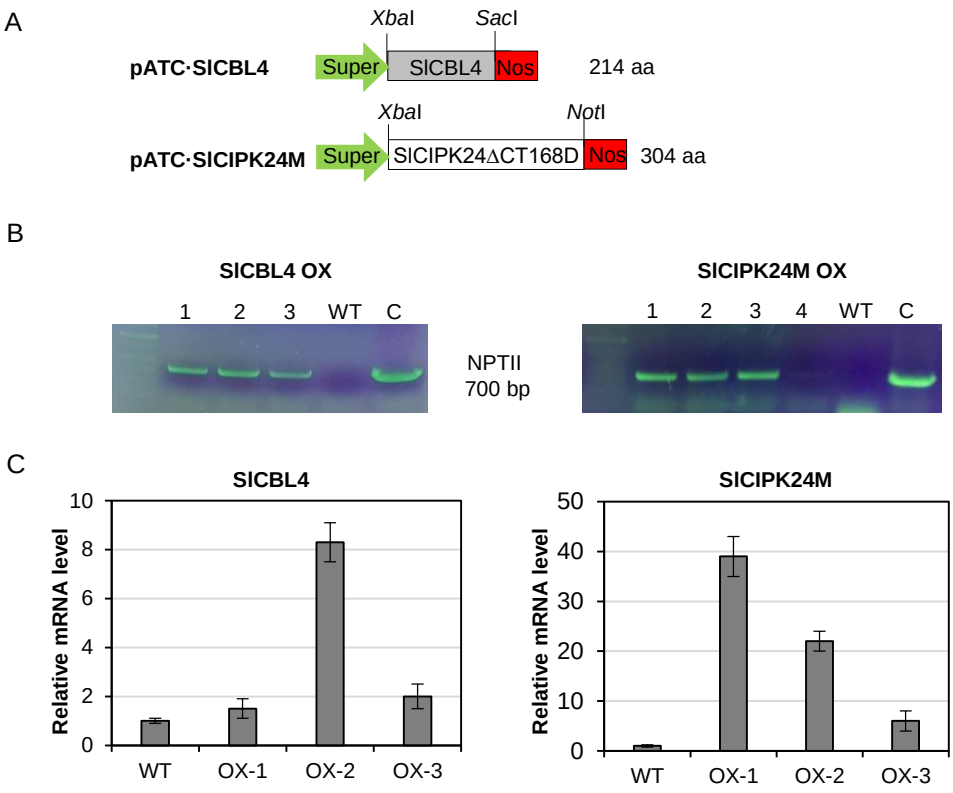
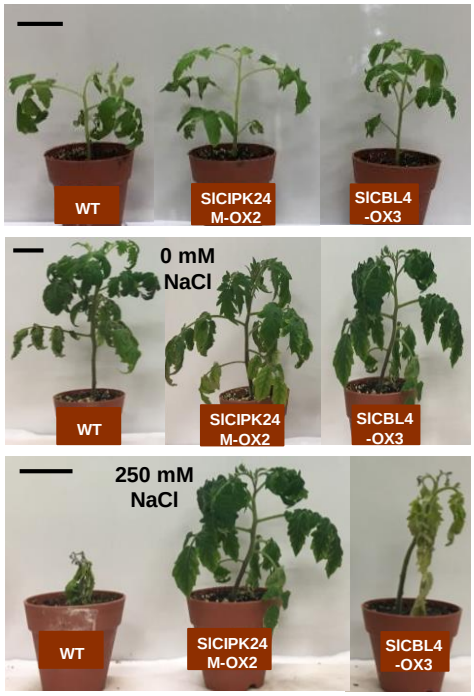


Supplemental Figure S1. Isolation of transgenic tomato (cv. Moneymaker) plants overexpressing SlCBL4 or superactive SlCIPK24 mutant. (A) Schematic diagrams of the pATC·SlCBL4 and pATC·SlCIPK24M. (B) PCR analysis showing the presence of the overexpression constructs in the transgenic lines. Genomic DNAs prepared from the indicated plants was amplified with a pair of primers (NPTII-F: 5'- GCTGCTCTGATGCCGCCGTGTTCCG-3', NPTII-R: 5'-AACTCGTCAAGAAGGCGATAGAAGGC -3) to produce a 700 bp fragment of the kanamycin resistance gene (NPTII). Lane C contains the PCR product from the plasmid template and serves as a positive control. (C) Real-time qRT-PCR analysis of the transcript levels in the WT and transgenic tomato lines. Total RNA was extracted from 3-week-old tomato plants. *SlActin7* was employed as an internal control for transcript normalization. The expression levels of *SlCBL4* and *SlCIPK24* in WT plants were respectively set to 1.0 to show their relative abundance in the transgenic plants. Error bars denote standard deviation (SD) of three technical replicates. Values are significantly different (the Student's *t*-test, $P \leq 0.05$).



Supplemental Figure S2. Salt stress tolerance of the mature transgenic plants. Three-week-old plants (top panel) of the indicated lines were treated with 0 mM (middle panel) or 250 mM NaCl solution (bottom panel). The photos were taken on the 21st day after treatment. *Scale bars*, 5 cm. All the experiments were performed in three biological replicates.

Supplemental Table S1. Primers used for plasmid construction. Restriction enzyme sites were underlined. Three additional bases were included at the 5′ end of the primers for efficient restriction enzyme digestion.

Primers	Sequences 5' to 3' (Restriction enzyme site)
SlCBL4-1	TATGAATTCGATGGGCTGCTTTCCCTCAA (EcoRI)
SlCBL4-2	TTTGTCGACCTAGACTTCCGAATCTTCCA (SalI)
SlCBL4-3	AAACCATGGTAATGGGCTGCTTTCCCTCA (NcoI)
SlCBL4-4	AAAACTAGTGACTTCCGAATCTTCCACCT (SpeI)
c-Myc-2	AAAGCGGCCGCAAGATCCTCCTCAGAAATCA (NotI)
SlCBL4-5	TATGTCGACATGGGCTGCTTTCCCTCAAA (SalI)
SlCBL4-6	TTACTCGAGGACTTCCGAATCTTCCACCT (XhoI)
SlCBL4-7	ATAACTAGTATGGGCTGCTTTCCCTCAAA (SpeI)
SlCBL4-8	TAAGAGCTCCTAGACTTCCGAATCTTCCA (SacI)
SlCIPK24-1	AAAGGATCCCATGAAGAAAGTGAAGAGAA (BamHI)
SlCIPK24-2	AATGTCGACTCAGCGAGTCCTTGTCCTAA (SalI)
SlCIPK24-3	TTTGTCGACTTACTCACTATCTTCTGATTTTT (SalI)
SlCIPK24-4	TCTGGATCCGAGTGGTCCCTTGGTAATGA (BamHI)
SlCIPK24-5	AAAGGATCCATGAAGAAAGTGAAGAGAAA (BamHI)
SlCIPK24-6	AAATCTAGAATGAAGAAAGTGAAGAGAAA (XbaI)
SlCIPK24-7	AATGGATCCGCGAGTCCTTGTCCTAAGCA (BamHI)
SlCIPK24-8	AAACCATGGTAATGAAGAAAGTGAAGAGA (NcoI)
SlCIPK24-9	AAAACTAGTGCGAGTCCTTGTCCTAAGCA (SpeI)
SlCIPK24-10	TTTGCGGCCGCACTCACTATCTTCTGATT (NotI)
SlCIPK24-TD1	GTCGAGCTCCTCTATGACACTTGTGGGACTCC
SlCIPK24-TD2	GGAGTCCCACAAGTGTCATAGAGGAGCTCGAC

Supplemental Table S2. Primers used for quantitative real-time RT-PCR.

Primers	Sequence 5’ to 3’	Size
SlCBL4 Forward	CTGCTGGCTGCTGAGACCGCTTTTA	181 bp
SlCBL4 Reverse	CAAATAGATCAAAAATCCTGTCTGC	
SlCIPK24 Forward	TAAGCATAGAATGGTTGAACAGATC	179 bp
SlCIPK24 Reverse	CAGAAAGCCTACCTAGATGAACAAT	
SlRD29B Forward	GTCACCGATCCAACAGGCGCCAACA	224 bp
SlRD29B Reverse	CCAACCGAGGGGAAAACAGTAGGCG	
SlRD22 Forward	GAGAATGTGAAGAGCCAGGTATTA	191 bp
SlRD22 Reverse	GTCATTGTTGTTTCCCATTTTCTTG	
TAS14 Forward	TCGATCCGACAGCTCTAGCTCGTCG	172 bp
TAS14 Reverse	AGGGATCTTGTCCTTGATTTTGTCC	
SlActin7 Forward	TGCTAGCGGTCGTACCACTGGTATT	181 bp
SlActin7 Reverse	TACAATTTCCCGTTCAGCAGTAGTG	
SlTubulin Forward	AACCTCCATTCAGGAGATGTTTAGA	180 bp
SlTubulin Forward	TCTGCTGTAGCATCCTGGTATTGAA	