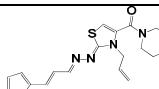
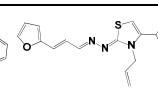
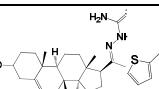
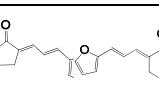
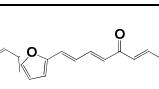


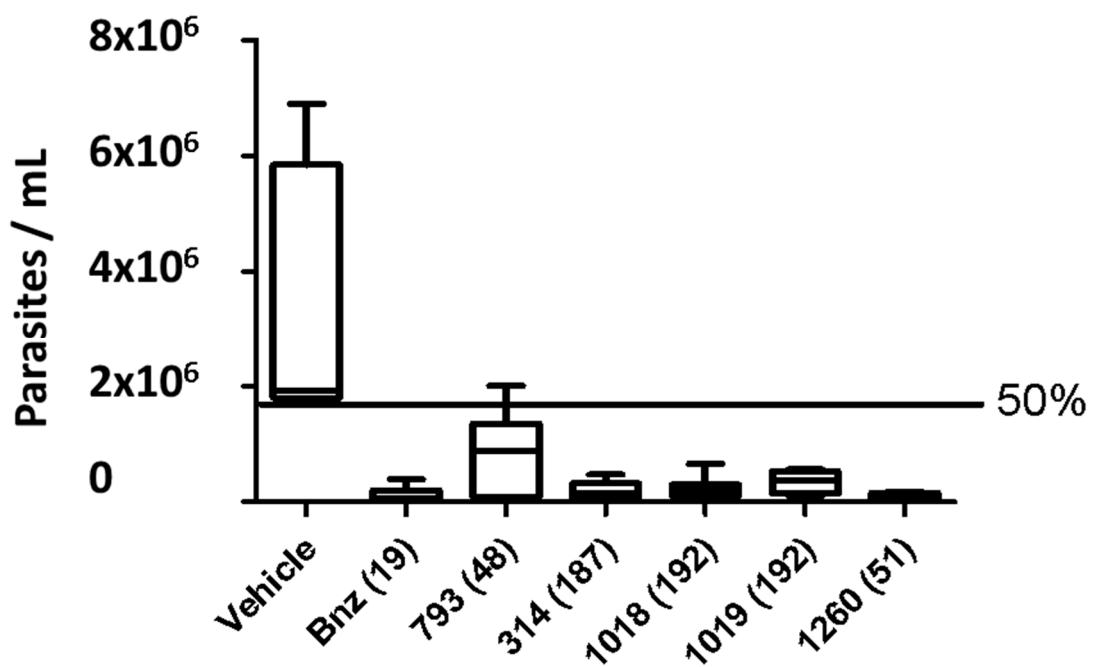
Supporting material

Table S1. Activity background. The anti-trypanosomatid activity identified inhibition in *T. cruzi* bio-system and toxicological profile of the best hits selected from our in-house chemical collection[33–35,44].

HITS					
314 ^a	266	1260	1018	1019	793
					
<i>In vitro</i> anti-<i>T. cruzi</i> activity (multiple strains) (IC₅₀)^b					
0.72 μM amastigotes	<0.25 μM amastigotes	0.1 μM amastigotes	40 nM epimastigotes	0.6 μM epimastigotes	5.0 μM epimastigotes
<i>In vitro</i> anti-<i>T. brucei</i> activity (IC₅₀)					
5.0 μM	5.0 μM	unknown	15 μM	22 μM	17 μM
<i>In vitro</i> anti-<i>Leishmania</i> spp (cutaneous) activity (IC₅₀)					
12 μM promastigotes	7 μM promastigotes	<20 μM promastigote s	>100 μM promastigote s	7.0 μM promastigotes	36 μM promastigotes
<i>In vitro</i> anti-<i>Leishmania infantum</i> (visceral) activity (IC₅₀)					
4.0 μM promastigotes	2.0 μM promastigotes	<0.2 μM amastigotes	11 μM promastigotes	13 μM promastigotes	31 μM promastigotes
Selectivity index >10 (IC₅₀ mammalian cell/IC₅₀ parasites)					
Identified <i>T. cruzi</i> bio-system inhibited by compounds					
Cruzipain inhibitor	unknown	Unknown	Triosephosphat e isomerase inhibitor	Triosephosphat e isomerase inhibitor	Triosephosphat e isomerase inhibitor
IC ₅₀ 4.3 μM			IC ₅₀ 4.7 μM	IC ₅₀ 86 nM	IC ₅₀ 3.0 μM
<i>In vitro</i> stability (microsomal, plasma, other solutions)					
high	high	High	Unknown	unknown	unknown
Toxicology and efficacy					
Ames test (mutagenicity)					
NO	NO	NO	Unknown	unknown	NO
Micronucleus test in mice (Genotoxicity)					
NO	NO	NO	Unknown	unknown	unknown
Teratogenicity in zebrafish (LD₅₀, μM)^c					
>25	>50	>15	Unknown	unknown	unknown
Acute oral toxicity in mice (Up and Down test, LD₅₀, mg/kg of body weight)					
>2000	>2000	>2000	Unknown	unknown	unknown
Full control of the parasitemia <i>in vivo</i> at 50 mg/kg in the murine model of Chagas disease and cutaneous model of Leishmaniasis					

a-The numbers are referred to the chemical collection. b- Is the concentration that inhibited the 50% of the biological response. c- the dose that kills 50% of the animals

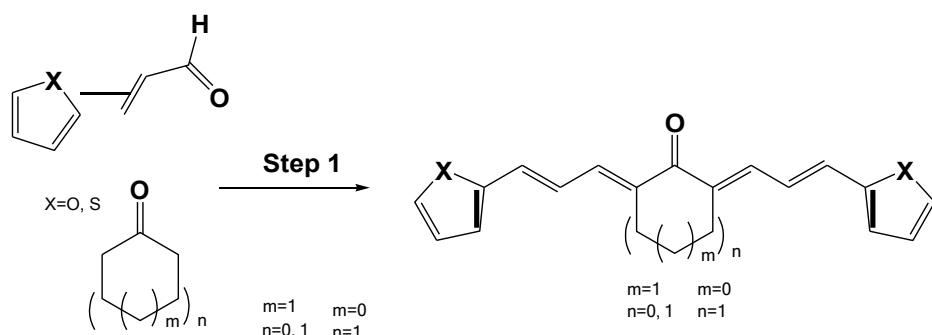
Figure S1. *In vivo* monotherapy activity of selected molecules in the Chagas disease murine model. The black bar showed 50 % of the parasitemia pick at day 25 post-infection. The doses, in $\mu\text{moles/kg/day}$ for 14 days, are shown in parenthesis at the side of the compound code[33–35,44].



1: Preparation details of the compounds

Compounds Characterizations. All of the synthesized compounds were chemically characterized by thin layer chromatography (TLC), nuclear magnetic resonance (¹H NMR, ¹³C NMR), and elemental microanalyses (CHN). Alugram SIL G/UV254 (Layer: 0.2 mm) (Macherey-Nagel GmbH & Co. KG., Düren, Germany) was used for TLC, and silica gel 60 (0.040–0.063 mm, Merck) was used for flash column chromatography. The NMR spectra were recorded on a Bruker DPX 400 (400 MHz for ¹H and 100 MHz for ¹³C), using TMS as the internal standard and with the indicated deuterated solvent; the chemical shifts are reported in ppm (δ) and coupling constants (J) values are given in Hertz (Hz). Signal multiplicities are represented by s (singlet), d (doublet), dd (doublet), t (triplet), tt (triple triplet), and m (multiplet). Structural assignments were corroborated by HMBC and HSQC experiments. Mass spectrometry experiments were performed on a HEWLETT PACKARD MSD 5973 or a LC/MSD-Serie 100 using electronic impact (EI) or electrospray ionization (ESI), respectively. To determine the purity of the compounds, elemental microanalyses obtained on a Carlo Erba Model EA1108 elemental analyzer from vacuum-dried samples were used. The analytical results for C, H, and N were within ± 0.4 of the theoretical values. Melting points were recorded on ELECTROTHERMAL IA-9100 equipment, and they were not corrected. Samples of all product of this work are available from the authors.

Preparation of the compounds **793**, **1018** and **1019**



A mixture of the ketone reactant (2.2 mmol) and the corresponding aldehyde (4.5 mmol) were dissolved in 4.4 mL of water and 3.5 mL of ethanol in a 50 mL flask equipped with a magnetic stirrer. Then NaOH (11.2 mmol) was added and the reaction mixture was stirred for 24 h at room temperature. The reaction was monitored using TLC with silica as stationary phase and a mixture of hexane:ethyl acetate (7:3) as the mobile phase. The precipitated solid was filtered under vacuum, washed with water and recrystallized from ethanol.

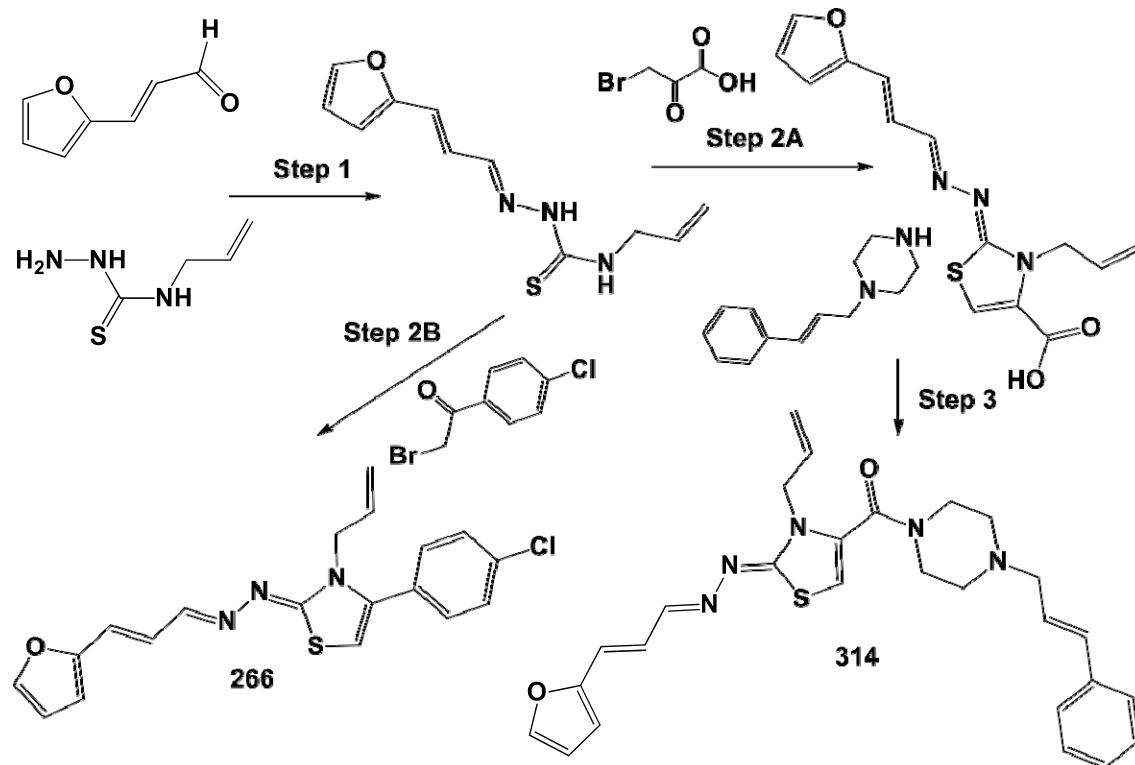
(*1E,3E,6E,8E*)-1,9-Di(furan-2-yl)nona-1,3,6,8-tetraen-5-one (**793**): orange solid, yield =98%, m.p.= 121-124 °C, ¹H-NMR ($CDCl_3$) δ (ppm): 7.46 (2H, d, $J=1.6$, H15,18), 7.42 (2H, d, $J=11.0$, H3,8), 6.87 (2H, d, $J=11.1$, H4,9), 6.75 (2H, d, $J=11.1$, H6,11), 6.55 (2H, d, $J=1.6$, H14,19), 6.52 (2H, d, $J=11.0$, H5,10), 6.47 (2H, d, $J=1.8$, H13,20). ¹³C-

NMR (CDCl_3) δ (ppm): 100 (C13,20), 113 (C14,19), 125 (C6,11), 128 (C3,8), 143 (C4,9), 153 (C15,18), 189 (C1). MS (EI) m/z (%): 266 (M^+ , 100); 185 ($\text{C}_1\text{H}_1\text{O}^+$ 18.49); 81 ($\text{C}_5\text{H}_4\text{O}^+$, 38.62). Elemental Analysis: C, 76.68; H, 5.30; O, 18.02

(*2E,6E*)-2,6-Bis[*(E*)-3-(furan-2-yl)allylydene]cyclohexanone (**1018**): brown solid, yield =60%, m.p.= 156-158 °C, ^1H -NMR (CDCl_3) δ (ppm): 7.65 (2H, d, $J=6.8$, H18,21), 7.35 (2H, s, H7,8), 7.05 (2H,d, $J=12.2$, H9,12), 6.75 (2H, d, $J=12.2$, H10,13), 6.55 (2H, m, H16,23), 6.41 (2H, m, H17,22), 2.79 (4H, m, H2,4), 1.88 (2H, m, H3). ^{13}C -NMR (CDCl_3) δ (ppm): 193 (C6), 149 (C11,14), 145 (C18,21), 142 (C1,5), 137 (C7,8), 130 (C9,12), 129 (C10,13), 113 (C17,22), 111 (C16,23), 28 (C2,4), 24 (C3). MS (EI) m/z (%): 306(M^+ , 100); 184($\text{C}_{13}\text{H}_{12}\text{O}_4^+$; 10). Elemental Analysis: C, 78.41; H, 5.92; O, 15.67.

(*2E,5E*)-2,5-Bis[*(E*)-3-(thien-2-yl)allylydene]cyclopentanone (**1019**): orange solid, yield =100%, m.p.= 168-170 °C, ^1H -NMR (CDCl_3) δ (ppm): 7.32 (2H, d, $J=5.0$, H17,20), 7.49 (2H, s, H6,7), 7.20 (2H, d, $J=3.5$, H15,22), 7.15 (2H, d, $J=4.5$, H16,21), 7.11 (2H, d, $J=15.3$, H8,10), 6.78 (2H, d, $J=15$, H9,11), 2.90 (2H, s, H2,3). ^{13}C -NMR (CDCl_3) δ (ppm): 198 (C5), 141 (C12,13), 140 (C1,4), 132 (C6,7), 130 (C9,11), 129 (C8,10), 128 (C17,20), 127 (C16,21), 126 (C15,22), 30 (C2,3). MS (EI) m/z (%): 324.06 (M^+ , 100). Elemental Analysis: C, 70.33; H, 4.97; O, 4.93; S, 19.77.

Preparation of **266** and **314**



Scheme S2. Synthetic steps for the preparation of **314** and **266**.

Step 1. A mixture of the corresponding aldehyde (1.05 equiv.), the corresponding thiosemicarbazide (1.00 equiv.), catalytic amount of *p*-toluenesulfonic acid, and dry

toluene (1 mL per 100 mg of aldehyde) was stirred at room temperature until disappearance of the aldehyde (12–24 h, checked by TLC, SiO₂, using as a mobile phase petroleum ether:ethyl acetate (70:30)). After that, the precipitate was filtered off and washed with petroleum ether. The solid was crystallized from ethanol.

Step 2. A mixture of the corresponding thiosemicarbazone (1.0 equiv.), the corresponding α -haloketone (1.2 equiv.), and ethanol 98% (1 mL per 100 mg of thiosemicarbazone) was heated at reflux until disappearance of the thiosemicarbazone (4–10 h, checked by TLC, SiO₂, and petroleum ether:ethyl acetate (70:30)). After that, the mixture was cooled at room temperature, and the precipitate was filtered off and washed with ethanol:water (80:20). The solid was crystallized from ethanol or ethanol:water.

1Z-[3-Allyl-4-(4-chlorophenyl)thiazol-2(3H)-ylidene]-2E-[3-(2-furyl)-2E-propenylidene] hydrazine hydrobromide hydrate, (**266**). Orange solid, 97 %; mp: 200 °C (d), ¹H NMR (CDCl₃) δ (ppm): 4.71 (2H s), 5.05 (2H d J17), 6.76 (1H d J16), 6.47 (m, 2H); 6.89 (1H dd J16/5.8), 7.36 (2H d 8), 7.44 (2H d 8), 7.46 (1H bs), 8.48 (1H bs), 8.25 (1H s). ¹³C NMR (CDCl₃): δ 48, 117, 103, 111, 112, 123, 126, 129, 130, 131, 136, 140, 144, 152, 154, 168. MS (EI) m/z (abundance, %): 369 (M⁺, 100), 235 (42); UV: λ_{max} : 379 nm, $\epsilon = 14.7 \pm 0.5 \text{ cm}^{-1} \text{ mM}^{-1}$. Found: C, 49.0; H, 3.9; N, 9.1; S, 6.9. C₁₉H₁₉BrClN₃O₂S Exact Mass: 467.01 C, 48.68; H, 4.09; Br, 17.04; Cl, 7.56; N, 8.96; O, 6.83; S, 6.84.

Step 3. A mixture of the corresponding acid (1.0 equiv.), thionyl chloride (1.2 equiv.) and dry toluene (1 mL per 100 mg of acid) was heated at 100 °C for 1 h. After that, a solution of the corresponding amine (1.0 equiv.) and triethylamine (5.0 equiv.) were added dropwise during 30 min to the reaction cooled at 0 °C. The mixture of reaction was stirred at room temperature until disappearance of the activated acid (12–24 h, checked by TLC, Al₂O₃, petroleum ether:ethyl acetate (70:30)). After that, the solvent was evaporated *in vacuo* and the residue was partitioned between methylene dichloride and saturated aqueous solution of sodium bicarbonate. The organic layer was washed with aqueous phosphate buffer (pH 4–5), dried with anhydrous sodium sulfate and evaporated *in vacuo*. The desired product was purified, from the residue of evaporation, by column chromatography (Al₂O₃, petroleum ether:ethyl acetate (0 to 40 %)).

(2E,2Z)-3-Allyl-4-[(*E*)-4-cinnamylpiperazin-1-yl)carbonyl]-2-[2-((*E*)-3-(furan-2-yl)propenylidene)hydrazono]-2,3-dihydrothiazole (**314**): yellow solid, yield =60 %, m.p.= 157–159 °C. ¹H NMR (CDCl₃) δ (ppm): 2.52 (bs, 4H), 3.2 (d, J= 6.6 Hz, 2H), 3.68 (bs, 4H), 4.66 (d, J= 5.6 Hz, 2H), 5.19 (m, 2H), 5.91 (m, 1H), 6.15 (s, 1H), 6.25 (m, 1H), 6.46 (m, 1H), 6.56 d, J=16Hz, 1H), 6.64 (d, J=16, 1H), 6.92 (dd, J=9.9/16, 1H), 7.38 (m, 4H), 7.44 (m, 1H), 8.07 (d, J=9.9, 1H). ¹³C NMR (CDCl₃): δ 142, 110, 132, 125, 152, 168, 104, 142, 170, 42, 47, 60, 124, 134, 128, 118. MS (EI) m/z (abundance, %): 487 (M⁺, 13), 407 (10), 117 (100). UV: 387 nm ($\epsilon = 27.2 \pm 0.4 \text{ cm}^{-1} \text{ mM}^{-1}$). Found: C, 66.6; H, 6.0; N, 14.1; S, 6.4.

Preparation of **1260**

Step 1. A mixture of pregnolone (1.05 equiv.), thiosemicarbazide (1.00 equiv.), catalytic amount of *p*-toluenesulfonic acid, and dry toluene (1 mL per 100 mg of ketone) was stirred at room temperature until disappearance of the aldehyde (12–24 h, checked by TLC, SiO₂, using as a mobile phase petroleum ether:ethyl acetate (70:30)). After that, the precipitate was filtered off and washed with petroleum ether. The solid was crystallized from ethanol.

1-(3β-Hydroxy-pregn-5-ene-20*E*-ylydene)thiosemicarbazide (1260**):** White powder, yield: 95%, white powder, m.p. 245–248 °C; ¹H RMN (CDCl₃) δ (ppm): 0.63 (s, 3H), 0.99 (s, 3H), 2.83 (t, 1H, J = 8.5 Hz), 3.52 (m, 1H), 5.35 (t, 1H, J = 2.5 Hz), 8.56 (s, 2H). ¹³C RMN (CDCl₃) δ (ppm): 13.2, 19.5, 21.2, 23.5, 25.2, 31.4, 32.6, 32.9, 37.2, 37.8, 39.5, 42.5, 45.5, 50.7, 57.5, 62.4, 71.2, 121.4, 165.4, 181.2 C₂₂H₃₅N₃OS. ESI-MS (m/z): 389.25 (100.0%), 390.25 (25.7%), 391.25 (10.0%), 392.25 (1.1%). Elemental analysis: C, 67.82; H, 9.05; N, 10.79; O, 4.11; S, 8.23.

2: Pan assay interference compound: Virtual check

Interference Compounds check. The active compounds (**793, 1019, 1018, 314** and **1260**) were tested online fallowing the recommendation on the Ecstasy and Agony of Assay Interference Compounds. The screenshot for the compounds is showed above, was made in four different on line software (<http://zinc15.docking.org/patterns/home>, <http://www.cbligand.org/PAINS/>, <https://fafdrugs4.rpbs.univ-paris-diderot.fr/links>, <http://advisor.docking.org>), for PAINS and aggregator compounds. Any of them show problem in those test. Also these compound were used in many other enzymatic assay, and were not active, demonstration their non-interference status.ii,iii,iv,v

Compound **793**:

Bulk Pattern Checker

Upload a list of compounds and receive a report on PAINS and/or aggregators

Paste SMILES

```
O=C1/C=C/C=C/C=C=O2\O(CCC1)=C/C=C/C3=CC=C=O3
O=C/C1=C/C=C/C2=C/C=C/S2\O(CC1)=C/C=C/C3=CC=C=S3
```

Upload a File

Choose File No file chosen

Check For

Check PAINS Check Aggregators

Results

Output Format View Only Check

Note! The Bulk Pattern Checker will only process the first 1000 lines of your file. To process more, please split it up into multiple uploads.

Only compounds matching PAINS or similar to known aggregators are shown. An empty result means, "No problems found".

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False Positive Remover

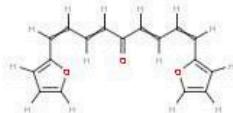
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- Protonated Accepted Molecules (pH7.4): [SDF]
- Intermediate Molecules: [SDF]
- Protonated Intermediate Molecules (pH7.4): [SDF]
- Rejected Molecules: [SDF]
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- Mixtures: *None*
- Empty Structures: *None*
- Isotopes and Inorganics: *None*
- Large Compounds: *None*
- PAINS: [SDF]
- Covalent Inhibitors: *None*
- Descriptors: [CSV]
- Groups: *None*
- PAINS: [CSV]

Filtering statistics

- Input molecules: 1
- Duplicates: 0
- Mixtures: 0
- Large Compounds: 0
- Isotopes and Inorganics: 0
- Empty Structures: 0
- Filtered molecules: 1
- Rejected molecules: 0
- Accepted molecules: 1
- Intermediate molecules: 0
- PAINS (Pan Assays Interferences Compounds): 0
- Covalent Inhibitors: 0

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Library of compounds computed descriptors:

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<p>M</p> <p>R R</p> <p>ot i</p> <p>at g</p> <p>l 1</p> <p>1 1</p> <p>IMoo</p> <p>Wgg</p> <p>PD S</p> <p>AB o</p> <p>w o n</p> <p>ds s</p>	<p>T C H</p> <p>a o e a e</p> <p>H x t r t</p> <p>B R C a a b e</p> <p>B D A S h 1 v</p> <p>B ib n i i a</p> <p>ep H z r A</p> <p>o cc n o</p> <p>n n</p> <p>to B s e g h</p> <p>rs A Re a o t</p> <p>o rs A i r m o</p> <p>g</p>	<p>Li S</p> <p>R pi ol</p> <p>a ns u</p> <p>bi</p> <p>Solu</p> <p>bility</p> <p>Forec</p> <p>astIn</p> <p>Hla (</p> <p>t / ti m</p> <p>C o g/</p> <p>mm</p> <p>ns l)</p>	<p>Or Or T</p> <p>al al r</p> <p>Bi Bi a</p> <p>oa oa f</p> <p>vai vai fi</p> <p>sph s reo</p> <p>olip s Ce</p> <p>Fri a</p> <p>ido p nte</p> <p>sis 3 dl u</p> <p>rs y s</p>
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<p>^{1v1}</p> <p>R R</p> <p>ot i</p> <p>at g</p> <p>l 1</p> <p>1 1</p> <p>IMoo</p> <p>Wgg</p> <p>PD S</p> <p>AB o</p> <p>w o n</p> <p>ds s</p>	<p>T C H</p> <p>a o e a e</p> <p>H x t r t</p> <p>B R C a a b e</p> <p>B D A S h 1 v</p> <p>B ib n i i a</p> <p>ep H z r A</p> <p>o cc n o</p> <p>n n</p> <p>to B s e g h</p> <p>rs A Re a o t</p> <p>o rs A i r m o</p> <p>g</p>	<p>Li S</p> <p>R pi ol</p> <p>a ns u</p> <p>bi</p> <p>Solu</p> <p>bility</p> <p>Forec</p> <p>astIn</p> <p>Hla (</p> <p>t / ti m</p> <p>C o g/</p> <p>mm</p> <p>ns l)</p>	<p>Or Or T</p> <p>al al r</p> <p>Bi Bi a</p> <p>oa oa ¹ 4</p> <p>vai vai fi</p> <p>sph s reo</p> <p>olip Ce</p> <p>Fri a</p> <p>ido p nte</p> <p>sis 3 dl u</p> <p>rs y s</p>
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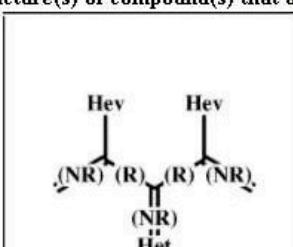
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inf_psa sup_psa inf_logp sup_logp
inf_cf sup_cf inf_sc sup_sc
inf_c sup_c inf_h sup_h
inf_ch sup_ch

Compound 1018:

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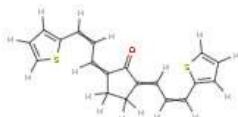


163:ene_one_ene_A(57)

1 Compound(s) have the above features:

1 Compound(s) have the above features:

C=[!R]C(Hev)-[R]C(=[!R]Het)-[R]C(=[!R]C)Hev



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1

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- Accepted Molecules: [\[SDF\]](#)
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- Intermediate Molecules: [\[SDF\]](#)
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- Rejected Molecules: [\[SDF\]](#)
- Protonated Rejected Molecules: [\[SDF\]](#)
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- Large Compounds: *None*
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- Covalent Inhibitors: *None*
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Filtering statistics

- Input molecules: 1
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- Mixtures: 0
- Large Compounds: 0
- Isotopes and Inorganics: 0
- Empty Structures: 0
- Filtered molecules: 1
- Rejected molecules: 0
- Accepted molecules: 0
- Intermediate molecules: 1
- PAINS (Pan Assays Interferences Compounds): 1
- Covalent Inhibitors: 0

Main PhysChem Descriptors Analysis

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Library of compounds computed descriptors:

<p>M H C H</p> <p>R T H C H</p> <p>Ro R H a T H C H</p> <p>ta ig B H x o ear e Li So</p> <p>I I t ta i Fl D B H R S C t a b R pi</p> <p>Mo o P bl d ex Ac B i i h al vo e a ns lu</p> <p>Dw g S e B ibi o ce D n z a C y n r ti ki</p> <p>Dg g S e B ibi o ce D n z a C A A o o Vi</p> <p>P D S A Bo o lit pt H g e rg h (ecastl</p> <p>w nd n y or or BA s R e a t t A H ol</p> <p>s d s s s i rg m m o C on g/</p> <p>s s s s n e ss m m)</p> <p>g s s g s s s</p>	<p>Ora Ora T</p> <p>I I r</p> <p>Bio Bio a 4</p> <p>ava ava ff -</p> <p>ilab ilab ic 4</p> <p>ber an h</p> <p>) ts</p>
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Loading data from server

<p>M H</p> <p>R T H C H</p> <p>Ro R H a T H C H</p> <p>ta ig B H x o ear e Li So</p> <p>I I t ta i Fl D B H R S C t a b R pi</p> <p>Mo o P bl d ex Ac B i i h al vo e a ns lu</p> <p>Dw g S e B ibi o ce D n z a C y n r ti ki</p> <p>Dg g S e B ibi o ce D n z a C A A o o Vi</p> <p>P D S A Bo o lit pt H g e rg h (ecastl</p> <p>w nd n y or or BA s R e a t t A H ol</p> <p>s d s s s i rg m m o C on g/</p> <p>s s s s n e ss m m)</p> <p>g s s g s s s</p>	<p>Ora Ora T</p> <p>I I r</p> <p>Bio Bio a 4</p> <p>ava ava ff -</p> <p>ilab ilab ic 4</p> <p>ber an h</p> <p>) ts</p>
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Filtering input parameters:

Your input parameters correspond to the **nophyschem** parameters (see the [documentation](#)).

Click on a parameter to get related informations in the official FAF-Drugs4 documentation.

[max_lipinski](#) [max_ring](#) [inf_r](#) [sup_r](#)

[inf_mw](#) [sup_mw](#) [inf_hb](#) [sup_hb](#)

[inf_hbd](#) [sup_hbd](#) [inf_hba](#) [sup_hba](#)

[inf_hva](#) [sup_hva](#) [inf_nc](#) [sup_nc](#)

[inf_nbb](#) [sup_nbb](#) [inf_nrb](#) [sup_nrb](#)

[inf_psa](#) [sup_psa](#) [inf_logp](#) [sup_logp](#)

[inf_cf](#) [sup_cf](#) [inf_sc](#) [sup_sc](#)

[inf_c](#) [sup_c](#) [inf_h](#) [sup_h](#)

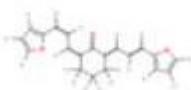
[inf_ch](#) [sup_ch](#)

Compound 1019:

 **False Positive Remover**

Welcome Created 12-2009
[Main Page](#) >> [Retrieve Structure](#) [Return](#)

Totally 1 Compound(s) were filtered out by the filter. [Download \(Smiles format\)](#) [View Detailed](#)
The following are the structure(s) of compound(s) that are filtered out by the filter


[Return](#)

1

FAFDugs4 Results

Downloads

- Accepted Molecules: [\[SDF\]](#)
- Protonated Accepted Molecules (pH7.4): [\[SDF\]](#)
- Intermediate Molecules: [\[SDF\]](#)
- Protonated Intermediate Molecules (pH7.4): [\[SDF\]](#)
- Rejected Molecules: [\[SDF\]](#)
- Protonated Rejected Molecules: [\[SDF\]](#)
- Duplicates: *None*
- Mixtures: *None*
- Empty Structures: *None*
- Isotopes and Inorganics: *None*
- Large Compounds: *None*
- PAINS: [\[SDF\]](#)
- Covalent Inhibitors: *None*
- Descriptors: [\[CSV\]](#)
- Groups: *None*
- PAINS: [\[CSV\]](#)

Filtering statistics

- Input molecules: 1
- Duplicates: 0
- Mixtures: 0
- Large Compounds: 0
- Isotopes and Inorganics: 0
- Empty Structures: 0
- Filtered molecules: 1
- Rejected molecules: 0
- Accepted molecules: 0
- Intermediate molecules: 1
- PAINS (Pan Assays Interferences Compounds): 1
- Covalent Inhibitors: 0

Main PhysChem Descriptors Analysis

Click on a plot to enlarge the picture

Your bank contains less than 30 molecules, thus histograms were not generated...

Library of compounds computed descriptors:

Loading data from server

Filtering input parameters:

Your input parameters correspond to the **nophyschem** parameters (see the [documentation](#)).

Click on a parameter to get related informations in the official FAF-Drugs4 documentation.

<u>max_lipinski</u>	<u>max_ring</u>	<u>inf_r</u>	<u>sup_r</u>
<u>inf_mw</u>	<u>sup_mw</u>	<u>inf_hb</u>	<u>sup_hb</u>
<u>inf_hbd</u>	<u>sup_hbd</u>	<u>inf_hba</u>	<u>sup_hba</u>
<u>inf_hva</u>	<u>sup_hva</u>	<u>inf_nc</u>	<u>sup_nc</u>
<u>inf_nbb</u>	<u>sup_nbb</u>	<u>inf_nrb</u>	<u>sup_nr</u>
<u>inf_psa</u>	<u>sup_psa</u>	<u>inf_logp</u>	<u>sup_logp</u>
<u>inf_cf</u>	<u>sup_cf</u>	<u>inf_sc</u>	<u>sup_sc</u>

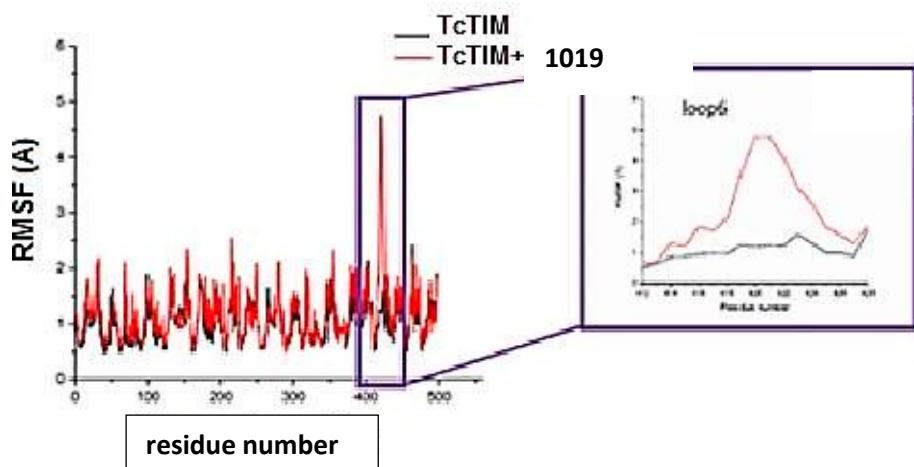
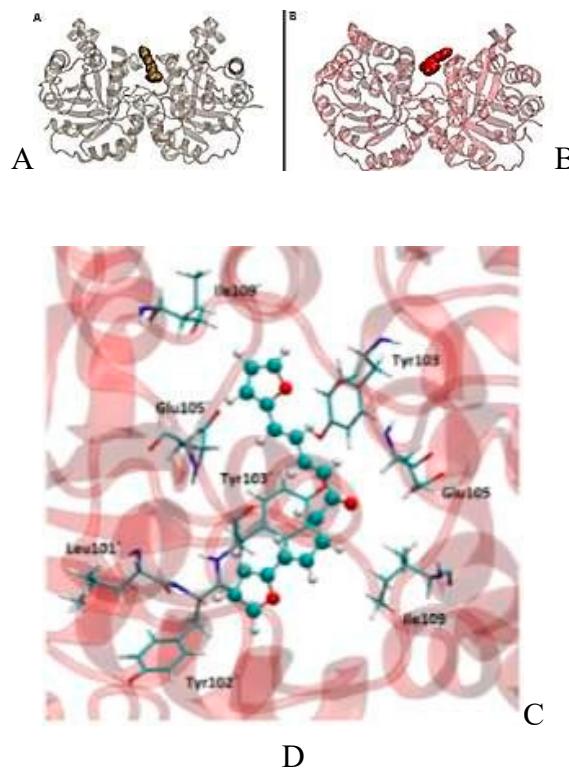
inf_c sup_c inf_h sup_h

inf_ch sup_ch

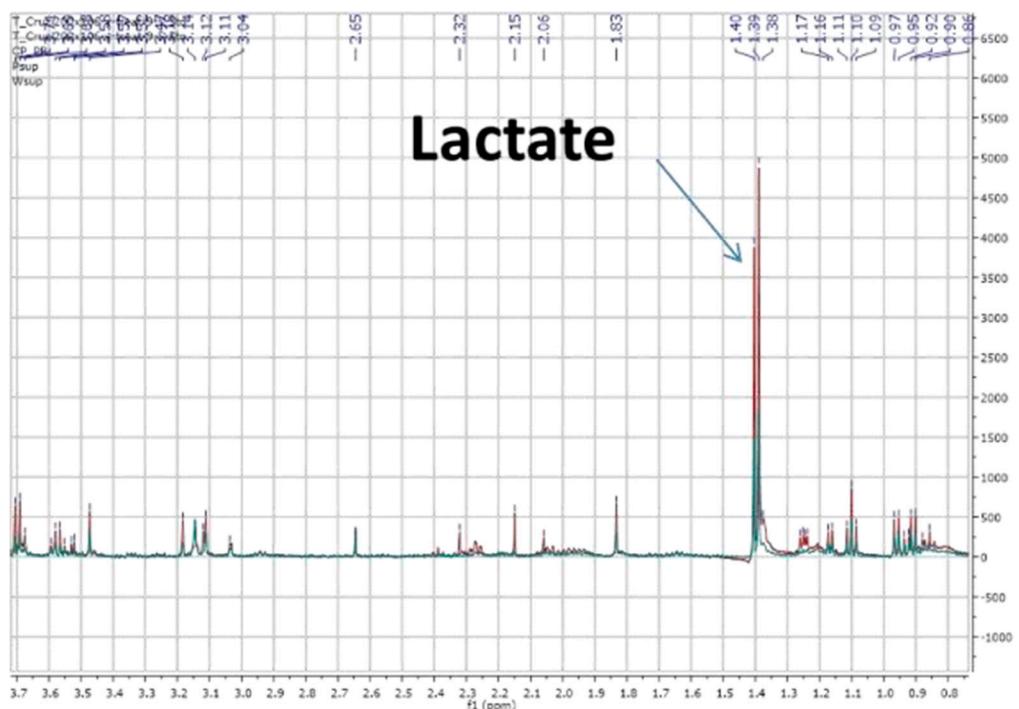
Table S2. Ames test using *Salmonella thyphimunum* strain TA98, TA100, TA102, TA1535, TA1537 in the absence of metabolic activation for compound 793, 1018 and 1019.

Compound	Doses (μg/plate)	TA98	TA100	TA102	TA1535	TA1537
793	0	6±1	109±5	204±8	11±3	31±7
	500	5±1	69±1	222±6	20±1	30 ± 3
	166	3 ± 1	58±3	203±2	18±1	28± 3
	55	3±1	74±6	194±3	14± 3	24± 4
	18	9±3	76±5	176±1	12±1	18±1
	6	5±2	89±3	171±1	8±1	16 ± 1
1018	0	6±1	60±5	204±8	11±3	31±7
	800	7±2	125±4	243±1	22±1	40.5±1.5
	266	6±3	113±3	237±1	16±1	36±1
	88	11±2	101±3	231±2	9±1	32±1
	29	25±1	90±3	222 ±2	8±1	28 ±2
	9	4±1	80±1	216±1	3±1	25±3
1019	0	15±2	109±5	204±8	11±3	31±7
	400	24 ± 3	81 ± 3	230±2	15±1	60 ± 2
	133	20 ± 1	74±6	217±3	17±1	51± 3
	44	19 ± 2	62±11	207±2	14±1	44 ± 1
	14	27 ± 7	79±2	201±2	14± 2	36± 3
	4	15 ± 1	60±5	187±2	9± 1	29± 1

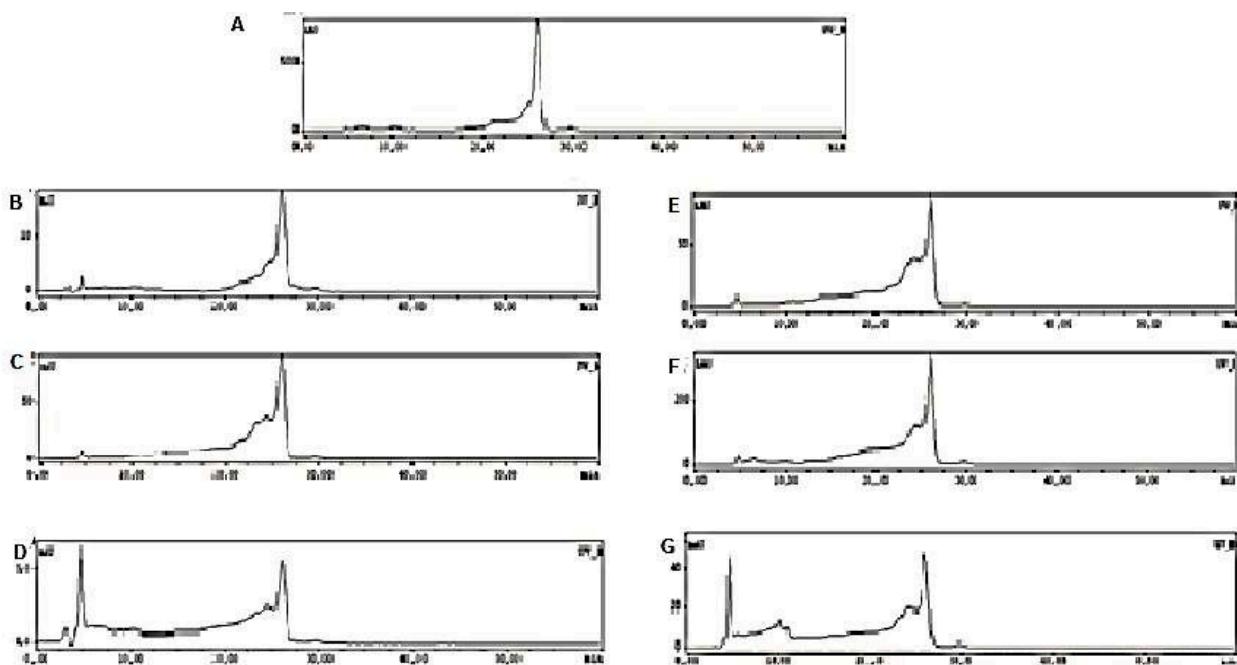
Supporting material 3. Docking and molecular dynamic figures with structural secondary details. Molecular dynamics and docking studies. Site of interaction of compound 793 in TcTIM after molecular docking (A) and after molecular dynamics (B). C) Compound 793 interaction site at the TcTIM interface. In sky-blue are represented the carbons; hydrogens in blank; in blue the nitrogen and in red the oxygen. D) RMSF of the residues according to the Ca. The greatest shift is observed in the loop6 of monomer B for compound 1019.



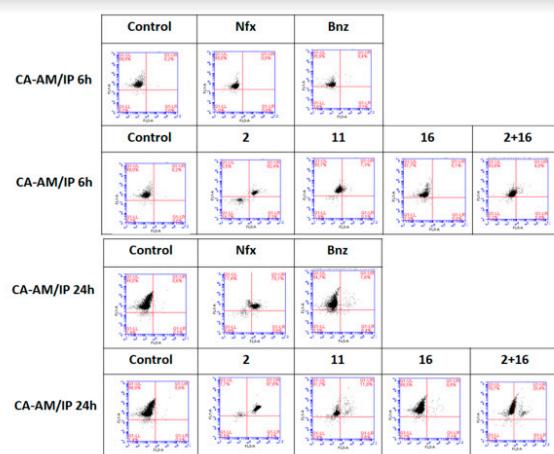
Supporting material 4. ^1H NMR spectrum from the metabolomics analysis showing the lactate accumulation. This spectrum correspond to the internal metabolites under **1019** treatment. In red are the spectrum for the condition with inhibition (with **1019**) and in blue the control parasites, (10 spectrum overlapping).



Supporting material 5. Chromatograms from the metabolism analysis. Study of metabolism of compound **793** by HPLC. A) **793** (solution in DMSO at a concentration of 0.4 mM) measured at 440 nm, using as mobile phase water and trifluoroacetic acid (0.05%):acetonitrile (50:50). B) FC + **793** at 0.4 mM (incubation time: 0 min). C) FC + **793** at 0.4 mM (incubation time: 30 minutes). D) FC + **793** at 0.4 mM (incubation time 60 min hour). E) FM + **793** at 0.4 mM (incubation time 0 min). F) FM + **793** at 0.4 mM (incubation time 30 minutes). G) FM + **793** at 0.4 mM (incubation time 60 min).



Supporting material 6. Analysis of the viability and mechanism of death by flow cytometry in parasites at 6 and 24 h of incubation at a 20 × IC₅₀ concentration of Nfx, Bnz, 793 (2), 1019 (11), 1018 (16) and 793+1018 (2+16) A) CA-AM/IP labeling. B) AV/IP labeling



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