. <https://doi.org/10.1161/CIRCHEARTFAILURE.110.960096>.
39. Ottesen, A.H.; Carlson, C.R.; Louch, W.E.; Dahl, M.B.; Sandbu, R.A.; Johansen, R.F.; Jarstadmarken, H.; Bjørås, M.; Høiseth, A.D.; Brynildsen, J.; et al. Glycosylated Chromogranin A in Heart Failure: Implications for Processing and Cardiomyocyte Calcium Homeostasis. *Circ. Heart Fail.* **2017**, *10*. <https://doi.org/10.1161/CIRCHEARTFAILURE.116.003675>.
40. Avci, A.; Ozturk, B.; Demir, K.; Akyürek, F.; Altunkeser, B.B. The Prognostic Utility of Plasma NGAL Levels in ST Segment Elevation in Myocardial Infarction Patients. *Adv. Prev. Med.* **2020**, *2020*, 4637043. <https://doi.org/10.1155/2020/4637043>.

41. Hou, Y.; Deng, Y.; Hu, L.; He, L.; Yao, F.; Wang, Y.; Deng, J.; Xu, J.; Wang, Y.; Xu, F.; Chen, C. Assessment of 17 clinically available renal biomarkers to predict acute kidney injury in critically ill patients. *J. Transl. Intern. Med.* **2021**, *9*, 273–284. <https://doi.org/10.2478/jtim-2021-0047>.
42. Yang, C.-H.; Chang, C.-H.; Chen, T.-H.; Fan, P.-C.; Chang, S.-W.; Chen, C.-C.; Chu, P.-H.; Chen, Y.-T.; Yang, H.-Y.; Yang, C.-W.; Chen, Y.-C. Combination of Urinary Biomarkers Improves Early Detection of Acute Kidney Injury in Patients With Heart Failure. *Circ. J.* **2016**, *80*, 1017–1023. <https://doi.org/10.1253/circj.CJ-15-0886>.
43. Pöss, J.; Mahfoud, F.; Seiler, S.; Heine, G.H.; Fliser, D.; Böhm, M.; Link, A. FGF-23 is associated with increased disease severity and early mortality in cardiogenic shock. *Eur. Heart J. Acute Cardiovasc. Care* **2013**, *2*, 211–218. <https://doi.org/10.1177/2048872613494025>.
44. McDonagh, T.A.; Metra, M.; Adamo, M.; Gardner, R.S.; Baumbach, A.; Böhm, M.; Burri, H.; Butler, J.; Čelutkienė, J.; Chioncel, O.; et al. 2021 ESC Guidelines for the Diagnosis and Treatment of Acute and Chronic Heart Failure. *Eur. Heart J.* **2021**, *42*, 3599–3726. <https://doi.org/10.1093/EURHEARTJ/EHAB368>.
45. Ordu, S.; Ozhan, H.; Alemdar, R.; Aydin, M.; Caglar, O.; Yuksel, H.; Kandis, H. Carbohydrate antigen-125 and N-terminal pro-brain natriuretic peptide levels: Compared in heart-failure prognostication. *Tex. Heart Inst. J.* **2012**, *39*, 30–35.
46. Lundberg, O.H.M.; Bergenzaun, L.; Rydén, J.; Rosenqvist, M.; Melander, O.; Chew, M.S. Adrenomedullin and endothelin-1 are associated with myocardial injury and death in septic shock patients. *Crit. Care* **2016**, *20*, 178. <https://doi.org/10.1186/s13054-016-1361-y>.
47. el shafey, W.E.D.H.; Ahmedy, I.A. Diagnostic values of Copeptin as a novel cardiac marker in relation to traditional markers in acute myocardial infarction. *Clin. Trials Regul. Sci. Cardiol.* **2016**, *19*, 13–19. <https://doi.org/10.1016/j.ctrsc.2016.05.003>.
48. Cruz, D.N.; Fard, A.; Clementi, A.; Ronco, C.; Maisel, A. Role of Biomarkers in the Diagnosis and Management of Cardio-Renal Syndromes. *Semin. Nephrol.* **2012**, *32*, 79–92. <https://doi.org/10.1016/J.SEMNEPHROL.2011.11.011>.
49. Cernaro, V.; Bolignano, D.; Donato, V.; Lacquaniti, A.; Buemi, A.; Crasci, E.; Lucisano, S.; Buemi, M. NGAL Is a Precocious Marker of Therapeutic Response. *Curr. Pharm. Des.* **2011**, *17*, 844–849. <https://doi.org/10.2174/138161211795428939>.
50. van Deursen, V.M.; Damman, K.; Voors, A.A.; van der Wal, M.H.; Jaarsma, T.; van Veldhuisen, D.J.; Hillege, H.L. Prognostic Value of Plasma Neutrophil Gelatinase-Associated Lipocalin for Mortality in Patients with Heart Failure. *Circ. Heart Fail.* **2014**, *7*, 35–42. <https://doi.org/10.1161/CIRCHEARTFAILURE.113.000242>.
51. Nawrocka-Millward, S.; Biegus, J.; Hurkacz, M.; Guzik, M.; Rosiek-Biegus, M.; Jankowska, E.A.; Ponikowski, P.; Zymliński, R. Differences in the Biomarker Profile of De Novo Acute Heart Failure versus Decompensation of Chronic Heart Failure. *Bio-molecules* **2021**, *11*, 1701. <https://doi.org/10.3390/BIOM11111701>.
52. Aghel, A.; Shrestha, K.; Mullens, W.; Borowski, A.; Tang, W.H.W. Serum Neutrophil Gelatinase-Associated Lipocalin (NGAL) in Predicting Worsening Renal Function in Acute Decompensated Heart Failure. *J. Card. Fail.* **2010**, *16*, 49–54. <https://doi.org/10.1016/J.CARDFAIL.2009.07.003>.
53. Alvelos, M.; Pimentel, R.; Pinho, E.; Gomes, A.; Lourenço, P.; Teles, M.J.; Almeida, P.; Guimarães, J.T.; Bettencourt, P. Neutrophil Gelatinase-Associated Lipocalin in the Diagnosis of Type 1 Cardio-Renal Syndrome in the General Ward. *Clin. J. Am. Soc. Nephrol.* **2011**, *6*, 476–481. <https://doi.org/10.2215/CJN.06140710>.
54. Helanova, K.; Spinar, J.; Parenica, J. Diagnostic and Prognostic Utility of Neutrophil Gelatinase-Associated Lipocalin (NGAL) in Patients with Cardiovascular Diseases—Review. *Kidney Blood Press. Res.* **2014**, *39*, 623–629. <https://doi.org/10.1159/000368474>.
55. Siasos, G.; Tousoulis, D.; Michalea, S.; Oikonomou, E.; Vavuranakis, M.; Athanasiou, D.; Tourikis, P.; Gouliopoulos, N.; Miliou, A.; Mourouzis, K.; et al. Novel Biomarkers Assessing Renal Function in Heart Failure: Relation to Inflammatory Status and Cardiac Remodelling. *Curr. Med. Chem.* **2014**, *21*, 3976–3983. <https://doi.org/10.2174/0929867321666140826114656>.
56. Damman, K.; Masson, S.; Hillege, H.L.; Voors, A.A.; van Veldhuisen, D.J.; Rossignol, P.; Proietti, G.; Barbuzzi, S.; Nicolosi, G.L.; Tavazzi, L.; et al. Tubular Damage and Worsening Renal Function in Chronic Heart Failure. *JACC Heart Fail.* **2013**, *1*, 417–424. <https://doi.org/10.1016/J.JCHF.2013.05.007>.
57. Murray, P.T.; Wettersten, N.; van Veldhuisen, D.J.; Mueller, C.; Filippatos, G.; Nowak, R.; Hogan, C.; Kontos, M.C.; Cannon, C.M.; Müller, G.A.; et al. Utility of Urine Neutrophil Gelatinase-Associated Lipocalin for Worsening Renal Function during Hospitalization for Acute Heart Failure: Primary Findings of the Urine N-Gal Acute Kidney Injury N-Gal Evaluation of Symptomatic Heart Failure Study (AKINESIS). *J. Card. Fail.* **2019**, *25*, 654–665. <https://doi.org/10.1016/J.CARDFAIL.2019.05.009>.
58. Maisel, A.S.; Wettersten, N.; van Veldhuisen, D.J.; Mueller, C.; Filippatos, G.; Nowak, R.; Hogan, C.; Kontos, M.C.; Cannon, C.M.; Müller, G.A.; et al. Neutrophil Gelatinase-Associated Lipocalin for Acute Kidney Injury During Acute Heart Failure Hospitalizations: The AKINESIS Study. *J. Am. Coll. Cardiol.* **2016**, *68*, 1420–1431. <https://doi.org/10.1016/J.JACC.2016.06.055>.
59. Damman, K.; Valente, M.A.E.; van Veldhuisen, D.J.; Cleland, J.G.F.; O'Connor, C.M.; Metra, M.; Ponikowski, P.; Cotter, G.; Davison, B.; Givertz, M.M.; et al. Plasma Neutrophil Gelatinase-Associated Lipocalin and Predicting Clinically Relevant Worsening Renal Function in Acute Heart Failure. *Int. J. Mol. Sci.* **2017**, *18*. <https://doi.org/10.3390/IJMS18071470>.
60. Bailly, V.; Zhang, Z.; Meier, W.; Cate, R.; Sanicola, M.; Bonventre, J. v. Shedding of Kidney Injury Molecule-1, a Putative Adhesion Protein Involved in Renal Regeneration. *J. Biol. Chem.* **2002**, *277*, 39739–39748. <https://doi.org/10.1074/JBC.M200562200>.
61. Vaidya, V.S.; Ramirez, V.; Ichimura, T.; Bobadilla, N.A.; Bonventre, J.V. Urinary Kidney Injury Molecule-1: A Sensitive Quantitative Biomarker for Early Detection of Kidney Tubular Injury. *Am. J. Physiol. Renal. Physiol.* **2006**, *290*, F517–F529. <https://doi.org/10.1152/AJPRENAL.00291.2005>.

62. Ichimura, T.; Hung, C.C.; Yang, S.A.; Stevens, J.L.; Bonventre, J.V. Kidney Injury Molecule-1: A Tissue and Urinary Biomarker for Nephrotoxicant-Induced Renal Injury. *Am. J. Physiol. Renal. Physiol.* **2004**, *286*, F552–F563. <https://doi.org/10.1152/AJPRENAL.00285.2002>.
63. Ichimura, T.; Asselton, E.J.P.V.; Humphreys, B.D.; Gunaratnam, L.; Duffield, J.S.; Bonventre, J. v. Kidney Injury Molecule-1 Is a Phosphatidylserine Receptor That Confers a Phagocytic Phenotype on Epithelial Cells. *J. Clin. Investig.* **2008**, *118*, 1657–1668. <https://doi.org/10.1172/JCI34487>.
64. Bonventre, J. v. Kidney Injury Molecule-1 (KIM-1): A Urinary Biomarker and Much More. *Nephrol. Dial. Transpl.* **2009**, *24*, 3265–3268. <https://doi.org/10.1093/NDT/GFP010>.
65. Geng, J.; Qiu, Y.; Qin, Z.; Su, B. The Value of Kidney Injury Molecule 1 in Predicting Acute Kidney Injury in Adult Patients: A Systematic Review and Bayesian Meta-Analysis. *J. Transl. Med.* **2021**, *19*. <https://doi.org/10.1186/S12967-021-02776-8>.
66. Pang HM, Qin XL, Liu TT, Wei WX, Cheng DH, Lu H, Guo Q, Jing L. Urinary kidney injury molecule-1 and neutrophil gelatinase-associated lipocalin as early biomarkers for predicting vancomycin-associated acute kidney injury: a prospective study. *Eur Rev Med Pharmacol Sci.* **2017**, *21*:4203–4213. PMID: 29028077.
67. Jungbauer, C.G.; Birner, C.; Jung, B.; Buchner, S.; Lubnow, M.; von Bary, C.; Endemann, D.; Banas, B.; MacK, M.; Böger, C.A.; et al. Kidney Injury Molecule-1 and N-Acetyl- β -D-Glucosaminidase in Chronic Heart Failure: Possible Biomarkers of Cardiorenal Syndrome. *Eur. J. Heart Fail.* **2011**, *13*, 1104–1110. <https://doi.org/10.1093/EURJHF/HFR102>.
68. Damman, K.; van Veldhuisen, D.J.; Navis, G.; Vaidya, V.S.; Smilde, T.D.J.; Westenbrink, B.D.; Bonventre, J.V.; Voors, A.A.; Hillege, H.L. Tubular Damage in Chronic Systolic Heart Failure Is Associated with Reduced Survival Independent of Glomerular Filtration Rate. *Heart* **2010**, *96*, 1297. <https://doi.org/10.1136/HRT.2010.194878>.
69. Tavazzi, L.; Maggioni, A.P.; Marchioli, R.; Barlera, S.; Franzosi, M.G.; Latini, R.; Lucci, D.; Nicolosi, G.L.; Porcu, M.; Tognoni, G.; et al. Effect of rosuvastatin in patients with chronic heart failure (the GISSI-HF trial): A randomised, double-blind, placebo-controlled trial. *Lancet* **2008**, *372*, 1231–1239. [https://doi.org/10.1016/S0140-6736\(08\)61240-4](https://doi.org/10.1016/S0140-6736(08)61240-4).
70. Sarhene, M.; Wang, Y.; Wei, J.; Huang, Y.; Li, M.; Li, L.; Acheampong, E.; Zhengcan, Z.; Xiaoyan, Q.; Yunsheng, X.; et al. Biomarkers in Heart Failure: The Past, Current and Future. *Heart Fail. Rev.* **2019**, *24*, 867–903. <https://doi.org/10.1007/S10741-019-09807-Z>.
71. Levey, A.S.; Inker, L.A. Assessment of Glomerular Filtration Rate in Health and Disease: A State of the Art Review. *Clin. Pharmacol. Ther.* **2017**, *102*, 405–419. <https://doi.org/10.1002/CPT.729>.
72. Costanzo, M.R.; Barasch, J. Creatinine and Cystatin C: Not the Troponin of the Kidney. *Circulation* **2018**, *137*, 2029–2031. <https://doi.org/10.1161/CIRCULATIONAHA.118.033343>.
73. Ix, J.H.; Shlipak, M.G.; Chertow, G.M.; Whooley, M.A. Association of Cystatin C with Mortality, Cardiovascular Events, and Incident Heart Failure among Persons with Coronary Heart Disease: Data from the Heart and Soul Study. *Circulation* **2007**, *115*, 173–179. <https://doi.org/10.1161/CIRCULATIONAHA.106.644286>.
74. Chen, S.; Tang, Y.; Zhou, X. Cystatin C for Predicting All-Cause Mortality and Rehospitalization in Patients with Heart Failure: A Meta-Analysis. *Biosci. Rep.* **2019**, *39*. <https://doi.org/10.1042/BSR20181761>.
75. Huerta, A.; López, B.; Ravassa, S.; José, G.S.; Querejeta, R.; Beloqui, Ó.; Zubillaga, E.; Rábago, G.; Brugnolaro, C.; Díez, J.; et al. Association of Cystatin C with Heart Failure with Preserved Ejection Fraction in Elderly Hypertensive Patients: Potential Role of Altered Collagen Metabolism. *J. Hypertens* **2016**, *34*, 130–138. <https://doi.org/10.1097/HJH.0000000000000757>.
76. Berezin, A.; Berezin, A.E. Up-to-Date Clinical Approaches of Biomarkers' Use in Heart Failure. *Biomed. Res. Ther.* **2017**, *4*, 1344–1373. <https://doi.org/10.15419/bmrat.v4i06.178>.
77. Yao, Z.; Li, G.; Li, G. Correlation between Serum Urea Nitrogen, Cystatin C, Homocysteine, and Chronic Heart Failure. *Am. J. Transl. Res.* **2021**, *13*, 3254.
78. Selcuk, H.; Selcuk, M.T.; Maden, O.; Balci, K.G.; Balci, M.M.; Tekeli, S.; Çetin, E.H.; Temizhan, A.; Balci, M.; Karabiber, N. The Impact of Admission Cystatin C Levels on In-Hospital and Three-Year Mortality Rates in Acute Decompensated Heart Failure. *Cardiovasc J. Afr.* **2018**, *29*, 305–309. <https://doi.org/10.5830/CVJA-2018-035>.
79. Bazzi, C.; Petrini, C.; Rizza, V.; Arrigo, G.; Napodano, P.; Paparella, M.; D'Amico, G. Urinary N-Acetyl-Beta-Glucosaminidase Excretion Is a Marker of Tubular Cell Dysfunction and a Predictor of Outcome in Primary Glomerulonephritis. *Nephrol. Dial. Transplant.* **2002**, *17*, 1890–1896. <https://doi.org/10.1093/NDT/17.11.1890>.
80. Damman, K.; Ng Kam Chuen, M.J.; MacFadyen, R.J.; Lip, G.Y.H.; Gaze, D.; Collinson, P.O.; Hillege, H.L.; van Oeveren, W.; Voors, A.A.; van Veldhuisen, D.J. Volume Status and Diuretic Therapy in Systolic Heart Failure and the Detection of Early Abnormalities in Renal and Tubular Function. *J. Am. Coll. Cardiol.* **2011**, *57*, 2233–2241. <https://doi.org/10.1016/J.JACC.2010.10.065>.
81. Jungbauer, C.G.; Uecer, E.; Stadler, S.; Birner, C.; Buchner, S.; Maier, L.S.; Luchner, A. N-Acetyl- β -D-Glucosaminidase and Kidney Injury Molecule-1: New Predictors for Long-Term Progression of Chronic Kidney Disease in Patients with Heart Failure. *Nephrology* **2016**, *21*, 490–498. <https://doi.org/10.1111/NEP.12632>.
82. Hu, M.C.; Shiizaki, K.; Kuro-o, M.; Moe, O.W. Fibroblast Growth Factor 23 and Klotho: Physiology and Pathophysiology of an Endocrine Network of Mineral Metabolism. *Annu. Rev. Physiol.* **2013**, *75*, 503–533. <https://doi.org/10.1146/ANNUREV-PHYSIOL-030212-183727>.
83. Vázquez-Sánchez, S.; Poveda, J.; Navarro-García, J.A.; González-Lafuente, L.; Rodríguez-Sánchez, E.; Ruilope, L.M.; Ruiz-Hurtado, G. An Overview of FGF-23 as a Novel Candidate Biomarker of Cardiovascular Risk. *Front. Physiol.* **2021**, *12*, 268. <https://doi.org/10.3389/FPHYS.2021.632260>.

84. Navarro-García, J.A.; Fernández-Velasco, M.; Delgado, C.; Delgado, J.F.; Kuro-o, M.; Ruilope, L.M.; Ruiz-Hurtado, G. PTH, Vitamin D, and the FGF-23-Klotho Axis and Heart: Going beyond the Confines of Nephrology. *Eur. J. Clin. Investig.* **2018**, *48*, e12902. <https://doi.org/10.1111/ECI.12902>.
85. Silva, A.P.; Mendes, F.; Carias, E.; Gonçalves, R.B.; Fragoso, A.; Dias, C.; Tavares, N.; Café, H.M.; Santos, N.; Rato, F.; et al. Plasmatic Klotho and FGF23 Levels as Biomarkers of CKD-Associated Cardiac Disease in Type 2 Diabetic Patients. *Int. J. Mol. Sci.* **2019**, *20*, 1536. <https://doi.org/10.3390/IJMS20071536>.
86. Andersen, I.A.; Huntley, B.K.; Sandberg, S.S.; Heublein, D.M.; Burnett, J.C. Elevation of Circulating but Not Myocardial FGF23 in Human Acute Decompensated Heart Failure. *Nephrol. Dial. Transplant.* **2016**, *31*, 767–772. <https://doi.org/10.1093/NDT/GFV398>.
87. Gruson, D.; Lepoutre, T.; Ketelslegers, J.M.; Cumps, J.; Ahn, S.A.; Rousseau, M.F. C-Terminal FGF23 Is a Strong Predictor of Survival in Systolic Heart Failure. *Peptides* **2012**, *37*, 258–262. <https://doi.org/10.1016/J.PEPTIDES.2012.08.003>.
88. Scialla, J.J.; Xie, H.; Rahman, M.; Anderson, A.H.; Isakova, T.; Ojo, A.; Zhang, X.; Nessel, L.; Hamano, T.; Grunwald, J.E.; et al. Fibroblast Growth Factor-23 and Cardiovascular Events in CKD. *J. Am. Soc. Nephrol.* **2014**, *25*, 349–360. <https://doi.org/10.1681/ASN.2013050465/-DCSUPPLEMENTAL>.
89. de Buyzere, M.L.; Delanghe, J.R. Fibroblast Growth Factor 23 and the Quest for the Holy Grail in Heart Failure: Will the Crusaders Be Forced to Surrender? *Eur. J. Heart Fail.* **2020**, *22*, 710–712. <https://doi.org/10.1002/EJHF.1786>.
90. Stöhr, R.; Brandenburg, V.M.; Heine, G.H.; Maeder, M.T.; Leibundgut, G.; Schuh, A.; Jeker, U.; Pfisterer, M.; Sanders-van Wijk, S.; Brunner-la Rocca, H.P. Limited Role for Fibroblast Growth Factor 23 in Assessing Prognosis in Heart Failure Patients: Data from the TIME-CHF Trial. *Eur. J. Heart Fail.* **2020**, *22*, 701–709. <https://doi.org/10.1002/EJHF.1749>.
91. Biegus, J.; Zymlinski, R.; Testani, J.; Marciniak, D.; Zdanowicz, A.; Jankowska, E.A.; Banasiak, W.; Ponikowski, P. Renal Profiling Based on Estimated Glomerular Filtration Rate and Spot Urine Sodium Identifies High-Risk Acute Heart Failure Patients. *Eur. J. Heart Fail.* **2020**, *23*, 729–739. <https://doi.org/10.1002/ejhf.2053>.
92. Biegus, J.; Nawrocka-Millward, S.; Zymlinski, R.; Fudim, M.; Testani, J.; Marciniak, D.; Rosiek-Biegus, M.; Ponikowska, B.; Guzik, M.; Garus, M.; et al. Distinct Renin/Aldosterone Activity Profiles Correlate with Renal Function, Natriuretic Response, Decongestive Ability and Prognosis in Acute Heart Failure. *Int. J. Cardiol.* **2021**, *345*, 54–60. <https://doi.org/10.1016/j.ijcard.2021.10.149>.
93. Biegus, J.; Zymlinski, R.; Sokolski, M.; Todd, J.; Cotter, G.; Metra, M.; Jankowska, E.A.; Banasiak, W.; Ponikowski, P. Serial Assessment of Spot Urine Sodium Predicts Effectiveness of Decongestion and Outcome in Patients with Acute Heart Failure. *Eur. J. Heart Fail.* **2019**, *21*, 624–633. <https://doi.org/10.1002/EJHF.1428>.
94. Biegus, J.; Zymlinski, R.; Fudim, M.; Testani, J.; Sokolski, M.; Marciniak, D.; Ponikowska, B.; Guzik, M.; Garus, M.; Urban, S.; et al. Spot Urine Sodium in Acute Heart Failure: Differences in Prognostic Value on Admission and Discharge. *ESC Heart Fail.* **2021**, *8*, 2597–2602. <https://doi.org/10.1002/EHF2.13372>.
95. Zymlinski, R.; Sierpiński, R.; Metra, M.; Cotter, G.; Sokolski, M.; Siwołowski, P.; Garus, M.; Gajewski, P.; Tryba, J.; Samorek, M.; et al. Elevated Plasma Endothelin-1 Is Related to Low Natriuresis, Clinical Signs of Congestion, and Poor Outcome in Acute Heart Failure. *ESC Heart Fail.* **2020**, *7*, 3536–3544. <https://doi.org/10.1002/ehf2.13064>.
96. Kashani, K.; Al-Khafaji, A.; Ardiles, T.; Artigas, A.; Bagshaw, S.M.; Bell, M.; Bihorac, A.; Birkhahn, R.; Cely, C.M.; Chawla, L.S.; et al. Discovery and validation of cell cycle arrest biomarkers in human acute kidney injury. *Crit. Care* **2013**, *17*, R25. <https://doi.org/10.1186/cc12503>.
97. Cui, M.; Han, Y.; Yang, J.; Li, G.; Yang, C. A Narrative Review of the Research Status of Exosomes in Cardiovascular Disease. *Ann. Palliat. Med.* **2022**, *11*, 363–377. <https://doi.org/10.21037/APM-21-3364/COIF>.
98. Tschuschke, M.; Kocherova, I.; Bryja, A.; Mozdziak, P.; Volponi, A.A.; Janowicz, K.; Sibiak, R.; Piotrowska-Kempisty, H.; Iżycki, D.; Bukowska, D.; et al. Inclusion Biogenesis, Methods of Isolation and Clinical Application of Human Cellular Exosomes. *J. Clin. Med.* **2020**, *9*, 436. <https://doi.org/10.3390/JCM9020436>.
99. Zheng, D.; Huo, M.; Li, B.; Wang, W.; Piao, H.; Wang, Y.; Zhu, Z.; Li, D.; Wang, T.; Liu, K. The Role of Exosomes and Exosomal MicroRNA in Cardiovascular Disease. *Front. Cell Dev. Biol.* **2021**, *8*, 1810. <https://doi.org/10.3389/FCELL.2020.616161/BIBTEX>.
100. Raposo, G.; Nijman, H.W.; Stoorvogel, W.; Leijendekker, R.; Harding, C.V.; Melief, C.J.M.; Geuze, H.J. B Lymphocytes Secrete Antigen-Presenting Vesicles. *J. Exp. Med.* **1996**, *183*, 1161–1172. <https://doi.org/10.1084/JEM.183.3.1161>.
101. Asgarpour, K.; Shojaei, Z.; Amiri, F.; Ai, J.; Mahjoubin-Tehran, M.; Ghasemi, F.; Arefnezhad, R.; Hamblin, M.R.; Mirzaei, H. Exosomal MicroRNAs Derived from Mesenchymal Stem Cells: Cell-to-Cell Messages. *Cell Commun. Signal.* **2020**, *18*, 1–16. <https://doi.org/10.1186/S12964-020-00650-6/TABLES/2>.
102. Biasucci, L.M.; Maino, A.; Grimaldi, M.C.; Cappannoli, L.; Aspromonte, N. Novel Biomarkers in Heart Failure: New Insight in Pathophysiology and Clinical Perspective. *J. Clin. Med.* **2021**, *10*, 2771. <https://doi.org/10.3390/JCM10132771>.
103. Bang, C.; Batkai, S.; Dangwal, S.; Gupta, S.K.; Foinquinos, A.; Holzmann, A.; Just, A.; Remke, J.; Zimmer, K.; Zeug, A.; et al. Cardiac Fibroblast-Derived MicroRNA Passenger Strand-Enriched Exosomes Mediate Cardiomyocyte Hypertrophy. *J. Clin. Investig.* **2014**, *124*, 2136–2146. <https://doi.org/10.1172/JCI70577>.
104. Fang, X.; Stroud, M.J.; Ouyang, K.; Fang, L.; Zhang, J.; Dalton, N.D.; Gu, Y.; Wu, T.; Peterson, K.L.; Huang, H. da; et al. Adipocyte-Specific Loss of PPAR γ Attenuates Cardiac Hypertrophy. *JCI Insight* **2016**, *1*. <https://doi.org/10.1172/JCI.INSIGHT.89908>.
105. Xue, R.; Tan, W.; Wu, Y.; Dong, B.; Xie, Z.; Huang, P.; He, J.; Dong, Y.; Liu, C. Role of Exosomal MiRNAs in Heart Failure. *Front. Cardiovasc. Med.* **2020**, *7*, 347. <https://doi.org/10.3389/FCVM.2020.592412/BIBTEX>.

106. Zou, M.; Wang, F.; Gao, R.; Wu, J.; Ou, Y.; Chen, X.; Wang, T.; Zhou, X.; Zhu, W.; Li, P.; et al. Autophagy Inhibition of Hsa-MiR-19a-3p/19b-3p by Targeting TGF- β R II during TGF-B1-Induced Fibrogenesis in Human Cardiac Fibroblasts. *Sci. Rep.* **2016**, *6*, 24747. <https://doi.org/10.1038/SREP24747>.
107. Qiao, L.; Hu, S.; Liu, S.; Zhang, H.; Ma, H.; Huang, K.; Li, Z.; Su, T.; Vandergriff, A.; Tang, J.; et al. MicroRNA-21-5p Dysregulation in Exosomes Derived from Heart Failure Patients Impairs Regenerative Potential. *J. Clin. Investig.* **2019**, *129*, 2237–2250. <https://doi.org/10.1172/JCI123135>.
108. Goren, Y.; Kushnir, M.; Zafir, B.; Tabak, S.; Lewis, B.S.; Amir, O. Serum Levels of MicroRNAs in Patients with Heart Failure. *Eur. J. Heart Fail.* **2012**, *14*, 147–154. <https://doi.org/10.1093/EURJHF/HFR155>.
109. dos Reis Schneider, S.I.; Silvello, D.; Martinelli, N.C.; Garbin, A.; Biolo, A.; Clausell, N.; Andrades, M.; dos Santos, K.G.; Rohde, L.E. Plasma Levels of MicroRNA-21, -126 and -423-5p Alter during Clinical Improvement and Are Associated with the Prognosis of Acute Heart Failure. *Mol. Med. Rep.* **2018**, *17*, 4736–4746. <https://doi.org/10.3892/MMR.2018.8428>.
110. Endo, K.; Naito, Y.; Ji, X.; Nakanishi, M.; Noguchi, T.; Goto, Y.; Nonogi, H.; Ma, X.; Weng, H.; Hirokawa, G.; et al. MicroRNA 210 as a Biomarker for Congestive Heart Failure. *Biol. Pharm. Bull.* **2013**, *36*, 48–54. <https://doi.org/10.1248/BPB.B12-00578>.
111. Ovchinnikova, E.S.; Schmitter, D.; Vegter, E.L.; ter Maaten, J.M.; Valente, M.A.E.; Liu, L.C.Y.; van der Harst, P.; Pinto, Y.M.; de Boer, R.A.; Meyer, S.; et al. Signature of Circulating MicroRNAs in Patients with Acute Heart Failure. *Eur. J. Heart Fail.* **2016**, *18*, 414–423. <https://doi.org/10.1002/EJHF.332>.
112. Wu, T.; Chen, Y.; Du, Y.; Tao, J.; Li, W.; Zhou, Z.; Yang, Z. Circulating Exosomal MiR-92b-5p Is a Promising Diagnostic Biomarker of Heart Failure with Reduced Ejection Fraction Patients Hospitalized for Acute Heart Failure. *J. Thorac. Dis.* **2018**, *10*, 6211–6220. <https://doi.org/10.21037/JTD.2018.10.52>.
113. Sun, C.; Ni, M.; Song, B.; Cao, L. Circulating Circular RNAs: Novel Biomarkers for Heart Failure. *Front. Pharmacol.* **2020**, *11*, 1649. <https://doi.org/10.3389/FPHAR.2020.560537/BIBTEX>.
114. Castiglione, V.; Aimo, A.; Vergaro, G.; Saccaro, L.; Passino, C.; Emdin, M. Biomarkers for the Diagnosis and Management of Heart Failure. *Heart Fail. Rev.* **2021**, *27*, 625–664. <https://doi.org/10.1007/S10741-021-10105-W>.
115. Yancy, C.W.; Jessup, M.; Bozkurt, B.; Butler, J.; Casey, D.E.; Colvin, M.M.; Drazner, M.H.; Filippatos, G.S.; Fonarow, G.C.; Givertz, M.M.; et al. 2017 ACC/AHA/HFSA Focused Update of the 2013 ACCF/AHA Guideline for the Management of Heart Failure: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines and the Heart Failure Society of America. *Circulation* **2017**, *136*, e137–e161. <https://doi.org/10.1161/CIR.0000000000000509/-/DC2>.
116. Chirinos, J.A.; Orlenko, A.; Zhao, L.; Basso, M.D.; Cvijic, M.E.; Li, Z.; Spires, T.E.; Yarde, M.; Wang, Z.; Seiffert, D.A.; et al. Multiple Plasma Biomarkers for Risk Stratification in Patients With Heart Failure and Preserved Ejection Fraction. *J. Am. Coll. Cardiol.* **2020**, *75*, 1281–1295. <https://doi.org/10.1016/J.JACC.2019.12.069>.
117. He, T.; Mischak, M.; Clark, A.L.; Campbell, R.T.; Delles, C.; Díez, J.; Filippatos, G.; Mebazaa, A.; McMurray, J.J.V.; González, A.; et al. Urinary Peptides in Heart Failure: A Link to Molecular Pathophysiology. *Eur. J. Heart Fail.* **2021**, *23*, 1875–1887. <https://doi.org/10.1002/EJHF.2195>.
118. Zeillemaker, A.M.; Verbrugh, H.A.; Hoyneck van Papendrecht, A.A.; Leguit, P. CA 125 Secretion by Peritoneal Mesothelial Cells. *J. Clin. Pathol.* **1994**, *47*, 263–265. <https://doi.org/10.1136/jcp.47.3.263>.
119. Yin, B.W.T.; Lloyd, K.O. Molecular Cloning of the CA125 Ovarian Cancer Antigen. *J. Biol. Chem.* **2001**, *276*, 27371–27375. <https://doi.org/10.1074/jbc.M103554200>.
120. Vizzard, E.; D'Aloia, A.; Curnis, A.; Dei Cas, L. Carbohydrate Antigen 125. *Cardiol. Rev.* **2013**, *21*, 23–26. <https://doi.org/10.1097/CRD.0b013e318265f58f>.
121. Bulska-Będkowska, W.; Chelmecka, E.; Owczarek, A.J.; Mizia-Stec, K.; Witek, A.; Szybalska, A.; Grodzicki, T.; Olszan-ecka-Glinianowicz, M.; Chudek, J. CA125 as a Marker of Heart Failure in the Older Women: A Population-Based Analysis. *J. Clin. Med.* **2019**, *8*, 607. <https://doi.org/10.3390/jcm8050607>.
122. SEO, T.; IKEDA, Y.; ONAKA, H.; HAYASHI, T.; KAWAGUCHI, K.; KOTAKE, C.; TODA, T.; KOBAYASHI, K. Usefulness of Serum CA125 Measurement for Monitoring Pericardial Effusion. *Jpn. Circ. J.* **1993**, *57*, 489–494. <https://doi.org/10.1253/jcj.57.489>.
123. Núñez, J.; Bayés-Genís, A.; Revuelta-López, E.; ter Maaten, J.M.; Miñana, G.; Barallat, J.; Cserkóová, A.; Bodi, V.; Fernández-Cisnal, A.; Núñez, E.; et al. Clinical Role of CA125 in Worsening Heart Failure. *JACC Heart Fail.* **2020**, *8*, 386–397. <https://doi.org/10.1016/j.jchf.2019.12.005>.
124. Pan, C.; Zhou, M.; Jian, Y.; Zeng, Y.; Wang, M.; Chen, F. CA125: An Increasingly Promising Biomarker of Heart Failure. *Curr. Pharm. Des.* **2021**, *27*, 3871–3880. <https://doi.org/10.2174/1381612827666210118122521>.
125. Wussler, D.; Bayes-Genis, A.; Belkin, M.; Strebel, I.; Kozhuharov, N.; Revuelta-Lopez, E.; Nowak, A.; Lupon, J.; Gualandro, D.M.; Shrestha, S.; Breidhardt, T.; Nunez, J.; Mueller, C. CA 125 in the diagnosis and risk stratification of acute heart failure. *Eur. Heart J.* **2021**, *42*(Suppl.1), ehab724.1021. <https://doi.org/10.1093/eurheartj/ehab724.1021>.
126. Yilmaz, M.B.; Zorlu, A.; Tandogan, I. Plasma CA-125 Level Is Related to Both Sides of the Heart: A Retrospective Analysis. *Int. J. Cardiol.* **2011**, *149*, 80–82. <https://doi.org/10.1016/j.ijcard.2009.12.003>.
127. Núñez, J.; Núñez, E.; Bayés-Genís, A.; Fonarow, G.C.; Miñana, G.; Bodí, V.; Pascual-Figal, D.; Santas, E.; Garcia-Blas, S.; Chorro, F.J.; et al. Long-Term Serial Kinetics of N-Terminal pro B-Type Natriuretic Peptide and Carbohydrate Antigen 125 for Mortality Risk Prediction Following Acute Heart Failure. *Eur. Heart J. Acute Cardiovasc. Care* **2017**, *6*, 685–696. <https://doi.org/10.1177/2048872616649757>.

128. Monteiro, S.; Franco, F.; Costa, S.; Monteiro, P.; Vieira, H.; Coelho, L.; Oliveira, L.; Providência, L.A. Prognostic Value of CA125 in Advanced Heart Failure Patients. *Int. J. Cardiol.* **2010**, *140*, 115–118. <https://doi.org/10.1016/j.ijcard.2008.11.023>.
129. Frigy, A.; Belényi, B.; Kirchmaier, Á.; Fekete, N.; Szabó, I.A. Elevated CA-125 as Humoral Biomarker of Congestive Heart Failure: Illustrative Cases and a Short Review of Literature. *Case Rep. Cardiol.* **2020**, *2020*, 1–5. <https://doi.org/10.1155/2020/1642914>.
130. Soler, M.; Miñana, G.; Santas, E.; Núñez, E.; de la Espriella, R.; Valero, E.; Bodí, V.; Chorro, F.J.; Fernández-Cisnal, A.; D'Ascoli, G.; et al. CA125 Outperforms NT-ProBNP in Acute Heart Failure with Severe Tricuspid Regurgitation. *Int. J. Cardiol.* **2020**, *308*, 54–59. <https://doi.org/10.1016/j.ijcard.2020.03.027>.
131. Núñez, J.; Miñana, G.; González, M.; García-Ramón, R.; Sanchis, J.; Bodí, V.; Núñez, E.; Chorro, F.J.; Llàcer, A.; Miguel, A. Antigen Carbohydrate 125 in Heart Failure: Not Just a Surrogate for Serosal Effusions? *Int. J. Cardiol.* **2011**, *146*, 473–474. <https://doi.org/10.1016/j.ijcard.2010.12.027>.
132. Miñana, G.; Palau, P.; Núñez, J.; Sanchis, J. The Tumor Marker CA125 and Heart Failure. *Rev. Española De Cardiol.* **2010**, *63*, 1213–1214. [https://doi.org/10.1016/S1885-5857\(10\)70240-1](https://doi.org/10.1016/S1885-5857(10)70240-1).
133. Kumric, M.; Ticinovic Kurir, T.; Bozic, J.; Glavas, D.; Saric, T.; Marcellius, B.; D'Amaro, D.; Borovac, J.A. Carbohydrate Antigen 125: A Biomarker at the Crossroads of Congestion and Inflammation in Heart Failure. *Cardiac Failure Review* **2021**, *7*. <https://doi.org/10.15420/cfr.2021.22>.
134. Fonarow, G.C.; Albert, N.M.; Curtis, A.B.; Stough, W.G.; Gheorghiade, M.; Heywood, J.T.; McBride, M.L.; Inge, P.J.; Mehra, M.R.; O'Connor, C.M.; et al. Improving Evidence-Based Care for Heart Failure in Outpatient Cardiology Practices: Primary Results of the Registry to Improve the Use of Evidence-Based Heart Failure Therapies in the Outpatient Setting (IMPROVE HF). *Circulation* **2010**, *122*, 585–596. <https://doi.org/10.1161/CIRCULATIONAHA.109.934471>.
135. Hartupee, J.; Mann, D.L. Neurohormonal Activation in Heart Failure with Reduced Ejection Fraction. *Nat. Rev. Cardiol.* **2017**, *14*, 30–38. <https://doi.org/10.1038/nrcardio.2016.163>.
136. Ge, Z.; Li, A.; McNamara, J.; dos Remedios, C.; Lal, S. Pathogenesis and Pathophysiology of Heart Failure with Reduced Ejection Fraction: Translation to Human Studies. *Heart Fail. Rev.* **2019**, *24*, 743–758. <https://doi.org/10.1007/s10741-019-09806-0>.
137. Nishikimi, T.; Nakao, K.; Kangawa, K. Adrenomedullin in Heart Failure: Molecular Mechanism and Therapeutic Implication. *Curr. Hypertens. Rev.* **2011**, *7*, 273–283. <https://doi.org/10.2174/157340211799304752>.
138. Voors, A.A.; Kremer, D.; Geven, C.; ter Maaten, J.M.; Struck, J.; Bergmann, A.; Pickkers, P.; Metra, M.; Mebazaa, A.; Düngen, H.-D.; et al. Adrenomedullin in Heart Failure: Pathophysiology and Therapeutic Application. *Eur. J. Heart Fail.* **2019**, *21*, 163–171. <https://doi.org/10.1002/ehf.1366>.
139. Mehmood, M. Adrenomedullin: A Double-Edged Sword in Septic Shock and Heart Failure Therapeutics? *Am. J. Respir. Crit. Care Med.* **2020**, *201*, 1164–1165. <https://doi.org/10.1164/rccm.201912-2412LE>.
140. Krzeminski, K. The Role of Adrenomedullin in Cardiovascular Response to Exercise—A Review. *J. Hum. Kinet.* **2016**, *53*, 127–142. <https://doi.org/10.1515/hukin-2016-0017>.
141. Nishikimi, T.; Yoshihara, F.; Mori, Y.; Kangawa, K.; Matsuoka, H. Cardioprotective Effect of Adrenomedullin in Heart Failure. *Hypertens. Res.* **2003**, *26* (Suppl.), S121–S127. <https://doi.org/10.1291/hypres.26.s121>.
142. Tsuruda, T.; Kato, J.; Hatakeyama, K.; Masuyama, H.; Cao, Y.-N.; Imamura, T.; Kitamura, K.; Asada, Y.; Eto, T. Antifibrotic Effect of Adrenomedullin on Coronary Adventitia in Angiotensin II-Induced Hypertensive Rats. *Cardiovasc. Res.* **2005**, *65*, 921–929. <https://doi.org/10.1016/j.cardiores.2004.11.004>.
143. Rademaker, M.T.; Cameron, V.A.; Charles, C.J.; Lainchbury, J.G.; Nicholls, M.G.; Richards, A.M. Adrenomedullin and Heart Failure. *Regul. Pept.* **2003**, *112*, 51–60. [https://doi.org/10.1016/s0167-0115\(03\)00022-3](https://doi.org/10.1016/s0167-0115(03)00022-3).
144. Cockcroft, J.R.; Noon, J.P.; Gardner-Medwin, J.; Bennett, T. Haemodynamic Effects of Adrenomedullin in Human Resistance and Capacitance Vessels. *Br. J. Clin. Pharmacol.* **1997**, *44*, 57–60. <https://doi.org/10.1046/j.1365-2125.1997.00622.x>.
145. Diaz, E.; Israel, A. Effect of Adrenomedullin Receptor and Calcitonin Gene-Related Peptide Receptor Antagonists on Centrally Mediated Adrenomedullin Renal Action. *Brain. Res. Bull.* **2001**, *55*, 29–35. [https://doi.org/10.1016/s0361-9230\(01\)00461-0](https://doi.org/10.1016/s0361-9230(01)00461-0).
146. Brain, S.D.; Grant, A.D. Vascular Actions of Calcitonin Gene-Related Peptide and Adrenomedullin. *Physiol. Rev.* **2004**, *84*, 903–934. <https://doi.org/10.1152/physrev.00037.2003>.
147. Ebara, T.; Miura, K.; Okumura, M.; Matsuura, T.; Kim, S.; Yukimura, T.; Iwao, H. Effect of Adrenomedullin on Renal Hemodynamics and Functions in Dogs. *Eur. J. Pharmacol.* **1994**, *263*, 69–73. [https://doi.org/10.1016/0014-2999\(94\)90524-x](https://doi.org/10.1016/0014-2999(94)90524-x).
148. Geven, C.; Kox, M.; Pickkers, P. Adrenomedullin and Adrenomedullin-Targeted Therapy As Treatment Strategies Relevant for Sepsis. *Front. Immunol.* **2018**, *9*, 292. <https://doi.org/10.3389/fimmu.2018.00292>.
149. Möckel, M.; Searle, J.; Hartmann, O.; Anker, S.D.; Peacock, W.F.; Wu, A.H.B.; Maisel, A. Mid-regional pro-adrenomedullin improves disposition strategies for patients with acute dyspnoea: Results from the BACH trial. *Emerg. Med. J.* **2013**, *30*, 633–637. <https://doi.org/10.1136/emmermed-2012-201530>.
150. Peacock, W.F. Novel biomarkers in acute heart failure: MR-pro-adrenomedullin. *Clin. Chem. Lab. Med.* **2014**, *52*, 1433–1435. <https://doi.org/10.1515/cclm-2014-0222>.
151. Czajkowska, K.; Zbroch, E.; Bielach-Bazyluk, A.; Mitrosz, K.; Bujno, E.; Kakareko, K.; Rydzewska-Rosolowska, A.; Hryszko, T. Mid-Regional Proadrenomedullin as a New Biomarker of Kidney and Cardiovascular Diseases—Is It the Future? *J. Clin. Med.* **2021**, *10*, 524. <https://doi.org/10.3390/jcm10030524>.

152. Koyama, T.; Kuriyama, N.; Suzuki, Y.; Saito, S.; Tanaka, R.; Iwao, M.; Tanaka, M.; Maki, T.; Itoh, H.; Ihara, M.; Shindo, T.; Uehara, R. Mid-regional pro-adrenomedullin is a novel biomarker for arterial stiffness as the criterion for vascular failure in a cross-sectional study. *Sci. Rep.* **2021**, *11*, 305. <https://doi.org/10.1038/s41598-020-79525-2>.
153. Pandhi, P.; ter Maaten, J.M.; Emmens, J.E.; Struck, J.; Bergmann, A.; Cleland, J.G.; Givertz, M.M.; Metra, M.; O'Connor, C.M.; Teerlink, J.R.; et al. Clinical Value of Pre-Discharge Bio-Adrenomedullin as a Marker of Residual Congestion and High Risk of Heart Failure Hospital Readmission. *Eur. J. Heart Fail.* **2020**, *22*, 683–691. <https://doi.org/10.1002/ehf.1693>.
154. van Kimmenade, R.R.J.; Januzzi, J.L. Emerging Biomarkers in Heart Failure. *Clin. Chem.* **2012**, *58*, 127–138. <https://doi.org/10.1373/clinchem.2011.165720>.
155. Cuzzo, B.; Padala, S.A.; Lappin, S.L. *Physiology, Vasopressin*; 2022.
156. Bankir, L. Antidiuretic Action of Vasopressin: Quantitative Aspects and Interaction between V1a and V2 Receptor-Mediated Effects. *Cardiovasc. Res.* **2001**, *51*, 372–390. [https://doi.org/10.1016/S0008-6363\(01\)00328-5](https://doi.org/10.1016/S0008-6363(01)00328-5).
157. Gilotra, N.A. Arginine Vasopressin as a Target in the Treatment of Acute Heart Failure. *World J. Cardiol.* **2014**, *6*, 1252. <https://doi.org/10.4330/wjc.v6.i12.1252>.
158. Goldsmith, S.R. Arginine Vasopressin Antagonism in Heart Failure: Current Status and Possible New Directions. *J. Cardiol.* **2019**, *74*, 49–52. <https://doi.org/10.1016/j.jjcc.2019.03.001>.
159. Goldsmith, S.R. The Role of Vasopressin in Congestive Heart Failure. *Cleve Clin. J. Med.* **2006**, *73* (Suppl. 3), S19–S23. https://doi.org/10.3949/ccjm.73.suppl_3.s19.
160. Chirinos, J.A.; Sardana, M.; Oldland, G.; Ansari, B.; Lee, J.; Hussain, A.; Mustafa, A.; Akers, S.R.; Wei, W.; Lakatta, E.G.; et al. Association of Arginine Vasopressin with Low Atrial Natriuretic Peptide Levels, Left Ventricular Remodelling, and Outcomes in Adults with and without Heart Failure. *ESC Heart Fail.* **2018**, *5*, 911–919. <https://doi.org/10.1002/ehf2.12319>.
161. Jia, J.; Chang, G.-L.; Qin, S.; Chen, J.; He, W.-Y.; Lu, K.; Li, Y.; Zhang, D.-Y. Comparative evaluation of copeptin and NT-proBNP in patients with severe acute decompensated heart failure, and prediction of adverse events in a 90-day follow-up period: A prospective clinical observation trial. *Exp. Ther. Med.* **2017**, *13*, 1554–1560.
162. Nickel, C.H.; Bingisser, R.; Morgenthaler, N.G. The role of copeptin as a diagnostic and prognostic biomarker for risk stratification in the emergency department. *BMC Med.* **2012**, *10*, 7. <https://doi.org/10.1186/1741-7015-10-7>.
163. Kim, H.N.; Yang, D.H.; Park, B.E.; Park, Y.J.; Kim, H.J.; Jang, S.Y.; Bae, M.H.; Lee, J.H.; Park, H.S.; Cho, Y.; Chae, S.C. Prognostic impact of chromogranin A in patients with acute heart failure. *Yeungnam Univ. J. Med.* **2021**, *38*, 337–343. <https://doi.org/10.12701/yujm.2020.00843>.
164. Wohlschlaeger J.; von Winterfeld, M.; Milting, H.; El Banayosy, A.; Schmitz, K.J.; Takeda, A.; Takeda, N.; Azhari, P.; Schmid, C.; August, C.; Schmid, K.W.; Baba, H.A. Decreased myocardial chromogranin a expression and colocalization with brain natriuretic peptide during reverse cardiac remodeling after ventricular unloading. *J Heart Lung Transplant.* **2008**;27(4):442-9.doi: 10.1016/j.healun.2008.01.017.
165. Goetze, J.P.; Alehagen, U.; Flyvbjerg, A.; Rehfeld, J.F. Chromogranin A as a Biomarker in Cardiovascular Disease. *Biomark Med.* **2014**, *8*, 133–140. <https://doi.org/10.2217/bmm.13.102>.
166. Goetze, J.P.; Hilsted, L.M.; Rehfeld, J.F.; Alehagen, U. Plasma chromogranin A is a marker of death in elderly patients presenting with symptoms of heart failure. *Endocr. Connect.* **2014**, *3*, 47–56. <https://doi.org/10.1530/EC-14-0017>.