

## SUPPLEMENTARY MATERIAL

**Table S1. Results of real-time RT-qPCR detection of SARS-CoV-2 in the tested samples**

Sample ID	Recovery (%)	g.c. / $\mu$ L of RNA	g.c. / L of sewage	PCR ID 980
4158	1.71%	6.09	$1.52 \times 10^4$	+
4159	1.28%	6.04	$1.51 \times 10^4$	+
4160	5.31%	2.40	$6.00 \times 10^3$	-
4161	2.52%	not detected	-	-
4162	1.61%	5.09	$1.27 \times 10^4$	+
4163	2.11%	36.26	$9.06 \times 10^4$	+
4170	2.12%	3.50	$8.75 \times 10^3$	-
4171	2.35%	2.11	$5.27 \times 10^3$	-
4172	3.06%	19.39	$4.85 \times 10^4$	+
4173	1.91%	27.77	$6.94 \times 10^4$	+
4174	6.16%	1.07	$2.68 \times 10^3$	-
4175	4.52%	0.78	$1.94 \times 10^3$	-

**Table S2. Number of raw and final reads selected after filtering steps**

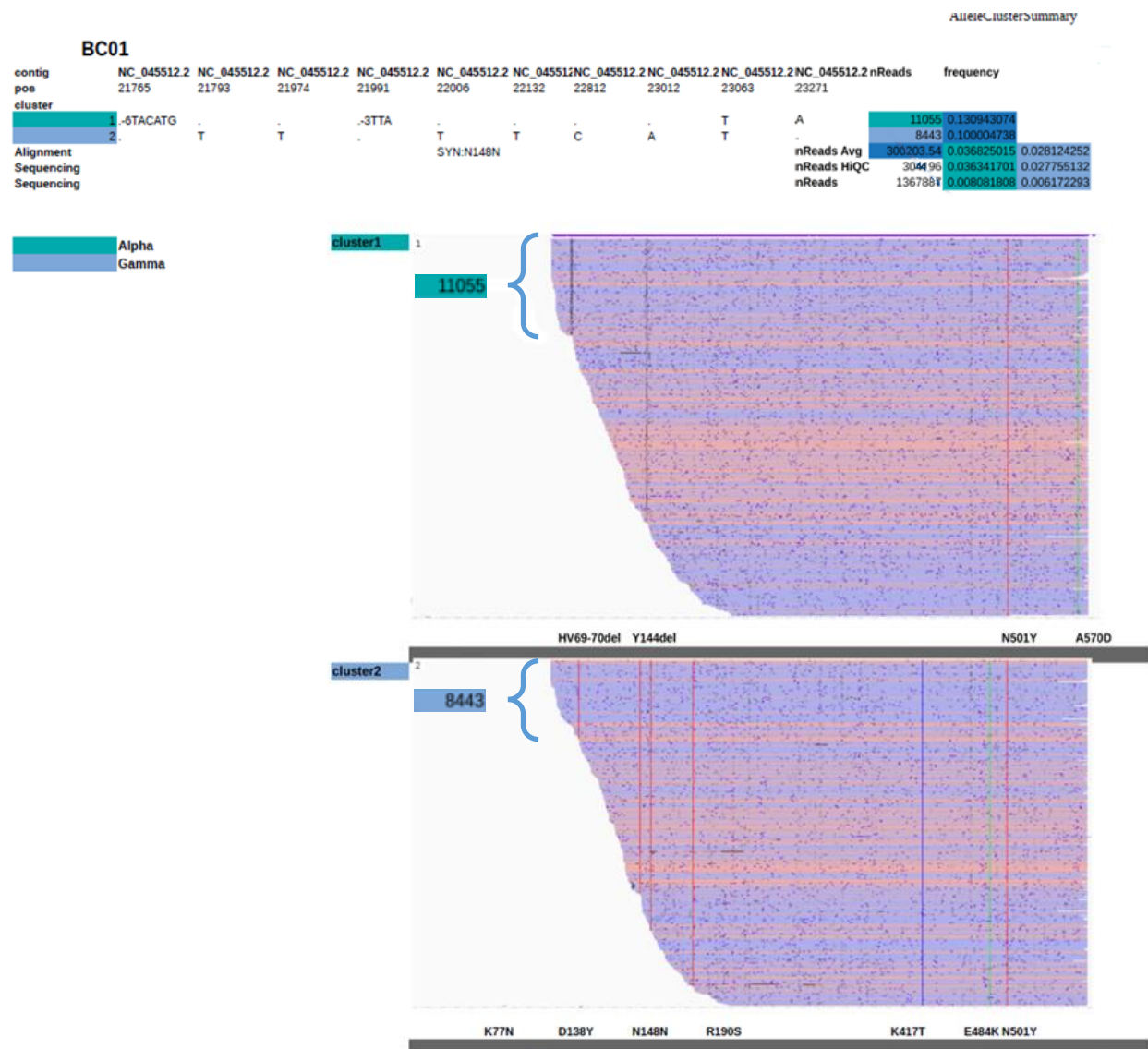
Sample ID	Raw Data	dual-barcode filtering	HiFi (q 8. trim end 20) + 1400-1700 nt	Aligned to SARS-CoV-2 reference sequence NC_045512
BC01	1367887	370776	312766	312697
BC02	1925893	668332	328851	328701
BC03	764517	188387	331399	330950

**Table S3. Concatenated mutations queried using the BAMQL tool v1.6 for the detection of each VOC/VOI**

<b>Variant</b>	<b>Mutations</b> (aa substitutions or deletions)	<b>Queries</b>
<b>Alpha</b>	69-70del 144del N501Y A570D	21764.A and 21771.T and exclude(21765.T. 21769.T) 21990.T and 21994.T and exclude(21992.T) 23063.T 23271.A
<b>Beta</b>	D80A D215G K417N E484K N501Y	21801.C 22206.G 22813.T 23012.A 23063.T
<b>Gamma</b>	D138Y R190S K417T E484K N501Y	21794.T 22132.T 22812.C 23012.A 23063.T
<b>Delta</b>	156-157-158del L452R T478K	22028.G and 22035.G and exclude(22029.A. 22034.A) 22917.G 22995.A
<b>Eta</b>	A67V 69-70del 144del E484K	21762.T 21764.A and 21771.T and exclude(21765.T. 21769.T) 21990.T and 21994.T and exclude(21992.T) 23012.A

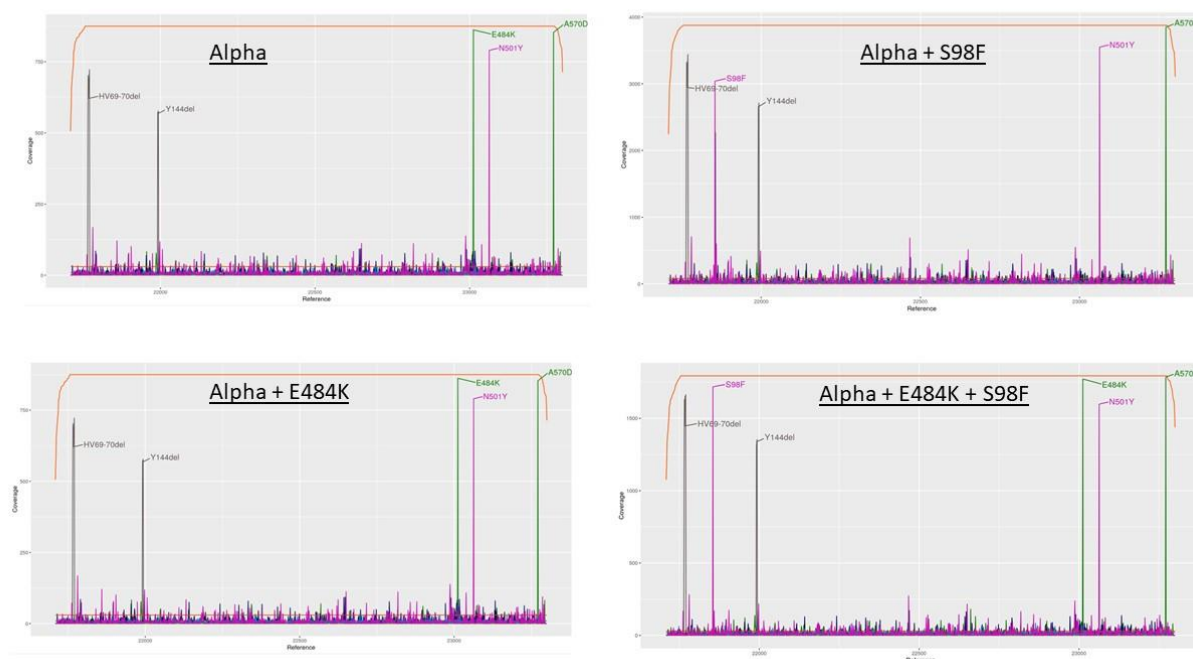
Numbers represent nucleotide positions referred to SARS-CoV-2 reference genome NC\_045512. Reads were queried for concatenated mutations by: a) selecting defined nucleotide mutations; b) for deletions, by searching for the absence of a wild type nucleotide within the deleted region, in combination with the presence of the wild type nucleotide upstream (5') and downstream (3') of the deletion. This latter search criterion was included to minimize selection of random deletions generated by nanopore sequencing.

Figure S1. AlleleClusterSummary for pool BC01 (Clustering unsupervised results)

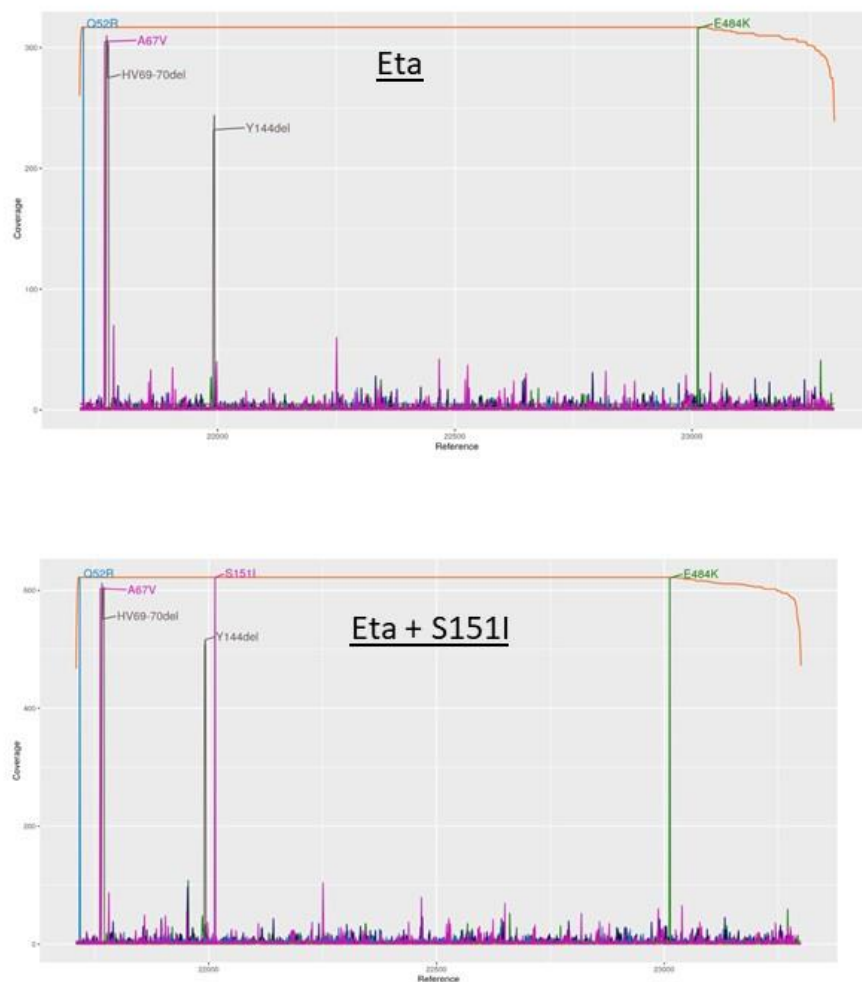


**Mutation panel of SARS-CoV-2 Alpha, Gamma and Eta variants in wastewaters in Italy detected by nanopore technology long-read amplicon sequencing** La Rosa G., Brandtner D., Mancini P., Veneri C., Bonanno Ferraro G., Bonadonna L., Lucentini L., Suffredini E.  
*Water*

**Figure S2. Mutations detected in the alpha cluster**

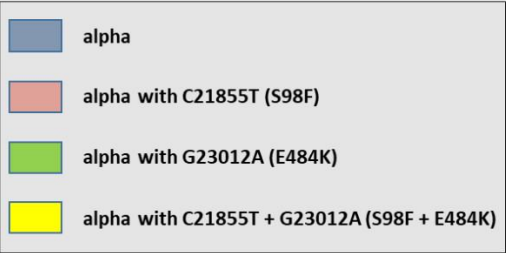


**Figure S3. Mutations detected in the eta cluster**

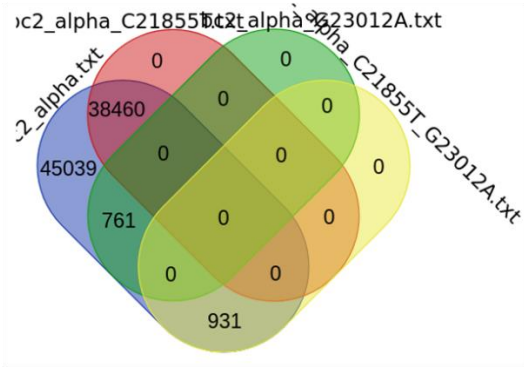


**Figure S4. Venn diagrams for Alpha and Eta reads in pool BC02**

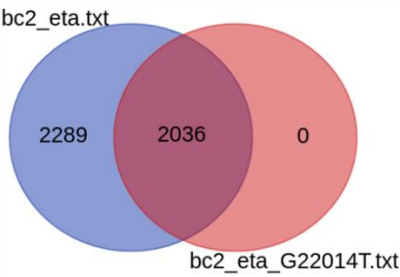
**BARCODE02**



**ALPHA BAMQL**



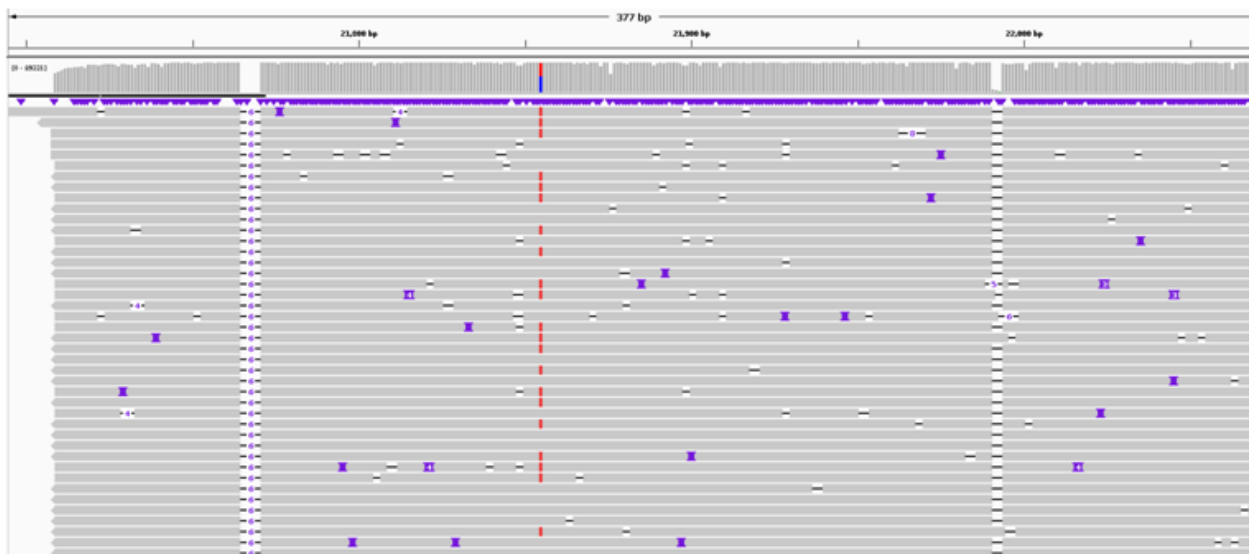
**ETA BAMQL**



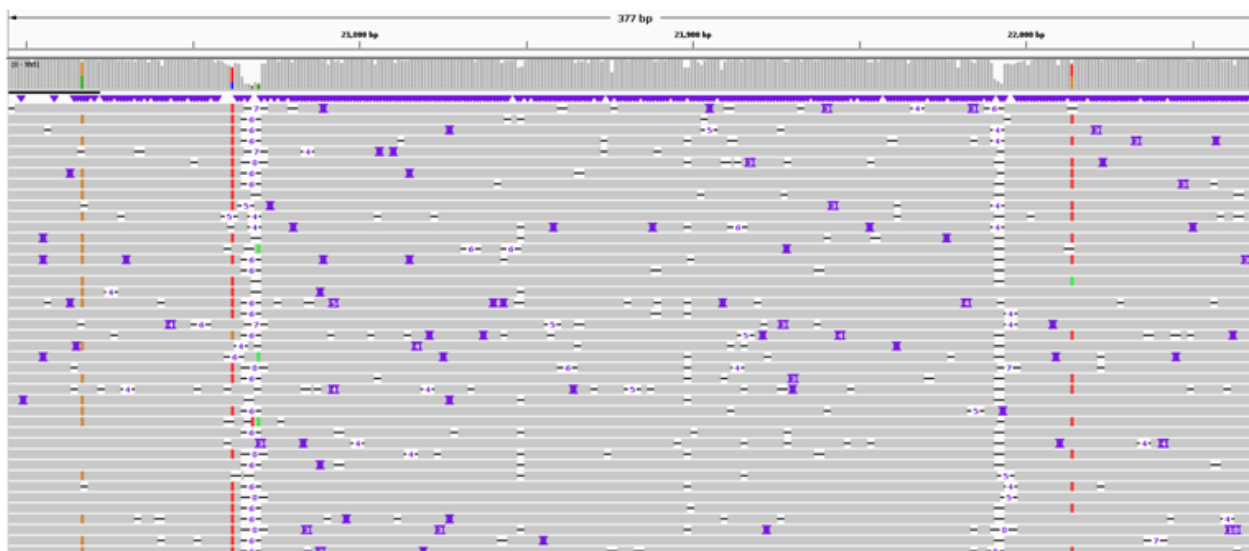
**Figure S5. Distribution of small deletions in assigned reads (A) and in unassigned reads (B): small deletions are significantly more frequent in unassigned reads**

(Snapshots from IGV alignment viewer)

**A**



**B**



**Figure S6. Insertion/Deletion analysis in assigned reads (A) and unassigned reads (B): in unassigned reads Ins/Del ratio is exceedingly low in the range of 10 bases indicating an abundance of small deletions in comparison to assigned reads**

