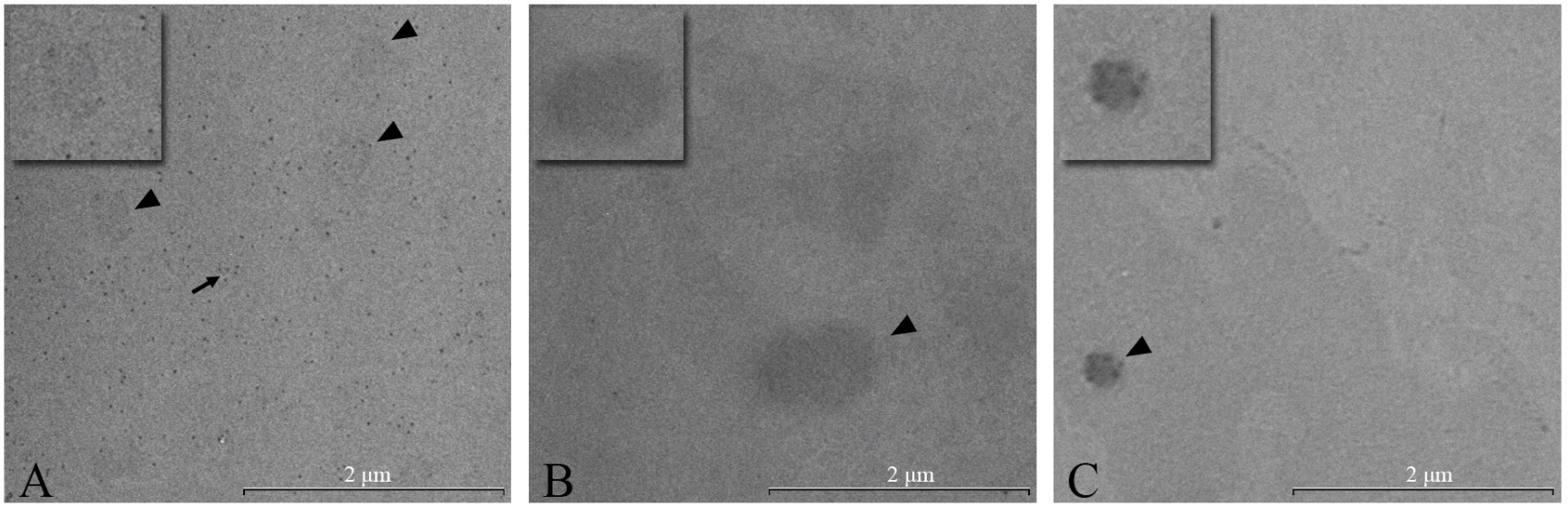


Figure S 1 Representative transmission electron microscopy images of natamycin-loaded liposomal vesicles: (A) 90H + nat, proliposome, (B) S100 + nat, proliposome, and (C) S100+ nat, thin film; bar - 2 µm. The arrow points to the natamycin particles; whereas the arrowhead shows the liposomal vesicle.

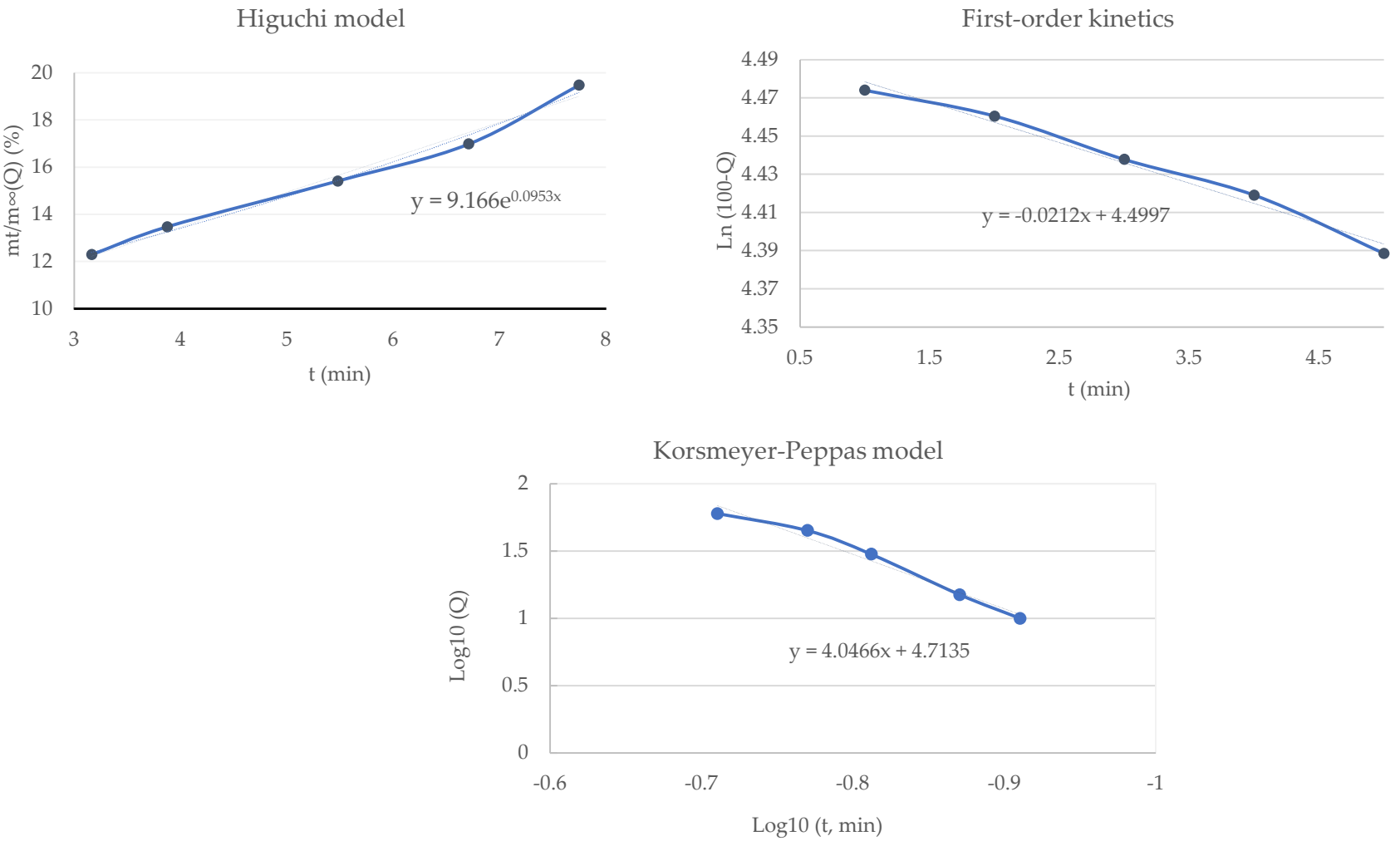


Figure S2 Kinetic fitting of natamycin release from the natamycin solution, using the Higuchi, First-order, and Korsmeyer-Peppas models. Fitting data are represented by lines, while symbols refer to experimental data of cumulative natamycin release from liposomes.

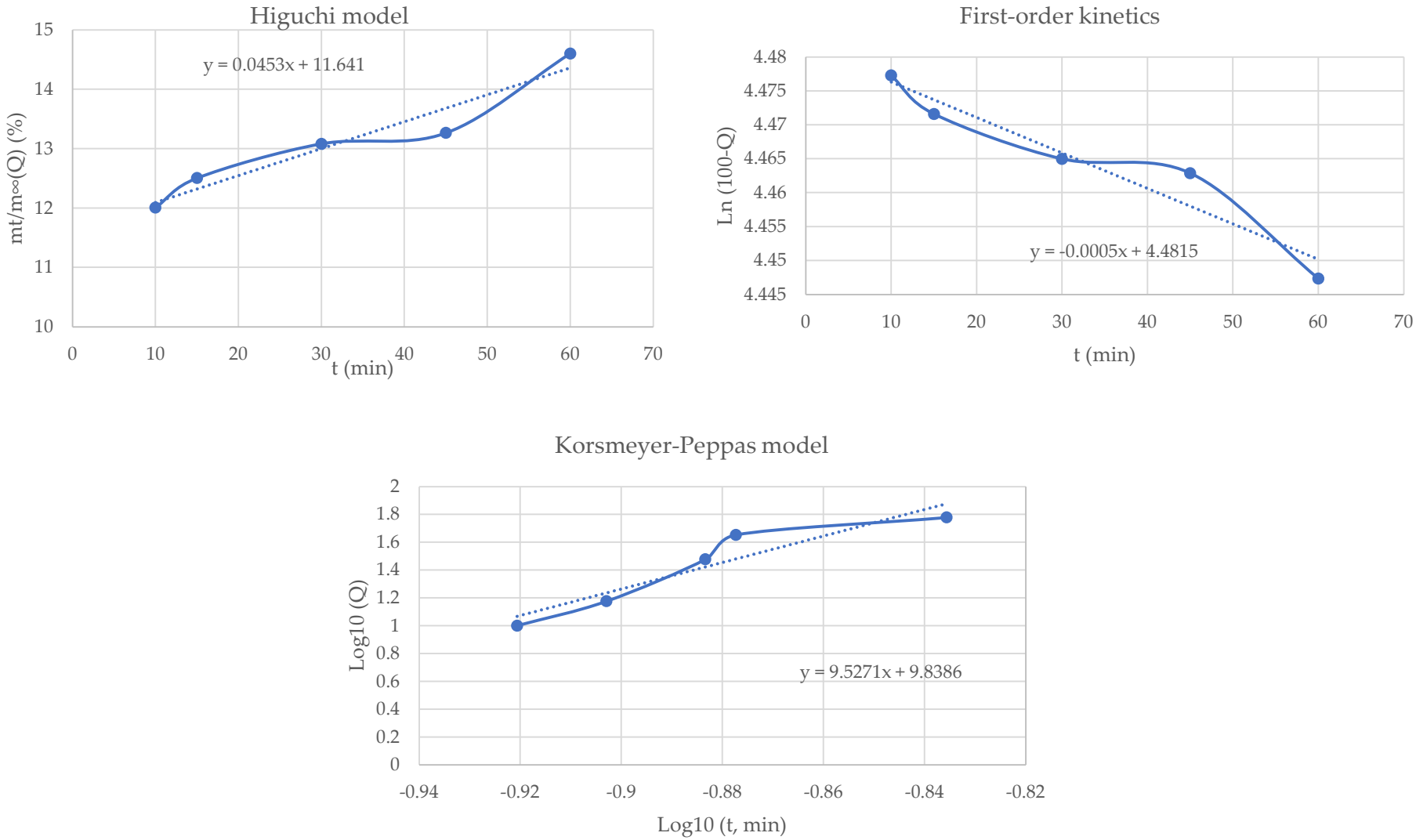


Figure S 3 Kinetic fitting of natamycin release from the tested liposomal formulations prepared by the proliposome method and Lipoid S100 phospholipid, using the Higuchi, First-order, and Korsmeyer-Peppas models. Fitting data are represented by lines, while symbols refer to experimental data of cumulative natamycin release from liposomes.

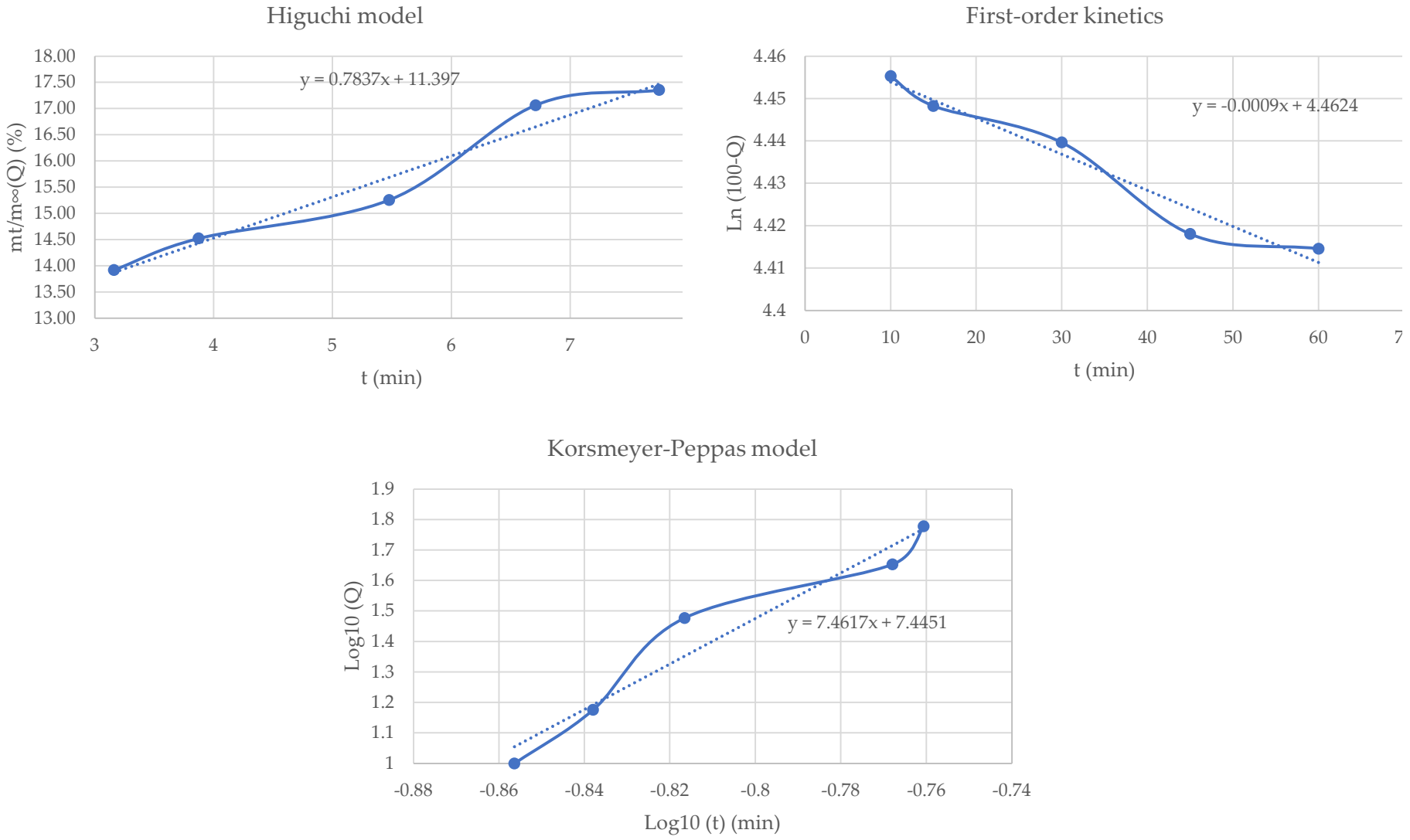


Figure S 4 Kinetic fitting of natamycin release from the tested liposomal formulations prepared by the thin-film method and Lipoid S100 phospholipid, using the Higuchi, First-order, and Korsmeyer-Peppas models. Fitting data are represented by lines, while symbols refer to experimental data of cumulative natamycin release from liposomes.

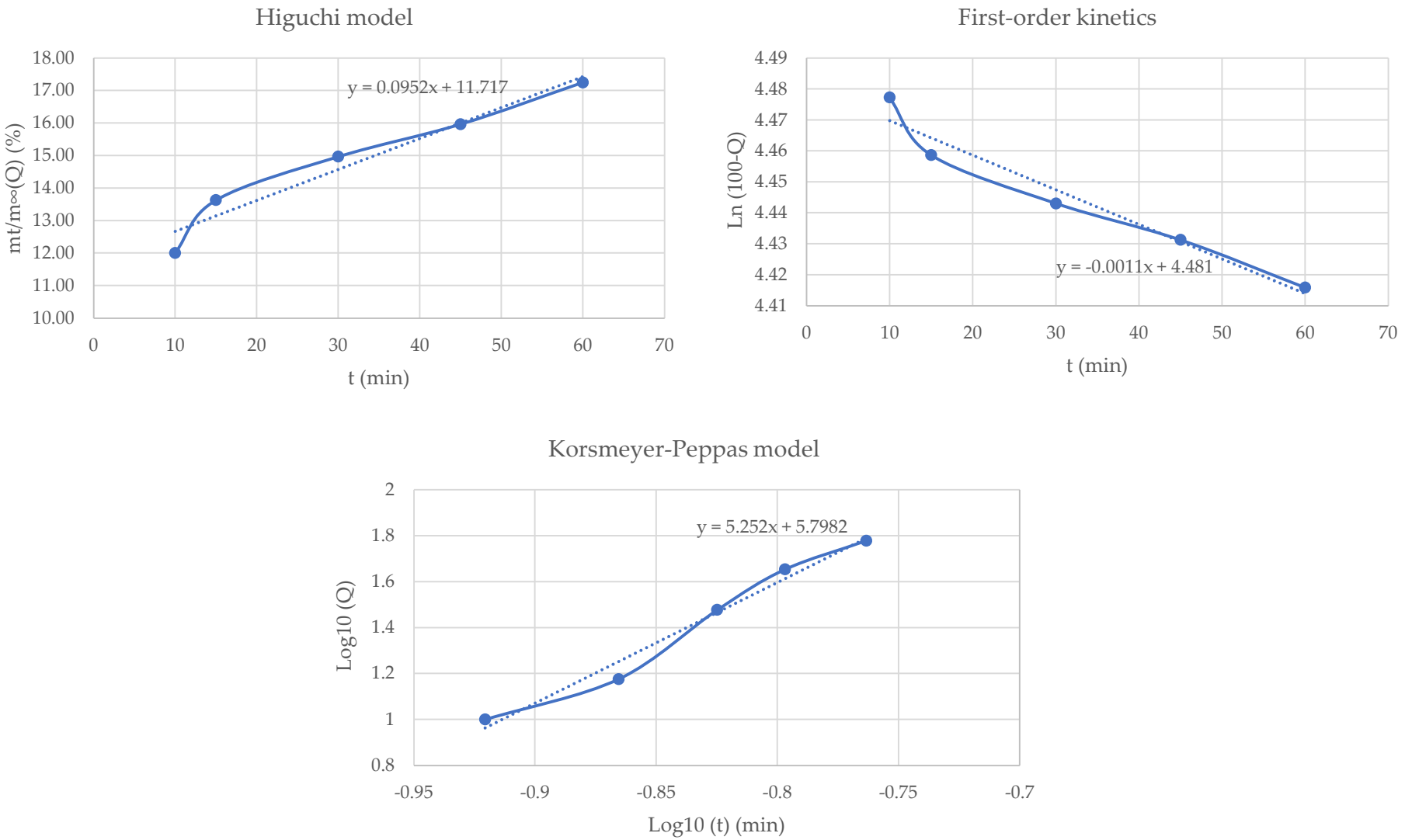


Figure S 5 Kinetic fitting of natamycin release from the tested liposomal formulations prepared by the proliposome method and Phospholipon 90H phospholipid, using the Higuchi, First-order, and Korsmeyer-Peppas models. Fitting data are represented by lines, while symbols refer to experimental data of cumulative natamycin release from liposomes.

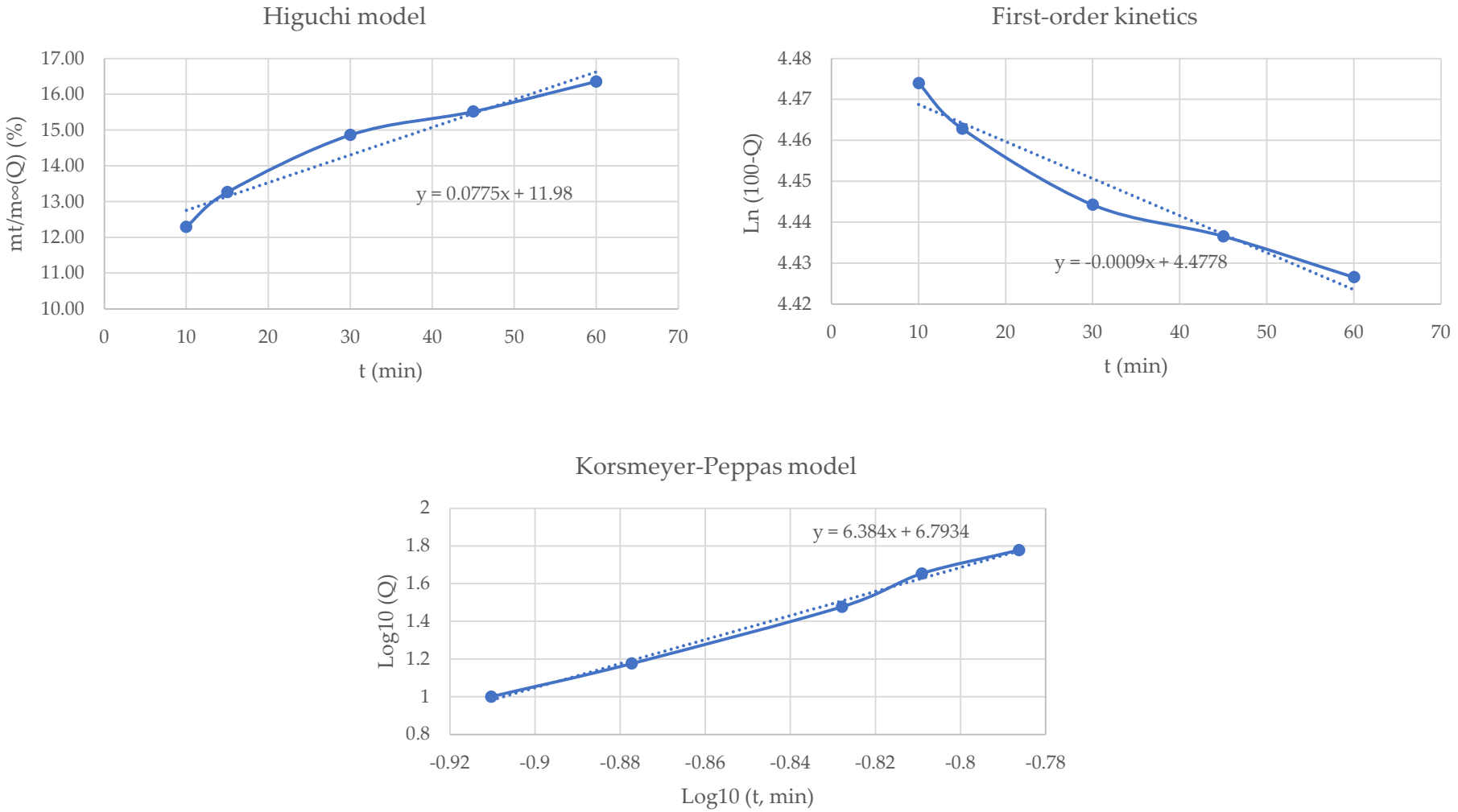


Figure S 6 Kinetic fitting of natamycin release from the tested liposomal formulations prepared by the thin-film method and Phospholipon 90H phospholipid, using the Higuchi, First-order, and Korsmeyer-Peppas models. Fitting data are represented by lines, while symbols refer to experimental data of cumulative natamycin release from liposomes.

Table S1 Polydispersity index (PDI) and zeta potential of blank (control) and natamycin-loaded liposomal vesicles prepared using thin film and proliposome techniques, and Lipoid S100 and Phospholipon 90H phospholipids, determined using photon correlation spectroscopy

Sample	PDI	Zeta potential (mV)
90H + natamycin, thin film	0.387±0.115	-37.98±0.98
90H, thin film	0.546±0.114	-41.00±1.57
S100 + natamycin, thin film	0.321±0.112	-32.90±2.07
S100, thin film	0.356±0.089	-28.54±0.98
90H + natamycin, proliposome	0.654±0.115	-32.15±1.15
90H, proliposome	0.721±0.118	-31.24±2.20
S100 + natamycin, proliposome	0.367±0.067	-30.23±2.56
S100, proliposome	0.836±0.110	-35.32±1.59

Table S2 Model parameters for First-order kinetics of natamycin release from liposomes and natamycin solution into pH=5.5 phosphate-buffered saline medium

Formulation	**Equation (First-order kinetics)**	**Parameter R^2**
90H + natamycin, proliposome	$\frac{m_t}{m_\infty} = 100 \cdot [1 - e^{-kt}]$	0.951
S100 + natamycin, thin film		0.955
90H + natamycin, thin film		0.947
S100 + natamycin, proliposome		0.926
Natamycin solution, 1 mg mL^{-1}		0.976

Table S3 Model parameters for Higuchi kinetics of natamycin release from liposomes and natamycin solution into pH=5.5 phosphate-buffered saline medium

Formulation	**Equation (Higuchi)**	**Parameter R^2**
90H + natamycin, proliposome	$\frac{m_t}{m_\infty} = \mathrm{k} \cdot t^{1/2}$	0.983
S100 + natamycin, thin film		0.944
90H + natamycin, thin film		0.947
S100 + natamycin, proliposome		0.959
Natamycin solution, 1 mg mL^{-1}		0.927

Table S4 Model parameters for Korsmeyer-Peppas kinetics of natamycin release from liposomes and natamycin solution into pH=5.5 phosphate-buffered saline medium

Formulation	Equation (Korsmeyer-Peppas)	Parameter R^2
90H + natamycin, proliposome	$\frac{m_t}{m_\infty} = \mathrm{k} \cdot t^n$	0.978
S100 + natamycin, thin film		0.946
90H + natamycin, thin film		0.995
S100 + natamycin, proliposome		0.879
Natamycin solution, 1 mg mL^{-1}		0.976