

Supplementary Data: A Validation Study on Immunophenotypic Differences in T-lymphocyte Chromosomal Radiosensitivity between Newborns and Adults in South Africa

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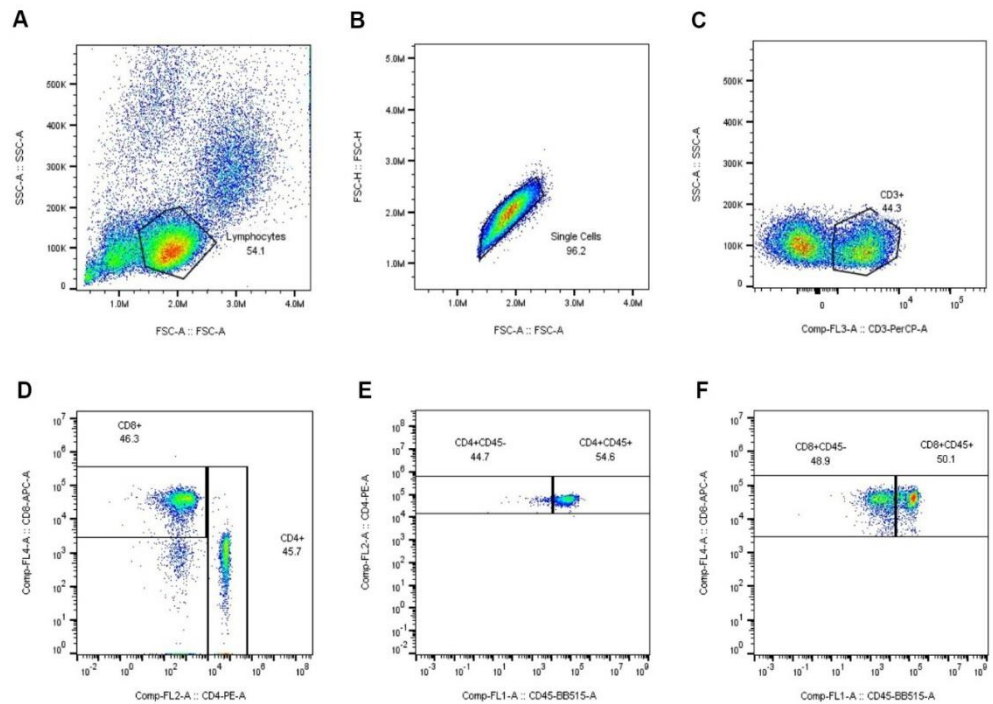


Figure S1. The gating strategy for the expression of naïve (CD45RA) markers on peripheral blood CD4⁺ and CD8⁺ T-lymphocytes in adults (APB). The peripheral blood T-lymphocytes were gated on forward (FSC) versus side scatter (SSC) to select the lymphocyte cell population (A). The cells were gated on FSC-Height (FSC-H) vs FSC-Area (FSC-A) to exclude all the doublets and to generate the singlets gate (B). Subsequently, the cells were gated on CD3-PerCP vs SSC-A to include all CD3⁺ cells (C), then gated on the CD4⁺ or CD8⁺ population (D). All the subpopulations were analysed on the CD4-PE-A (E) or CD8-APC-A (F) versus CD45-BB515-A. Data were analysed using FlowJo v10.6.1 software, and population frequencies expressed as percent of the CD4⁺ or CD8⁺ parent population.

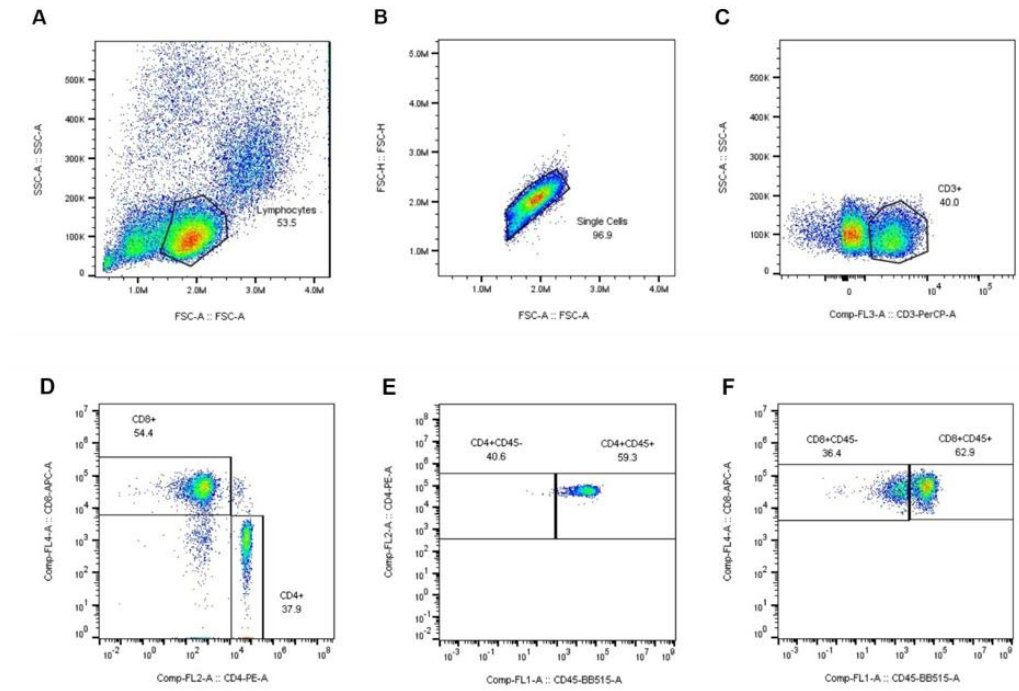


Figure S2. The gating strategy for the expression of memory (CD45RO) markers on peripheral blood CD4⁺ and CD8⁺ T-lymphocytes in adults (APB). The peripheral blood T-lymphocytes were gated on forward (FSC) versus side scatter (SSC) to select the lymphocyte cell population (A). The cells were gated on FSC-Height (FSC-H) vs FSC-Area (FSC-A) to exclude all the doublets and to generate the singlets gate (B). Subsequently, the cells were gated on CD3-PerCP vs SSC-A to include all CD3⁺ cells (C), then gated on the CD4⁺ or CD8⁺ population (D). All the subpopulations were analysed on the CD4-PE-A (E) or CD8-APC-A (F) versus CD45-BB515-A. Data were analysed using FlowJo v10.6.1 software, and population frequencies expressed as percent of the CD4⁺ or CD8⁺ parent population.

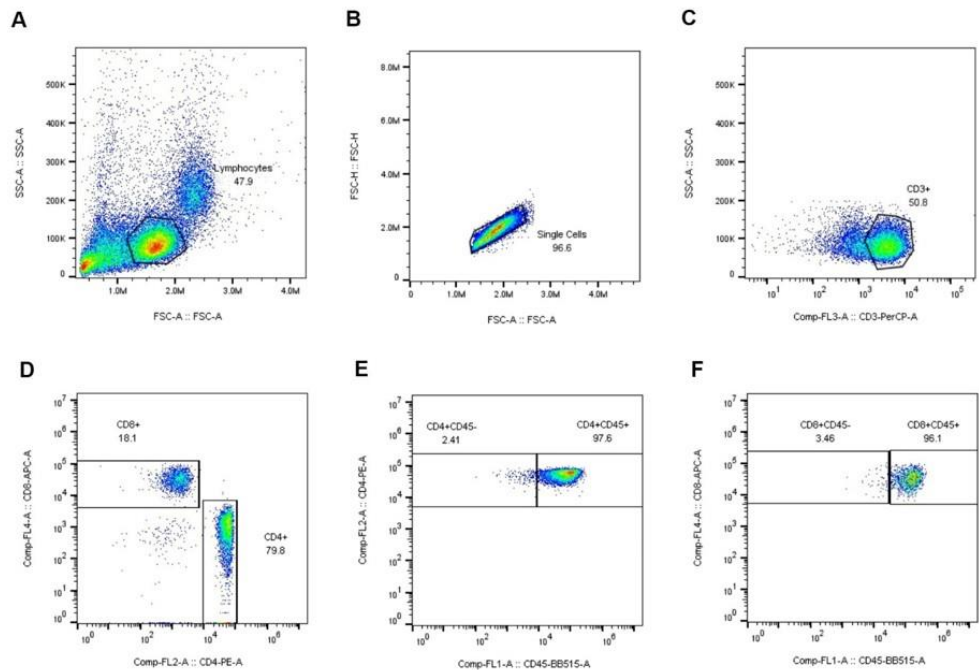


Figure S3. The gating strategy for the expression of naïve (CD45RA) markers on peripheral blood CD4⁺ and CD8⁺ T-lymphocytes in newborns (UCB). The peripheral blood T-lymphocytes were gated on forward (FSC) versus side scatter (SSC) to select the lymphocyte cell population (A). The cells were gated on FSC-Height (FSC-H) vs FSC-Area (FSC-A) to exclude all the doublets and to generate the singlets gate (B). Subsequently, the cells were gated on CD3-PerCP vs SSC-A to include all CD3⁺ cells (C), then gated on the CD4⁺ or CD8⁺ population (D). All the subpopulations were analysed on the CD4-PE-A (E) or CD8-APC-A (F) versus CD45-BB515-A. Data were analysed using FlowJo v10.6.1 software, and population frequencies expressed as percent of the CD4⁺ or CD8⁺ parent population.

the singlets gate (B). Subsequently, the cells were gated on CD3-PerCP vs SSC-A to include all CD3⁺ cells (C), then gated on the CD4⁺ or CD8⁺ population (D). All the subpopulations were analysed on the CD4-PE-A (E) or CD8-APC-A (F) versus CD45-BB515-A. Data were analysed using FlowJo v10.6.1 software, and population frequencies expressed as percent of the CD4⁺ or CD8⁺ parent population.

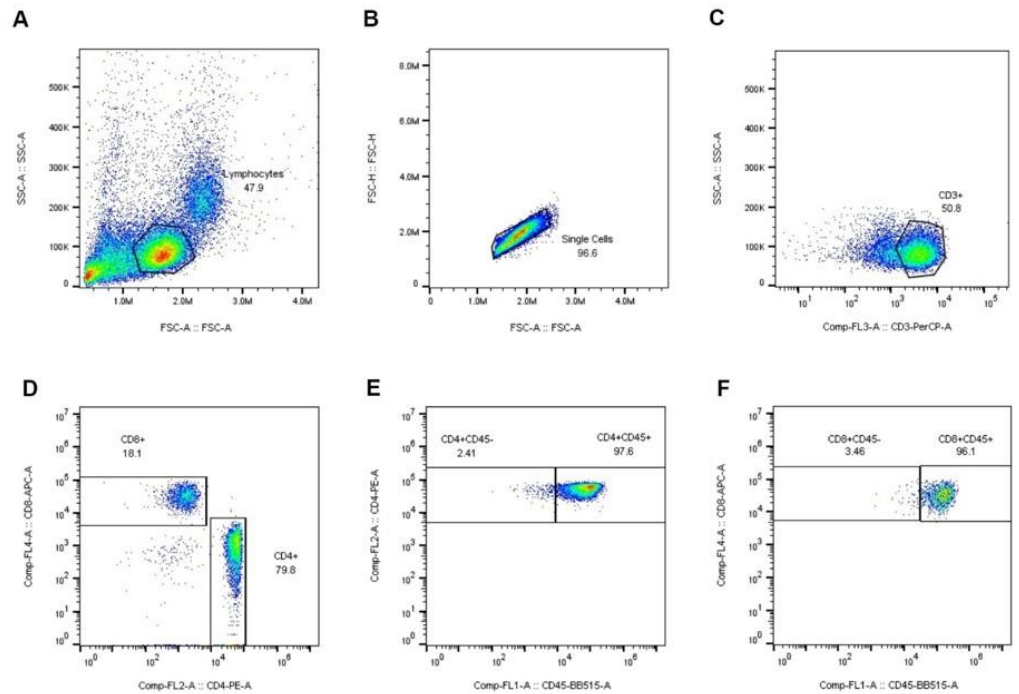


Figure S4. The gating strategy for the expression of memory (CD45RO) markers on peripheral blood CD4⁺ and CD8⁺ T-lymphocytes in newborns (UCB). The peripheral blood T-lymphocytes were gated on forward (FSC) versus side scatter (SSC) to select the lymphocyte cell population (A). The cells were gated on FSC-Height (FSC-H) vs FSC-Area (FSC-A) to exclude all the doublets and to generate the singlets gate (B). Subsequently, the cells were gated on CD3-PerCP vs SSC-A to include all CD3⁺ cells (C), then gated on the CD4⁺ or CD8⁺ population (D). All the subpopulations were analysed on the CD4-PE-A (E) or CD8-APC-A (F) versus CD45-BB515-A. Data were analysed using FlowJo v10.6.1 software, and population frequencies expressed as percent of the CD4⁺ or CD8⁺ parent population.