Supplementary Material

Chemodiversity of Arctic Plant *Dryas oxyodonta*: LC-MS Profile and Antioxidant Activity

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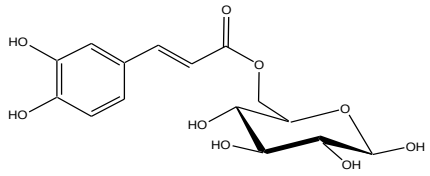
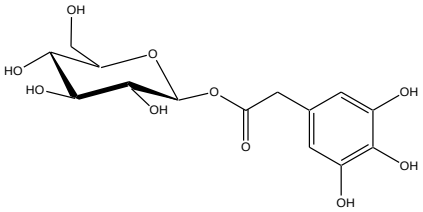
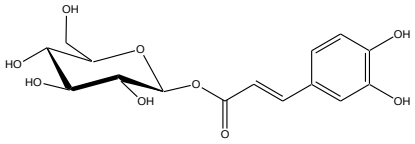


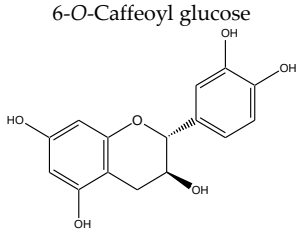
6-*O*-Caffeoyl glucose



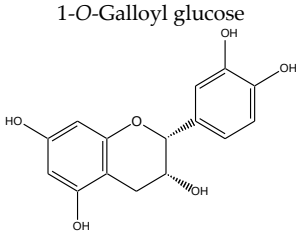
1-*O*-Galloyl glucose



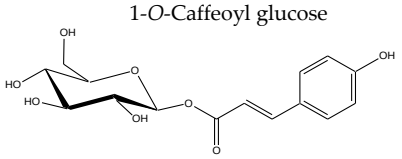
1-*O*-Caffeoyl glucose



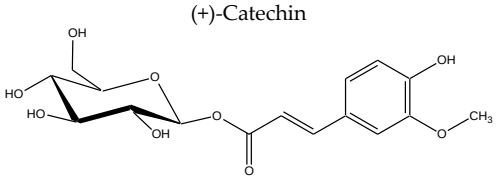
(+)-Catechin



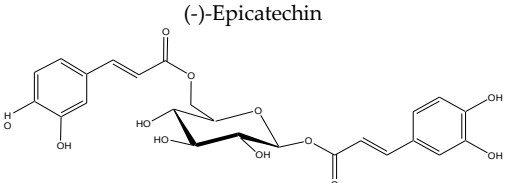
(-)-Epicatechin



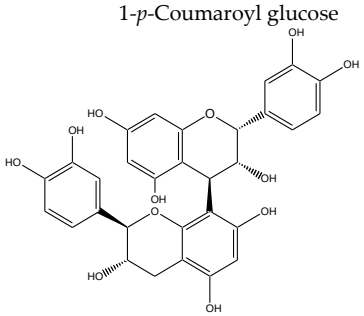
1-*p*-Coumaroyl glucose



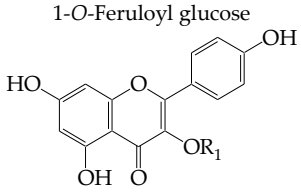
1-*O*-Feruloyl glucose



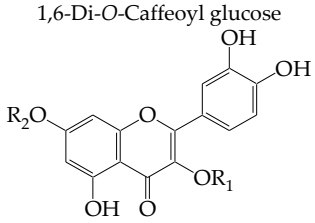
1,6-Di-*O*-Caffeoyl glucose



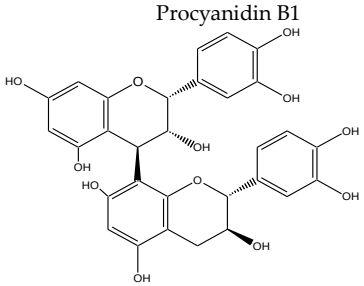
Procyanidin B1



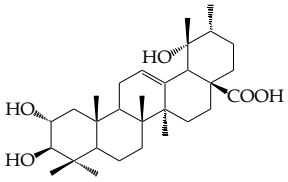
Kaempferol 3-*O*-arabinoside: R_1 = Ara; R_2 = H
Kaempferol 3-*O*-galactoside: R_1 = Gal; R_2 = H



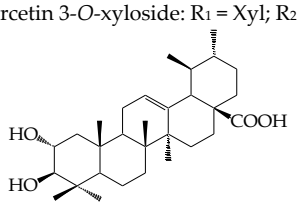
Quercetin 3-*O*-galactoside: R_1 = Gal; R_2 = H
Quercetin 3-*O*-arabinoside: R_1 = Ara; R_2 = H
Quercetin 3-*O*-xyloside: R_1 = Xyl; R_2 = H



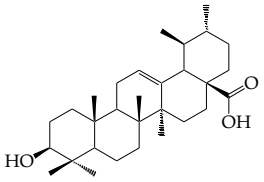
Procyanidin B2



Tormentic acid



Corosolic acid



Ursolic acid

Figure S1. Structures of compounds identified in *Dryas oxyodonta*. Abbreviation used: Gal – galactose; Ara – arabinose; Xyl – xylose.

Table S1. Conditions for liquid chromatography mass spectrometry detection of metabolites in *Dryas oxyodonta* extracts

Liquid chromatograph	LC-20 Prominence liquid chromatograph (Shimadzu, Columbia, MD, USA)
Photodiode array detector	SPD-M30A
Mass spectrometer	LCMS 8050 triple quadrupole
Column	GLC Mastro column (2.1 × 150 mm, 3 μm; Shimadzu, Kyoto, Japan)
Eluents	0.25% formic acid in water (A) and 0.25% formic acid in acetonitrile (B)
Gradient elution program	0–3 min (5–9% B), 3–6 min (9–13% B), 6–10 min (13–35% B), 10–15 min (35–58% B), 15–18 min (58–79% B), 18–20 min (79–95% B), 20–25 min (95–5% B)
Injection volume	1 μL
Flow rate	100 μL/min
Column temperature	28 °C
Spectral range of photodiode array detection	200–600 nm
Mass spectrometric mode detection	Negative electrospray ionization
Source voltage	3 kV
Collision energy	−35 eV
Scanning range	100–2000 *m*/*z*
ESI interface temperature	300 °C
Desolvation line temperature	250 °C
Heat block temperature	400 °C
Flow rate of nebulazing gas N_2	3 L/min
Flow rate of heating gas	10 L/min
Flow rate of collision-induced dissociation gas Ar	0.3 mL/min