



Review

The Activity of Substance P (SP) on the Corneal Epithelium

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Abstract: In 1931, Von Euler and Gaddum isolated substance P (SP), an undecapeptide from the tachykinin family, from equine brain and intestine tissue extracts. Numerous types of cells, including neurons, astrocytes, microglia, epithelial, and endothelial cells, as well as immune cells including T-cells, dendritic cells, and eosinophils, are responsible for its production. The corneal epithelium, immune cells, keratocytes, and neurons all express the two isoforms of NK1R, which has the highest affinity for SP. The most recent research supports SP's contribution to corneal healing by encouraging epithelial cell migration and proliferation. Additionally, when applied to the eyes, SP has proinflammatory effects that result in miosis, intraocular inflammation, and conjunctival hyperemia. In this review article, we examine the role of substance P within the eye. We focus on the role of SP with regards to maintenance and healing of the corneal epithelium.

Keywords: substance P; cornea; corneal epithelium; ophthalmology



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1. Introduction

In 1931, Von Euler and Gaddum isolated substance P (SP), an undecapeptide from the tachykinin family, from equine brain and intestine tissue extracts [1]. Numerous types of cells, including neurons, astrocytes, microglia, epithelial and endothelial cells, as well as immune cells including T-cells, dendritic cells (DCs), and eosinophils, are responsible for its production. Neurokinin 1 receptor (NK1R), Neurokinin 2 receptor (NK2R), and Neurokinin 3 receptor (NK3R) are members of the class I (rhodopsin-like) family of G-protein-coupled neurokinin receptors that SP uses to exert its biological effects [1]. The corneal epithelium, immune cells, keratocytes, and neurons all express the two isoforms of NK1R. The trigeminal ganglion ophthalmic branch fibers are primarily responsible for producing SP on the ocular surface. Healthy people's tears have been found to contain SP and its metabolites. SP levels in tears are much lower in people with diabetic keratopathy and corneal hypoesthesia, which disturbs the homeostasis of the ocular surface. In cases of neurotrophic keratopathy, topical use of SP-derived peptide accelerates healing [1]. The most recent research supports SP's contribution to corneal healing by encouraging epithelial cell migration and proliferation. Additionally, when applied to the eyes, SP has proinflammatory effects that result in miosis, intraocular inflammation, and conjunctival hyperemia [1]. In this review article, we examine the role of substance P within the eye. We focus on the role of SP with regards to maintenance and healing of the corneal epithelium.

2. Discovery of Substance P

Neuronal and non-neuronal cells generate SP, which is an 11-amino-acid-long neuropeptide that regulates tissue homeostasis, wound healing, and ocular inflammation. Substance P binds to the Neurokinin 1 receptor (NK1R), NK2R, and NK3R, which are G-protein-coupled receptors that SP binds to [2–4]. Among the receptors, NK1R has the most affinity for SP of the three. There is an abundance of data showing that SP controls immune cell activity and microbial infection immunity. Substance P was first detected

in extracts from equine brain and intestine, and was found to induce transient hypotension and to facilitate muscle contraction in the intestine when injected intravenously into anesthetized rabbits. The sequence of SP remained elusive until Leeman and coworkers discovered a peptide that stimulated salivary secretion when injected into anesthetized rats while attempting to isolate corticotropin-releasing factor [5]. This peptide was referred to as sialogen until its physical and chemical characteristics were compared to those of SP. It was later found to be identical to SP. Further purification led to the sequence of SP, reported in 1971 by Chang, Leeman, and Niall: Arg-Pro-Lys-Pro-Gln-Gln-Phe-Phe-Gly-Leu-Met-NH₂. Substance P belongs to a family of peptides known as tachykinins. There have been five mammalian tachykinins identified thus far: Substance P, neurokinin A (NKA), neurokinin B (NKB), neurokinin K (NPK), and neuropeptide Y (NPY). Tachykinins have a common carboxy-terminal sequence Phe-X-Gly-Leu-Met-NH₂, where X is an aliphatic or aromatic amino acid [6,7].

3. Molecular Biology of Substance P

The nucleic acid sequences for SP, NKA, NPK, and NPY are encoded on a single gene, while the gene encoding NKB lies in a separate location [8–10]. The gene containing the SP sequence is approximately eight kilobases in length and contains seven exons [11]. The transcribed SP encoding gene undergoes alternative splicing to produce three distinct preprotachykinin (PPT) mRNA's that are designated as α -, P- and γ -PPT mRNA [11]. A fourth PPT mRNA, 6, has been identified in rats [12]. Even though all PPT mRNA's contain the SP sequence, the mRNAs are subject to differential exon usage. Interestingly, differences in the splicing patterns seem to be tissue-specific [13,14]. In the adult rat, γ -PPT mRNA is the most abundant of the three PPT mRNAs. However, in bovines, different PPT mRNAs are more prevalent in different tissues. The exact mechanism for determining differential mRNA splicing is unknown, but the formation of secondary structures in the RNA may be important [15]. These PPT mRNAs, once translated, undergo post-translational processing to form the desired tachykinins.

4. Vasoactive Effects of Substance P

The systemic hypotensive effect of SP was one of its defining characteristics and has been shown to result from peripheral vasodilation. When endothelial cells are subjected to physiological changes, such as hypoxia or shear stress, SP is released from these cells [16–19]. Nitric oxide (NO) has been shown to mediate the SP-induced relaxation of porcine coronary arterioles [20]. Jin et al. demonstrated that SP induces vasoactive intestinal peptide (VIP) release and NO production [20,21]. Other results obtained by Sharma and Davis suggest that in porcine coronary artery endothelial cells, SP induces a rise in intracellular Ca²⁺. The resulting increase in calcium activates an intermediate-conductance Ca²⁺-activated K⁺ channel, which produces hyperpolarization. This ultimately produces a sustained Ca²⁺ entry, which is necessary for endothelium-derived nitric oxide production. SP has also been shown to have profound effects on vasomotion, where a regular, fluctuating flow pattern in skeletal muscles was seen after intravenous injections [22].

In addition, SP causes aggregation of leukocytes and platelets [23]. The effect of SP on vasodilation, vasomotion, leukocytes, and platelets indicates a complex role for this peptide in response to tissue trauma. SP causes a process called neurogenic inflammation, the vasodilation and plasma extravasation that occurs following the release of SP from capsaicin-sensitive nerves [24]. This process was shown to be mediated by the NK1 receptor by direct measurement of protein release from blood vessels [24]. Substance P, released by proinflammatory stimuli, causes plasma protein extravasation by increasing endothelial permeability, indicating a potential role for SP in the inflammatory process. The SP antagonist CP-96,345 was shown to block Evans blue dye leakage, a measure of plasma protein extravasation, induced by SP and releasers of endogenous SP [25]. This result indicates that SP controls endothelial permeability by binding at the NK1 receptor. Recently SP was also found to cause altered vascular permeability in diabetic rats [26].

5. Secretory Effects of Substance P

Substance P may help regulate the release of anterior pituitary hormones. Substance P applied intracerebroventricularly in rats inhibits the release of growth hormone, but increases levels of plasma prolactin [27–29]. Intrathecally applied SP in rats stimulated the release of epinephrine and norepinephrine [30,31]. It has also been suggested that SP interacts with the release of opioid peptides into the circulatory system [32]. In addition, SP has been shown to increase parotid gland secretion through an inositol trisphosphate (IP3)-mediated mechanism, where its effects were measured by the release of labeled protein [33]. Measuring amylase release from dispersed acinar cells has also shown that SP stimulates pancreatic secretion [34]. In the gastrointestinal tract, SP induces secretion of pepsinogen in dogs and inhibition of gastric acid secretion [35,36]. Contradictory information on insulin and glucagon secretion stems from the fact that SP stimulates the release of these two substances in dogs, but has an inhibitory effect on these substances in rats [37,38]. Substance P also induces histamine release by human mast cells and causes eosinophil cationic protein release by human eosinophils [39,40].

6. Mitogenic Effects of Substance P

A new and intriguing development is the discovery that regulatory peptides can also act as mitogens for cells in culture. A direct growth-promoting effect of SP and substance K (NKA) has been reported in smooth muscle cells and human skin fibroblasts [41–44]. Substance P also enhances the proliferation of human blood T-lymphocytes by specific receptors for this peptide [41]. Substance P was also reported to stimulate release of PGE2 and proliferation in rheumatoid synoviocytes, and to stimulate neovascularization [44,45]. Recently, Reid et al. showed that SP at picomolar levels is mitogenic for ocular epithelial cells. These findings are in accord with other evidence which indirectly suggests that tachykinins released by sensory nerves in the skin, joints, and other peripheral tissues may function as mediators of local inflammatory and wound healing responses [46]. It is interesting to note that SP stimulates mitogenesis of embryonic rat aorta cells, but fails to induce significant contraction of these cells. In contrast, SP induced contraction of cultured adult rat vascular smooth muscle cells, but failed to stimulate mitogenesis [47]. Thus, the differentiation state of the cell modulates the mode of action of SP. Substance P may interact with the immune system by partaking in the regulation of lymphocytes. Concentrations in the nanomolar range are capable of stimulating human and mouse lymphocyte proliferation [41]. Substance P has been proposed to be involved in the regulation of glial cell response to injury in the central nervous system [48].

7. Muscle Contraction with Substance P Stimulation

Substance P can induce contractions in smooth muscle cells from many different tissues. Muscle strips from guinea pig renal pelvis, rat and guinea pig bladder, rabbit pulmonary artery, rat portal vein, and most parts of the gastrointestinal tract are just some of the tissues that contract in response to SP doses [49–57]. Capsaicin-sensitive primary afferents induce contraction through a direct action on smooth muscle by SP and an indirect action through cholinergic enteric neurons [55,58,59].

8. Substance P as a Neurotransmitter

For a compound to be considered a neurotransmitter, three criteria must be satisfied. A neurotransmitter must: (1) be in the proper pre-synaptic structures; (2) be released when appropriately stimulated; and (3) induce the postsynaptic effect or effects in the amounts released. In 1953, Lembeck proposed that SP might be a neurotransmitter. At that time, SP was known to be a vasodilator and able to contract smooth muscle. In addition, several groups had shown that this physiological activity was demonstrated to be much higher in dorsal root tissues rather than in ventral root tissues [60–63]. Substance P was later proven to have an excitatory effect on spinal motor neurons by an extract from bovine dorsal root tissue. When this extract was compared with synthetic SP, the

chemical and pharmacological tests proved to be the same material [64,65]. Several lines of research support the hypothesis of SP as a neurotransmitter. Radioimmunoassay and bioassay concluded that SP levels are approximately 20 times higher in the dorsal horn than in the ventral root [65]. Electron microscopy has determined that SP is associated with large granular vesicles located in primary afferent terminals [66,67]. In addition, immunohistochemistry of rat lumbar dorsal root ganglia revealed that primary afferent C-fibers contain the highest amount of SP immunoreactivity [68]. Stimulation of C-afferent fibers induced an increased release of SP from rat spinal cord [69–71]. Application of SP also produces a dose-dependent depolarization of motor neurons in the spinal cord of newborn rats [64,72,73].

9. Ocular Distribution of Substance P

SP is ubiquitous in all ocular tissues, but levels of SP immunoreactivity vary among different species. Elbadri et al. compared SP immunoreactivity levels in cornea, iris/ciliary body, retinal tissues, and choroid/sclera tissues in the cow, sheep, rabbit, and rat [74]. Substance P levels were found to be highest in the retina of cow and rat, whereas in rabbit and sheep, the iris/ciliary body contained the highest degree of SP immunoreactivity. In cat and rabbit retina, most SP-containing cells are located in the ganglion cell layer [75]. Brecha et al. used retrograde labeling from the superior colliculus and optic nerve section to determine that the SP immunoreactive cells in the rabbit retina are ganglion cells [75,76]. Vaney et al., however, determined that the SP immunoreactivity in cats stems from displaced amacrine cells. Immunocytochemistry performed in rats, frogs, lizards, and chicks demonstrated that SP immunoreactivity is prevalent in the amacrine cells of the inner nuclear layer, neurons of the ganglion cell layer, and two distinct layers of processes in the inner plexiform layer [77,78]. These discrepancies in the localization of SP immunoreactivity in the retina may be due to species differences, but the reason for this variability, if it exists, is not known.

In 1984, Osborne presented evidence to implicate SP as a potential neurotransmitter in the bovine retina. He showed that SP binds to retinal membrane preparations with a K_d value of 0.32 nM, suggesting the presence of substance P receptors, and while the retina would not take up exogenous SP, it would release endogenous SP when external K⁺ concentrations are increased. In more recent studies, Zalutsky and Miller showed that SP at concentrations less than 1 nM excited 78% of rabbit ganglion cells and depolarized some amacrine cells [79]. However, tachykinin antagonists and SP desensitization did not alter characteristics of receptive field properties. Studies conducted by Otori et al. determined that SP levels in the suprachiasmatic nucleus (SCN) are unaffected by changes in lighting conditions or ocular enucleation [80]. Otori et al. also concluded that SP levels in the SCN come from an area other than the retina [80].

Immunoreactivity studies have also determined that the iris, cornea, and ciliary body of several species contain SP [81–86]. Substance P immunoreactive (SPI) fibers enter the cornea from two levels: one from the middle layer of the sclera and the other from the episcleral. From the sclera, a thick SPI fiber trunk, extending to the central part, subdivides into smaller SPI fiber bundles and approaches the epithelium. The SP immunoreactive fiber bundles from the episclera are smaller than those from the sclera. However, both fiber bundles form a dense network in the uppermost part of the stroma [87]. After denervation of the trigeminal nerve, a complete disappearance of all SPI axons in the iris occurs [88]. Blood vessels in the anterior uvea are often surrounded by SP fibers [86]. The SP level in the cornea of the adult mouse is reduced 40% by a similar procedure, and neonatal capsaicin treatment results in an 80% reduction of the SP level in the cornea [89]. Surgical denervation of the trigeminal nerve decreases the level of SP in these areas [82,85,89,90]. This information serves as the basis for believing that the SP levels in the anterior segment of the eye arise from the trigeminal nerve. Murphy et al. reported high concentrations (3 nM) of SP in tears of dogs [91]. The degradation, rather than the release, of SP was recently examined by Igc et al. in rabbit and dog aqueous humor [92]. He concluded that

SP is inactivated by a serine protease in the aqueous humor. A possible serine protease for this function is deamidase, which deamidates SP and other tachykinins [93]. The extensive research on the presence of SP in the eye and the multiple physiological effects of this neurotransmitter make ocular tissue a suitable model for the analysis of SP's interaction with its receptor and pathways involved in SP receptor activation.

10. Ocular Smooth Muscle Contraction

The large distribution of SP immunoreactivity in the rabbit and sheep iris/ciliary body gives support to the hypothesis that the physiological role for SP in the eye is to control pupillary diameter [69,94–96]. The iris sphincter muscle is subject to dose-dependent contraction by SP in several species including rabbit, bovine, and pig [97]. Application of tachykinin antagonists reduces the miotic response in these systems [98–100]. Wang and Hakanson, and more recently Kunitomo et al., have shown that the electrically evoked tachykinin-mediated contractile response of the isolated rabbit iris sphincter muscle involves only the SP (NK1) receptor [101]. However, Unger and Tighe reported species differences in the effects of SP on the contractile response of the iris sphincter, and Tachado et al. discovered that while the rabbit, bovine, and pig iris sphincters respond to administered SP by contracting, dog, cat, and human iris sphincter muscles do not [97,102]. However, Anderson et al. contradict this finding by presenting data that demonstrate that human iris sphincter muscle is subject to SP-induced muscle contraction in the eye cup model with an EC₅₀ value of 93 nM [103]. Tachado further exemplified these species differences by measuring cAMP formation and IP₃ accumulation in these species [97]. The bovine, rabbit, and pig, which were subject to SP-induced muscle contraction, demonstrated a significant increase in IP₃ accumulation, but no increase in cAMP formation in the presence of 1 μM SP. Consequently, the dog, cat, and human, which were insensitive to contraction by SP, showed no increase in IP₃ accumulation despite a significant increase in cAMP formation. This inter-species variability may suggest that the functional role of SP may be species-dependent.

Other iris smooth muscle studies with non-peptide SP antagonists found that pretreatment with the NK1 antagonist CP96,345 produced a right shift in the dose-response curve, with an IC₅₀ of approximately 4 mM [104]. While several different laboratories have used different antagonists to implicate the NK1 receptor in the SP-stimulated contraction of rabbit iris smooth muscle (Wang and Hakanson used CP96,345; Kunitomo et al. used spantide and L668169; Hall et al. used GR82334), the pA₂ (or IC₅₀) values were quite high compared with those found for other tissues [101,104,105]. Since the IC₅₀ value reported for CP-96,345 binding to rabbit brain is 0.54 nM, compared to the value for the rabbit iris sphincter of 4 nM, these data also appear consistent with the presence of a different NK1 receptor subtypes for smooth muscle contraction [106].

Varying data have been reported on the effect of calcitonin gene-related peptide (CGRP) in conjunction with SP upon the induction of miosis [107]. In 1991, Anders et al. summarized the synergistic effects of several neuropeptides with SP. Anders et al. concluded that miosis is due to SP, but not CGRP. Contrary to this information, Andersson and Almegard showed that CGRP fragment 8–37 and fragment 32–37 were functional at inducing a dose-dependent iris sphincter muscle contraction in rabbits [108]. Immunohistochemical analysis of mouse trigeminal ganglia, which express SP, indicated that they also express CGRP. Not all neurons that expressed CGRP contained SP, but the SP-containing neurons also contained detectable CGRP. It is unknown whether CGRP assists in facilitating the functional role of SP in the eye or if CGRP elicits a response of its own that is not mediated by SP. However, the fact that CGRP is found in all of the nerves that contain SP and that CGRP is quite synergistic with SP for cell growth stimulation would seem to suggest that, although CGRP may function alone, it probably also functions in association with SP [109].

11. Substance P and Corneal Pain

The cornea is the most sensitive and innervated tissue on the ocular surface. Myelinated and unmyelinated sensory fibers sandwiched between the various layers of the corneal epithelium provide the only innervation to the cornea. Therefore, preserving the integrity of the cornea from potential damage remains important for maintaining the integrity of the eye structure and function. Specifically, corneal mechanical, chemical, or thermal stimulation produces aversive or nociceptive sensations, except for the purely cold sensations elicited by low-temperature stimuli [110–115]. Moreover, direct corneal nerve terminal stimulation triggers defensive reflexes such as tear production, eye blinking, and endocrine and cardiovascular responses [110–115]. The trigeminal nerve (ophthalmic branch, V1), whose ganglion includes the somas of the primary sensory neurons, conducts primary afferent fibers that innervate the cornea. These fibers are of the myelinated A and unmyelinated C types. The trigeminal spinal nucleus (Sp5) within the brainstem acts as the center, whereby the central branches of corneal afferents touch the second order sensory neurons, which are represented by both projection and local circuit neurons. Many ophthalmic and systemic illnesses have a deleterious effect on the structure and function of the corneal nerve. A long-lasting noxious stimulus, injury to the ocular surface, ocular neurosensory abnormalities, or damage to the ocular surface can all cause persistent ocular pain (neuropathic pain). Pain sensitization, which can show as spontaneous pain, hyperalgesia, and allodynia, can be brought on by persistent and aberrant activation of corneal nociceptors [116,117]. Damaged corneal tissue and immune cells release several molecules and inflammatory mediators that interact with the membrane receptors/channels of the nociceptor ending membrane, including ATP, hydrogen ions, SP, neurokinin A, tumor necrosis factor alpha, prostaglandin E2, and interleukins.

Trigeminal fibers release neuropeptides, which have well-known roles in neuroinflammatory processes and the control of nociceptive signal processing. Many effects of SP include neurogenic inflammation and nociception (pain perception) [118–126]. According to current research, diabetic mice have significantly fewer SP nerves than wild-type mice, which accounts for around 59% of the total epithelial innervation in the mouse cornea [118–126]. An ion channel known as transient receptor potential melastatin (TRPM8) is found in sensory neurons and functions as a key cold and osmolarity sensor [118–126]. Several studies have shown that TRPM8 regulates the wetting of the ocular surface and that altered expression of the TRPM8 channel contributes to cold allodynia and neuropathic pain [118–126]. It has recently been shown that abnormal SP expression, low concentration of TRPM8 at nerve terminals, and hypersensitive nerve response occur long after the injury, and changes in gene expression in the trigeminal nerves may contribute to the pathogenesis of corneal surgery-induced dry-eye-like pain [127].

12. Substance P and Corneal Epithelium Healing

One of the first experiments to demonstrate the link between SP and corneal epithelium healing was done by the French physiologist, François Magendie, while examining patients with neurotrophic keratopathy or “neuro-paralytic keratitis” [128]. Neurotrophic keratitis is a rare degenerative corneal disease characterized by lack of or decreased corneal sensation, corneal epithelial breakdown, and impaired healing, resulting in increased susceptibility of the corneal surface to injury and compromised healing. Without treatment, this can lead to stromal melting, corneal ulceration, and corneal perforation. While documenting and treating patients with neurotrophic keratopathy, Magendie hypothesized that the presence of trophic nerve fibers in the trigeminal nerve may be important for regulating tissue metabolism and healing. Subsequent studies later identified the substance involved in the pathogenesis of neurotrophic keratopathy and corneal healing was SP [128].

Over the last few decades, there have been several pre-clinical and clinical studies to show the importance of SP with corneal epithelium healing. Bee et al. showed that collagen type IV intrastromal fibers are orthogonal to the epithelial basement membrane in the cornea [129]. The neurons that innervate the epithelium display SP immunoreactivity

abundantly. On the twelfth day of development, SP immunoreactive nerves are first discovered, coinciding with the beginning of epithelial innervation rather than the extension of nerves through the stroma [129]. Such nerve fibers exhibited substantial connection with both basal and superficial epithelial cells, and increase in number as the body develops. Therefore, SP primary afferents are abundantly supplied to the avian cornea [129]. Furthermore, the expression of SP immunoreactivity correlates directly with the initiation of innervation of the corneal epithelium. Another study by Miller et al. found a dense network of SP immunoreactive axons in the cornea's substantia propria, subepithelial layer, and corneal epithelium [82].

After the superior cervical ganglion was removed, most of the SP immunoreactive axons in the iris developed. Therefore, Miller et al. determined that sensory neurons with SP positivity supply the iris and cornea [82]. A subsequent study by Tervo et al. examining human corneas and irises obtained from corneal surgery or sector iridectomies showed that varicose fibers of the corneal epithelium and nerve trunks in the corneal stroma both exhibited SP immunofluorescence [85]. Substance P immunoreactive nerve fibers in the iris were primarily found near the pupillary edge in both dilator and sphincter areas [85]. Additional radioisotope studies further confirmed that specific binding of SP was found in the iris sphincter muscle, choroid, and retina [130]. In rats, detectable amounts of SP-binding sites were also expressed in the corneal epithelium and iridial stroma [130]. Subsequently, it was discovered that SP has important functions in corneal re-epithelialization after injury (Figure 1) [91,131,132].

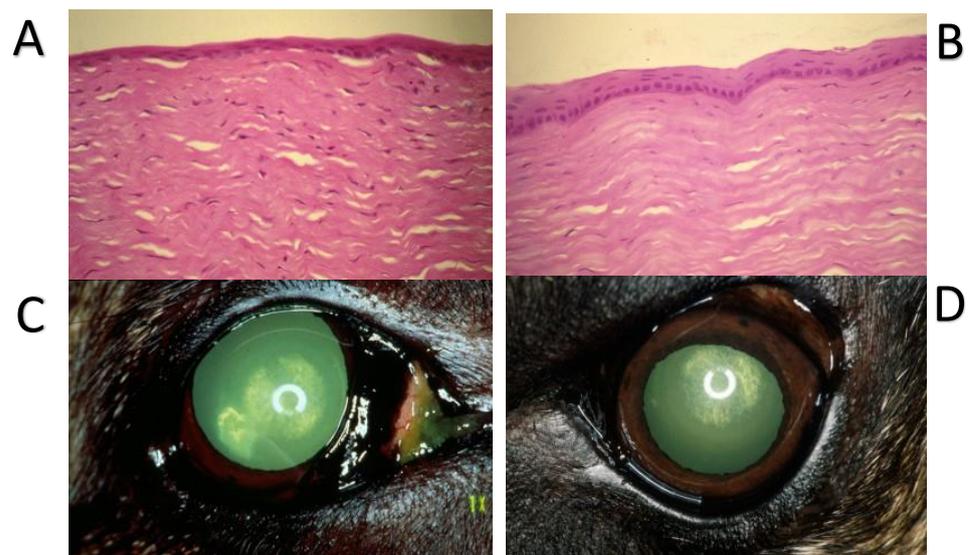


Figure 1. Corneal healing using SP in dogs. (A) Cornea of capsaicin rabbit after healing—not treated with Substance P (note epithelium is not well attached). (B) Cornea of capsaicin rabbit after healing—treated with Substance P (note epithelium is well attached). (C) Dog eye with corneal ulcer in the lower left corner (haze in the background is a cataract). (D) Dog cornea after one week treatment with substance P, which shows resolution of the corneal ulcer.

Further investigation demonstrates that neither SP nor IGF-1 alone affects corneal epithelial wound closure *in vivo*, but that they act synergistically to stimulate corneal re-epithelialization by activating the NK1 receptor [133–135]. Specifically, Nishida et al. showed that the addition of either SP or IGF-1 alone did not affect epithelial migration, while the combination of SP and IGF-1 stimulated epithelial migration in a dose-dependent fashion [136]. Their study also showed that the synergistic effects of SP and IGF-1 on corneal epithelial migration were inhibited using an SP antagonist or enkephalinase [136]. Interestingly, the combination of SP and IGF-1 did not affect the incorporation of ³H-thymidine into corneal epithelial cells [136]. Instead, Nishida et al. showed that SP with

IGF-1 caused an increase in attachment of corneal epithelial cells to fibronectin, collagen type IV, and laminin matrices [136]. A similar improvement in corneal epithelial healing was observed when an NK-1 receptor agonist was used, which suggests that the synergistic effect of SP and IGF-1 might be mediated through the NK-1 receptor system [136]. These results suggest that the maintenance of the normal integrity of the corneal epithelium might be regulated by both humoral and neural factors [136].

Another study by Ofuji et al. suggested that other pathways, namely the tyrosine kinase and protein kinase C, may also be involved in corneal healing with SP activation [137]. Once activated, SP induces corneal healing through downstream effects in the integrins, zonula occludens-1, focal adhesion kinase, paxillin systems, E-cadherins, calmodulin-dependent protein kinase II, and p38 mitogen-activated protein kinase, all of which have important roles in epithelial remodeling and healing [138–142]. This follows closely with a previous study by Kingsley et al., which showed that topical application of SP or its NK1 receptor antagonist has no significant effect on the rate of corneal epithelial wound closure in the rabbit [143]. However, these were normal rabbits and nothing had been done to deplete their normal reserves of SP found in their corneal sensory nerves. The effects of SP on corneal epithelial healing was demonstrated in both mouse model and human studies, which showed improvements in neurotrophic keratopathy, postsurgical superficial punctate keratopathy, and recurrent corneal erosion in diabetic patients with native SP and SP analogs, such as phenylalanineglycine-leucine-methionine amide (FGLM), in conjunction with insulin-like growth factor-1 [144–154]. An example of the effects of Substance P corneal healing is shown in Figure 2 [155]. The patient in Figure 2 was born with no corneal sensory nerves and a severe corneal ulcer in the right eye, which was about to perforate. A skin flap was pulled to cover the right eye. In addition, the patient later developed a corneal ulcer on the left eye that was also close to perforating. Patient was treated with SP plus IGF-1 in an eye drop twice daily (one drop every 15 min for two hours in the morning and at night. The drops contained normal saline). Substance P is added over two hours since it must be present in the wound for that time. The patient showed complete healing of the ulcer in two weeks. Other studies found that adjunct therapies, such as tetrapeptide (SSSR; Ser33-Ser-Ser-Arg) derived from the C domain of IGF-1 and capsaicin, improved both corneal epithelial along with SP and IGF-1 [156–158]. Through the activation of Akt and ROS scavenging through the NK-1 receptor, SP protects corneal epithelial cells from the apoptosis that is brought on by hyperosmotic stress [159].



Figure 2. The effect of Substance P and IGF-1 on corneal ulcer healing in a human patient. (A) Two-year-old patient with skin flap over ulcer in right eye. (B) Same patient with large corneal ulcer in left eye. (C) Left eye after two weeks treatment with SP with no ulcer remaining. (D) Right eye, six months after cornea transplant and treatment with SP.

13. Therapeutic Applications of SP in Cornea Epithelium

Currently, SP research has created several NK-1R/substance P antagonists. Research is underway on substance P/NK1R antagonists as antidepressants, anxiolytics, and anti-inflammatory medications [160]. The first studies on NK-1R/substance P antagonists were originally tested as antidepressants, but subsequent research revealed them to have antiemetic effects. Specifically, NK-1R/substance P antagonists were shown to be effective antiemetic drugs for chemotherapy-induced vomiting by inhibiting substance P from binding to NK1 receptors in the region postrema, which regulates emesis. NK-1 receptor antagonists, such as aprepitant and its prodrug fosaprepitant, are utilized as antiemetic medications. Both oral and intravenous (IV) delivery of aprepitant are accessible; however, only the intravenous version of aprepitant is available. These two medications are helpful in preventing nausea and vomiting brought on by chemotherapy. Hepatic enzymes in the body transform fosaprepitant, a prodrug of aprepitant, into aprepitant, the chemical that is physiologically active [161,162]. Netupitant is also used as an antiemetic in conjunction with Palonosetron as a preventative measure before chemotherapy to lessen nausea and vomiting [163,164]. It has been demonstrated that the chile pepper compound capsaicin reduces the concentration of SP at the terminal and peripheral nerve ends of afferent neurons. Specifically, capsaicin reduces the perception of painful stimuli because of substance P's function in pain transmission [165–167]. Nerve growth factor, a chemical required to produce substance P, is interfered with by capsaicin. Arthritis, post-herpetic neuralgia, shingles, fibromyalgia, and peripheral diabetic neuropathy are treated with capsaicin cream to reduce pain [168–170].

As described in the previous sections, SP has been shown to be an important component for neurogenic inflammation, as it can modulate functions of various immune cells like microglia and dendritic cells [152,159]. Substance P has been shown to be important in aspects of chronic pruritus and was shown to activate Mas Related G-protein-coupled receptor MrgprX2, present on mast cells, besides NK1 receptor [171]. Since SP has been shown to activate NK1 receptors, aprepitant, serlepitant, and other NK1 antagonists have been investigated in clinical trials in treating chronic severe pruritus [172]. In the cornea, SP has been shown to cause avascular hemangiogenesis and lymphangiogenesis in mice models. Topical application of NK1 antagonist, Lanepitant, was shown to reduce the angiogenesis within the inflamed cornea, and this was also shown to occur with topical Fosaprepitant [173,174]. In a recent paper, ocular mast cells were also shown to cause increased inflammatory angiogenesis or neovascularization due to release of VEGF-A in mouse models of corneal inflammation [175]. They were able to show inhibition of mast cells activation resulting in decreased angiogenesis. In Figure 3, we summarize the potential role of SP in different regions of the eye where it is expressed with possible functions.

In mouse models of *Pseudomonas* corneal infection, it was shown that high levels of SP caused an increase in inflammatory cytokines, resulting in more adverse reactions in the cornea. The same group also showed that using NK1 antagonist Spantide 1 improved outcomes in the *Pseudomonas*-infected cornea since the levels of cytokines were reduced due to inhibition of SP-mediated NK1 activation [176]. Since SP mediates angiogenesis via NK1 activation in cancer, NK1 antagonists have been studied in animal models of hepatoblastoma and small cell lung cancer as potential therapeutic options to reduce tumor burden using in vivo and in vitro models due to their anti-angiogenic effect as well as potential for increasing apoptosis [177,178]. There have also been cases where topical application of SP was able to show corneal epithelial wound healing in diabetic mice models as well as model of alkali burn in mouse and rabbit eyes [152,179]. In 1997, there was a study to test topical application of SP on rabbit eyes for wound closure which did not show any statistical significance in wound closure; however, these were normal rabbits and nothing had been done to deplete their normal reserves of SP found in their corneal sensory nerves [143]. In the section above, we discussed the beneficial effect of FGLM-amide peptide with IGF1 in corneal neurotrophic keratopathy. Currently there are no therapies of direct topical SP application for corneal healing that are approved by the FDA. Most substance

P antagonists (e.g., Spantide-1 and -2, Lanepitant, L-733,0660, L-732,138, L-733,060, and L-732,138) have been tested in vitro or in vivo [180]. These substances have demonstrated several effects on reducing inflammation, apoptosis, hemangiogenesis, corneal sensitivity, and cell migration. However, further clinical studies are needed to assess the efficacy and safety in human patients [180].

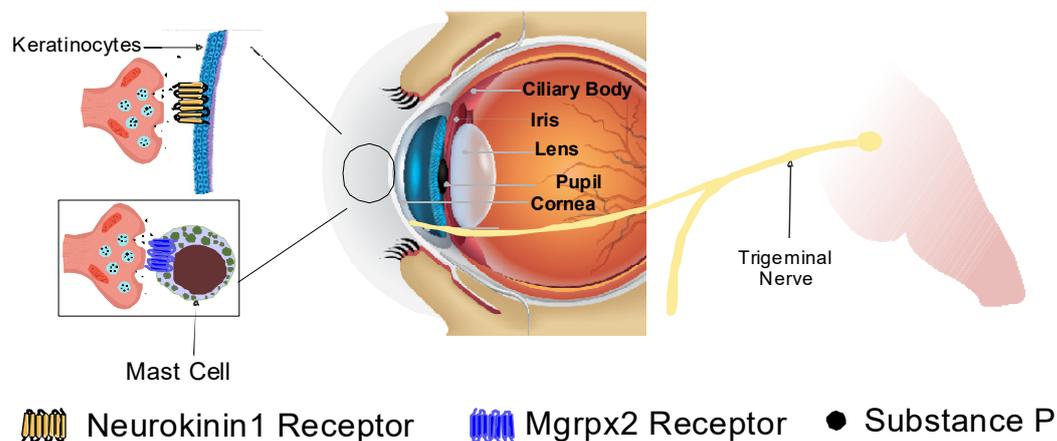


Figure 3. Substance P release from trigeminal nerve: Trigeminal nerve (V1) innervates the corneal epithelium, and it releases SP from its axonal terminals. Keratinocytes have been shown to express Neurokinin 1 (Nk1) receptors that promote inflammation and are important for wound regeneration. Mast cells have been shown to express MgrpX2 receptor that stimulates mast cells to release cytokines and promote inflammation.

14. Conclusions

Substance P is an important neuropeptide that results in its effects via activation of neurokinin receptors as well as MgrpX2 receptors [180]. The current field of study in ophthalmology for SP has been progressing to target SP combined with IGF1 and other molecules to promote wound healing in damaged corneas. On the other hand, efforts are underway to use SP antagonists that prevent binding of SP to its target effectors in controlling inflammation and neovascularization for various pathologies, including keratopathies. As we learn more about the complex nature of SP in various organ systems and the results from clinical trials in different fields like dermatology, they will guide us to start potential trials using SP or its antagonists in helping patients with corneal inflammation and wound healing.

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References

1. Singh, R.B.; Naderi, A.; Cho, W.; Ortiz, G.; Musayeva, A.; Dohlman, T.H.; Chen, Y.; Ferrari, G.; Dana, R. Modulating the tachykinin: Role of substance P and neurokinin receptor expression in ocular surface disorders. *Ocul. Surf.* **2022**, *25*, 142–153. [[CrossRef](#)] [[PubMed](#)]
2. Suvas, S. Role of Substance P Neuropeptide in Inflammation, Wound Healing, and Tissue Homeostasis. *J. Immunol.* **2017**, *199*, 1543–1552. [[CrossRef](#)] [[PubMed](#)]

3. Maggi, C.A.; Patacchini, R.; Rovero, P.; Giachetti, A. Tachykinin receptors and tachykinin receptor antagonists. *J. Auton. Pharmacol.* **1993**, *13*, 23–93. [[CrossRef](#)] [[PubMed](#)]
4. Steinhoff, M.S.; von Mentzer, B.; Geppetti, P.; Pothoulakis, C.; Bunnett, N.W. Tachykinins and their receptors: Contributions to physiological control and the mechanisms of disease. *Physiol. Rev.* **2014**, *94*, 265–301. [[CrossRef](#)]
5. Chang, M.M.; Leeman, S.E. Isolation of a Sialogogic Peptide from Bovine Hypothalamic Tissue and Its Characterization as Substance P. *J. Biol. Chem.* **1970**, *245*, 4784–4790. [[CrossRef](#)]
6. Stone, R.A.; Kuwayama, Y.; Laties, A.M. Regulatory peptides in the eye. *Experientia* **1987**, *43*, 791–800. [[CrossRef](#)]
7. Euler, U.S.V.; Gaddum, J.H. An unidentified depressor substance in certain tissue extracts. *J. Physiol.* **1931**, *72*, 74–87. [[CrossRef](#)]
8. Nawa, H.; Kotani, H.; Nakanishi, S. Tissue-specific generation of two preprotachykinin mRNAs from one gene by alternative RNA splicing. *Nature* **1984**, *312*, 729–734. [[CrossRef](#)]
9. Krause, J.E.; Blount, P.; Sachais, B.S. Molecular Biology of Receptors. In *The Tachykinin Receptors*; Humana Press: Totowa, NJ, USA, 1994; pp. 165–218.
10. Kotani, H.; Hoshimaru, M.; Nawa, H.; Nakanishi, S. Structure and gene organization of bovine neuromedin K precursor. *Proc. Natl. Acad. Sci. USA* **1986**, *83*, 7074–7078. [[CrossRef](#)]
11. Carter, M.S.; Krause, J.E. Structure, expression, and some regulatory mechanisms of the rat preprotachykinin gene encoding substance P, neurokinin A, neuropeptide K, and neuropeptide gamma. *J. Neurosci.* **1990**, *10*, 2203–2214. [[CrossRef](#)]
12. Harmar, A.J.; Hyde, V.; Chapman, K. Identification and cDNA sequence of δ -preprotachykinin, a fourth splicing variant of the rat substance P precursor. *FEBS Lett.* **1990**, *275*, 22–24. [[CrossRef](#)]
13. Krause, J.E.; Chirgwin, J.M.; Carter, M.S.; Xu, Z.S.; Hershey, A.D. Three rat preprotachykinin mRNAs encode the neuropeptides substance P and neurokinin A. *Proc. Natl. Acad. Sci. USA* **1987**, *84*, 881–885. [[CrossRef](#)]
14. Krause, J.E.; Cremins, J.D.; Carter, M.S.; Brown, E.R.; MacDonald, M.R. Solution hybridization-nuclease protection assays for sensitive detection of differentially spliced substance P- and neurokinin A-encoding messenger ribonucleic acids. In *Methods Enzymology*; Elsevier: Amsterdam, The Netherlands, 1989; pp. 634–652.
15. Krause, J.E.; Bu, J.Y.; Takeda, Y.; Blount, P.; Raddatz, R.; Sachais, B.S.; Chou, K.B.; Takeda, J.; McCarron, K.; DiMaggio, D. Structure, expression and second messenger-mediated regulation of the human and rat substance P receptors and their genes. *Regul. Pept.* **1993**, *46*, 59–66. [[CrossRef](#)] [[PubMed](#)]
16. Nowicki, M.; Ostalska-Nowicka, D.; Kondraciuk, B.; Miskowiak, B. The significance of substance P in physiological and malignant haematopoiesis. *J. Clin. Pathol.* **2007**, *60*, 749–755. [[CrossRef](#)] [[PubMed](#)]
17. Bény, J.L. Effect of substance P on the membrane potential of coronary arterial endothelial cells in situ. *Agents Actions* **1990**, *31*, 317–320. [[CrossRef](#)] [[PubMed](#)]
18. Milner, P.; Ralevic, V.; Hopwood, A.M.; Fehér, E.; Lincoln, J.; Kirkpatrick, K.A.; Burnstock, G. Ultrastructural localisation of substance P and choline acetyltransferase in endothelial cells of rat coronary artery and release of substance P and acetylcholine during hypoxia. *Experientia* **1989**, *45*, 121–125. [[CrossRef](#)] [[PubMed](#)]
19. Ralevic, V.; Milner, P.; Hudlická, O.; Kristek, F.; Burnstock, G. Substance P is released from the endothelium of normal and capsaicin-treated rat hind-limb vasculature, in vivo, by increased flow. *Circ. Res.* **1990**, *66*, 1178–1183. [[CrossRef](#)]
20. Kuo, L.; Chilian, W.M.; Davis, M.J. Interaction of pressure- and flow-induced responses in porcine coronary resistance vessels. *Am. J. Physiol.-Heart Circ. Physiol.* **1991**, *261*, H1706–H1715. [[CrossRef](#)] [[PubMed](#)]
21. Jin, J.G.; Misra, S.; Grider, J.R.; Makhoul, G.M. Functional difference between SP and NKA: Relaxation of gastric muscle by SP is mediated by VIP and NO. *Am. J. Physiol.-Gastrointest. Liver Physiol.* **1993**, *264*, G678–G685. [[CrossRef](#)]
22. Sandberg, B.E.B.; Iversen, L.L. Substance P. *J. Med. Chem.* **1982**, *25*, 1009–1015. [[CrossRef](#)] [[PubMed](#)]
23. Öhlén, A.; Thureson-Klein, Å.; Lindbom, L.; Persson, M.G.; Hedqvist, P. Substance P Activates Leukocytes and Platelets in Rabbit Microvessels. *J. Vasc. Res.* **1989**, *26*, 84–94. [[CrossRef](#)]
24. White, D.M.; Helme, R.D. Release of substance P from peripheral nerve terminals following electrical stimulation of the sciatic nerve. *Brain Res.* **1985**, *336*, 27–31. [[CrossRef](#)] [[PubMed](#)]
25. Lembeck, F.; Donnerer, J.; Tsuchiya, M.; Nagahisa, A. The non-peptide tachykinin antagonist, CP-96,345, is a potent inhibitor of neurogenic inflammation. *Br. J. Pharmacol.* **1992**, *105*, 527–530. [[CrossRef](#)] [[PubMed](#)]
26. Mathison, R.; Davison, J.S. Altered vascular permeability responses to substance P in diabetic rats: Interactions with a nitric oxide synthesis inhibitor. *Eur. J. Pharmacol.* **1993**, *240*, 163–168. [[CrossRef](#)] [[PubMed](#)]
27. Arisawa, M.; Snyder, G.D.; De Palatis, L.; Ho, R.H.; Xu, R.K.; Pan, G.; McCann, S.M. Role of substance P in suppressing growth hormone release in the rat. *Proc. Natl. Acad. Sci. USA* **1989**, *86*, 7290–7294. [[CrossRef](#)]
28. Chihara, K.; Arimura, A.; Coy, D.H.; Schally, A.V. Studies on the Interaction of Endorphins, Substance P, and Endogenous Somatostatin in Growth Hormone and Prolactin Release in Rats. *Endocrinology* **1978**, *102*, 281–290. [[CrossRef](#)]
29. Eckstein, N.; Wehrenberg, W.B.; Louis, K.; Carmel, P.W.; Zimmermann, E.A.; Frantz, A.G.; Ferin, M. Effects of Substance P on Anterior Pituitary Secretion in the Female Rhesus Monkey. *Neuroendocrinology* **1980**, *31*, 338–342. [[CrossRef](#)]
30. Hasséssian, H.; Couture, R.; de Champlain, J. Sympathoadrenal Mechanisms Underlying Cardiovascular Responses to Intrathecal Substance P in Conscious Rats. *J. Cardiovasc. Pharmacol.* **1990**, *15*, 736–744. [[CrossRef](#)]
31. Yashpal, K.; Gauthier, S.G.; Henry, J.L. Substance P given intrathecally at the spinal T9 level increases adrenal output of adrenaline and noradrenaline in the rat. *Neuroscience* **1985**, *15*, 529–536. [[CrossRef](#)]

32. Cridland, R.A.; Henry, J.L. An adrenal-mediated, naloxone-reversible increase in reaction time in the tail-flick test following intrathecal administration of substance p at the lower thoracic spinal level in the rat. *Neuroscience* **1988**, *26*, 243–251. [[CrossRef](#)]
33. Dreux, C.; Imhoff, V.; Mauduit, P.; Rossignol, B. Substance P Effect on the Secretory Process in Rat Parotid Gland. In *Substance P and Neurokinins*; Springer: New York, NY, USA, 1987; pp. 195–196.
34. Gardner, J.D.; Jackson, M.J. Regulation of amylase release from dispersed pancreatic acinar cells. *J. Physiol.* **1977**, *270*, 439–454. [[CrossRef](#)] [[PubMed](#)]
35. Vigna, S.R.; Mantyh, C.R.; Soll, A.H.; Maggio, J.E.; Mantyh, P.W. Substance P receptors on canine chief cells: Localization, characterization, and function. *J. Neurosci.* **1989**, *9*, 2878–2886. [[CrossRef](#)] [[PubMed](#)]
36. Yokotani, K.; Fujiwara, M. Effects of substance-P(SP) on parasympathetically stimulated gastric acid secretion and mucosal blood flow(MBF) in rats. *Jpn. J. Pharmacol.* **1984**, *36*, 250. [[CrossRef](#)]
37. Chiba, K.; Kontani, K.; Tadenuma, H.; Katada, T.; Hoshi, M. Induction of starfish oocyte maturation by the beta gamma subunit of starfish G protein and possible existence of the subsequent effector in cytoplasm. *Mol. Biol. Cell* **1993**, *4*, 1027–1034. [[CrossRef](#)] [[PubMed](#)]
38. Kaneto, A.; Kaneko, T.; Kajinuma, H.; Kosaka, K. Effects of Substance P and Neurotensin Infused Intrapancreatically on Glucagon and Insulin Secretion. *Endocrinology* **1978**, *102*, 393–401. [[CrossRef](#)] [[PubMed](#)]
39. Church, M.K.; Lowman, M.A.; Robinson, C.; Holgate, S.T.; Benyon, C. Interaction of Neuropeptides with Human Mast Cells. *Int. Arch. Allergy Immunol.* **1989**, *88*, 70–78. [[CrossRef](#)]
40. Iwamoto, I.; Nakagawa, N.; Yamazaki, H.; Kimura, A.; Tomioka, H.; Yoshida, S. Mechanism for substance P-induced activation of human neutrophils and eosinophils. *Regul. Pept.* **1993**, *46*, 228–230. [[CrossRef](#)] [[PubMed](#)]
41. Payan, D.G.; Brewster, D.R.; Missirlian-Bastian, A.; Goetzl, E.J. Substance P recognition by a subset of human T lymphocytes. *J. Clin. Investig.* **1984**, *74*, 1532–1539. [[CrossRef](#)] [[PubMed](#)]
42. Nilsson, K.O. Improved final height in girls with Turner’s syndrome treated with growth hormone and oxandrolone. *J. Clin. Endocrinol. Metab.* **1996**, *81*, 635–640. [[CrossRef](#)]
43. Nilsson, J.; von Euler, A.M.; Dalsgaard, C.-J. Stimulation of connective tissue cell growth by substance P and substance K. *Nature* **1985**, *315*, 61–63. [[CrossRef](#)]
44. Ziche, M.; Morbidelli, L.; Pacini, M.; Dolara, P.; Maggi, C.A. NK1-receptors mediate the proliferative response of human fibroblasts to tachykinins. *Br. J. Pharm.* **1990**, *100*, 11–14. [[CrossRef](#)] [[PubMed](#)]
45. Lotz, M.; Vaughan, J.H.; Carson, D.A. Effect of Neuropeptides on Production of Inflammatory Cytokines by Human Monocytes. *Science* **1988**, *241*, 1218–1221. [[CrossRef](#)] [[PubMed](#)]
46. Lotz, M.; Carson, D.A.; Vaughan, J.H. Substance P Activation of Rheumatoid Synoviocytes: Neural Pathway in Pathogenesis of Arthritis. *Science* **1987**, *235*, 893–895. [[CrossRef](#)] [[PubMed](#)]
47. Mitsuhashi, M.; Payan, D.G. The mitogenic effects of vasoactive neuropeptides on cultured smooth muscle cell lines. *Life Sci.* **1987**, *40*, 853–861. [[CrossRef](#)]
48. Matthews, M.R.; Connaughton, M.; Cuello, A.C. Ultrastructure and distribution of substance P-immunoreactive sensory collaterals in the guinea pig prevertebral sympathetic ganglia. *J. Comp. Neurol.* **1987**, *258*, 28–51. [[CrossRef](#)]
49. Maggi, C.A.; Giuliani, S.; Santicioli, P.; Abelli, L.; Regoli, D.; Meli, A. Further studies on the mechanisms of the tachykinin-induced activation of micturition reflex in rats: Evidence for the involvement of the capsaicin-sensitive bladder mechanoreceptors. *Eur. J. Pharmacol.* **1987**, *136*, 189–205. [[CrossRef](#)]
50. Maggi, C.A. The role of peptides in the regulation of the micturition reflex: An update. *Gen. Pharmacol. Vasc. Syst.* **1991**, *22*, 1–24. [[CrossRef](#)]
51. Maggi, C.A.; Theodorsson, E.; Santicioli, P.; Giuliani, S. Tachykinins and calcitonin gene-related peptide as co-transmitters in local motor responses produced by sensory nerve activation in the guinea-pig isolated renal pelvis. *Neuroscience* **1992**, *46*, 549–559. [[CrossRef](#)]
52. Shinkai, M.; Takayanagi, I. Characterization of Tachykinin Receptors in Urinary Bladder from Guinea Pig. *Jpn. J. Pharmacol.* **1990**, *54*, 241–243. [[CrossRef](#)]
53. D’Orléans-Juste, P.; Dion, S.; Drapeau, G.; Regoli, D. Different receptors are involved in the endothelium-mediated relaxation and the smooth muscle contraction of the rabbit pulmonary artery in response to substance and related neurokinins. *Eur. J. Pharmacol.* **1986**, *125*, 37–44. [[CrossRef](#)]
54. Mastrangelo, D.; Mathison, R.; Huggel, H.J.; Dion, S.; D’Orléans-Juste, P.; Rhaleb, N.E.; Drapeau, G.; Rovero, P.; Regoli, D. The rat isolated portal vein: A preparation sensitive to neurokinins, particularly neurokinin B. *Eur. J. Pharmacol.* **1987**, *134*, 321–326. [[CrossRef](#)] [[PubMed](#)]
55. Barthó, L.; Holzer, P.; Lembeck, F.; Szolcsányi, J. Evidence that the contractile response of the guinea-pig ileum to capsaicin is due to release of substance P. *J. Physiol.* **1982**, *332*, 157–167. [[CrossRef](#)]
56. Barthó, L.; Holzer, P. Search for a physiological role of substance P in gastrointestinal motility. *Neuroscience* **1985**, *16*, 1–32. [[CrossRef](#)] [[PubMed](#)]
57. Daniel, E.E.; Collins, S.M.; Fox, J.A.E.T.; Huizinga, J.D. Pharmacology of neuroendocrine peptides. In *Comprehensive Physiology*; Wiley: New York, NY, USA, 1989; pp. 759–816.
58. Chahl, L.A. Evidence that the contractile response of the guinea-pig ileum to capsaicin is due to substance P release. *Naunyn-Schmiedeberg’s Arch. Pharmacol.* **1982**, *319*, 212–215. [[CrossRef](#)] [[PubMed](#)]
59. Björkroth, U.; Rosell, S.; Xu, J.-C.; Folkers, K. Pharmacological characterization of four related substance P antagonist. *Acta Physiol. Scand.* **1982**, *116*, 167–173. [[CrossRef](#)]

60. Amin, A.H.; Crawford, T.B.B.; Gaddum, J.H. The distribution of substance P and 5-hydroxytryptamine in the central nervous system of the dog. *J. Physiol.* **1954**, *126*, 596–618. [[CrossRef](#)]
61. Kopera, H.; Lazarini, W. Zur Frage der zentralen Übertragung afferenter Impulse. *Naunyn-Schmiedebergs Arch. Exp. Pathol. Pharmacol.* **1953**, *219*, 197–213. [[CrossRef](#)]
62. Lembeck, F. Zur Frage der zentralen Übertragung afferenter Impulse. *Naunyn-Schmiedebergs Arch. Exp. Pathol. Pharmacol.* **1953**, *219*, 214–222. [[CrossRef](#)]
63. Pernow, B. Substance P. *Pharmacol. Rev.* **1983**, *35*, 85–141.
64. Otsuka, M.; Konishi, S.; Takahashi, T. A Further Study of the Motoneuron-Depolarizing Peptide Extracted from Dorsal Roots of Bovine Spinal Nerves. *Proc. Jpn. Acad.* **1972**, *48*, 747–752. [[CrossRef](#)]
65. Takahashi, T.; Konishi, S.; Powell, D.; Leeman, S.E.; Otsuka, M. Identification of the motoneuron-depolarizing peptide in bovine dorsal root as hypothalamic substance P. *Brain Res.* **1974**, *73*, 59–69. [[CrossRef](#)]
66. De Biasi, S.; Rustioni, A. Glutamate and substance P coexist in primary afferent terminals in the superficial laminae of spinal cord. *Proc. Natl. Acad. Sci. USA* **1988**, *85*, 7820–7824. [[CrossRef](#)] [[PubMed](#)]
67. Ribeiro-da-silva, A.; Tagari, P.; Cuello, A.C. Morphological characterization of substance P-like immunoreactive glomeruli in the superficial dorsal horn of the rat spinal cord and trigeminal subnucleus caudalis: A quantitative study. *J. Comp. Neurol.* **1989**, *281*, 497–515. [[CrossRef](#)] [[PubMed](#)]
68. McCarthy, P.W.; Lawson, S.N. Cell type and conduction velocity of rat primary sensory neurons with substance p-like immunoreactivity. *Neuroscience* **1989**, *28*, 745–753. [[CrossRef](#)]
69. Gamse, R.; Molnar, A.; Lembeck, F. Substance P release from spinal cord slices by capsaicin. *Life Sci.* **1979**, *25*, 629–636. [[CrossRef](#)]
70. Theriault, E.; Otsuka, M.; Jessell, T. Capsaicin-evoked release of substance P from primary sensory neurons. *Brain Res.* **1979**, *170*, 209–213. [[CrossRef](#)] [[PubMed](#)]
71. Yaksh, T.L.; Jessell, T.M.; Gamse, R.; Mudge, A.W.; Leeman, S.E. Intrathecal morphine inhibits substance P release from mammalian spinal cord in vivo. *Nature* **1980**, *286*, 155–157. [[CrossRef](#)] [[PubMed](#)]
72. Konishi, S.; Otsuka, M. Excitatory action of hypothalamic substance P on spinal motoneurons of newborn rats. *Nature* **1974**, *252*, 734–735. [[CrossRef](#)]
73. Otsuka, M.; Konishi, S. Substance P and Excitatory Transmitter of Primary Sensory Neurons. *Cold Spring Harb. Symp. Quant. Biol.* **1976**, *40*, 135–143. [[CrossRef](#)]
74. Elbadri, A.A.; Shaw, C.; Johnston, C.F.; Archer, D.B.; Buchanan, K.D. The distribution of neuropeptides in the ocular tissues of several mammals: A comparative study. *Comp. Biochem. Physiol. C Comp. Pharmacol.* **1991**, *100*, 625–627. [[CrossRef](#)]
75. Brecha, N.; Sharma, S.C.; Karten, H.J. Localization of substance P-like immunoreactivity in the adult and developing goldfish retina. *Neuroscience* **1981**, *6*, 2737–2746. [[CrossRef](#)]
76. Brecha, N.; Johnson, D.; Bolz, J.; Sharma, S.; Parnavelas, J.G.; Lieberman, A.R. Substance P-immunoreactive retinal ganglion cells and their central axon terminals in the rabbit. *Nature* **1987**, *327*, 155–158. [[CrossRef](#)] [[PubMed](#)]
77. Fukuda, M.; Kuwayama, Y.; Shiosaka, S.; Ishimoto, I.; Shimizu, Y.; Takagi, H.; Inagaki, S.; Sakanaka, M.; Semba, E.; Takatsuki, K.; et al. Demonstration of a substance P-like immunoreactivity in retinal cells of the rat. *Neurosci. Lett.* **1981**, *23*, 239–242. [[CrossRef](#)] [[PubMed](#)]
78. Osborne, N.N. Substance P in the bovine retina: Localization, identification, release, uptake and receptor analysis. *J. Physiol.* **1984**, *349*, 83–93. [[CrossRef](#)] [[PubMed](#)]
79. Zalutsky, R.A.; Miller, R.F. The physiology of substance P in the rabbit retina. *J. Neurosci.* **1990**, *10*, 394–402. [[CrossRef](#)] [[PubMed](#)]
80. Otori, Y.; Tominaga, K.; Fukuhara, C.; Yang, J.; Yamazaki, S.; Cagampang, F.R.A.; Okamura, H.; Inouye, S.-I.T. Substance P-like immunoreactivity in the suprachiasmatic nucleus of the rat. *Brain Res.* **1993**, *619*, 271–277. [[CrossRef](#)] [[PubMed](#)]
81. Gibbins, I.L.; Morns, J.L. Co-existence of neuropeptides in sympathetic, cranial autonomic and sensory neurons innervating the iris of the guinea-pig. *J. Auton. Nerv. Syst.* **1987**, *21*, 67–82. [[CrossRef](#)]
82. Miller, A.; Costa, M.; Furness, J.B.; Chubb, I.W. Substance P immunoreactive sensory nerves supply the rat iris and cornea. *Neurosci. Lett.* **1981**, *23*, 243–249. [[CrossRef](#)]
83. Shimizu, Y.; Ishimoto, I.; Shiosaka, S.; Kuwayama, Y.; Fukuda, M.; Inagaki, S.; Sakanaka, M.; Tohyama, M. A direct contact of substance P-containing nerve fibers with pupillary sphincter muscle of the rat: An immunohistochemical analysis. *Neurosci. Lett.* **1982**, *33*, 25–28. [[CrossRef](#)]
84. Stone, R.A.; Laties, A.M.; Brecha, N.C. Substance P-like immunoreactive nerves in the anterior segment of the rabbit, cat and monkey eye. *Neuroscience* **1982**, *7*, 2459–2468. [[CrossRef](#)]
85. Tervo, K.; Tervo, T.; Eränkö, L.; Vannas, A.; Cuello, A.C.; Eränkö, O. Substance P-immunoreactive nerves in the human cornea and iris. *Investig. Ophthalmol. Vis. Sci.* **1982**, *23*, 671–674.
86. Tornqvist, K.; Mandahl, A.; Leander, S.; Lorén, I.; Håkanson, R.; Sundler, F. Substance P-immunoreactive nerve fibres in the anterior segment of the rabbit eye. *Cell Tissue Res.* **1982**, *222*, 467–477. [[CrossRef](#)] [[PubMed](#)]
87. Sasaoka, A.; Ishimoto, I.; Kuwayama, Y.; Sakiyama, T.; Manabe, R.; Shiosaka, S.; Inagaki, S.; Tohyama, M. Overall distribution of substance P nerves in the rat cornea and their three-dimensional profiles. *Investig. Ophthalmol. Vis. Sci.* **1984**, *25*, 351–356.
88. Unger, W.G.; Butler, J.M.; Cole, D.F.; Bloom, S.R.; McGregor, G.P. Substance P, vasoactive intestinal polypeptide (VIP) and somatostatin levels in ocular tissue of normal and sensorily denervated rabbit eyes. *Exp. Eye Res.* **1981**, *32*, 797–801. [[CrossRef](#)]

89. Keen, P.; Tullo, A.B.; Blyth, W.A.; Hill, T.J. Substance P in the mouse cornea: Effects of chemical and surgical denervation. *Neurosci. Lett.* **1982**, *29*, 231–235. [[CrossRef](#)] [[PubMed](#)]
90. Butler, J.M.; Powell, D.; Unger, W.G. Substance P levels in normal and sensorily denervated rabbit eyes. *Exp. Eye Res.* **1980**, *30*, 311–313. [[CrossRef](#)]
91. Murphy, C.J.; Marfurt, C.F.; McDermott, A.; Bentley, E.; Abrams, G.A.; Reid, T.W.; Campbell, S. Spontaneous Chronic Corneal Epithelial Defects (SCCED) in Dogs: Clinical Features, Innervation, and Effect of Topical SP, with or without IGF-1. *Investig. Ophthalmol. Vis. Sci.* **2001**, *42*, 2252–2261.
92. Igić, R. Substance P Inactivation by Aqueous Humor. *Exp. Eye Res.* **1993**, *57*, 415–417. [[CrossRef](#)]
93. Jackman, H.L.; Tan, F.L.; Tamei, H.; Beurling-Harbury, C.; Li, X.Y.; Skidgel, R.A.; Erdős, E.G. A peptidase in human platelets that deamidates tachykinins. Probable identity with the lysosomal “protective protein”. *J. Biol. Chem.* **1990**, *265*, 11265–11272. [[CrossRef](#)]
94. Gamse, R.; Leeman, S.E.; Holzer, P.; Lembeck, F. Differential effects of capsaicin on the content of somatostatin, substance P, and neurotensin in the nervous system of the rat. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1981**, *317*, 140–148. [[CrossRef](#)]
95. Mochizuki-Oda, N.; Nakajima, Y.; Nakanishi, S.; Ito, S. Substance P-induced elevation of intracellular calcium in transfected Chinese hamster ovary cells: Role of inositol trisphosphate. *Regul. Pept.* **1993**, *46*, 450–452. [[CrossRef](#)] [[PubMed](#)]
96. Mochizuki-Oda, N.; Nakajima, Y.; Nakanishi, S.; Ito, S. Characterization of the substance P receptor-mediated calcium influx in cDNA transfected Chinese hamster ovary cells. A possible role of inositol 1,4,5-trisphosphate in calcium influx. *J. Biol. Chem.* **1994**, *269*, 9651–9658. [[CrossRef](#)]
97. Tachado, S.D.; Akhtar, R.A.; Yousofzai, S.Y.K.; Abdel-Latif, A.A. Species differences in the effects of substance P on inositol trisphosphate accumulation and cyclic AMP formation, and on contraction in isolated iris sphincter of the mammalian eye: Differences in receptor density. *Exp. Eye Res.* **1991**, *53*, 729–739. [[CrossRef](#)]
98. Holmdahl, G.; Håkanson, R.; Leander, S.; Rosell, S.; Folkers, K.; Sundler, F. A Substance P Antagonist, [D-Pro2, D-Trp 7,9]SP, Inhibits Inflammatory Responses in the Rabbit Eye. *Science* **1981**, *214*, 1029–1031. [[CrossRef](#)] [[PubMed](#)]
99. Oksala, O.; Stjernschantz, J.; von Dickhoff, K. Characterization of the Mechanism of Acute Ocular Irritation to YAG Laser Capsulotomy in Rabbits: Effects of Substance P Antagonists, Met-Enkephalin, Tetracaine and Tetrodotoxin. *Ophthalmic Res.* **1989**, *21*, 360–368. [[CrossRef](#)]
100. Mandahl, A.; Bill, A. Effects of the substance P antagonist (D-Arg1, D-Pro2, D-Trp7, 9, Leu11)-SP on the miotic response to substance P, antidromic trigeminal nerve stimulation, capsaicin, prostaglandin E1, compound 48/80 and histamine. *Acta Physiol. Scand.* **1984**, *120*, 27–35. [[CrossRef](#)] [[PubMed](#)]
101. Kunitomo, M.; Imaizumi, N.; Sameshima, E.; Fujiwara, M. Pharmacological analysis of receptors involved in tachykinergic contraction induced by electrical transmural stimulation in the rabbit iris sphincter muscle. *Regul. Pept.* **1993**, *46*, 282–284. [[CrossRef](#)] [[PubMed](#)]
102. Unger, W.G.; Tighe, J. The response of the isolated iris sphincter muscle of various mammalian species to substance P. *Exp. Eye Res.* **1984**, *39*, 677–684. [[CrossRef](#)]
103. Anderson, J.A.; Malfroy, B.; Richard, N.R.; Kullerstrand, L.; Lucas, C.; Binder, P.S. Substance P contracts the human iris sphincter: Possible modulation by endogenous enkephalinase. *Regul. Pept.* **1990**, *29*, 49–58. [[CrossRef](#)]
104. Wang, Z.-Y.; Håkanson, R. The rabbit iris sphincter contains NK1 and NK3 but not NK2 receptors: A study with selective agonists and antagonists. *Regul. Pept.* **1993**, *44*, 269–275. [[CrossRef](#)]
105. Hall, J.M.; Mitchell, D.; Morton, I.K.M. Tachykinin receptors mediating responses to sensory nerve stimulation and exogenous tachykinins and analogues in the rabbit isolated iris sphincter. *Br. J. Pharmacol.* **1993**, *109*, 1008–1013. [[CrossRef](#)]
106. Gitter, B.D.; Waters, D.C.; Bruns, R.F.; Mason, N.R.; Nixon, J.A.; Howbert, J.J. Species differences in affinities of non-peptide antagonists for substance p receptors. *Eur. J. Pharmacol.* **1991**, *197*, 237–238. [[CrossRef](#)] [[PubMed](#)]
107. Muramatsu, I.; Nakanishi, S.; Fujiwara, M. Comparison of the Responses to the Sensory Neuropeptides, Substance P, Neurokinin A, Neurokinin B and Calcitonin Gene-Related Peptide and to Trigeminal Nerve Stimulation in the Iris Sphincter Muscle of the Rabbit. *Jpn. J. Pharmacol.* **1987**, *44*, 85–92. [[CrossRef](#)] [[PubMed](#)]
108. Andersson, S.E.; Almegård, B. CGRP(8–37) and CGRP(32–37) contract the iris sphincter in the rabbit eye: Antagonism by spantide and GR82334. *Regul. Pept.* **1993**, *49*, 73–80. [[CrossRef](#)]
109. Boscan, P.; Paton, J.F. Integration of cornea and cardiorespiratory afferents in the nucleus of the solitary tract of the rat. *Am. J. Physiol. Heart Circ. Physiol.* **2002**, *282*, H1278–H1287. [[CrossRef](#)]
110. Bereiter, D.A.; Bereiter, D.F.; Hathaway, C.B. The NMDA receptor antagonist MK-801 reduces Fos-like immunoreactivity in central trigeminal neurons and blocks select endocrine and autonomic responses to corneal stimulation in the rat. *Pain* **1996**, *64*, 179–189. [[CrossRef](#)]
111. Acosta, M.C.; Tan, M.E.; Belmonte, C.; Gallar, J. Sensations evoked by selective mechanical, chemical, and thermal stimulation of the conjunctiva and cornea. *Investig. Ophthalmol. Vis. Sci.* **2001**, *42*, 2063–2067.
112. Belmonte, C.; Acosta, M.C.; Schmelz, M.; Gallar, J. Measurement of corneal sensitivity to mechanical and chemical stimulation with a CO2 esthesiometer. *Investig. Ophthalmol. Vis. Sci.* **1999**, *40*, 513–519.
113. Beuerman, R.W.; Tanelian, D.L. Corneal pain evoked by thermal stimulation. *Pain* **1979**, *7*, 1–14. [[CrossRef](#)]
114. Kenshalo, D.R. Comparison of thermal sensitivity of the forehead, lip, conjunctiva and cornea. *J. Appl. Physiol.* **1960**, *15*, 987–991. [[CrossRef](#)]
115. Guerrero-Moreno, A.; Baudouin, C.; Melik Parsadaniantz, S.; Réaux-Le Goazigo, A. Morphological and Functional Changes of Corneal Nerves and Their Contribution to Peripheral and Central Sensory Abnormalities. *Front. Cell. Neurosci.* **2020**, *14*, 610342. [[CrossRef](#)] [[PubMed](#)]

116. Galor, A.; Moein, H.R.; Lee, C.; Rodriguez, A.; Felix, E.R.; Sarantopoulos, K.D.; Levitt, R.C. Neuropathic pain and dry eye. *Ocul. Surf.* **2018**, *16*, 31–44. [[CrossRef](#)] [[PubMed](#)]
117. De Felipe, C.; Herrero, J.F.; O'Brien, J.A.; Palmer, J.A.; Doyle, C.A.; Smith, A.J.; Laird, J.M.; Belmonte, C.; Cervero, F.; Hunt, S.P. Altered nociception, analgesia and aggression in mice lacking the receptor for substance P. *Nature* **1998**, *392*, 394–397. [[CrossRef](#)] [[PubMed](#)]
118. Motterle, L.; Diebold, Y.; Enriquez de Salamanca, A.; Saez, V.; Garcia-Vazquez, C.; Stern, M.E.; Calonge, M.; Leonardi, A. Altered expression of neurotransmitter receptors and neuromediators in vernal keratoconjunctivitis. *Arch. Ophthalmol.* **2006**, *124*, 462–468. [[CrossRef](#)]
119. He, J.; Bazan, H.E. Neuroanatomy and Neurochemistry of Mouse Cornea. *Investig. Ophthalmol. Vis. Sci.* **2016**, *57*, 664–674. [[CrossRef](#)]
120. He, J.; Pham, T.L.; Kakazu, A.; Bazan, H.E.P. Recovery of Corneal Sensitivity and Increase in Nerve Density and Wound Healing in Diabetic Mice After PEDF Plus DHA Treatment. *Diabetes* **2017**, *66*, 2511–2520. [[CrossRef](#)]
121. Bautista, D.M.; Siemens, J.; Glazer, J.M.; Tsuruda, P.R.; Basbaum, A.I.; Stucky, C.L.; Jordt, S.E.; Julius, D. The menthol receptor TRPM8 is the principal detector of environmental cold. *Nature* **2007**, *448*, 204–208. [[CrossRef](#)]
122. Quallo, T.; Vastani, N.; Horridge, E.; Gentry, C.; Parra, A.; Moss, S.; Viana, F.; Belmonte, C.; Andersson, D.A.; Bevan, S. TRPM8 is a neuronal osmosensor that regulates eye blinking in mice. *Nat. Commun.* **2015**, *6*, 7150. [[CrossRef](#)]
123. Parra, A.; Madrid, R.; Echevarria, D.; del Olmo, S.; Morenilla-Palao, C.; Acosta, M.C.; Gallar, J.; Dhaka, A.; Viana, F.; Belmonte, C. Ocular surface wetness is regulated by TRPM8-dependent cold thermoreceptors of the cornea. *Nat. Med.* **2010**, *16*, 1396–1399. [[CrossRef](#)]
124. Proudfoot, C.J.; Garry, E.M.; Cottrell, D.F.; Rosie, R.; Anderson, H.; Robertson, D.C.; Fleetwood-Walker, S.M.; Mitchell, R. Analgesia mediated by the TRPM8 cold receptor in chronic neuropathic pain. *Curr. Biol.* **2006**, *16*, 1591–1605. [[CrossRef](#)]
125. Liu, B.; Fan, L.; Balakrishna, S.; Sui, A.; Morris, J.B.; Jordt, S.E. TRPM8 is the principal mediator of menthol-induced analgesia of acute and inflammatory pain. *Pain* **2013**, *154*, 2169–2177. [[CrossRef](#)]
126. He, J.; Pham, T.L.; Kakazu, A.H.; Bazan, H.E.P. Remodeling of Substance P Sensory Nerves and Transient Receptor Potential Melastatin 8 (TRPM8) Cold Receptors After Corneal Experimental Surgery. *Investig. Ophthalmol. Vis. Sci.* **2019**, *60*, 2449–2460. [[CrossRef](#)] [[PubMed](#)]
127. Magendie, P. De l'influence de la cinquieme paire de nerfs sur la nutrition et les fonctions de l'oeil. *J. Physiol.* **1824**, *4*, 176–177.
128. Bee, J.A.; Kuhl, U.; Edgar, D.; von der Mark, K. Avian corneal nerves: Co-distribution with collagen type IV and acquisition of substance P immunoreactivity. *Investig. Ophthalmol. Vis. Sci.* **1988**, *29*, 101–107.
129. Denis, P.; Fardin, V.; Nordmann, J.P.; Elena, P.P.; Laroche, L.; Saraux, H.; Rostene, W. Localization and characterization of substance P binding sites in rat and rabbit eyes. *Investig. Ophthalmol. Vis. Sci.* **1991**, *32*, 1894–1902.
130. Reid, T.W.; Murphy, C.J.; Iwahashi, C.K.; Foster, B.A.; Mannis, M.J. Stimulation of epithelial cell growth by the neuropeptide substance P. *J. Cell. Biochem.* **1993**, *52*, 476–485. [[CrossRef](#)]
131. Bentley, E.; Abrams, G.A.; Covitz, D.; Cook, C.S.; Fischer, C.A.; Hacker, D.; Stuhr, C.M.; Reid, T.W.; Murphy, C.J. Morphology and immunohistochemistry of spontaneous chronic corneal epithelial defects (SCCED) in dogs. *Investig. Ophthalmol. Vis. Sci.* **2001**, *42*, 2262–2269.
132. Nakamura, M.; Ofuji, K.; Chikama, T.; Nishida, T. Combined effects of substance P and insulin-like growth factor-1 on corneal epithelial wound closure of rabbit in vivo. *Curr. Eye Res.* **1997**, *16*, 275–278. [[CrossRef](#)]
133. Nakamura, M.; Ofuji, K.; Chikama, T.; Nishida, T. The NK1 receptor and its participation in the synergistic enhancement of corneal epithelial migration by substance P and insulin-like growth factor-1. *Br. J. Pharmacol.* **1997**, *120*, 547–552. [[CrossRef](#)]
134. Nakamura, M.; Chikama, T.; Nishida, T. Synergistic effect with Phe-Gly-Leu-Met-NH₂ of the C-terminal of substance P and insulin-like growth factor-1 on epithelial wound healing of rabbit cornea. *Br. J. Pharmacol.* **1999**, *127*, 489–497. [[CrossRef](#)]
135. Nishida, T.; Nakamura, M.; Ofuji, K.; Reid, T.W.; Mannis, M.J.; Murphy, C.J. Synergistic effects of substance P with insulin-like growth factor-1 on epithelial migration of the cornea. *J. Cell. Physiol.* **1996**, *169*, 159–166. [[CrossRef](#)]
136. Ofuji, K.; Nakamura, M.; Nishida, T. Signaling regulation for synergistic effects of substance P and insulin-like growth factor-1 or epidermal growth factor on corneal epithelial migration. *Jpn. J. Ophthalmol.* **2000**, *44*, 1–8. [[CrossRef](#)]
137. Nakamura, M.; Nagano, T.; Chikama, T.; Nishida, T. Up-regulation of phosphorylation of focal adhesion kinase and paxillin by combination of substance P and IGF-1 in SV-40 transformed human corneal epithelial cells. *Biochem. Biophys. Res. Commun.* **1998**, *242*, 16–20. [[CrossRef](#)] [[PubMed](#)]
138. Nakamura, M.; Chikama, T.; Nishida, T. Participation of p38 MAP kinase, but not p44/42 MAP kinase, in stimulation of corneal epithelial migration by substance P and IGF-1. *Curr. Eye Res.* **2005**, *30*, 825–834. [[CrossRef](#)] [[PubMed](#)]
139. Araki-Sasaki, K.; Aizawa, S.; Hiramoto, M.; Nakamura, M.; Iwase, O.; Nakata, K.; Sasaki, Y.; Mano, T.; Handa, H.; Tano, Y. Substance P-induced cadherin expression and its signal transduction in a cloned human corneal epithelial cell line. *J. Cell. Physiol.* **2000**, *182*, 189–195. [[CrossRef](#)]
140. Ko, J.A.; Yanai, R.; Nishida, T. Up-regulation of ZO-1 expression and barrier function in cultured human corneal epithelial cells by substance P. *FEBS Lett.* **2009**, *583*, 2148–2153. [[CrossRef](#)]
141. Yamada, N.; Yanai, R.; Inui, M.; Nishida, T. Sensitizing effect of substance P on corneal epithelial migration induced by IGF-1, fibronectin, or interleukin-6. *Investig. Ophthalmol. Vis. Sci.* **2005**, *46*, 833–839. [[CrossRef](#)]
142. Kingsley, R.E.; Marfurt, C.F. Topical substance P and corneal epithelial wound closure in the rabbit. *Investig. Ophthalmol. Vis. Sci.* **1997**, *38*, 388–395.
143. Chikama, T.; Fukuda, K.; Morishige, N.; Nishida, T. Treatment of neurotrophic keratopathy with substance-P-derived peptide (FGLM) and insulin-like growth factor I. *Lancet* **1998**, *351*, 1783–1784. [[CrossRef](#)]

144. Nagano, T.; Nakamura, M.; Nakata, K.; Yamaguchi, T.; Takase, K.; Okahara, A.; Ikuse, T.; Nishida, T. Effects of substance P and IGF-1 in corneal epithelial barrier function and wound healing in a rat model of neurotrophic keratopathy. *Investig. Ophthalmol. Vis. Sci.* **2003**, *44*, 3810–3815. [[CrossRef](#)]
145. Chikamoto, N.; Chikama, T.; Yamada, N.; Nishida, T.; Ishimitsu, T.; Kamiya, A. Efficacy of substance P and insulin-like growth factor-1 peptides for preventing postsurgical superficial punctate keratopathy in diabetic patients. *Jpn. J. Ophthalmol.* **2009**, *53*, 464–469. [[CrossRef](#)] [[PubMed](#)]
146. Yamada, N.; Matsuda, R.; Morishige, N.; Yanai, R.; Chikama, T.I.; Nishida, T.; Ishimitsu, T.; Kamiya, A. Open clinical study of eye-drops containing tetrapeptides derived from substance P and insulin-like growth factor-1 for treatment of persistent corneal epithelial defects associated with neurotrophic keratopathy. *Br. J. Ophthalmol.* **2008**, *92*, 896–900. [[CrossRef](#)] [[PubMed](#)]
147. Nishida, T.; Chikama, T.; Morishige, N.; Yanai, R.; Yamada, N.; Saito, J. Persistent epithelial defects due to neurotrophic keratopathy treated with a substance p-derived peptide and insulin-like growth factor 1. *Jpn. J. Ophthalmol.* **2007**, *51*, 442–447. [[CrossRef](#)] [[PubMed](#)]
148. Nakamura, M.; Kawahara, M.; Morishige, N.; Chikama, T.; Nakata, K.; Nishida, T. Promotion of corneal epithelial wound healing in diabetic rats by the combination of a substance P-derived peptide (FGLM-NH2) and insulin-like growth factor-1. *Diabetologia* **2003**, *46*, 839–842. [[CrossRef](#)]
149. Lee, C.H.; Whiteman, A.L.; Murphy, C.J.; Barney, N.P.; Taylor, P.B.; Reid, T.W. Substance P, Insulinlike Growth Factor 1, and Surface Healing. *Arch. Ophthalmol.* **2002**, *120*, 215–217.
150. Benitez-Del-Castillo, J.M.; Rodriguez-Bayo, S.; Fontan-Rivas, E.; Martinez-de-la-Casa, J.M.; Garcia-Sanchez, J. Treatment of recurrent corneal erosion with substance P-derived peptide and insulin-like growth factor I. *Arch. Ophthalmol.* **2005**, *123*, 1445–1447. [[CrossRef](#)]
151. Yang, L.; Di, G.; Qi, X.; Qu, M.; Wang, Y.; Duan, H.; Danielson, P.; Xie, L.; Zhou, Q. Substance P promotes diabetic corneal epithelial wound healing through molecular mechanisms mediated via the neurokinin-1 receptor. *Diabetes* **2014**, *63*, 4262–4274. [[CrossRef](#)]
152. Yanai, R.; Nishida, T.; Hatano, M.; Uchi, S.H.; Yamada, N.; Kimura, K. Role of the Neurokinin-1 Receptor in the Promotion of Corneal Epithelial Wound Healing by the Peptides FGLM-NH2 and SSSR in Neurotrophic Keratopathy. *Investig. Ophthalmol. Vis. Sci.* **2020**, *61*, 29. [[CrossRef](#)]
153. Ko, J.A.; Murata, S.; Nishida, T. Up-regulation of the tight-junction protein ZO-1 by substance P and IGF-1 in A431 cells. *Cell Biochem. Funct.* **2009**, *27*, 388–394. [[CrossRef](#)]
154. Brown, S.M.; Lamberts, D.W.; Reid, T.W.; Nishida, T.; Murphy, C.J. Neurotrophic and Anhidrotic Keratopathy Treated With Substance P and Insulinlike Growth Factor 1. *Arch. Ophthalmol.* **1997**, *115*, 926–927. [[CrossRef](#)]
155. Nakamura, M.; Kawahara, M.; Nakata, K.; Nishida, T. Restoration of corneal epithelial barrier function and wound healing by substance P and IGF-1 in rats with capsaicin-induced neurotrophic keratopathy. *Investig. Ophthalmol. Vis. Sci.* **2003**, *44*, 2937–2940. [[CrossRef](#)]
156. Yamada, N.; Yanai, R.; Kawamoto, K.; Nagano, T.; Nakamura, M.; Inui, M.; Nishida, T. Promotion of corneal epithelial wound healing by a tetrapeptide (SSSR) derived from IGF-1. *Investig. Ophthalmol. Vis. Sci.* **2006**, *47*, 3286–3292. [[CrossRef](#)]
157. Yamada, N.; Yanai, R.; Nakamura, M.; Inui, M.; Nishida, T. Role of the C domain of IGFs in synergistic promotion, with a substance P-derived peptide, of rabbit corneal epithelial wound healing. *Investig. Ophthalmol. Vis. Sci.* **2004**, *45*, 1125–1131. [[CrossRef](#)] [[PubMed](#)]
158. Yang, L.; Sui, W.; Li, Y.; Qi, X.; Wang, Y.; Zhou, Q.; Gao, H. Substance P Inhibits Hyperosmotic Stress-Induced Apoptosis in Corneal Epithelial Cells through the Mechanism of Akt Activation and Reactive Oxygen Species Scavenging via the Neurokinin-1 Receptor. *PLoS ONE* **2016**, *11*, e0149865. [[CrossRef](#)] [[PubMed](#)]
159. Wang, S.Y.; Yang, Z.J.; Zhang, Z.; Zhang, H. Aprepitant in the prevention of vomiting induced by moderately and highly emetogenic chemotherapy. *Asian Pac. J. Cancer Prev.* **2014**, *15*, 10045–10051. [[CrossRef](#)] [[PubMed](#)]
160. Candelario, N.; Lu, M.L. Fosaprepitant dimeglumine for the management of chemotherapy-induced nausea and vomiting: Patient selection and perspectives. *Cancer Manag. Res.* **2016**, *8*, 77–82. [[CrossRef](#)]
161. Jordan, K. Neurokinin-1-receptor antagonists: A new approach in antiemetic therapy. *Onkologie* **2006**, *29*, 39–43. [[CrossRef](#)]
162. Keating, G.M. Netupitant/Palonosetron: A Review in the Prevention of Chemotherapy-Induced Nausea and Vomiting. *Drugs* **2015**, *75*, 2131–2141. [[CrossRef](#)]
163. He, A.; Alhariri, J.M.; Sweren, R.J.; Kwatra, M.M.; Kwatra, S.G. Aprepitant for the Treatment of Chronic Refractory Pruritus. *BioMed Res. Int.* **2017**, *2017*, 4790810. [[CrossRef](#)]
164. Anand, P.; Bley, K. Topical capsaicin for pain management: Therapeutic potential and mechanisms of action of the new high-concentration capsaicin 8% patch. *Br. J. Anaesth.* **2011**, *107*, 490–502. [[CrossRef](#)]
165. Sharma, S.K.; Vij, A.S.; Sharma, M. Mechanisms and clinical uses of capsaicin. *Eur. J. Pharmacol.* **2013**, *720*, 55–62. [[CrossRef](#)]
166. Roosterman, D.; Goerge, T.; Schneider, S.W.; Bunnett, N.W.; Steinhoff, M. Neuronal control of skin function: The skin as a neuroimmunoendocrine organ. *Physiol. Rev.* **2006**, *86*, 1309–1379. [[CrossRef](#)] [[PubMed](#)]
167. Burks, T.F.; Buck, S.H.; Miller, M.S. Mechanisms of depletion of substance P by capsaicin. *Fed. Proc.* **1985**, *44*, 2531–2534. [[PubMed](#)]
168. Fernandes, E.S.; Cerqueira, A.R.; Soares, A.G.; Costa, S.K. Capsaicin and Its Role in Chronic Diseases. *Adv. Exp. Med. Biol.* **2016**, *929*, 91–125. [[CrossRef](#)]
169. Derry, S.; Rice, A.S.; Cole, P.; Tan, T.; Moore, R.A. Topical capsaicin (high concentration) for chronic neuropathic pain in adults. *Cochrane Database Syst. Rev.* **2017**, *1*, Cd007393. [[CrossRef](#)]
170. Chompunud Na Ayudhya, C.; Roy, S.; Thapaliya, M.; Ali, H. Roles of a Mast Cell-Specific Receptor MRGPRX2 in Host Defense and Inflammation. *J. Dent. Res.* **2020**, *99*, 882–890. [[CrossRef](#)] [[PubMed](#)]

171. Ständer, S.; Spellman, M.C.; Kwon, P.; Yosipovitch, G. The NK1 receptor antagonist serlopitant for treatment of chronic pruritus. *Expert Opin. Investig. Drugs* **2019**, *28*, 659–666. [[CrossRef](#)]
172. Bignami, F.; Giacomini, C.; Lorusso, A.; Aramini, A.; Rama, P.; Ferrari, G. NK1 receptor antagonists as a new treatment for corneal neovascularization. *Investig. Ophthalmol. Vis. Sci.* **2014**, *55*, 6783–6794. [[CrossRef](#)]
173. Bignami, F.; Lorusso, A.; Rama, P.; Ferrari, G. Growth inhibition of formed corneal neovascularization following Fosaprepitant treatment. *Acta Ophthalmol.* **2017**, *95*, e641–e648. [[CrossRef](#)]
174. Cho, H.J.; Jeon, Y.J.; Yoon, W.; Yoon, J.; Kim, J.; Kim, J.W. Neovascular age-related macular degeneration without exudative recurrence over 24 months after initial remission. *Sci. Rep.* **2022**, *12*, 15662. [[CrossRef](#)]
175. Foldenauer, M.E.; McClellan, S.A.; Barrett, R.P.; Zhang, Y.; Hazlett, L.D. Substance P affects growth factors in *Pseudomonas aeruginosa*-infected mouse cornea. *Cornea* **2012**, *31*, 1176–1188. [[CrossRef](#)] [[PubMed](#)]
176. Muñoz, M.; Coveñas, R. Involvement of substance P and the NK-1 receptor in pancreatic cancer. *World J. Gastroenterol.* **2014**, *20*, 2321–2334. [[CrossRef](#)]
177. Muñoz, M.; Coveñas, R. Involvement of substance P and the NK-1 receptor in cancer progression. *Peptides* **2013**, *48*, 1–9. [[CrossRef](#)] [[PubMed](#)]
178. Hong, H.S.; Lee, J.; Lee, E.; Kwon, Y.S.; Lee, E.; Ahn, W.; Jiang, M.H.; Kim, J.C.; Son, Y. A new role of substance P as an injury-inducible messenger for mobilization of CD29+ stromal-like cells. *Nat. Med.* **2009**, *15*, 425–435. [[CrossRef](#)]
179. Bignami, F.; Rama, P.; Ferrari, G. Substance P and its Inhibition in Ocular Inflammation. *Curr. Drug Targets* **2016**, *17*, 1265–1274. [[CrossRef](#)]
180. Green, D.P.; Limjunyawong, N.; Gour, N.; Pundir, P.; Dong, X. A Mast-Cell-Specific Receptor Mediates Neurogenic Inflammation and Pain. *Neuron* **2019**, *101*, 412–420.e3. [[CrossRef](#)] [[PubMed](#)]

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