



# **Communication Phytotoxic Effects of Three** *Origanum* Species Extracts and Essential Oil on Seed Germinations and Seedling Growths of Four Weed Species

Saban Kordali <sup>1</sup>, Gulbahar Kabaagac <sup>2</sup>, İsmail Sen <sup>3</sup>, Ferah Yilmaz <sup>1</sup>, and Agnieszka Najda <sup>4,\*</sup>

- <sup>1</sup> Department of Plant Protection, Fethiye Faculty of Agriculture, Muğla Sıtkı Koçman University, 48300 Fethiye, Türkiye
- <sup>2</sup> Department of Plant Protection, Faculty of Agriculture, Atatürk University, 25240 Erzurum, Türkiye
- <sup>3</sup> Department of Research and Development, Demirsoy Agriculture Company, 48360 Seydikemer, Türkiye
- <sup>4</sup> Department of Vegetable and Herbal Crops, University of Life Sciences in Lublin, 50A Doświadczalna Street, 20-280 Lublin, Poland
- \* Correspondence: agnieszka.najda@up.lublin.pl

Abstract: The use of chemical pesticides to protect agricultural products is a global concern because of their adverse effects on the environment and public health. To avoid the dangers of synthetic herbicides, research has turned to natural alternatives. This study was conducted to evaluate the allelopathic effect of essential oil (EO) extracted from Origanum syriacum, Origanum onites, and Origanum majorana. In addition, the chemical composition of the essential oil was elucidated by gas chromatography and mass spectrometry (GC-MS) analysis. A total of 11 different components of O. syriacum were identified, and the main components were carvacrol (88.49), p-Cymene (5.71),  $\gamma$ -Terpinene (1.63),  $\beta$ -Caryoplhyllene (1.48), and Terpinen-4-ol (0.65), respectively. For O. onites, 10 different compounds were identified, and the main components were carvacrol (58.65), Thymol (30.97), Linalool (4.17), p-Cymene (1.94), and  $\beta$ -Caryoplhyllene (0.98), respectively. Finally, for O. majorana, 14 different compounds were identified, and the main components were carvacrol (40.57), α-Terpineol (29.28), *p*-Cymene (9.02), γ-Terpinene (5.80), and carvacrol methyl ether (3.46). Finally, 14 compounds from the Origanum majorana species were identified, with carvacrol (40.57), -Terpineol (29.28), p-Cymene (9.02), and -Terpinene (5.80) as the parent compound (3.46). Oxygenated monoterpenes were the highest in all species' EO content. EOs and plant extracts were tested at 5, 10, and 20 L/Petri concentrations against seed germination and seedling growth in four weed species (Thlaspi arvense, Amaranthus retroflexus, Rumex cripus, and Lactuca serriola). The concentrations of essential oil were set as 5, 10, and 20  $\mu$ L/Petri dishes for seed germination. In the greenhouse experiment, the final concentration of solutions was set as 20 µL and the solutions were directly sprayed on the surface of the weeds, and the mortality rates were noted after 24 and 48 h of application. It was observed that increasing the application decreased seed germination. The phytotoxic effects on the seedling germination in the greenhouse were observed, resulting in 48.76–94% mortality rates. Consequently, the essential oil from Origanum species could be considered as an alternative bio-herbicide to tested weeds.

Keywords: oregano; herbicidal effect; essential oil; extract

## 1. Introduction

Crop production has been increasing yearly to supply the food demand, which is a consequence of the fast expansion of the world population [1,2]. Unfortunately, weeds in the modern agricultural systems are one of the major problems worldwide because of yield and crop loss [3]. Globally, around 1800 weed species cause a 31.5% reduction in crop production [4]. Therefore, farmers have tended to use more herbicides to improve yields. Many studies have shown that less than 10% of conventional pesticides target the plant [5]



Citation: Kordali, S.; Kabaagac, G.; Sen, İ.; Yilmaz, F.; Najda, A. Phytotoxic Effects of Three *Origanum* Species Extracts and Essential Oil on Seed Germinations and Seedling Growths of Four Weed Species. *Agronomy* 2022, *12*, 2581. https:// doi.org/10.3390/agronomy12102581

Academic Editor: Antonios Chrysargyris

Received: 24 August 2022 Accepted: 12 October 2022 Published: 20 October 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). and only 0.1% of these remain long enough to reach the plant, while the rest are dispersed directly into the environment [6,7]. However, intensive usage of synthetic herbicides can contaminate soil and groundwater [8–11], and weeds could also gain resistance against these synthetic herbicides [12,13]. Therefore, the awareness of chemical-free weed control methods has gained importance to keep society healthy. As a result, research on bioherbicides obtained from aromatic plants and their selective herbicidal mechanism has been conducted in recent years [14–20].

Essential oils (EOs) by nature (as plant secondary metabolites) represent a safer alternative in many applications such as food preservation, biomedicine, cosmetics or agriculture [20,21]. Because of their allelopathic effects, they are often used for biopesticide production [22–27].

Because of their inherent allelopathic effects, essential oils (EOs) are often used for the production of biopesticides [20,21,23–27]. At the same time, because of their specific composition, they are rapidly degraded in the soil and are considered environmentally safe [25–27].

The genus *Origanum* (oregano), which belongs to the *Lamiaceae* family, is widespread worldwide and comprises about 900 species. One of the most common genera of the Laminaceae family, *Origanum* L. has 21 species (24 taxa) and 13 hybrids in Turkey (5–7). There are about 20 species of the *Origanum* genus in Turkish flora [27–31]. The *Origanum* species has traditionally been used as a spicy additive for food instead of thyme in Turkey. This genus is rich in essential oils and bitter substances [32]. The *Origanum* species has several medicinal properties such as a sedative, diuretics, sweaters, antiseptics, and additionally in the treatment of gastrointestinal diseases and constipation, and it is used traditionally in Turkey [32]. The *Origanum* species is commonly known as "Oregano", and Turkey is the world's largest supplier. Approximately 15,000 tons of *Origanum* species were harvested and exported as raw material and essential oil in 2019 [33,34].

The *Origanum* species has been shown to have anti-diabetic, anti-obesity, antihyperlipidemic, hepatoprotective, anti-urolytic, anti-microbial, antioxidant, antiproliferative, anti-nociceptive, anti-platelet, anti-melanogenic, anxiolytic, anti-inflammatory, memory-enhancing, and cytotoxic properties [35,36]. The chemical properties of the Origanum species were found to be rich in phenolic acids, flavonoids, sesquiterpenes, monocyclic monoterpenes, bicyclic monoterpenes, diterpenoids, and triterpenoids [35]. There also are numerous reports on the chemical composition and the various biological activities of the *Origanum* species [37–42].

The several biological activities of *Origanum* species such as antioxidant, antimicrobial, antifungal, phytotoxic, and insecticidal effects were described previously [43–51]. However, it could be considered that the essential oils and extracts obtained from several *Origanum* species could be a useful synthetic herbicide alternative in modern agriculture.

The purpose of this study was to determine the herbicidal properties of essential oils and extracts isolated from *Origanum syriacum* L., *Origanum onites* L., and *Origanum majorana* L. on weed species that cause significant crop losses in agricultural production.

## 2. Materials and Methods

## 2.1. Plant Materials and Isolation of Essential Oils

Fresh plant materials of *Origanum syriacum*, *Origanum onites*, and *Origanum majorana* were collected at the flowering stage from the production areas located in the Kahramanmaraş region of Turkey. The plant materials were identified morphologically and voucher specimens were deposited in the herbarium of Atatürk University, Erzurum. The plant materials were dried in shadow at room temperature and ground to 0.1–0.4 mm by using a grinder.

To obtain the essential oils from the selected plants, the dried plant samples (100 g for each cycle, n = 5) were subjected to hydro-distillation using a Clevenger-type apparatus (DURAN<sup>®</sup>, Mainz, Germany) for 4 h. The obtained essential oils were extracted into chloroform and the water was removed by using dry sodium sulfate. Then chloroform was

removed by using a rotary evaporator (DURAN<sup>®</sup>, Mainz, Germany) under low temperature and pressure conditions.

In addition to the essential oils, the n-hexane and acetone extracts were obtained. To obtain the extracts from grounded plants, 100 g of dried plant materials were placed in a volumetric flask and 500 mL of n-hexane and acetone were added for each extraction process. The extractions process kept going for 48 h and the processes were repeated 4 times [52–57]. The supernatants were united and the organic solvent was evaporated under low temperature and pressure conditions by using a rotary evaporator.

The essential oils and the extracts were stored at 4 °C until further experiments. The yields of the essential oils and extracts (% referred to dry plant materials) are given in Table 1.

Species	Essential Oils —	Extracts	
		Acetone	<i>n</i> -Hexane
Origanum syriacum	4.0	13.8	14.3
Origanum onites	4.5	14.2	14.7
Origanum majorana	5.0	14.3	14.8

**Table 1.** The yields of essential oils and extracts (g/100 dried plant materials).

## 2.2. GC–MS Analysis

The essential oils were analyzed using a Thermofinnigan Trace GC/A1300 (E.I.) equipped with SGE/BPX5 MS capillary column (30 m  $\times$  0.25 mm i.d., 0.25 µm). Diluted samples (1/100, v/v, in methylene chloride) of 1.0 µL were injected in the splitless mode. Helium was used as the carrier gas, at a flow rate of 1 mL/min. The injector temperature was set at 220 °C. The program used was 50–150 °C at a rate of 3 °C/min, held isothermal for 10 min and finally raised to 250 °C at 10 °C/min [58,59].

## 2.3. Seed Germination and Seedling Growth Experiments In Vivo and In Vitro Conditions

The bio-herbicidal effects of the essential oil and extracts obtained from the *Origanum* specimen were tested against *Amaranthus retroflexus*, *Rumex crispus*, *Lactuca serriola*, and *Thlaspi arvense*. The seeds of weeds were collected in the Erzurum region (Turkey). Empty and undeveloped seeds were discarded by floating in tap water and the healthy seeds were selected to use in the experiments. To avoid possible inhibition caused by toxins from fungi or bacteria, the seeds were surface-sterilized with 15% sodium hypochlorite for 20 min [60] and then rinsed with abundant distilled water.

To determine herbicidal effects, the essential oils and extracts were dissolved in a 10% dimethyl sulfoxide (DMSO) (Sigma-Aldrich<sup>®</sup>, Darmstadt, Germany)-water solution, and the final concentrations of stock solutions were set as 5, 10, and 20  $\mu$ L/Petri dishes. The emulsions were transferred to a Petri dish (9 cm diameter) and placed on the bottom two layers of filter paper (10 mL/Petri dishes). Afterward, 50 disinfected seeds were placed on the filter paper [14,54]. Petri dishes were covered with adhesive tape to prevent volatile compounds from escaping. The Petri dishes were incubated at 23  $\pm$  2 °C and 80% humidity for 12 h consecutive dark and light periods in a growth chamber [61-63]. After 10 days, the number of germinated seeds was determined and the length of the seedling (root and shoot of seedlings) was measured by using a caliper. The germination rates were calculated as a percentage. Additionally, trifluralin (Maga-Tref 48 EC) (5, 10, and 20 µL/Petri) was used as the positive control. Petri dishes containing 10 mL dimethyl sulfoxide-water solution without the essential oils and extracts solutions were used as the negative control. A seed was considered as germinated when the emerging radicle elongated to 2 mm. Germination percentages were recorded every 24 h for 7 days. Rate of germination inhibition was calculated by using following formula:

$$GI = [GC - TG/GC] \times 100$$

where GI is rate of germination inhibition (%); GC is germination rate of control treatment; TG is germination rate in respective essential oil treatment of wheat genotypes or weed species.

The experimental design was a completely randomized design and all experiments were conducted thrice, including controls.

# 2.4. In Greenhouse Conditions

To test the herbicidal effects in in vitro conditions, twenty  $\mu L/pot$  dosage of essential oil and each extract obtained from O. syriacum, O. onites, and O. majorana were tested against the weeds that had 3–4 leaf stage and were growing in the greenhouse. The pots  $(10 \times 10 \text{ cm})$  were filled with 550 g of sterile soil (organic material ratio: 2.02%; cation change capacity: 43.34 me/100 g; pH = 7.5). Then, 50 seeds of the weeds were sown into the pots and kept under photoperiod conditions ( $23 \pm 2 \ ^{\circ}C$ , 12 h consecutive light and dark period) and relative humidity ( $80\% \pm 5$ ) in a growth chamber to allow germination and growth of the plant samples [16,64,65]. The pots were irrigated with tap water when necessary. The number of germinated seeds of the respective weed samples in each pot was counted. Afterward, the oil and extracts were emulsified in 10 mL of dimethyl sulfoxidewater solution (10% v/v). The final concentration of the treatments was 20  $\mu$ L/Pot. These emulsions were sprayed uniformly with a glass atomizer on the surface of whole plants in each pot in the stage of 2–4 real leaves. The plants in each pot sprayed uniformly with 10 mL of dimethyl sulfoxide-water solution (1%) were used as negative control groups. The plants sprayed with trifluralin (20 µL for each pot) were used as the positive control. Dead plants were counted and recorded at the 24th and 48th hour after sample applications. The treatments were arranged in a completely randomized design with three replications, including controls. The phytotoxicity of the treatments was expressed as the percent mean of dead plants [43].

The results are reported as the lethality percentage (LP) using the following formula:

$$LP = [N - n/N] \times 100$$

where N is number of healthy individuals before treatment; n is number of alive individuals after treatment.

#### 2.5. Statistical Analysis

The SPSS 10.0 software package was used to carry out the statistical test. The statistically significant differences among the herbicidal activity assays were analyzed in the Analysis of Variances (ANOVAs) test. When the statistical significance was observed, the Duncan test was used as a posthoc test.

#### 3. Results

#### 3.1. Chemical Composition of the Essential Oils

The essential oil components of *Origanum syriacum*, *O. onites* and *O. majorana* are given in Table 2. The major components of the essential oils were found as carvacrol representing 88.49% in *O. syriacum*, 58.65% in *O. onites*, 40.57% in *O. majorana*. *Origanum majorana* had a higher level of  $\alpha$ -Terpineol with 29.28% while *O. onites* included thymol with 30.97% (Table 2). Additionally, each essential oil contained relatively high amounts of oxygenated monoterpenes.

	<b>Essential Oil Compounds (%)</b>		
Compound	O. syriacum	O. onites	O. majorana
α-Pinene	0.30	-	0.40
Myrcene	0.21	-	0.57
3-Octanol	0.28	-	0.42
3-Octanone	-	-	0.17
α-Terpinene	0.37	0.19	0.85
<i>p</i> -Cymene	5.71	1.94	9.02
1,8-Čineole	-	-	2.20
γ-Terpinene	1.63	0.95	5.80
Terpinen-4-ol	0.65	0.58	2.15
α-Terpineol	0.27	0.21	29.28
Thymol	0.35	30.97	-
Carvacrol	88.49	58.65	40.57
Carvacrol methyl ether	-	-	3.46
β-Caryoplhyllene	1.48	0.98	1.76
Linalool	-	4.17	0.85
Borneol	-	0.64	-
	Class composition (	%)	
Monoterpene hydrocarbons	8.22	3.08	16.64
Oxygenated monoterpenes	89.76	95.22	78.51
Sesquiterpene hydrocarbons	1.48	0.98	1.76
Oxygenated hydrocarbons	-	-	-
Aliphatic compounds	0.28	-	0.59
Total	99.74	99.28	97.50

 Table 2. Chemical compositions of essential oils.

The essential oil rate of *Origanum syriacum* was found to be 99.74%. Eleven compounds from the essential oil of *O. syriacum* were identified, and these compounds are: 88.49% carvacrol; 5.71% *p*-Cymene; 1.63%  $\gamma$ -Terpinene; 1.48%  $\beta$ -Caryoplhyllene; 0.65% terpinen-4-ol; 0.37%  $\alpha$ -Terpinene; 0.35% thymol; 0.30%  $\alpha$ -Pinene; 0.28% 3-octanol; 0.27%  $\alpha$ -Terpineol; 0.21% Myrcene agents. The class composition of the essential oil was observed as monoterpene hydrocarbons 8.22%, oxygenated monoterpenes, 89.76%, sesquiterpene hydrocarbons 1.48%, aliphatic compound 0.28% (Table 2).

The essential oil contents of *Origanum onites* were found as 99.28%. The main compounds were 58.65% carvacrol; 30.97% thymol; 4.17% linalool; 1.94% *p*-Cymene; 0.98%  $\beta$ -Caryoplhyllene; 0.95%  $\gamma$ -Terpinene; borneol 0.64%; 0.58% terpinen-4-ol; 0.21%  $\alpha$ -Terpineol; 0.19%  $\alpha$ -Terpinene agents. The class compositions were monoterpene hydrocarbons 3.08%, oxygenated monoterpenes 95.22%, and sesquiterpenes hydrocarbons 0.98% (Table 2).

The essential oil contents of *Origanum majorana* were found as 97.50%. Fourteen of the main compounds were identified, and these compounds were carvacrol 40.57%,  $\alpha$ -Terpineol 29.28%, *p*-Cymene 9.02%,  $\gamma$ -Terpinene 5.8%, carvacrol methyl ether 3.46%, 1,8-Cineole 2.20%, terpinen-4-ol 2.15%,  $\beta$ -Caryoplhyllene 1.76%,  $\alpha$  -Terpinene 0.85%, linalool 0.85%, Myrcene 0.57%, 3-octanol 0.42%,  $\alpha$ -Pinene 0.40%, and 3-octanol 0.17%. The compound class was monoterpene hydrocarbons 16.64%, oxygenated monoterpenes 78.51%, and sesquiterpene hydrocarbons 1.76% (Table 2).

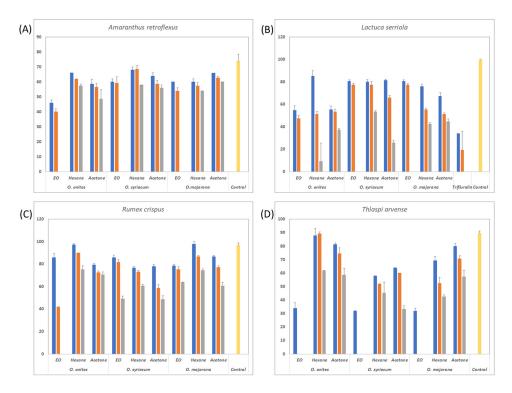
#### 3.2. Herbicidal Effects of the Oil and Extracts

The essential oils, *n*-hexane, and acetone of the extracts isolated from *O. syriacum*, *O. onites* and *O. majorana* were tested on seed germinations and seedling growths of *A. retroflexus*, *L. serriola*, *R. crispus*, and *T. arvense*, important weeds in cultivated areas in agriculture. Different degrees of the inhibition of germinations and seedling growths of the weeds were observed when compared with control groups.

The results showed that, in particular, the oils have a potent inhibitory effect on the seed germinations and seedling growths of all weeds tested. The current results also showed that the *n*-hexane and acetone extracts have a low herbicidal effect against the weeds tested as compared with those of the essential oils.

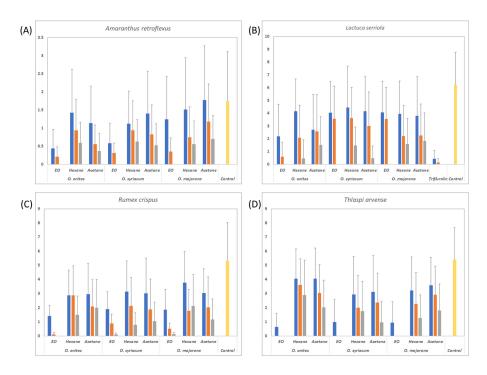
In general, the toxic effects of the extracts isolated from the aerial parts on the germinations and seedling growths of the weeds increased with an increase in the application concentrations of the extracts.

The essential oils and extracts obtained from the *Origanum* species and the application dosages have statistically significant effects on the seed germination rate of all tested weeds (p < 0.01). The seeds of *A. retroflexus*, *Lactuca serriola*, and *Rumex crispus* could not germinate with the 20 µL of essential oils of each tested *Origanum* species, whereas the seeds of *Thlaspi arvense* were not germinated with 10 µL essential oil application (Figure 1). The n-hexane and acetone extraction of all *Origanum* species were not effective as the tested essential oils (Figure 1), and none of them totally inhibited the seed germination of weeds.

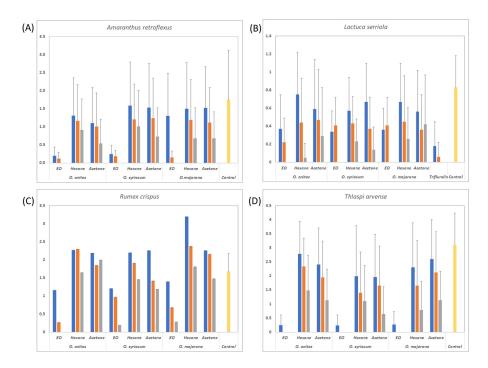


**Figure 1.** The seed germination rates (%) of (**A**) *Amaranthus retroflexus*, (**B**) *Lactuca serriola*, (**C**) *Rumex crispus*, (**D**) *Thlaspi arvense*. Blue bars indicate 5  $\mu$ L dose application while red bars 10  $\mu$ L, grey bars 20  $\mu$ L. The yellow bars indicate control application.

The essential oils obtained from all *Origanum* species inhibited root and shoot development (p < 0.01). In particular, the 20 µL dose of essential oils was effective for root and shoot development control. In addition, 10 µL of essential oils showed the best results for *Thlapsi arvense* root and shoot control (Figures 2 and 3). All extracts did not inhibit the root and shoot development.



**Figure 2.** The root development of the weeds (cm). (A) *Amaranthus retroflexus*, (B) *Lactuca serriola*, (C) *Rumex crispus*, (D) *Thlaspi arvense*. Blue bars indicate 5  $\mu$ L dose application while red bars 10  $\mu$ L, grey bars 20  $\mu$ L. The yellow bars indicate control application.



**Figure 3.** The shoot development of the weeds (cm). (A) *Amaranthus retroflexus*, (B) *Lactuca serriola*, (C) *Rumex crispus*, (D) *Thlaspi arvense*. Blue bars indicate 5  $\mu$ L dose application while red bars 10  $\mu$ L, grey bars 20  $\mu$ L. The yellow bars indicate control application.

In addition, the positive control of the experiment, Trifularin, inhibited totally the seed germination, root and shoot development of the weeds except for *Lactuca serriola* (the Trifularin was not shown in the Figure except for *L. serriola*). It was observed that only a 20  $\mu$ L dose of Trifularin totally inhibited the seed germination, and root and shoot development of *Lactuca serriola*.

In the nursery conditions, the essential oils and the extract can control weed development (p < 0.01). The tested essential oils showed better control against *Amaran*thus retroflexus after 24 - and 48 h application (Table 3). After 48 h of Trifluralin application, the mortality rate was observed as 76.67%, while *O. majorana* essential oils killed 79.33% of *A. retroflexus*. The essential oil obtained from *O. syriacum* and *O. onites* killed 73.33% of weeds. The extracts prepared with acetone yielded the best results from *O. onites and O. syriacum* species, while the extracts prepared with n-hexane yielded the best results from *O. majorana*.

Species	Applications	24 h	48 h
Origanum onites	Essential oil Acetone Hexane	$\begin{array}{c} 50.00 \pm 4.00  ^{\rm bcd} \ast \\ 37.33 \pm 4.16  ^{\rm ef} \\ 36.00 \pm 2.00  ^{\rm f} \end{array}$	$\begin{array}{c} \textbf{73.33} \pm 1.15 \ ^{\text{b}} \\ \textbf{61.33} \pm 2.31 \ ^{\text{d}} \\ \textbf{58.67} \pm 2.31 \ ^{\text{d}} \end{array}$
Origanum syriacum	Essential oil Acetone Hexane	$\begin{array}{l} 55.33 \pm 1.15 \; {}^{\rm bc} \\ 44.00 \pm 4.00 \; {}^{\rm de} \\ 42.00 \pm 4.00 \; {}^{\rm ef} \end{array}$	$\begin{array}{c} \textbf{79.33} \pm 1.15 \text{ a} \\ 68.00 \pm 2.00 \text{ c} \\ 66.67 \pm 1.15 \text{ c} \end{array}$
Origanum majorana	Essential oil Acetone Hexane	$\begin{array}{c} 56.67 \pm 0.58 \ ^{\rm b} \\ 49.33 \pm 1.15 \ ^{\rm cd} \\ 50.00 \pm 2.00 \ ^{\rm bcd} \end{array}$	$\begin{array}{c} \textbf{79.33} \pm 1.15 \text{ a} \\ 68.67 \pm 1.15 \text{ c} \\ 69.33 \pm 1.15 \text{ c} \end{array}$
	Control Trifluralin	$0.00 \pm 0.00$ g 69.33 $\pm$ 3.06 <sup>a</sup>	$0.00 \pm 0.00 \ ^{ m e}$ <b>76.67</b> $\pm$ 3.06 $^{ m ab}$

Table 3. The mortality rate (%) of Amaranthus retroflexus in nursery.

\* The letters indicate the Duncan test results.

The mortality rate of *Lactuca serriola* was observed at 90% 48 h after *O. onites* essential application, while Trifluralin killed 94% of plants (Table 4). The essential oil of *O. onites* showed the best herbicidal effects against *L. serriola*. In addition, the essential oils of *O. syriacum* and *O. majorana* can be useful for controlling *L. serriola* with 87.33% and 84.67%, respectively. The use of extracts from Origanum species in *L. serriola* greenhouse trials was similarly to *A. retroflexus* greenhouse trials.

Table 4. The mortality rate (%) of Lactuca serriola in nursery.

Species	Applications	24 h	48 h
Origanum onites	Essential oil Acetone Hexane	$\begin{array}{c} 62.67 \pm 1.15 \ ^{\text{b}*} \\ 52.67 \pm 1.15 \ ^{\text{c}} \\ 49.33 \pm 1.15 \ ^{\text{cd}} \end{array}$	$\begin{array}{c} \textbf{90.00} \pm 0.00 \; ^{ab} \\ 74.00 \pm 2.00 \; ^{c} \\ 76.00 \pm 0.00 \; ^{c} \end{array}$
Origanum syriacum	Essential oil Acetone Hexane	$\begin{array}{c} 63.33 \pm 3.06 \ ^{\rm b} \\ 50.67 \pm 1.15 \ ^{\rm cd} \\ 45.33 \pm 2.31 \ ^{\rm d} \end{array}$	$\begin{array}{c} \textbf{87.33} \pm 5.03 \ ^{\text{b}} \\ 74.67 \pm 1.15 \ ^{\text{c}} \\ 70.00 \pm 0.00 \ ^{\text{c}} \end{array}$
Origanum majorana	Essential oil Acetone Hexane	$71.33 \pm 1.15^{a} \\ 49.33 \pm 1.15^{cd} \\ 26.00 \pm 5.29^{e}$	$\begin{array}{c} \textbf{84.67} \pm 1.15 \ ^{\text{b}} \\ 70.67 \pm 2.31 \ ^{\text{c}} \\ 48.67 \pm 6.11 \ ^{\text{d}} \end{array}$
	Control Trifluralin	$0.00 \pm 0.00$ f 70.67 $\pm$ 1.15 <sup>a</sup>	$0.00 \pm 0.00 \ ^{ m e}$ 94.00 $\pm$ 2.00 $^{ m a}$

\* The letters indicate the Duncan test results.

After 48 h of the essential oil and the application of the extract against *Rumex crispus*, *Origanum onites* essential oil showed the best results to control this weed with an 81.33% mortality rate in the greenhouse conditions (Table 5). The mortality rate of *R. crispus* was observed as 71.33% for *O. syriacum* essential oil and 67.33% for *O. majorana* essential oil application. In addition, the n-hexane extraction of *O. majorana* showed better results than the essential oils of it. When the negative control application, Trifluralin, was applied, the

mortality rate of *R. crispus* was observed as 91.33%. The extracts obtained with acetone yielded the best results from *O. syriacum* and *O. majorana* species, while the extracts prepared with n-hexane yielded the best results from *O. onites*. Additionally, acetone extraction of all *Origanum* species outperformed n-hexane extraction.

Species	Applications	24 h	48 h
Origanum onites	Essential oil Acetone Hexane	$56.67 \pm 0.58$ <sup>b</sup> * 49.33 $\pm$ 1.15 <sup>bc</sup> 47.33 $\pm$ 1.15 <sup>c</sup>	$egin{array}{c} {\bf 81.33} \pm 1.15\ {}^{ m b} \\ {\bf 68.67} \pm 1.15\ {}^{ m c} \\ {\bf 68.67} \pm 1.15\ {}^{ m c} \end{array}$
Origanum syriacum	Essential oil Acetone Hexane	$\begin{array}{l} 55.33 \pm 1.15 \ ^{\rm bc} \\ 52.67 \pm 4.16 \ ^{\rm bc} \\ 46.67 \pm 1.15 \ ^{\rm c} \end{array}$	$\begin{array}{c} \textbf{71.33} \pm 1.15 \text{ c} \\ \textbf{70.67} \pm 1.15 \text{ c} \\ \textbf{66.67} \pm 3.06 \text{ c} \end{array}$
Origanum majorana	Essential oil Acetone Hexane	$\begin{array}{c} 52.67 \pm 6.43 \ ^{\rm bc} \\ 51.67 \pm 4.73 \ ^{\rm bc} \\ 50.67 \pm 6.11 \ ^{\rm bc} \end{array}$	$67.33 \pm 6.11 \text{ c} \\ 69.33 \pm 3.06 \text{ c} \\ 58.67 \pm 2.31 \text{ d} \\ \end{array}$
	Control Trifluralin	$0.00 \pm 0.00$ <sup>d</sup> 77.33 $\pm$ 1.15 <sup>a</sup>	$0.00 \pm 0.00 \ ^{ m e}$ 91.33 $\pm$ 1.15 $^{ m a}$

Table 5. The mortality rate (%) of Rumex crispus in nursery.

\* The letters indicate the Duncan test results.

*Thlaspi arvense* was killed by using the essential oils and extracts of *Origanum* species (Table 6). Similarly, the essential oils showed better results by comparing the extracts. After 48 h of the application of Trifluralin, 83.33% of *T. arvense* plants were dead, while the mortality rate of the plants was observed at 80.67% for *O. onites* essential oil, 78% for *O. syriacum* essential oil, and 77.33% for *O. majorana* essential oil. The extracts obtained with acetone yielded the best results from *O. majorana*, while the extracts prepared with n-hexane yielded the best results from *O. syriacum*.

Table 6. The mortality rate (%) of *Thlaspi arvense* in nursery.

Species	Applications	24 h	48 h
Origanum onites	Essential oil Acetone Hexane	$\begin{array}{l} 56.67 \pm 0.58 \ ^{\rm b} \\ 46.67 \pm 2.31 \ ^{\rm ef} \\ 48.67 \pm 1.15 \ ^{\rm de} \end{array}$	$\begin{array}{l} \textbf{80.67} \pm 1.15 \text{ ab} \\ 66.67 \pm 1.15 \text{ f} \\ 67.33 \pm 1.15 \text{ f} \end{array}$
Origanum syriacum	Essential oil Acetone Hexane	$\begin{array}{c} 56.67 \pm 0.58 \ ^{\rm b} \\ 44.00 \pm 2.00 \ ^{\rm f} \\ 50.67 \pm 1.15 \ ^{\rm cde} \end{array}$	$\begin{array}{l} \textbf{78.00} \pm 2.00 \ ^{\rm bc} \\ 68.67 \pm 1.15 \ ^{\rm ef} \\ \textbf{71.33} \pm 1.15 \ ^{\rm de} \end{array}$
Origanum majorana	Essential oil Acetone Hexane	$\begin{array}{c} 54.67 \pm 1.15 \ ^{\rm bc} \\ 49.33 \pm 1.15 \ ^{\rm de} \\ 52.67 \pm 1.15 \ ^{\rm bcd} \end{array}$	$\begin{array}{c} \textbf{77.33} \pm 1.15 ^{\text{c}} \\ \textbf{72.67} \pm 2.31 ^{\text{d}} \\ \textbf{70.67} \pm 1.15 ^{\text{de}} \end{array}$
	Control Trifluralin	$0.00 \pm 0.00 \text{ g} \\ 72.00 \pm 4.00 \text{ a}$	$0.00 \pm 0.00$ g 83.33 $\pm$ 1.15 a

\* The letters indicate the Duncan test results.

#### 4. Discussion

In this study, essential oil and crude extract content analyses from three *Organum* species were performed, and the effects of the obtained essential oils on seed germination and development against weeds of *Thlaspi arvense*, *Amaranthus retroflexus*, *Rumex crispus*, and *Lactuca serriola* were investigated.

The major component obtained from *O. syriacum* was carvacrol (88.49%), followed by other components: *p*-Cymene (5.71%),  $\gamma$ -terpinene (1.63),  $\beta$ -caryophyllene (1.48), Terpinen-4-ol (0.65%) (Table 2). The most abundant carvacrol component was found to be in previous studies: 81.38% [66], 60.80% [67], 22.29% [68], 60.01% [69], 39.87% [42], 74.21–90.22% [70], 82.60% [71], 44.49% [72], 61.18% [73], 35.80% [74].

In our study, the other main components, carvacrol [74,75], *p*-Cymene [66–71,73,75,76],  $\gamma$ -terpinene [42,66–71,73–75],  $\beta$ -caryophyllene [67,72,75], and terpinen-4 ol [66], were in line with previous studies.

The major component of obtained from *Origanum majorana* was carvacrol (40.57%), followed by other components:  $\alpha$ -Terpineol (28.29%), *p*-Cymene (9.02%), carvacrol methyl ether (3.46%), y-Terpinene (5.80%), 1,8 Cineole (1.20%) (Table 2.). The most abundant carvacrol component was found to be the first most abundant compound in previous studies: 84.00% [51], 78.27% [77], 75.30% [51], 40.57% [78], 34.14% [79]. In our study, the other main components,  $\alpha$ -Terpineol [45,78,80–82], *p*-Cymene [51,80,83–86],  $\alpha$ -Terpineol [45,78,80–82], y-Terpinene [46,51,77–81,83–88], and 1,8 Cineole [45,80,83,89,90], were in line with previous studies.

The major component of obtained from *Origanum onites* was carvacrol (58.65%), followed by other components: Thymol (30.97%), Linalool (4.17%), *p*-Cymene (1.94%),  $\gamma$ -terpinene (0.95%) (Table 2). The most abundant carvacrol component was found to be the first most abundant compound in previous studies: 88.71% [47], 83.30% [91], 81.01% [92], 78.40% [50], 72.12% [93], 59.87% [94], 57.63% [49], 57.01% [79] 47.99% [95], 26.91% [96].

In our study, the other main components, Thymol [91,92,94,96,97] *p*-Cymene [47,50,79, 91–97] Linalool [50,79,93,96], and  $\gamma$ -terpinene [47,49,50,79,91,92,94–96], were in line with previous studies.

The reasons for the differences in EO content and rates are geographical, climatic (macroclimatic–microclimatic factors), soil properties, as well as plant collection time [98,99], drying conditions, analysis methods, and geographical or ontogenesis variations [21,100–104].

In this study, the herbicidal effects of the essential oils and the extracts obtained from *Origanum syriacum*, *O. onites*, and *O. majorana* were determined against *Amaranthus retroflexus*, *Lactuca serriola*, *Rumex crispus*, and *Thlaspi arvense*, which cause several detrimental effects in the cropland. All *Origanum* essential oils tested in this study showed herbicidal effects on seed germination, and root and shoot development. In particular, the application of 10  $\mu$ L for *T. arvense* and 20  $\mu$ L for other weeds completely inhibited the seed germination, root, and shoot development. Moreover, 5  $\mu$ L essential oil application can be useful for weed control.

The extracts obtained from *Origanum* species comparatively need a higher application dose for totally herbicidal effects. Even though the tested dose of these extracts was not inhibited totally, it was observed that the higher dose of these extracts can be useful for controlling these weeds. In addition, it was observed that the herbicidal effects of the extraction agent differed depending on the weeds. For instance, the acetone extract of *O. onites* showed the best results for *T. arvense*, while the n-hexane extract of *O. onites* was more suitable for seed germination and shoot development of *R. crispus*.

In the nursery experiment,  $20 \ \mu$ L of essential oils and  $20 \ mg$  of each extract were used because this dose was the best effective application in Petri experiments. It was observed that the essential oils obtained from *Origanum* species could be a good bio-herbicide candidate against the tested weed.

Tanacetum aucheranum and Tanacetum chiliophyllum var. chiliophyllum 30 mL/Petri [30], Zataria multiflora Boiss. 320–640 mL/L<sup>-1</sup> [105], Satureja hortensis L. essential oil nanoemulsion (NE) 1000  $\mu$ L/L<sup>-1</sup> [17] determined that Amaranthus retroflexus weed species inhibited seed germination, and root and shoot development by 100%. In another study, (*Ruta* graveolens L.) and Bergamot (*Citrus bergamia* Risso et Poiteau) essential oils were found to be more toxic at doses of *A. retroflexus* L., 20  $\mu$ L/mL<sup>-1</sup>, inhibiting seed germination, and root and shoot development 100% [19]. Kordali et al. [43] found that the essential oil obtained from *Origanum acutidens* and carvacrol and thymol compounds in *A. retroflexus* 10  $\mu$ L/Petri application dose, 9.8  $\mu$ L/Petri carvacrol and 10  $\mu$ L/Petri thymol petri doses of *Amaranthus retroflexus* completely inhibited seed germination and seedling growth of *Nepeta meyeri* Benth. Kordali et al. [16,52] reported that EO *A. retroflexus* L., which was obtained, completely affected the germination of the *Myrtus communis* seed and the effect was in parallel with the dose amount. It was reported that *Thymus kotschyanus* EO  $\geq$  500 ppm inhibited the germination and subsequent development of *A. retroflexus* seeds and showed an herbicidal effect [106]. An application of 600 and 800 mg/L<sup>-1</sup> of *Foeniculum vulgare* essential oil showed the highest inhibitory effect against the *A. retroflexus* weed [107]. *Tagetes minuta* essential oil showed the highest inhibition of Amaranthus retrofexus seed germination at 600  $\mu$ L/L<sup>-1</sup> concentrations [21]. Yilar et al. [94] reported that *Origanum onites* essential oil completely inhibited seed germination, and root and shoot growth in *Amaranthus retroflexus* L. at a concentration of 15 L/Petri.

Kordali et al. [43] reported that the essential oil obtained from Origanum acutidens and carvacrol and thymol compounds in Rumex crispus 10 µL/Petri application dose, 9.8 µL/Petri carvacrol and 10 µL/Petri thymol petri doses of *Rumex crispus* completely inhibited seed germination and seedling growth. Doses of 10, 15, 20 g/cm<sup>2</sup> of essential oil obtained from Allium sativum germination and root growth of *Rumex crispus* 100% have inhibited, as have all doses of Cuminum cyminum L., Mentha longifolian essential oils (10, 15, 20 g/cm<sup>2</sup>) 100% [62]. For *Tanacetum aucheranum* and *Tanacetum chiliophyllum* var. chiliophyllum 30 mL/Petri [30], Tursun et al. [108] used essential oils of thyme (Origanum syriacum L.) and laurel (Laurus no bilis L.) and their main components, carvacrol, 1,8-cineole, and pinene, to investigate the inhibitory effects of essential oils on three separate weeds. The seed germination tests showed that oregano essential oil and carvacrol completely inhibited weed germination at all concentrations ranging from 1 to 5  $\mu$ L/Petri dish, whereas seed germination of test weeds decreased significantly with increasing concentrations of laurel essential oil and its main components, 1,8-cineol and -pinene ranging from 5 to 20 L/Petri dish. In another study, the most efficient essential oil dosages for inhibiting Amaranthus palmeri seed germination were determined to be 2–4  $\mu$ L/Petri dish. The most successful (100%) application of all essential oils in suppressing seed germination was achieved with an Origanum syriacum x Origanum onites hybrid grown at 800 ppm [109].

According to Said et al. [110], the component of *Thymus capitatus* essential oil has an allelopathic effect on Lactuca sativa L. seed germination. Eucalyptus globulus essential oil has the potential to be examined as a biological pesticide with phytotoxic effects against *Lactuca* sativa weed [111]. Ruiz-Vásquez et al. [112] reported that EOs from Peruvian casapiense, P. reticulatum, P. sancti-felicis, and P. mituense, effectively inhibited the root growth of Lactuca sativa. The essential oil obtained from *P. soledadense* caused a decrease in root growth of Lactuca sativa. These results show strong selective herbicidal potential of the EOs tested against monocotyledonous plants. Azizan et al. [113] reported that applying Wedelia trilobata essential oil to Lactuca sativa L. caused changes in fatty acid compositions and could lead to root growth inhibition, as well as providing information on L. sativa metabolic responses and the potential mechanisms of action of W. trilobata EO as bioherbicides. In another study, Origanum vulgare subsp hirtum Letswaart (Greek thyme) essential oil evaluated the seed germination inhibition of *Lolium perenne* L. and *Trifolium pratense* L. weeds in the form of aqueous solutions at concentrations ranging from 0.5 to 3.0  $\mu$ L/mL, and a 1.5  $\mu$ L/mL dose application provided complete inhibition. They also evaluated thyme oil in terms of its inhibitory activity on seed germination under field conditions. It was observed that the weight of the plants decreased by 77% as a result of the observations made 1 month after the essential oil was applied as an aqueous solution at 3, 5 and 10  $\mu$ L/mL concentrations on the super absorbent Teravet, mixed with the seeds of the target plants and planted in the field. They concluded that teravet (super-absorbent) application of essential oil is a good way of herbicidal effect in field conditions [114].

It is known that the essential oils and the extracts have bio-herbicidal effects on the weeds. In particular, the other *Origanum* species have a bio-herbicidal effect. For instance, Kordali et al. [43] tested thymol and carvacrol obtained from *Origanum acutidens* against *Amaranthus retfollexus*, and these components of the essential oil inhibited the seed germination and seedling development. In the present study, we observed a similar effect. Even though thymol and carvacrol contents were observed from *Origanum syriacum* and *O. onites* (Table 2), thymol could not be isolated from *O. majorana* while for carvacrol contents it was 40.57%. However, the essential oils and extracts of *O. majorana* have bio

herbicidal effects. Therefore, the other components of the essential oils obtained from *O. majorana* could inhibit the seed and seedling development.

The use of EOs in weed control is based on the fact that they contain allelochemical compounds, primarily terpenoids, that can inhibit weed species germination and growth [115,116]. Terpenoids, at least those that can disrupt mitosis, are a type of mitotic disrupting bioherbicide [117]. Monoterpenes were found in high concentrations in all of the EOs tested in our study.

Essential oils have phytotoxic potential in the form of hydrocarbons, alcohols, aldehydes, ketones, ethers, esters, peroxides, and phenols. This is attributed to terpenoids (primarily mono and 14 sesquiterpenes) as the main constituents [26–104,115–118].

Therefore, even though EO monoterpenes are known to be phytotoxic and to cause membrane integrity, cell division, and elongation, we observed non-incidental damage and necrosis in shoot and root tissues [119–121].

During using allelochemicals, the most studied response variables are weed seed shoot and root length [122]. The shoot and root lengths in the control treatment are generally longer than those in the treated seeds.

The results of the allelopathic activities of EOs and their effects on shoot and root lengths varied. In addition, when compared to the control treatment, the germinated seeds did not develop normally. Our findings confirmed that sprout and root length can vary depending on the type of EOs, concentrations tested, environmental conditions, and weed species [118–123]. Although essential oils appear to perform well in controlled laboratory conditions, their practical use for weed control in the field is limited due to their low water solubility and high volatility [118,124].

To overcome these drawbacks and improve essential oil performance, researchers recommend using essential oils as solid emulsions to prevent the loss of biological properties of essential oil components [124].

#### 5. Conclusions

In recent years, essential oils have been considered a chemical alternative herbicide [62,117,120–122]. However, the herbicidal efficiency of essential oils and extracts is changing, depending on application dose and weeds. The main reasons for these differences in essential oil herbicide efficacy may be component differences and target weed species, but also application dose. Therefore, the dose and formulations should be designed to directly use these essential oils and extracts in agricultural fields [115]. However, in order to get the best results from field applications, more comprehensive research on the ways in which essential oils can be applied to the target plants is necessary. The limitation of our study is that we did not test individual or combinations of essential oil components to determine the exact source of the herbicide effect. We believe that if different essential oil blends or direct single composition applications can be made in future studies, weed control will become more effective, reducing product loss from weeds in agriculture. However, the herbicidal potential of the *Origanum* species was observed.

**Author Contributions:** Conceptualization, S.K. and A.N.; methodology, S.K., G.K. and F.Y.; software, G.K.; validation, F.Y.; formal analysis, S.K., G.K., F.Y., İ.S. and A.N.; resources, İ.S.; data curation, S.K.; writing—original draft preparation, G.K., F.Y. and İ.S.; writing—review and editing, S.K. and A.N.; visualization, G.K., F.Y. and İ.S.; supervision, S.K. and A.N.; project administration, S.K. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data sharing is not applicable to this article.

Conflicts of Interest: The authors declare no conflict of interest.

# References

- 1. Chauhan, B.S. Grand challenges in weed management. Front. Agron. 2020, 1, 1–4. [CrossRef]
- 2. Majrashi, A.A. Preliminary assessment of weed population in vegetable and fruit farms of Taif, Saudi Arabia. *Braz. J. Biol.* 2022, *82*, 1–9. [CrossRef]
- 3. Jabran, K.; Mahajan, G.; Sardana, V.; Chauhan, B.S. Allelopathy for weed control in agricultural systems. *Crop. Prot.* 2015, 72, 57–65. [CrossRef]
- 4. Kubiak, A.; Wolna-Maruwka, A.; Niewiadomska, A.; Pilarska, A.A. The Problem of Weed Infestation of Agricultural Plantations vs. the Assumptions of the European Biodiversity Strategy. *Agronomy* **2022**, *12*, 1808. [CrossRef]
- Zhao, X.; Cui, H.; Wang, Y.; Sun, C.; Cui, B.; Zeng, Z. Development strategies and prospects of nano-based smart pesticide formulation. J. Agric. Food Chem. 2018, 66, 6504–6512. [CrossRef]
- Liang, J.; Yu, M.; Guo, L.; Cui, B.; Zhao, X.; Sun, C.; Wang, Y.; Liu, G.; Cui, H.; Zeng, Z. Bioinspired development of P(St–MAA)– avermectin nanoparticles with high affinity for foliage to enhance folia retention. *J. Agric. Food Chem.* 2018, 66, 6578–6584. [CrossRef]
- 7. Sarkar, M.R.; Rashid, M.H.O.; Rahman, A.; Kafi, M.A.; Hosen, M.I.; Rahman, M.S.; Khan, M.N. Recent advances in nanomaterials based sustainable agriculture: An overview. *Environ. Nanotechnol. Monit. Manag.* **2022**, *18*, 100687.
- 8. Islam, F.; Wang, J.; Farooq, M.A.; Khan, M.S.; Xu, L.; Zhu, J.; Zhao, M.; Stéphane, M. Potential impact of the herbicide 2,4dichlorophenoxyacetic acid on human and ecosystems. *Environ. Int.* **2018**, *111*, 332–351. [CrossRef]
- 9. Böcker, T.; Möhring, N.; Finger, R. Herbicide free agriculture? A bio-economic modelling application to Swiss wheat production. *Agric. Syst.* **2019**, *173*, 378–392. [CrossRef]
- 10. Tandon, S.; Pant, R. Kinetics of diuron under aerobic condition and residue analysis in sugarcane under subtropical field conditions. *Environ. Technol.* **2019**, *40*, 86–93. [CrossRef]
- 11. Mohanty, S.S.; Jena, H.M. A systemic assessment of the environmental impacts and remediation strategies for chloroacetanilide herbicides. *J. Water Process Eng.* **2019**, *31*, 100860. [CrossRef]
- 12. Duke, S.O.; Dayan, F.E.; Romagni, J.G.; Rimando, A.M. Natural Products as Sources of Herbicides: Current Status and Future Trends. *Weed Res.* 2000, 40, 99–111. [CrossRef]
- 13. Chhokar, R.S.; Sharma, R.K.; Chauhan, D.S.; Mongia, A.D. Evaluation of herbicides against *Phalaris minor* in wheat in north-western. Indian plains. *Weed Res.* 2006, *46*, 40–49. [CrossRef]
- 14. Kordali, S.; Cakir, A.; Sutay, S. Inhibitory Effects of Monoterpenes on Seed Germination and Seedling Growth. Z. Naturforsch. C 2007, 62, 207–214. [CrossRef]
- Salamci, E.; Kordali, S.; Kotan, R.; Cakir, A.; Kaya, Y. Chemical Composition, Antimicrobial and herbicidal effects of essential oils isolated from Turkish *Tanacetum aucheranum* and *Tanacetum chiliophyllum* var. *chiliophyllum*. *Biochem. Syste. Ecol.* 2007, 35, 569–581. [CrossRef]
- 16. Kordali, S.; Usanmaz, A.; Cakir, A.; Komaki, A.; Ercisli, S. Antifungal and herbicidal effects of fruit essential oils of four *Myrtus communis* genotypes. *Chem. Biodivers.* **2016**, *13*, 77–84. [CrossRef]
- Hazrati, H.; Saharkhiz, M.J.; Niakousari, M.; Moein, M. Natural herbicide activity of *Satureja hortensis* L. essential oil nanoemulsion on the seed germination and morphophysiological features of two important weed species. *Ecotoxicol. Environ. Saf.* 2017, 142, 423–430. [CrossRef]
- Kaab, S.B.; Lins, L.; Hanafi, M.; Rebey, I.B.; Deleu, M.; Fauconnier, M.L.; Ksouri, R.; Jijacli, M.H.; De Clerck, C. *Cynara cardunculus* crude extract as a powerful natural herbicide and insight into the mode of action of its bioactive molecules. *Biomolecules* 2020, 10, 1–17.
- 19. Bozhuyuk, A.U. Herbicidal Activity and Chemical Composition of Two Essential Oils on Seed Germinations and Seedling Growths of Three Weed Species. *J. Essent. Oil Bear. Plants* **2020**, *23*, 821–831. [CrossRef]
- 20. De Mastro, G.; El Mahdi, J.; Ruta, C. Bioherbicidal potential of the essential oils from mediterranean lamiaceae for weed control in organic farming. *Plants* **2021**, *10*, 818. [CrossRef]
- 21. Taban, A.; Somayeh Rastegar, S.; Nasirzadeh, M.; Saharkhiz, M.J. Essential oil composition and comparative phytotoxic activity of fennel, summer savory, Mexican marigold and feverfew: A potential bioherbicide. *Vegetos* **2022**, *35*, 502–510. [CrossRef]
- 22. Ju, J.; Chen, X.; Xie, Y.; Yu, H.; Guo, Y.; Cheng, Y.; Qian, H.; Yao, W. Application of essential oil as a sustained release preparation in food packaging. *Trends Food Sci. Technol.* **2019**, *92*, 22–32. [CrossRef]
- Barton, A.F.M.; Dell, B.; Knight, A.R. Herbicidal activity of cineole derivatives. J. Agric. Food Chem. 2010, 58, 10147–10155. [CrossRef]
- 24. Jouini, A.; Verdeguer, M.; Pinton, S.; Araniti, F.; Palazzolo, E.; Badalucco, L.; Laudicina, V.A. Potential effects of essential oils extracted from mediterranean aromatic plants on target weeds and soil microorganisms. *Plants* **2020**, *9*, 1289. [CrossRef]
- Gokturk, T.; Chachkhiani-Anasashvili, N.; Kordali, S.; Dumbadze, G.; Bozhuyuk, A.U. Insecticidal effects of some essential oils against box tree moth (Cydalima perspectalis Walker (Lepidoptera: Crambidae)). *Int. J. Trop. Insect Sci.* 2020, 40, 637–643. [CrossRef]
- 26. Verdeguer, M.; Sanchez-Moreiras, A.M.; Araniti, F. Phytotoxic effects and mechanism of action of essential oils and terpenoids. *Plants* **2020**, *9*, 1571. [CrossRef]
- 27. Werrie, P.Y.; Durenne, B.; Delaplace, P.; Fauconnier, M.L. Phytotoxicity of essential oils: Opportunities and constraints for the development of biopesticides. A Review. *Foods* **2020**, *9*, 1291. [CrossRef]

- Dirmenci, T.; Yazici, T.; Özcan, T.; Çelenk, S.; Martin, E. A New Species and a New Natural Hybrid of *Origanum* L. (Lamiaceae) from the West of Turkey. *Turk. J. Bot.* 2018, 42, 73–90. [CrossRef]
- 29. Dirmenci, T.; Özcan, T.; Yazıcı, T.; Arabacı, T.; Martin, E. Morphological, cytological, palynological and molecular evidence on two new hybrids: An example of homoploid hybridization in *Origanum* (Lamiaceae). *Phytotaxa* **2018**, *371*, 145–167. [CrossRef]
- Dirmenci, T.; Özcan, T.; Acar, M.; Arabacı, T.; Yazıcı, T.; Martin, E. A rearranged homoploid hybrid species of *Origanum* (Lamiaceae): O. × *munzurense* Kit Tan & Sorger. *Bot. Lett.* 2019, *166*, 153–162.
- 31. Arabaci, T.; Dirmenci, T.; Yıldız, B. A New Hybrid of the Genus *Origanum* L. (Lamiaceae): Origanum × malatyanum. *Bagbahce Bilim Derg.* **2020**, *7*, 10–15.
- 32. Baytop, T. Türkiye'de Bitkiler ile Tedavi, Geçmişte ve Bugün; Nobel Tıp Kitabevleri: İstanbul, Türkiye, 1999.
- Bozdemir, Ç. Economic Importance and Usage Fields of Oregano Species Growing in Turkey. Yüzüncü Yil Üniv. J. Agr. Sci. 2019, 29, 583–594.
- TÜİK 2019. Available online: <a href="https://www.tuik.gov.tr">https://www.tuik.gov.tr</a> (accessed on 6 February 2020).
- 35. Marrelli, M.; Statti, G.A.; Conforti, F. *Origanum* spp.: An update of their chemical and biological profiles. *Phytochem. Rev.* **2018**, 17, 873–888. [CrossRef]
- 36. Naquvi, K.J.; Ahamad, J.; Salma, A.; Ansari, S.H.; Najmi, A.K. A critical review on traditional uses, phytochemistry and pharmacological uses of *Origanum vulgare* Linn. *Int. Res. J. Pharm.* **2019**, *10*, 7–11. [CrossRef]
- Gong, H.Y.; Liu, W.H.; LV, G.Y.; Zhou, X. Analysis of Essential Oils of *Origanum vulgare* from Six Production Areas of China and Pakistan. *Rev. Bras. Farmacogn.* 2014, 24, 25–32. [CrossRef]
- Sokmen, M.; Serkedjleva, J.; Dalerera, D.; Gulluce, M.; Pollsslou, M.; Tape, B.; Akpulat, H.A.; Sahin, F.; Sökmen, A. In vitro antioxidant, antimicrobial and antiviral activities of the essential oil and various extracts from herbal parts and callus cultures of Origanum acutidens. J. Agric. Food Chem. 2004, 52, 3309–3312. [CrossRef]
- Figueredo, G.; Özcan, M.M.; Chalchat, J.C.; Bağcı, Y.; Chalard, P. Chemical Composition of Essential Oil of Hyssopus officinalis L. and Origanum acutidens. J. Essent. Oil Bear. Plants 2012, 15, 300–306. [CrossRef]
- Kizil, S.; Hasimi, N.; Tolan, V. Biological Activities of Origanum, Satureja, Thymbra and Thymus Species Grown in Turkey. J. Essent. Oil Bear. Plants 2014, 17, 460–468. [CrossRef]
- 41. Aseyd Nezhad, S.; Es-haghi, A.; Tabrizi, M.H. Green synthesis of cerium oxide nanoparticle *using Origanum majorana L.* leaf extract, its characterization and biological activities. *Appl. Organomet. Chem.* **2019**, *34*, 1–10. [CrossRef]
- Shehadeh, M.; Jaradat, N.; Al-Masri, M.; Naser Zaid, A.; Hussein, F.; Khasati, A.; Darwish, R. Rapid, cost-effective and organic solvent-free production of biologically active essential oil from Mediterranean wild *Origanum Syr. Saudi Pharm. J.* 2019, 27, 612–618. [CrossRef]
- Kordali, S.; Cakir, A.; Ozer, H.; Cakmakcı, R.; Kesdek, M.; Mete, E. Antifungal, Phytotoxic, and Insecticidal Properties of Essential Oil Isolated from Turkish Origanum acutidens and Three Components, cavacrol, thymol and *p*-Cymene. *Bioresour. Technol.* 2008, 99, 8788–8795. [CrossRef]
- Tozlu, E.; Cakir, A.; Kordali, S.; Tozlu, G.; Ozer, H.; Aytas Akcin, T. Chemical compositions and insecticidal effects of essential oils isolated from *Achillea gypsicola*, *Satureja hortensis*, *Origanum acutidens* and *Hypericum scabrum* against broadbean weevil (*Bruchus dentipes*). Sci. Hortic. 2011, 130, 9–17. [CrossRef]
- 45. Baj, T.; Baryluk, A.; Sieniawska, E. Application of mixture design for optimum antioxidant activity of mixtures of essential oils from *Ocimum basilicum* L., *Origanum majorana* L. and *Rosmarinus officinalis* L. *Indus. Crops Prod.* **2018**, *115*, 52–61. [CrossRef]
- Dhaouadi, S.; Rouissi, W.; Mougou-Hamdane, A.; Hannachi, I.; Nasraoui, B. Antifungal activity of essential oils of *Origanum* majorana and Lavender angustifolia against Fusarium wilt and root rot disease of melon plants. *Tunis. J. Plant Prot.* 2018, 13, 39–55.
- Erenler, R.; Demirtas, I.; Karan, T.; Gul, F.; Kayir, O.; Karakoc, O.C. Chemical constituents, quantitative analysis and insecticidal activities of plant extract and essential oil from *Origanum onites* L. *Trends Phytochem. Res.* 2018, 2, 91–96.
- Della Pepa, T.; Elshafie, H.S.; Capasso, R.; De Feo, V.; Camele, I.; Nazzaro, F.; Scognamiglio, M.S.; Caputo, L. Antimicrobial and Phytotoxic Activity of *Origanum heracleoticum* and *O. majorana* Essential Oils Growing in Cilento (Southern Italy). *Molecules* 2019, 24, 2576. [CrossRef]
- 49. Karan, T.; Belguzar, S.; Selvi, B. Antibacterial Activity of Essential Oils of *Origanum bilgeri*, *Origanum onites*, *Satureja spicigera* Leaves Against Agricultural Plant Pathogenic Bacteria. J. Essent. Oil Bear. Plants **2021**, 24, 1159–1168. [CrossRef]
- Becer, E.; Altundag, E.M.; Başer, K.H.C.; Vatansever, H.S. Cytotoxic activity and antioxidant effects of *Origanum onites* essential oil and its two major contents, carvacrol and *p*-Cymene on human colorectal (HCT116) and hepatocelluler carcinoma (HepG2) cell lines. *J. Essent. Oil Res.* 2022, 34, 1–10. [CrossRef]
- Kaskatepe, B.; Aslan Erdem, S.; Ozturk, S.; Safi Oz, Z.; Subasi, E.; Koyuncu, M.; Vlainic, J.; Kosalec, I. Antifungal and Anti-Virulent Activity of *Origanum majorana* L. Essential Oil on Candida albicans and In Vivo Toxicity in the Galleria mellonella Larval Model. *Molecules* 2022, 27, 663. [CrossRef]
- 52. Kordali, S.; Tazegül, A.; Cakir, A. Phytotoxic Effects of *Nepeta meyeri* Benth. Extracts and Essential Oil on Seed Germinations and Seedling Growths of Four Weed Species. *Rec. Nat. Prod.* **2015**, *9*, 404–418.
- Özer, Z.; Gören, A.C.; Kılıc, T.; Öncü, M.; Çarıkçı, S.; Dirmenci, T. The phenolic contents, antioxidant and anticholinesterase activity of section *Amaracus* (Gled.) Vogel and Anatolican Ietsw. of *Origanum* L. species. *Arab. J. Chem.* 2020, 13, 5027–5039. [CrossRef]

- 54. Gokturk, T.S.; Kordali, S.; Usanmaz Bozhuyuk, A. Insecticidal effects of essential oils against nymphal and adult stage of Ricania simulans (Hemiptera: Ricanidae). *Nat. Prod. Commun.* **2017**, *12*, 973–976. [CrossRef]
- Kordali, S.; Cakir, A.; Akcin, T.A.; Mete, E.; Aydin, T.; Kilic, H. Antifungal and herbicidal properties of essential oils and n-hexane extracts of Achillea gypsicola Hub-Mor. and Achillea biebersteinii Afan. (Asteraceae). *Ind. Crop. Prod.* 2009, 29, 562–570. [CrossRef]
- 56. Ramluckan, K.; Moodley, K.G.; Bux, F. An evaluation of the efficacy of using selected solvents for the extraction of lipids from algal biomass by the soxhlet extraction method. *Fuel* **2014**, *116*, 103–108. [CrossRef]
- 57. Yildirim, B.A.; Kordali, S.; Yildirim, S.; Yildirim, F.; Ercisli, S. Antidiabetic and Antioxidant Effects of *Vitis Vinifera* L. Cv. 'Kara Erik' Seed Extract in Strept Ozotocin Diabetic Rats. *Oxid. Commun.* **2017**, *40*, 209–219.
- Sarikurkcu, C.; Ceylan, O.; Zeljković, S.Ć. Micromeria myrtifolia: Essential Oil Composition and Biological Activity. Nat. Prod. Commun. 2019, 14, 1934578X19851687. [CrossRef]
- 59. Cetin, B.; Cakmakci, S.; Cakmakci, R. The investigation of antimicrobial activity of thyme and oregano essential oils. *Turk. J. Agric. For.* **2011**, *35*, 145–154.
- Çakır, A.; Özer, H.; Aydın, T.; Kordali, Ş.; Çavuşoğlu, A.; Akçin, T.; Mete, E.; Akçin, A. Phytotoxic and Insecticidal Properties of Essential Oils and Extracts of Four Achillea Species. *Rec. Nat. Prod.* 2016, 10, 154–167.
- 61. Tworkoski, T. Herbicide effects of essential oils. Weed Sci. 2002, 50, 425–431. [CrossRef]
- 62. Üstüner, T.; Kordali, S.; Usanmaz Bozhüyük, A. Herbicidal and Fungicidal Effects of *Cuminum cyminum*, *Mentha longifolia* and *Allium sativum* Essential Oils on Some Weeds and Fungi. *Rec. Nat. Prod.* **2018**, *56*, 619–629. [CrossRef]
- 63. Fradi, A.J.; Al-Araji, A.M.Y. Effect of *Eucalyptus camaldulensis* terpenes, alkaloids and phenols against *Fusarium oxysporum*. *Iraqi J. Sci.* **2015**, *56*, 2807–2810.
- 64. Harčárová, M.; Čonková, E.; Proškovcová, M.; Váczi, P.; Marcinčáková, D.; Bujňák, L. Comparison of antifungal activity of selected essential oils against *Fusarium graminearum* in vitro. *Ann Agric Environ. Med.* **2021**, *28*, 414–418. [CrossRef]
- 65. Dudai, N.; Chaimovitsh, D.; Larkov, O.; Fischer, R.; Blaicher, Y.; Mayer, A. Allelochemicals released by leaf residues of *Micromeria fruticosa* in soils, their uptake and metabolism by inhibited wheat seed. *Plant Soil* **2009**, *314*, 311–317. [CrossRef]
- Gendy, A.N.E.; Leonardi, M.; Mugnaini, L.; Bertelloni, F.; Ebani, V.V.; Nardoni, S.; Mancianti, F.; Hendawy, S.; Omer, E.; Pistelli, L. Chemical composition and antimicrobial activity of essential oil of wild and cultivated *Origanum syriacum* plants grown in Sinai, Egypt. *Ind. Crops Prod.* 2015, 67, 201–207. [CrossRef]
- 67. Al Hafi, M.; El Beyrouthy, M.; Ouaini, N.; Stien, D.; Rutledge, D.; Chaillou, S. Chemical Composition and Antimicrobial Activity of *Origanum libanoticum*, *Origanum ehrenbergii*, and *Origanum syriacum* Growing Wild in Lebanon. *Chem. Biodivers* **2016**, *13*, 555–560. [CrossRef]
- Karan, T.; Simsek, S.; Yildiz, I.; Erenler, R. Chemical Composition and Insecticidal Activity of Origanum syriacum L. Essential Oil Against Sitophilus oryzae and Rhyzopertha dominica. Int. J. Second. Metab. 2018, 5, 87–93.
- El Khoury, R.; Michael Jubeli, R.; El Beyrouthy, M.; Baillet Guffroy, A.; Rizk, T.; Tfayli, A.; Lteif, R. Phytochemical screening and antityrosinase activity of carvacrol, thymoquinone, and four essential oils of Lebanese plants. J. Cosmet. Derm. 2019, 18, 944–952. [CrossRef]
- Badawy, A.A.; El-mohandes, M.A.; Algharib, A.M.; Hatab, B.E.; Omer, E.A. The essential oil and its main constituents of *Origanum* syriacum ssp. sinaicum grown wild in Saint Katherine Protectorate, South Sinai, Egypt. *Al-Azhar J. Agric. Res.* 2020, 45, 116–131. [CrossRef]
- 71. Kavallieratos, N.G.; Boukouvala, M.C.; Ntalli, N.; Skourti, A.; Karagianni, E.S.; Nika, E.P.; Kontodimasd, D.C.; Cappellaccie, L.; Petrellie, R.; Cianfaglionef, K.; et al. Effectiveness of eight essential oils against two key stored-product beetles, *Prostephanus truncatus* (Horn) and *Trogoderma granarium* Everts. *Food Chem. Toxicol.* 2020, *139*, 1–13. [CrossRef]
- 72. Alonazi, M.A.; Jemel, I.; Moubayed, N.; Alwhibi, M.; El-Sayed, N.N.E.; Ben Bacha, A. Evaluation of the in vitro anti-inflammatory and cytotoxic potential of ethanolic and aqueous extracts of *Origanum syriacum* and *Salvia lanigera* leaves. *Environ. Sci. Pollut Res. Int.* **2021**, *28*, 19890–19900. [CrossRef]
- Hassan, Y.A.; Khedr, A.I.M.; Alkabli, J.; Elshaarawy, R.F.M.; Nasr, A.M. Co-delivery of imidazolium Zn(II)salen and *Origanum* syriacum essential oil by shrimp chitosan nanoparticles for antimicrobial applications. *Carbohyd. Polym.* 2021, 260, 117834. [CrossRef]
- 74. El-Alam, I.; Zgheib, R.; Iriti, M.; El Beyrouthy, M.; Hattouny, P.; Verdin, A.; Fontaine, J.; Chahine, R.; Hadj Sahraoui, A.L.; Makhlouf, H. *Origanum syriacum* Essential Oil Chemical Polymorphism According to Soil Type. *Foods* **2019**, *8*, 90. [CrossRef]
- Zgheib, R.; Chaillou, S.; Ouaini, N.; Kassouf, A.; Rutledge, D.; El Azzi, D.; El Beyrouthy, M. Chemometric Tools to Highlight the Variability of the Chemical Composition and Yield of Lebanese *Origanum syriacum* L. *Essent. Oil. Chem. Biodivers.* 2016, 13, 1326–1347. [CrossRef]
- AL-Mariri, A.; Odeh, A.; Alobeid, B.; Boukai, H. In vitro antibacterial activity of *Origanum syriacum* L. essential oils against gram-negative bacteria. *Avicenna J. Clin. Microbiol. Infect* 2019, 6, 26–30. [CrossRef]
- 77. Jan, S.; Mir, J.I.; Shafi, W.; Faktoo, S.Z.; Singh, D.B.; Wijaya, L.; Alyemeni, M.N.; Ahmad, P. Divergence in tissue-specific expression patterns of genes associated with the terpeniod biosynthesis in two oregano species *Origanum vulgare* L., and *Origanum majorana*. *Ind. Crops Prod.* **2018**, *123*, 546–555. [CrossRef]
- 78. Abbasi-Maleki, S.; Kadkhoda, Z.; Taghizad-Farid, R. The antidepressant-like effects of *Origanum majorana* essential oil on mice through monoaminergic modulation using the forced swimming test. *J. Tradit Complement Med.* **2020**, *10*, 327–335. [CrossRef]

- 79. Demirel, N.; Erdoğan, C. Insecticidal effects of essential oils from Labiatae and Lauraceae families against cowpea weevil, *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae) in stored pea seeds. *Entomol. Appl. Sci. Lett.* **2017**, *4*, 13–19.
- 80. Waller, S.B.; Madrid, I.M.; Ferraz, V.; Picoli, T.; Cleff, M.B.; de Faria, R.O.; Meireles, M.C.A.; de Mello, J.R.B. *Cytotox* and anti-Sporothrix brasiliensis activity of the Origanum majorana Linn. oil. *Braz. J. Microbiol.* **2016**, *47*, 896–901. [CrossRef]
- Radaelli, M.; da Silvaa, B.P.; Weidlicha, L.; Hoehne, L.; Flach, A.; da Costac, L.A.M.; Ethur, E.M. Antimicrobial activities of six essential oils commonly used as condiments in Brazil against *Clostridium perfringens*. *Braz. J. Microbiol.* 2016, 47, 424–430. [CrossRef]
- Elmhalli, F.; Garboui, S.S.; Karlson, A.K.B.; Mozūraitis, R.; Baldauf, S.L.; Grandi, G. Acaricidal activity against *Ixodes ricinus* nymphs of essential oils from the Libyan plants *Artemisia herba* alba, *Origanum majorana* and *Juniperus phoenicea*. *Vet. Parasitol: Reg. Stud. Rep.* 2021, 24, 100575. [CrossRef]
- Waller, S.B.; Rıpoll, M.K.; Sılva, A.L.; Serra, E.F.; Dıas, T.P.; Neves, V.B.D.; Melo, L.P.D.; Lindemann, P.; De Almeida, M.O.; Gomes, A.; et al. Activities and mechanisms of oregano, marjoram and rosemary essential oils against *Malassezia pachydermatitis* isolates from canine and feline otitis. *Turk. J. Vet. Anim. Sci.* 2022, 46, 549–558.
- Dantas, A.; dos, S.; Klein-Júnior, L.C.; Machado, M.S.; Guecheva, T.N.; Santos dos, L.D.; Zanette, R.A.; de Mello, F.B.; Henriques, J.A.P.; de Mello, J.R.B. Origanum majorana Essential Oil Lacks Mutagenic Activity in the Salmonella/Microsome and Micronucleus Assays. Sci. World J. 2016, 2016, 1–7.
- Ouedrhiri, W.; Balouiri, M.; Bouhdid, S.; Moja, S.; Chahdi, F.O.; Taleb, M.; Greche, H. Mixture design of *Origanum compactum*, *Origanum majorana* and *Thymus serpyllum* essential oils: Optimization of their antibacterial effect. *Ind. Crops Prod.* 2019, 89, 1–9. [CrossRef]
- Ghazal, T.S.A.; Schelz, Z.; Vidács, L.; Szemerédi, N.; Veres, K.; Spengler, G.; Hohmann, J. Antimicrobial, Multidrug Resistance Reversal and Biofilm Formation Inhibitory Effect of *Origanum majorana* Extracts, Essential Oil and Monoterpenes. *Plants* 2022, 11, 1432. [CrossRef]
- Mossa, A.T.; Nawwar, G. Free radical scavenging and antiacetylcholinesterase activities of *Origanum majorana* L. essential oil. Hum. *Exp. Toxicol.* 2011, 30, 1501–1513. [CrossRef]
- Kimera, F.; Sewilam, H.; Fouad, W.M.; Suloma, A. Efficient utilization of aquaculture effluents to maximize plant growth, yield, and essential oils composition of Origanum majorana cultivation. *Ann. Agric. Sci.* 2021, 66, 1–7. [CrossRef]
- 89. Amor, G.; Caputo, L.; La Storia, A.; De Feo, V.; Mauriello, G.; and Fechtali, T. Chemical Composition and Antimicrobial Activity of *Artemisia herba-alba* and *Origanum majorana* Essential Oils from Morocco. *Molecules* **2019**, 24, 4021. [CrossRef]
- Mady, H.Y.; Ahmed, M.M.; El Namaky, A.H. Efficiency of Origanum majorana essential oil as insecticidal agent against Rhynchophorus ferrugineus the red palm weevil (Olivier) (Coleoptera: Curculionidae). J. Biopest 2021, 14, 32–40.
- 91. Stefanaki, A.; Cook, C.M.; Lanaras, T.; Kokkini, S. The Oregano plants of Chios Island (Greece): Essential oils of *Origanum onites* L. growing wild in different habitats. *Ind. Crops Prod.* **2016**, *82*, 107–113. [CrossRef]
- 92. Ozdemir, R.C.; Taştan, Y.; Guney, K. Prevention of Saprolegniasis in rainbow trout (*Oncorhynchus mykiss*) eggs using oregano (*Origanum onites*) and laurel (*Laurus nobilis*) essential oils. *J. Fish Dis.* **2022**, 45, 51–58. [CrossRef]
- 93. Yigit, N.O.; Kocaayan, H. Efficiency of thyme (*Origanum onites*) and coriander (*Coriandrum sativum*) essential oils on anesthesia and histopathology of rainbow trout (*Oncorhynchus mykiss*). Aquaculture **2023**, 52, 738813. [CrossRef]
- Yilar, M.; Bayar, Y.; Onaran, A. Chemical composition and allelopathic effect of *Origanum onites* L. essential oil. *Plant Prot. Bull.* 2019, 59, 71–78.
- Spyridopoulou, K.; Fitsiou, E.; Bouloukosta, E.; Tiptiri-Kourpeti, A.; Vamvakias, M.; Oreopoulou, A.; Papavassilopoulou, E.; Pappa, A.; Chlichlia, K. Extraction, Chemical Composition, and Anticancer Potential of *Origanum onites* L. Essential Oil. *Molecules* 2019, 24, 2612. [CrossRef]
- 96. Sarıkaya, A.G. Leaf and Flower Volatile Oil Components of Two Thyme Taxa *Origanum onites* L. and *Thymbra spicata* var. spicata L. in Turkey. *Eur. J. Sci. Technol.* **2019**, *17*, 346–350. [CrossRef]
- Ozdemir, N.; Ozgen, Y.; Kiralan, M.; Bayrak, A.; Arslan, N.; Ramadan, M.F. Effect of different drying methods on the essential oil yield, composition and antioxidant activity of *Origanum vulgare* L. and *Origanum onites* L. J. Food Meas. Charact. 2018, 12, 820–825. [CrossRef]
- 98. Senatore, F.; De Fusco, R.; De Feo, V. Essential oils from *Salvia spp*. (Lamiaceae). I. chemical composition of the essential oils from *Salvia glutinosa* L. growing wild in Southern Italy. *J. Essent. Oil Res.* **1997**, *9*, 151–157. [CrossRef]
- 99. Karimian, P.; Kavoosi, G.H.; Amirghofran, Z. Anti-oxidative and anti-infammatory efects of Tagetes minuta essential oil in activated macrophages. *Asian Pac. J. Trop. Biomed.* 2014, *4*, 219–227. [CrossRef]
- Morteza-Semnani, K.; Saeedi, M.; Changizi, S.; Vosoughi, M. Essential oil composition of *Salvia virgata* Jacq. from Iran. J. Essent. Oil-Bear. Plants 2005, 8, 330–333. [CrossRef]
- 101. Alizadeh, A. Essential Oil Constituents, Antioxidant and Antimicrobial Activities of *Salvia virgata* Jacq. from Iran. *J. Essent. Oil-Bear. Plants* **2013**, *16*, 172–182. [CrossRef]
- 102. Moadeli, S.N.; Rowshan, V.; Abotalebi, A. Comparison of the essential oil components in wild and cultivate population of *Salvia* virgata. Int. Res. J. Appl. Basic Sci. 2013, 4, 337–340.
- 103. Raut, J.S.; Karuppayil, S.M.; Raut, J.S.; Karuppayil, S.M.A. Status review on the medicinal properties of essential oils. *Ind. Crops Prod.* **2014**, *62*, 250–264. [CrossRef]

- Yilar, M.; Kadioglu, İ.; Telci, İ. Determination of Essential Oil Compositions of Some Salvia Species Naturally Growing in Tokat Province. *Turk. J. Agric. Nat. Sci.* 2015, 2, 313–319.
- Saharkhiz, J.M.; Smaeili, S.; Merikhi, M. Essential oil analysis and phytotoxic activity of two ecotypes of Zataria multiflora Boiss. growing in Iran. Nat. Prod. Res. 2010, 24, 1598–1609. [CrossRef]
- Ghasemi, G.; Alirezalu, A.; Ghosta, Y.; Jarrahi, A.; Safavi, S.A.; Mohammadi, M.A.; Barba, F.J.; Munekata, P.E.S.; Domínguez, R.; Lorenzo, J.M. Composition, Antifungal, Phytotoxic, and Insecticidal Activities of *Thymus kotschyanus* Essential Oil. *Molecules* 2020, 25, 1152. [CrossRef]
- 107. Gharibvandia, A.; Karimmojenia, H.; Ehsanzadeha, P.; Rahimmaleka, M.; Mastinu, A. Weed management by allelopathic activity of Foeniculum vulgare essential oil. *Plant Biosyst. Int. J. Deal. Asp. Plant Biol.* **2022**, *156*, 1–9. [CrossRef]
- 108. Tursun, N.; Işıkber, A.A.; Alma, M.H.; Bozhüyük, A.U. Inhibitory Effect of Oregano and Laurel Essential Oils and Their Main Components on Seed Germination of Some Weed and Crop Species. *Selcuk J. Agric. Food Sci. (SJAFS)* **2022**, *36*, 275–281.
- 109. Yasar, A.; Karaman, Y.; Gokbulut, I.; Tursun, A.O.; Tursun, N.; Uremis, I.; Arslan, M. Chemical Composition and Herbicidal Activities of Essential Oil from Aerial Parts of Origanum Hybrids Grown in Different Global Climate Scenarios on Seed Germination of Amaranthus palmeri. J. Essent. Oil Bear. Plants 2021, 24, 603–616. [CrossRef]
- Said, A.; Aoun, T.; Elhaji, N.; Marin, P.D.; Giweli, A. Allelopathic Effects on Seeds Germination of Lactuca sativa L. Seeds and Antibacterial Activity of Thymus capitatus Essential Oil from Zintan-Libya flora. *Am. Sci. Res. J. Eng. Technol. Sci. (ASRJETS)* 2016, 17, 121–131.
- 111. Flores-Macías, A.; Reyes-Zarate, G.G.; da Camara, C.A.G.; López-Ordaz, R.; C.Guillén, J.; Ramos-López, M.Á. Chemical composition and phytotoxic potential of *Eucalyptus globulus* essential oil against *Lactuca sativa* and two herbicide-resistant weeds: Avena fatua and Amaranthus hybridus. *TIP Rev. Espec. En Cienc. Quím.-Biol.* 2021, 24, 1–8. [CrossRef]
- 112. Ruiz-Vásquez, L.; Ruiz Mesia, L.; Caballero Ceferino, H.D.; Ruiz Mesia, W.; Andrés, M.F.; Díaz, C.E.; Gonzalez-Coloma, A. Antifungal and Herbicidal Potential of Piper Essential Oils from the Peruvian Amazonia. *Plants* **2022**, *11*, 1793. [CrossRef]
- Azizan, K.A.; Zamani, A.I.; Nor Muhammad, N.A.; Khairudin, K.; Yusoff, N.; Nawawi, M.F. Dose-Dependent Effect of *Wedelia* trilobata Essential Oil (EO) on Lettuce (*Lactuca sativa* L.) with Multivariate Analysis. *Chem. Biodivers.* 2022, 19, e202100833. [CrossRef]
- Nikolova, M.; Traykova, B.; Yankova-Tsvetkova, E.; Stefanova, T.; Dzhurmanski, A.; Aneva, I.; Berkov, S. Herbicide Potential of Selected Essential Oils From Plants of Lamiaceae and Asteraceae Families. *Acta Agrobot.* 2021, 74, 1–7. [CrossRef]
- 115. Dudai, N.; Poljakoff-Mayber, A.; Mayer, A.; Putievsky, E.; Lerner, H. Essential oils as allelochemicals and their potential use as bioherbicides. *J. Chem. Ecol.* **1999**, *25*, 1079–1089. [CrossRef]
- 116. Macías, F.A.; Marín, D.; Oliveros-Bastidas, A.; Varela, R.M.; Simonet, A.M.; Carrera, C.; Molinillo, J.M. Allelopathy as a new strategy for sustainable ecosystems development. *Biol. Sci. Space* 2003, *17*, 18–23. [CrossRef]
- Synowiec, A.; Lenart–Boron, A.; Kalemba, D. Effect of soil application of microencapsulated caraway oil on weed infestation and maize yield. *Int. J. Pest Manag.* 2018, 64, 315–323. [CrossRef]
- Verdeguer, M.; Castañeda, L.G.; Torres–Pagan, N.; Llorens–Molina, J.A.; Carrubba, A. Control of Erigeron bonariensis with Thymbra capitata, Mentha piperita, Eucalyptus camaldulensis, and Santolina chamaecyparissus Essential Oils. Molecules 2020, 25, 562. [CrossRef]
- Hamrouni, L.; Hanana, M.; Amri, I.; Romane, A.E.; Gargouri, S.; Jamoussi, B. Allelopathic effects of essential oils of Pinus halepensis Miller: Chemical composition and study of their antifungal and herbicidal activities. *Arch. Phytopathol. Pflanzenschutz* 2015, 48, 145–158. [CrossRef]
- 120. Ibanez, M.D.; Blazquez, M.A. Phytotoxicity of essential oils on selected weeds: Potential hazard on food crops. *Plants* **2018**, *7*, 79. [CrossRef]
- Nishida, N.; Tamotsu, S.; Nagata, N.; Saito, C.; Sakai, A. Allelopathic effects of volatile monoterpenoids produced by *Salvia leucophylla* inhibition of cell proliferation and DNA synthesis in the root apical meristem of Brassica campestris seedlings. *J. Chem. Ecol.* 2005, *31*, 1187–1203. [CrossRef]
- Calmasur, O.; Kordali, S.; Kaya, O.; Aslan, I. Toxicity of essential oil vapours obtained from *Achillea* spp. to *Sitophilus granarius* (L.) and *Tribolium confusum* (Jacquelin du Val). *J. Plant Dis. Prot.* 2006, 113, 37–41.
- Semerdjieva, I.; Atanasova, D.; Maneva, V.; Zheljazkov, V.; Radoukova, T.; Astatkie, T.; Dincheva, I. Allelopathic effects of Juniper essential oils on seed germination and seedling growth of some weed seeds. *Ind. Crops Prod.* 2022, 180, 114768. [CrossRef]
- 124. Synowiec, A.; Kalemba, D.; Drozdek, E.; Bocianowski, J. Phytotoxic potential of essential oils from temperate climate plants against the germination of selected weeds and crops. *J. Pest Sci.* 2017, *90*, 407–419. [CrossRef]