

Supplementary Materials

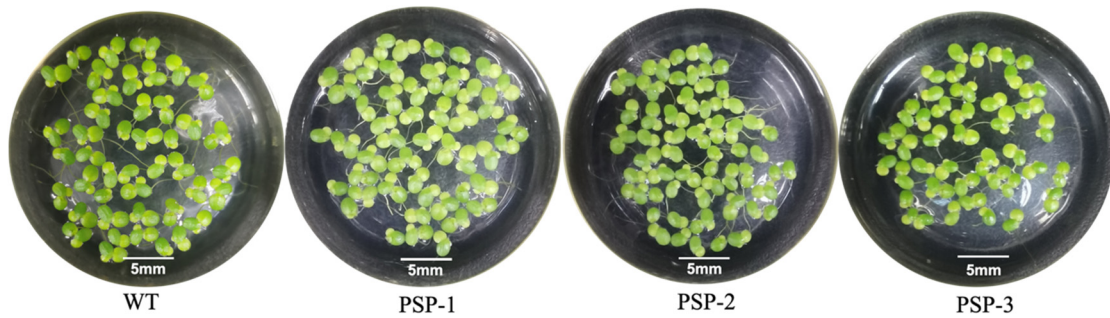


Figure S1. Phenotype of WT and three *AtPSP1* transgenic lines. The photos were taken after WT and *AtPSP1* transgenic lines were cultivated after full nutrition condition (Datko) for 9 days.

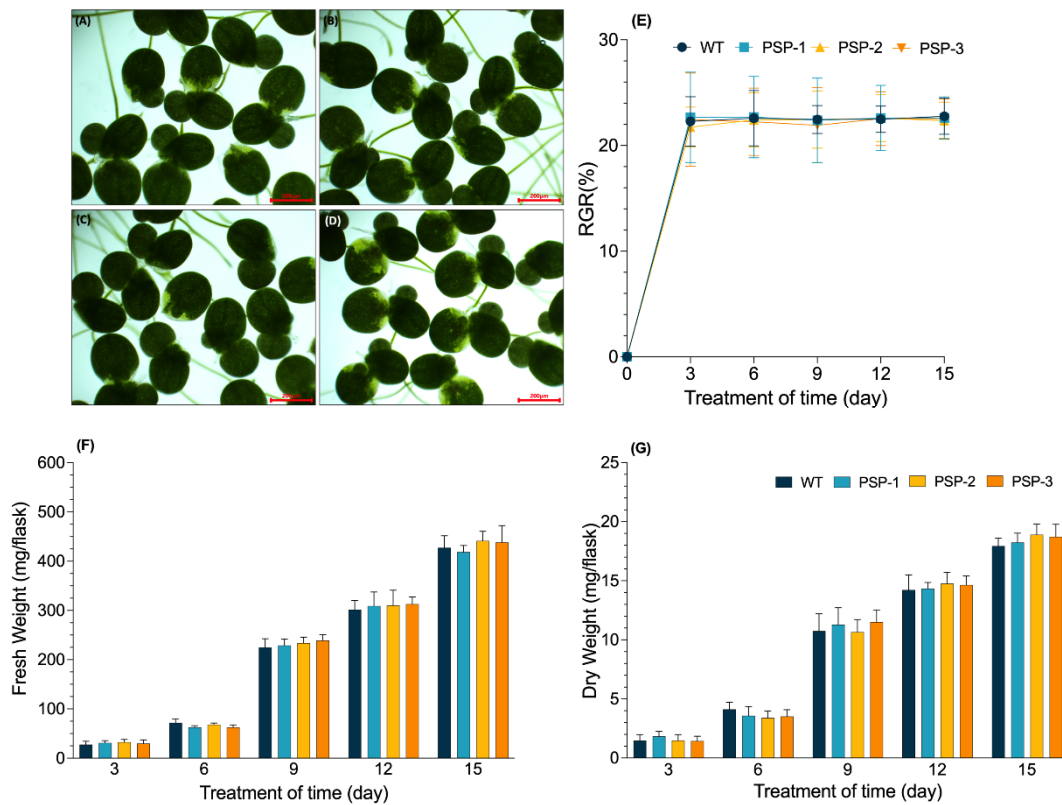


Figure S2. Analysis of phenotype, relative growth rate (RGR), fresh weight and dry weight in WT and three *AtPSP1* transgenic lines under Datko condition. **(A)** WT, **(B)** PSP-1, **(C)** PSP-2, **(D)** PSP-3 indicate the phenotype of WT and *AtPSP1* transgenic lines were cultivated under Datko condition for 9 days with dissecting microscope; **(E)** The RGR; **(F)** The fresh weight; **(G)** The dry weight. Each statistic is the mean \pm standard error ($n=9$, from three independent experiments with three repeats of each).

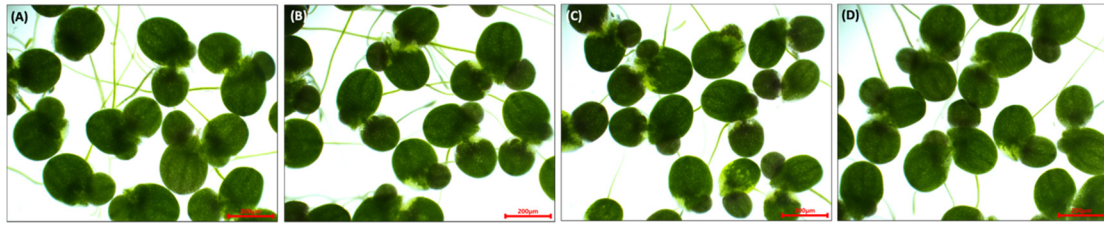


Figure S3. Phenotype of WT and three *AtPSP1* transgenic lines under sulfur-deficiency condition. **(A)** WT, **(B)** PSP-1, **(C)** PSP-2, **(D)** PSP-3 indicate the phenotype of WT and *AtPSP1* transgenic lines were cultivated under sulfur-deficiency condition for 9 days with dissecting microscope.

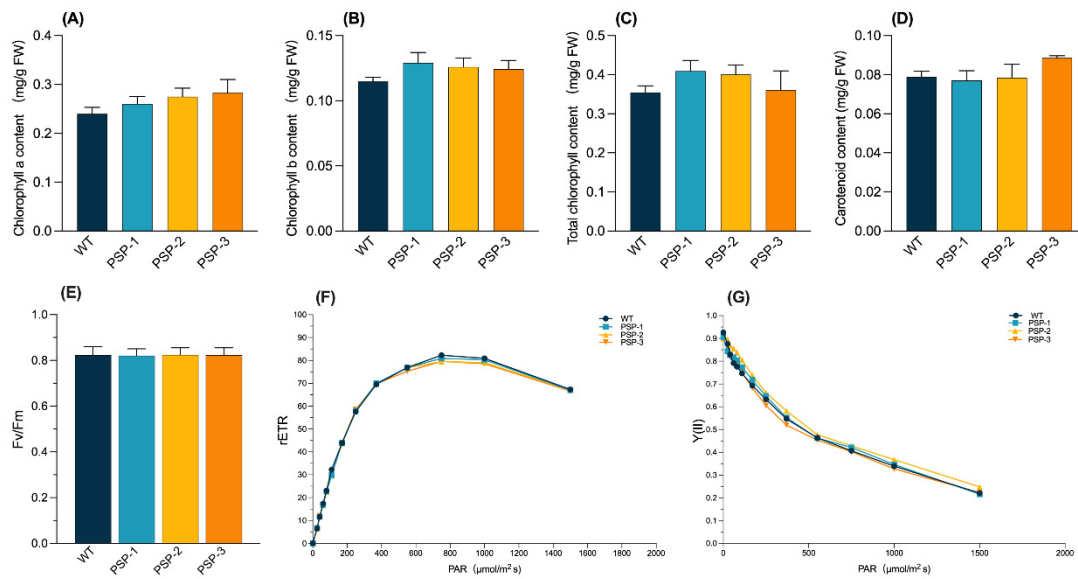


Figure S4. The analysis of photosynthetic pigment and chlorophyll fluorescence-related parameters in duckweed under full nutrition condition (Datko). **(A)** Content of Chlorophyll a; **(B)** Content of Chlorophyll b; **(C)** Total chlorophyll content; **(D)** Carotenoid content. The content of photosynthetic pigment was measured after the samples were cultivated under full nutrition condition for 9 days; **(E)** The value of Fv/Fm; **(F)** The value of rETR; **(G)** The value of Y(II), the chlorophyll fluorescence-related parameters were measured after the samples were cultivated under full nutrition condition for 6 days. Each statistic is the mean \pm standard error (n=9, from three independent experiments with three repeats of each, the data no average value in Figure F and G).

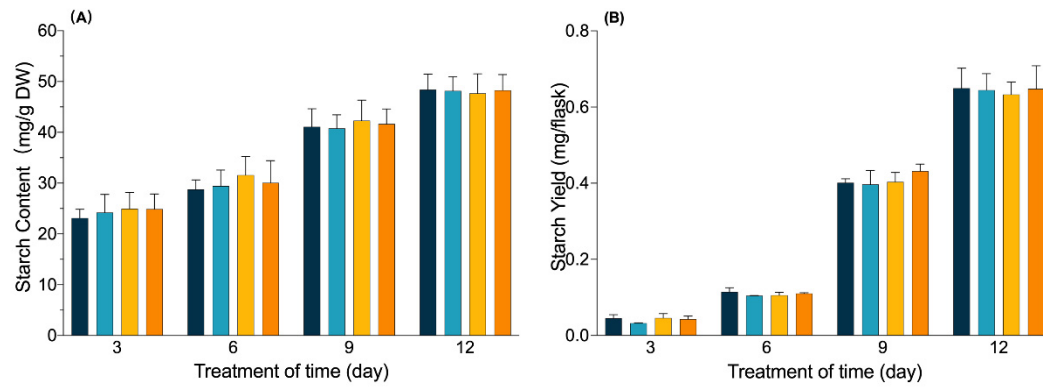


Figure S5. The content of starch , yield and the soluble protein in WT and three *AtPSP1* transgenic lines under full nutrition condition in different treatment of time. **(A)** The starch content; **(B)** The starch yield. Each statistic is the mean \pm standard error (n=9, from three independent experiments with three repeats of each).

Table S1 The primer sequences in the study.

Primer name	Primer sequence (5'-3')	Note	
<i>AtPSP1-F</i> <i>AtPSP1-R</i>	CCATGGGGATGGAAGCATTAAGTACTTCA GGGTTACCCTTAGTCCAATGAGTTTATGAG	The primers for genes amplification	
<i>AtPSP1-RT-F</i> <i>AtPSP1-RT-R</i> <i>Lt18S-RT-F</i> <i>Lt18S-RT-R</i>	GAAAGGCTGTTGCTGAAT TCGTTCTCATCAAATCCC ATACCGTCCTAGTCTCAACCA ACAAATCGCTCCACCAAC	The primers of genes for RT-PCR	
<i>AtPSP1-qRT-F</i> <i>AtPSP1-qRT-R</i>	GGTTGTGCCAGTTCAGGT ACGAAGCAATTTGGGATG	The primer of <i>AtPSP1</i>	The primers of genes for qRT-PCR
<i>Lt18S-qRT-F</i> <i>Lt18S-qRT-R</i>	ATAAACGATGCCGACCAG TCAGCCTTGCGACCATAC	The primer of <i>Lt18S</i>	
<i>LtAPK1-qRT-F</i> <i>LtAPK1-qRT-R</i> <i>LtAPR1-qRT-F</i> <i>LtAPR1-qRT-R</i> <i>LtPAPSS1-qRT-F</i> <i>LtPAPSS1-qRT-R</i> <i>LtSERAT1-qRT-F</i> <i>LtSERAT1-qRT-R</i> <i>LtSIR1-qRT-F</i> <i>LtSIR1-qRT-R</i> <i>LtSULTR1-qRT-F</i> <i>LtSULTR1-qRT-R</i> <i>LtSULTR2-qRT-F</i> <i>LtSULTR2-qRT-R</i> <i>LtSULTR3-qRT-F</i> <i>LtSULTR3-qRT-R</i> <i>LtSULTR4-qRT-F</i> <i>LtSULTR4-qRT-R</i> <i>LtSUOX1-qRT-F</i> <i>LtSUOX1-qRT-R</i>	GAAGGAGCCAGTGGTGAA AGTCGATACAGGGCGAAG TCGCCATAGCTTTCAGTG TTTCAGGGTTCAATCTTCC ATGTCTGTTCCCATCGTTC TCTTCTTTGTGGTGCTTGT GCGGCTAATCAAGTGCTG TTTCGGCGTAACTCCTCA GTGAGCGTCCAGGATTTA CCGATGCGATTAGTGAAG TTTCTTCGGAGTCATCTT CATTACACCAGGAATCTTT AGCTCTGGCTGCAAGACT ATGGCTGTAGGAGTGAAGTA TCGCCAACGCCTGCTACT CGCCCAAGTCCAAGACCACA TGGGTCTGGTGGACTATGA ACAAATGCAAGGGAGGAG GGCGTATGAAATGAATGG GTCAACTATGGGTGGGAAC	The primer of sulfur assimilation related genes	
<i>LtAPS1-qRT-F</i> <i>LtAPS1-qRT-R</i> <i>LtSSS1-qRT-F</i> <i>LtSSS1-qRT-R</i> <i>LtGBSS1-qRT-F</i> <i>LtGBSS1-qRT-R</i> <i>LtAPL1-qRT-F</i> <i>LtAPL1-qRT-R</i>	ATAGAGCATAGCGTCGTTG TTCATCCTCCGTCTCATAG TGTGGCTCTTTGCCTGTA TTCAACTTCGTGCTCTGC CGGGTAACAGGAAGGGTG TCCGGTTCTTTGGTGATTGA CACCGTATCCCAAGTCCC TCACCGTCTCGAAATCGT	The primer of starch metabolism related genes	

<i>LtISA1-qRT-F</i>	GCACGCCTCTGAGCAACCCCT	The primer of starch metabolism related genes	The primers of genes for qRT- PCR
<i>LtISA1-qRT-R</i>	TGCCCATTCCATTCTGACCACA		
<i>Ltα-Amy1-qRT-F</i>	ATAATCCGCTCAGAGTTCG		
<i>Ltα-Amy1-qRT-R</i>	ACCGATACCTTACCCTCC		
<i>Ltβ-Amy1-qRT-F</i>	GGCATTTCATTGGTGGTAC		
<i>Ltβ-Amy1-qRT-R</i>	GTCGTGCTTCTTGAGGGT		
<i>LtPFK1-qRT-F</i>	TCGGAATAATCGCTGTCTC	The primer of carbon metabolism related genes	
<i>LtPFK1-qRT-R</i>	CGTGAACCTTGTCCTTACC		
<i>LtFBA1-qRT-F</i>	AACCTCAATGCCATGAACC		
<i>LtFBA1-qRT-R</i>	TCCACCTTACCTGCCCAC		
<i>LtPK1-qRT-F</i>	TCAGGCAGAGCAGCAAAG		
<i>LtPK1-qRT-R</i>	CTGGGTTCCTCGAGAAAGT		
<i>LtPDC1-qRT-F</i>	GCCCAATCTTCAACGACTA		
<i>LtPDC1-qRT-R</i>	CTTCATCAGAACGCACCC		
<i>LtACO1-qRT-F</i>	GGAGTGACCGCAACAGAT		
<i>LtACO1-qRT-R</i>	GCACCGTATTGAGGAGACA		
<i>LtIDH1-qRT-F</i>	AACGACATACGGCTTCTA		
<i>LtIDH1-qRT-R</i>	TTACCCATTACCTGACAAC		
<i>Lt2OG-DH1-qRT-F</i>	ATTCACGGAGATGGGAGTT		
<i>Lt2OG-DH1-qRT-R</i>	CAGCGGTCCGATCAGTAG		
<i>LtSDH1-qRT-F</i>	AATGTAACCAAGGAGCCG		
<i>LtSDH1-qRT-R</i>	TTTGCGCCATGAACTGAT		
<i>LtFUM1-qRT-F</i>	AACGACATACGGCTTCTA		
<i>LtFUM1-qRT-R</i>	ATTACCCATTACCTGACAAC		
<i>LtMDH1-qRT-F</i>	GCTTCATCTGGGTCTGGT		
<i>LtMDH1-qRT-R</i>	CGTTCGTCATCGTCGTTA		
<i>LtCS1-qRT-F</i>	TTGGACTATGGCGGAAAT		
<i>LtCS1-qRT-R</i>	GCAGTATGGGCGCTAACA		

The sequence of genes

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Lt18S

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LtSERAT1

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LtSIR1

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LtSULTR1

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LtSULTR2

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LtSULTR3

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LtSULTR4

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LtSUOX1

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LtAPS1

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LtSSS1

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LtGBSS1

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LtAPL1

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LtISA1

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Lta-Amy1

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LtPFK1

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LtFBA1

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LtPK1

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LtPDC1

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LtACO1

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LtIDH1

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Lt2OG-DH1

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LtSDH1

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LtFUM1

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LtMDH1

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LtCS1

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