

**Introduction of a SiFA Moiety into the D-Glutamate Chain of DOTA-
PP-F11N Results in Radiohybrid-Based CCK-2R-Targeted Compounds
with Improved Pharmacokinetics *in vivo***

- Supplementary Materials -

Nadine Holzleitner^{1,*,} Thomas Günther^{1,#}, Roswitha Beck¹, Constantin Lapa² and Hans-Jürgen Wester¹

¹Chair of Pharmaceutical Radiochemistry, Department of Chemistry, Technical University of Munich, Garching, Germany

²Clinic for Nuclear Medicine, University Hospital Augsburg, Augsburg, Germany

Corresponding co-authors:

Nadine Holzleitner and Thomas Günther

Phone: +49.89.289.12203

Technical University of Munich,

Chair of Pharmaceutical Radiochemistry,

Walther-Meissner-Str. 3

85748 Garching

GERMANY

Fax: +49.89.289.12204

E-Mail: nadine.holzleitner@tum.de and thomas.guenther@tum.de

ORCID: <https://orcid.org/0000-0001-8258-3526> (NH) and <https://orcid.org/0000-0002-7412-0297> (TG)

Analytical data of $^{nat/177}\text{Lu}$ -labeled minigastrin analogues

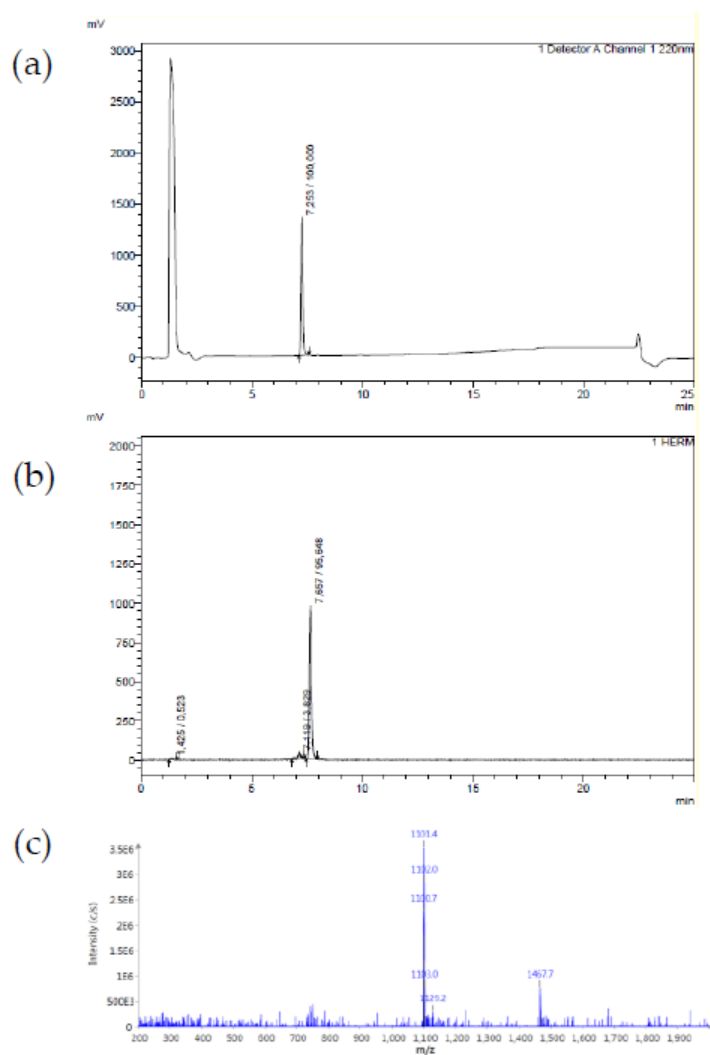


Figure S1. Confirmation of peptide identity and integrity for (a) ^{nat}Lu]-Lu-DOTA-PP-F11N and (b) ^{177}Lu]-Lu-DOTA-PP-F11N as analysed by analytical (radio-)RP-HPLC (MultoKrom 100-5 C18, 5 μm , 125 \times 4.6 mm, CS Chromatographie GmbH, Langerwehe, Germany; 10 \rightarrow 90% MeCN in H_2O + 0.1% TFA in 15 min). (c) Mass spectrum of ^{nat}Lu]-Lu-DOTA-PP-F11N.

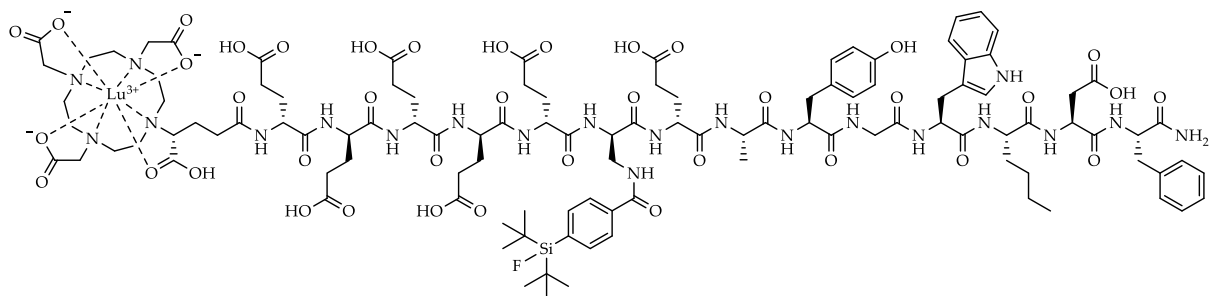
[^{nat}Lu]Lu-DOTA-PP-F11N: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 7.55 min, K' = 3.48; MS (ESI, positive): m/z calculated for C₉₀H₁₂₀LuN₁₉O₃₅: 2203.0, found: m/z = 1101.4 [M+2H]²⁺.

[^{nat}Lu]Lu-DOTA-PP-γ-F11N: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 7.41 min, K' = 3.39; MS (ESI, positive): m/z calculated for C₉₀H₁₂₀LuN₁₉O₃₅: 2203.0, found: m/z = 1101.9 [M+2H]²⁺.

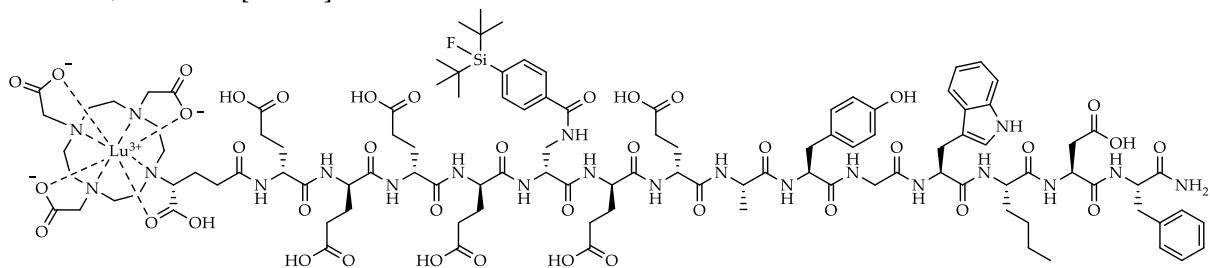
[^{nat}Lu]Lu-CP04: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 7.60 min, K' = 3.51; MS (ESI, positive): m/z calculated for C₈₉H₁₁₈LuN₁₉O₃₅S: 2219.7, found: m/z = 1110.3 [M+2H]²⁺.

[^{nat}Lu]Lu-γ-CP04: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 7.40 min, K' = 3.39; MS (ESI, positive): m/z calculated for C₈₉H₁₁₈LuN₁₉O₃₅S: 2219.7, found: m/z = 1110.8 [M+2H]²⁺.

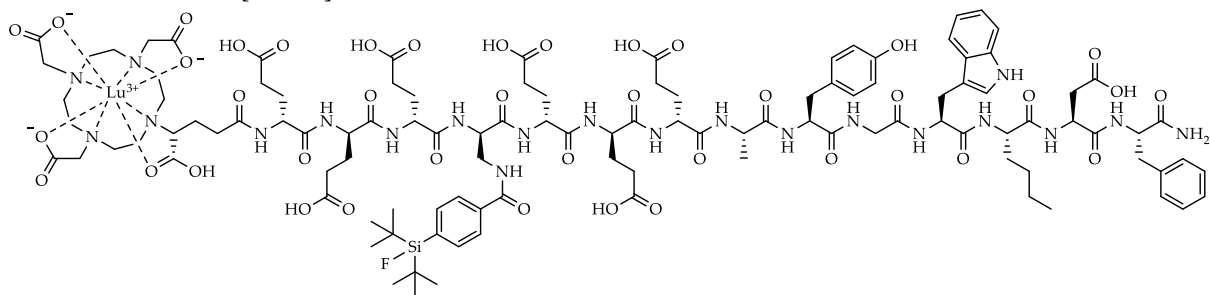
[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-1: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 10.1 min, K' = 4.99; MS (ESI, positive): m/z calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: m/z = 1312.8 [M+2H]²⁺.



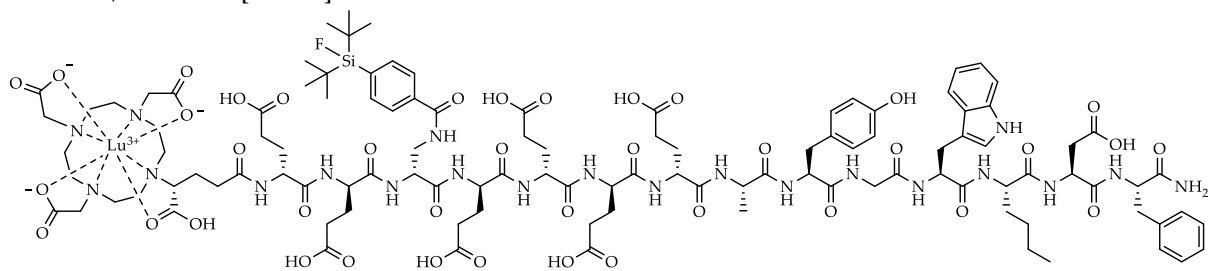
[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-2: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 9.84 min, K' = 4.83; MS (ESI, positive): m/z calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: m/z = 1312.7 [M+2H]²⁺.



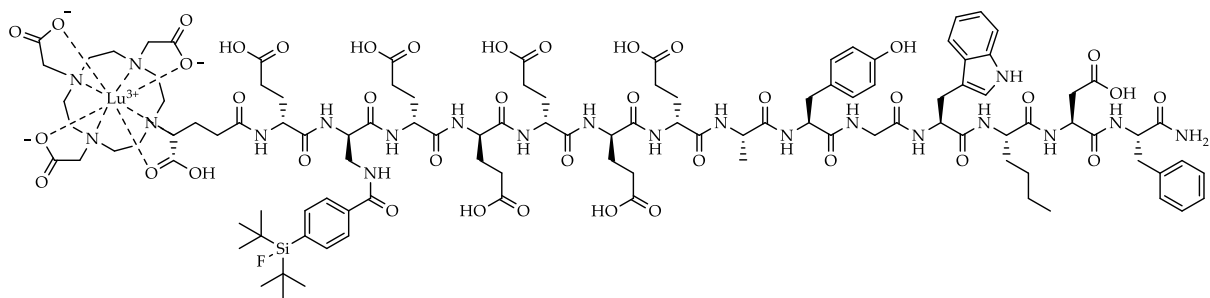
[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-3: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 9.85 min, K' = 4.84; MS (ESI, positive): m/z calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: m/z = 1312.3 [M+2H]²⁺.



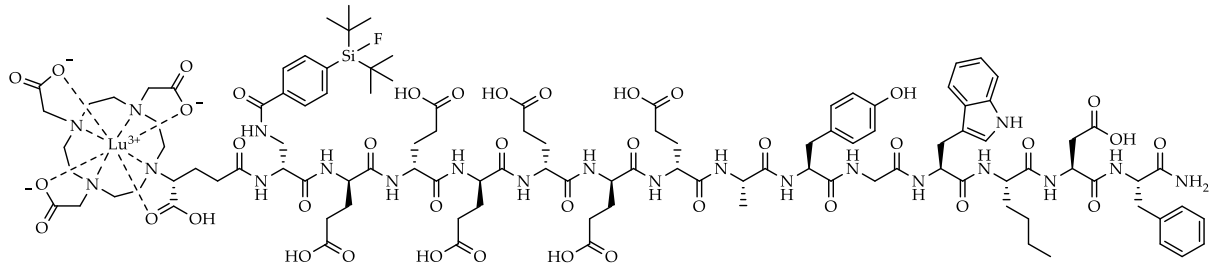
[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-4: RP-HPLC (10→70% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 11.9 min, K' = 6.05; MS (ESI, positive): m/z calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: m/z = 1313.0 [M+2H]²⁺.



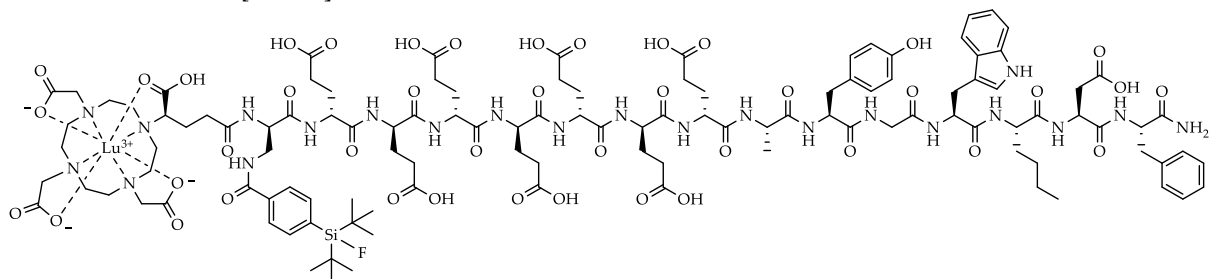
[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-5: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 9.67 min, K' = 4.73; MS (ESI, positive): m/z calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: m/z = 1313.3 [M+2H]²⁺.



[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-6: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): *t_R* = 9.71 min, *K'* = 4.76; MS (ESI, positive): *m/z* calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: *m/z* = 1312.9 [M+2H]²⁺.



[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-7: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): *t_R* = 9.62 min, *K'* = 4.70; MS (ESI, positive): *m/z* calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: *m/z* = 1313.0 [M+2H]²⁺.



[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-8: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): *t_R* = 9.77 min, *K'* = 4.79; MS (ESI, positive): *m/z* calculated for C₁₁₆H₁₅₈FLuN₂₂O₄₂Si: 2754.7, found: *m/z* = 1378.2 [M+2H]²⁺.

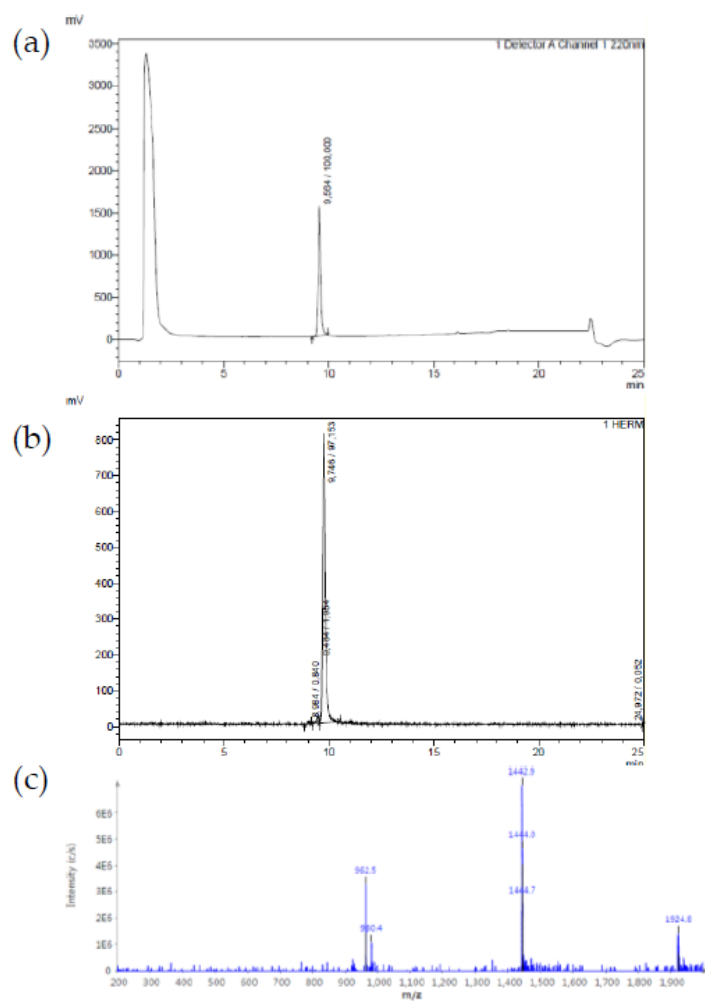
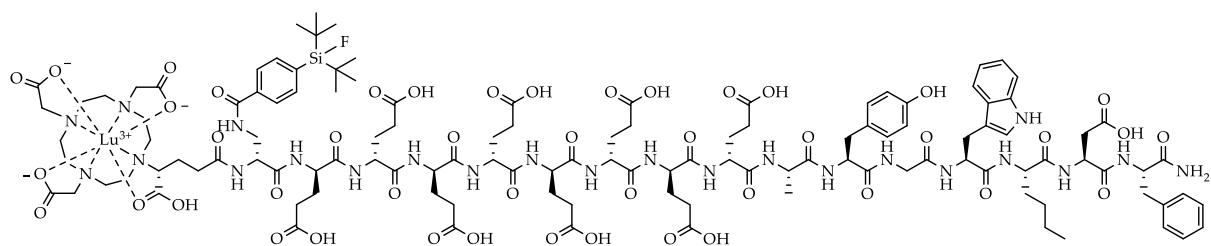
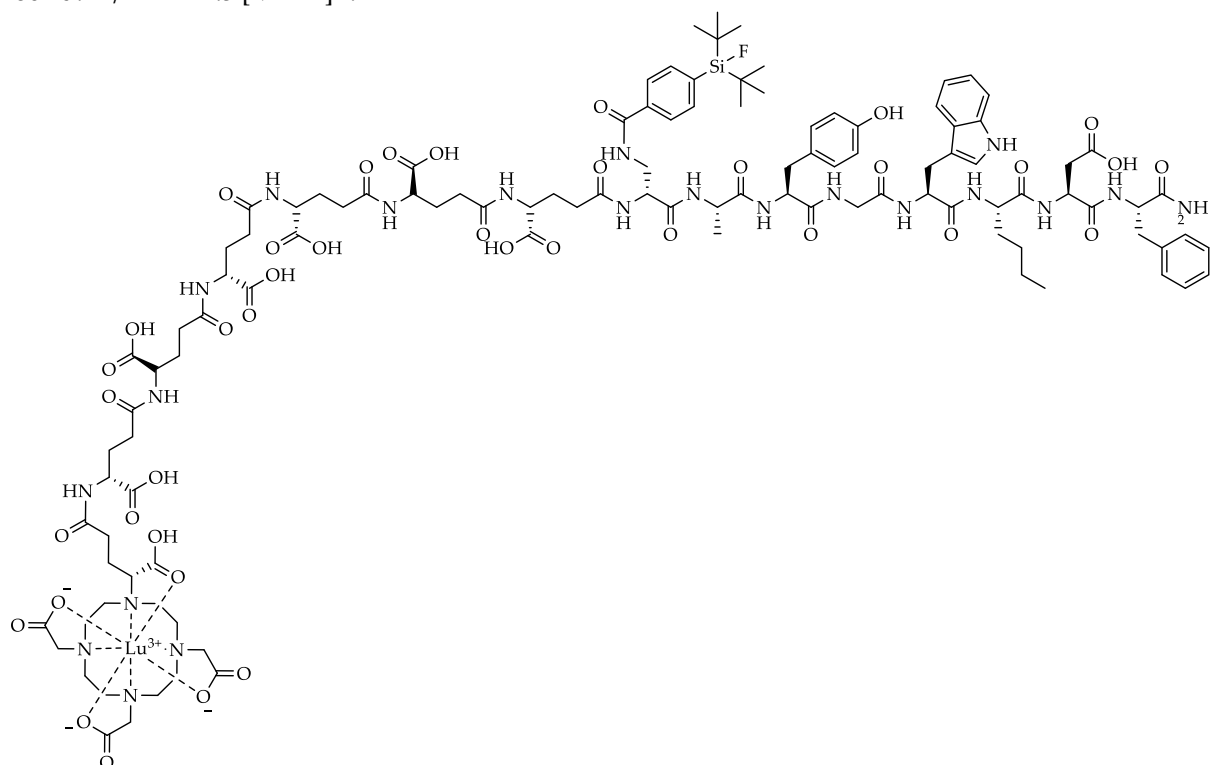


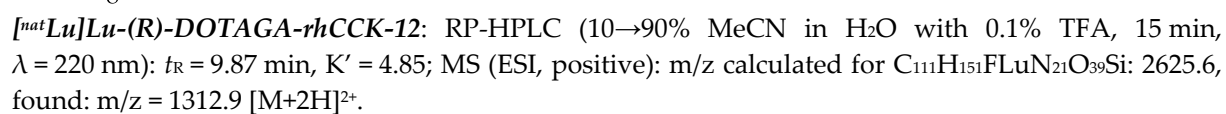
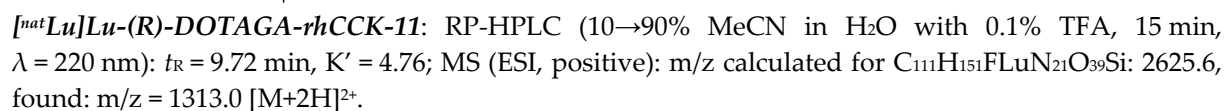
Figure S2. Confirmation of peptide identity and integrity for (a) $[\text{natLu}]$ Lu-(R)-DOTAGA-rhCCK-9 and (b) $[^{177}\text{Lu}]$ Lu-(R)-DOTAGA-rhCCK-9 as analysed by analytical (radio-)RP-HPLC (MultoKrom 100-5 C18, 5 μm , 125 \times 4.6 mm, CS Chromatographie GmbH, Langerwehe, Germany; 10 \rightarrow 90% MeCN in H_2O + 0.1% TFA in 15 min). (c) Mass spectrum of $[\text{natLu}]$ Lu-(R)-DOTAGA-rhCCK-9.

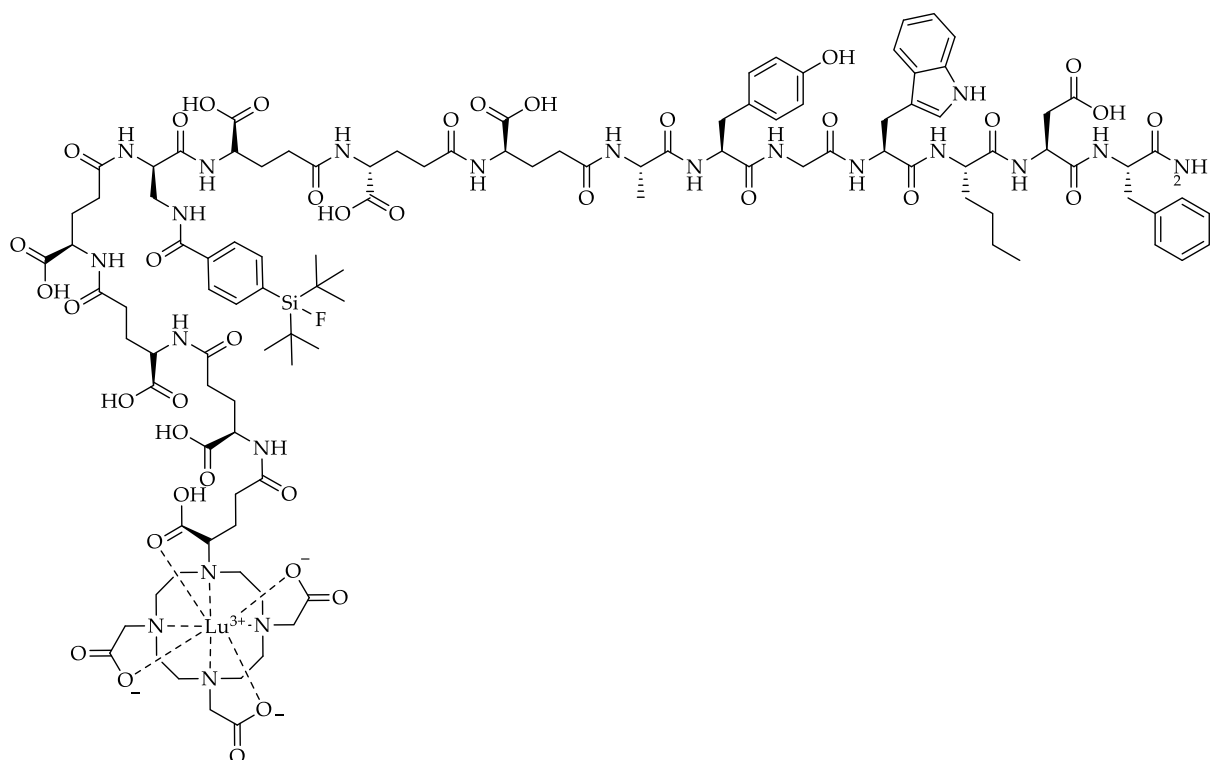


[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-9: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 9.56 min, K' = 4.67; MS (ESI, positive): m/z calculated for C₁₂₁H₁₆₅FLuN₂₃O₄₅Si: 2883.8, found: m/z = 1441.5 [M+2H]²⁺.

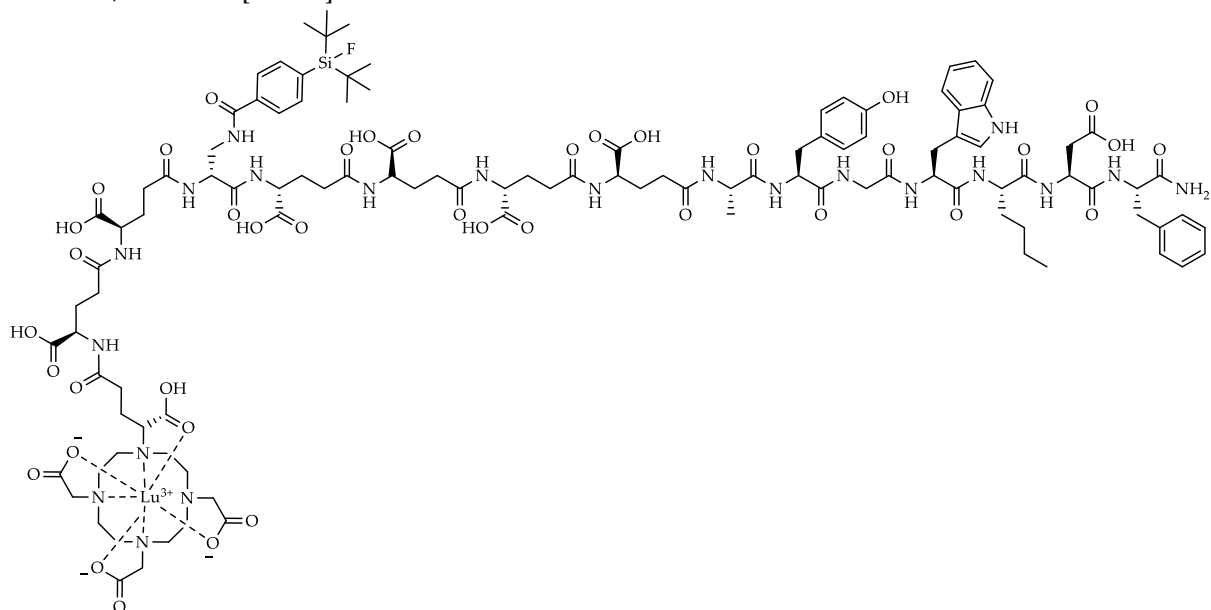


[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-10: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 10.2 min, K' = 5.05; MS (ESI, positive): m/z calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: m/z = 1313.6 [M+2H]²⁺.

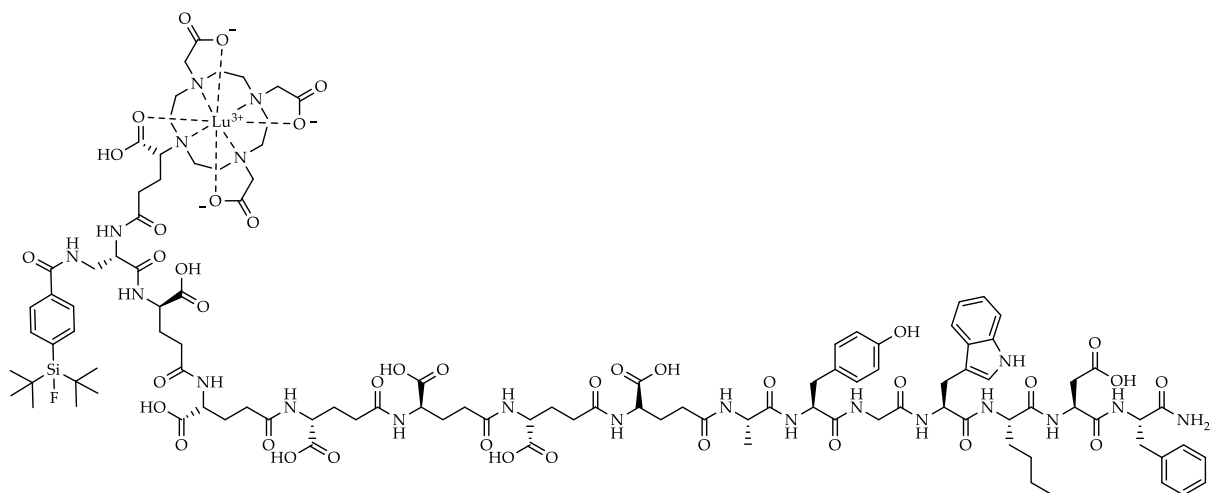




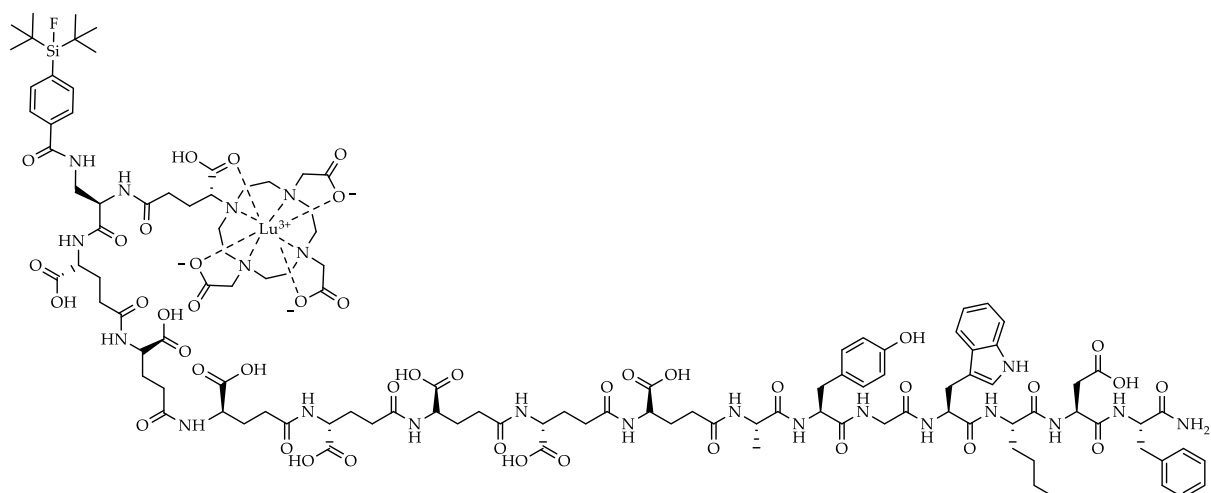
[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-13: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm); t_R = 9.61 min, K' = 4.70; MS (ESI, positive): m/z calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: m/z = 1313.2 [M+2H]²⁺.



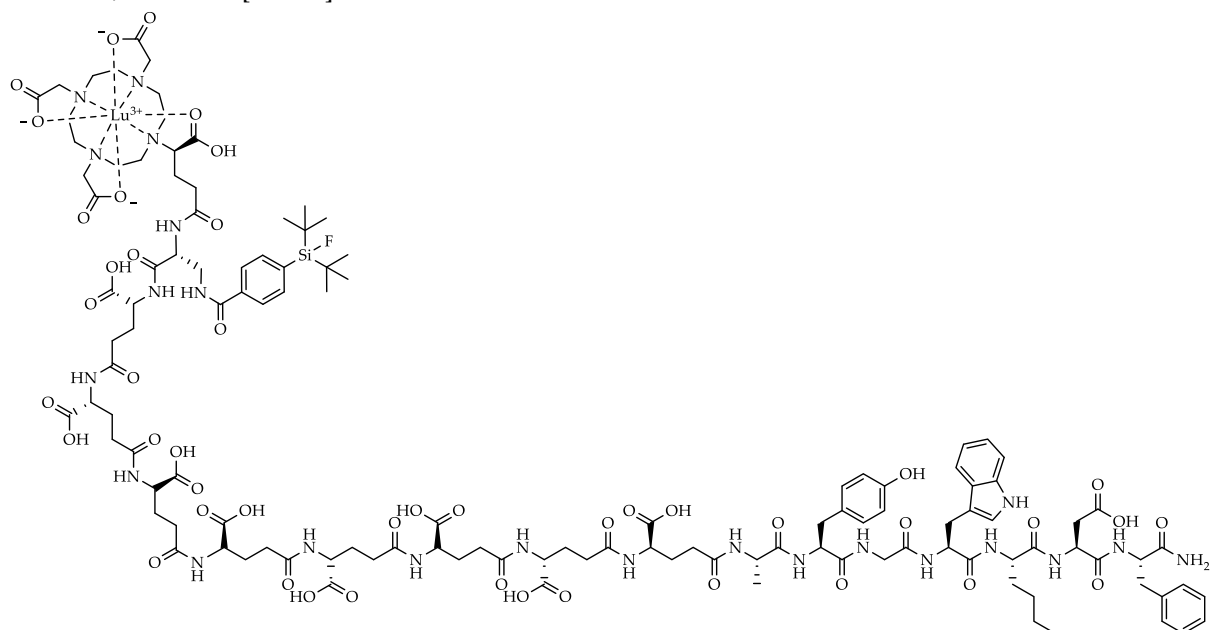
[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-14: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm); t_R = 9.60 min, K' = 4.69; MS (ESI, positive): m/z calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: m/z = 1313.9 [M+2H]²⁺.



[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-16: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 9.52 min, K' = 4.64; MS (ESI, positive): m/z calculated for C₁₁₁H₁₅₁FLuN₂₁O₃₉Si: 2625.6, found: m/z = 1314.4 [M+2H]²⁺.



[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-17: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): t_R = 9.48 min, K' = 4.62; MS (ESI, positive): m/z calculated for C₁₁₆H₁₅₈FLuN₂₂O₄₂Si: 2754.7, found: m/z = 1377.7 [M+2H]²⁺.



[^{nat}Lu]Lu-(R)-DOTAGA-rhCCK-18: RP-HPLC (10→90% MeCN in H₂O with 0.1% TFA, 15 min, λ = 220 nm): *t_R* = 9.50 min, *K'* = 4.63; MS (ESI, positive): *m/z* calculated for C₁₂₁H₁₆₅FLuN₂₃O₄₅Si: 2883.8, found: *m/z* = 1443.1 [M+2H]²⁺.

¹⁷⁷Lu-labelling

For ¹⁷⁷Lu-labelling experiments, [¹⁷⁷Lu]LuCl₃ dissolved in hydrochloric acid (0.04 M, 40 GBq/mL) was acquired from *ITM Isotope Technologies Munich SE* (Garching, Germany). Radiolabelling of the peptide precursor (1 nmol) was performed at 90 °C for 15 min in a NaOAc-buffered (1 M, pH = 5.5) hydrochloric acid (0.04 M) solution. After radiolabelling, a radiolysis quencher (sodium ascorbate, 1 M in H₂O) was added and radiochemical purity was determined via radio-RP-HPLC and radio-TLC (instant thin layer chromatography paper impregnated with silica gel (iTLC-SG, *Agilent Technologies Inc.*, Folsom, United States); sodium citrate*1.5 H₂O (0.1 M)).

In Vitro Experiments

Cell Culture. CCK-2R expressing rat pancreatic cancer cells AR42J (CLS GmbH, Eppelheim, Germany) were cultivated in monolayers in CELLSTAR® cell culture flasks acquired from Greiner Bio-One GmbH (Frickenhausen, Germany) at 37 °C in a humidified atmosphere (5% CO₂) using a HERAcell 150i-Incubator (Thermo Fisher Scientific Inc., Waltham, United States). As nutrient medium RPMI 1640 medium, supplemented with 5 mM L-Gln 5 mL non-essential amino acids (100×) and 10% FCS, was used. Furthermore, a Dulbecco's PBS solution with 0.1% EDTA (*v/v*) was applied to detach the cells for cell passaging. The detached cells were counted using a Neubauer hemocytometer (Paul Marienfeld, Lauda-Königshofen, Germany). In addition, all operations under sterile conditions were accomplished using a MSC-Advantage safety workbench (Thermo Fisher Scientific Inc., Waltham, United States).

Determination of IC₅₀. AR42J cells (2.0×10^5 cells/well) were seeded into 24-well plates 24 ± 2 h prior to testing, using 1 mL of nutrient medium (RPMI 1640, 5 mM L-Gln, 5 mL non-essential amino acids (100×), 10% FCS). Cells were incubated at 37 °C in a humidified atmosphere (5% CO₂).

After removal of the medium, each well was washed with 500 µL PBS. For the cell-based assay, 200 µL of nutrient medium (+5% BSA), [¹⁷⁷Lu]Lu-DOTA-PP-F11N (25 µL, 0.3 pmol) as a radiolabeled reference and 25 µL of the peptide of interest in increasing concentrations (10^{-10} to 10^{-4} M) in triplicate were added to the cells. Thereafter, the assay was incubated for 3 h at 37 °C and thereafter, the supernatant was collected. The cells were washed with 300 µL PBS and the collected supernatant fractions were unified. After lysis of the cells with NaOH (300 µL, 1 N) for 15 min, the respective wells were washed with NaOH (300 µL, 1 N) and both fractions were unified. The radioactivity of both, the supernatant and the lysed fractions was quantified using a γ-counter (PerkinElmer Inc., Waltham, United States). The obtained data were evaluated *via* the GraphPad PRISM software (GraphPad Software Inc., La Jolla, United States), which calculates the halfmaximal inhibitory concentration (IC₅₀) of the peptides.

Internalisation Studies. For the determination of the internalisation kinetics of the various peptides, AR42J cells (3.0×10^5 cells/well) were seeded into polylysine coated 24-well plates adding 1 mL of nutrient medium. Afterwards, the cells were incubated for 24 ± 2 h at 37 °C in a humidified atmosphere (5% CO₂).

On the day of the experiment, the medium was removed, and each well was washed with nutrient medium (300 µL). Afterwards 200 µL of nutrient medium, 25 µL of the ¹⁷⁷Lu-labeled peptide (0.3 pmol, *n* = 6) and either 25 µL of nutrient medium for internalisation studies (*n* = 3) or 25 µL of [^{nat}Lu]Lu-DOTA-PP-F11N (10 µmol) for competition studies (*n* = 3) were added. Thereafter, the assay was incubated for various time points (1, 2, 4 and 6 h) at 37 °C in a humidified atmosphere (5% CO₂). After incubation, the cells were put on ice for at least 1 min to stop internalisation kinetics and the supernatant was collected. Then, the cells were washed with an ice-cold nutrient medium (300 µL) and both fractions were unified. For the acid wash, 300 µL of an ice-cold glycine buffer (1 M, pH 1) was added. After lysis of the cells with NaOH (300 µL, 1 N) for 15 min, the respective wells were washed with NaOH (300 µL, 1 N) and both fractions were unified. The radioactivity of the supernatant, the acid wash and the lysed fractions were quantified using a γ-counter.

Table S1. Affinity and lipophilicity data of the compounds evaluated. Affinity data were determined on AR42J cells (2.0×10^5 cells/well) and [^{177}Lu]Lu-DOTA-PP-F11N (0.3 pmol/well) as radiolabeled reference (3 h, 37 °C, RPMI 1640, 5 mM L-Gln, 5 mL non-essential amino acids (100×), 10% FCS + 5% BSA (*v/v*)).

peptide	IC_{50} [nM]	$\log D_{7.4}$
[$^{nat/177}\text{Lu}$]Lu-DOTA-PP-F11N	12.8 ± 2.8	-4.75 ± 0.07
[$^{nat/177}\text{Lu}$]Lu-DOTA-PP- γ -F11N	7.55 ± 0.48	-4.38 ± 0.05
[$^{nat/177}\text{Lu}$]Lu-CP04	11.2 ± 0.2	-3.80 ± 0.10
[$^{nat/177}\text{Lu}$]Lu- γ -CP04	8.88 ± 0.67	-4.30 ± 0.08
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-1	629 ± 48	-1.71 ± 0.10
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-2	156 ± 18	-1.95 ± 0.09
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-3	156 ± 4	-2.03 ± 0.10
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-4	185 ± 14	-2.19 ± 0.08
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-5	90.0 ± 2.3	-2.67 ± 0.05
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-6	56.8 ± 6.4	-2.22 ± 0.08
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-7	75.7 ± 10.7	-2.63 ± 0.08
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-8	58.0 ± 11.0	-2.19 ± 0.08
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-9	55.3 ± 7.8	-2.84 ± 0.04
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-10	863 ± 148	-2.15 ± 0.08
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-11	111 ± 13	-2.23 ± 0.07
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-12	89.7 ± 14.5	-2.34 ± 0.05
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-13	68.5 ± 11.4	-2.57 ± 0.06
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-14	51.3 ± 9.3	-2.26 ± 0.05
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-15	33.0 ± 11.1	-2.50 ± 0.02
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-16	20.4 ± 2.7	-2.54 ± 0.05
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-17	29.5 ± 1.9	-2.10 ± 0.06
[$^{nat/177}\text{Lu}$]Lu-(R)-DOTAGA-rhCCK-18	20.4 ± 2.0	-2.16 ± 0.09

Table S2. Receptor-mediated internalisation values (37 °C, RPMI 1640, 5 mM L-Gln, 5 mL non-essential amino acids (100×), 10% FCS, 0.25 pmol/well) determined as percent (%) of the applied activity of [^{177}Lu]Lu-(R)-DOTAGA-rhCCK-9 and -16 as well as the references [^{177}Lu]Lu-DOTA-PP-F11N and [^{177}Lu]Lu-CP04 using AR42J cells (3.0×10^5 cells/well) at different time points (1, 2, 4 and 6 h). Data are corrected for non-specific binding (10 μmol /well, [^{nat}Lu]Lu-DOTA-PP-F11N).

Peptide	Internalisation Values (%)				Internalisation Values* (% of reference)
	1 h	2 h	4 h	6 h	6 h
[^{177}Lu]Lu-DOTA-PP-F11N	6.44 ± 0.32	10.1 ± 0.4	17.5 ± 1.0	22.4 ± 0.6	-
[^{177}Lu]Lu-CP04	5.16 ± 0.44	9.33 ± 0.36	15.4 ± 0.7	20.5 ± 0.5	91.6 ± 2.4
[^{177}Lu]Lu-(R)-DOTAGA-rhCCK-9	2.08 ± 0.12	3.44 ± 0.55	8.01 ± 0.53	13.2 ± 0.5	65.7 ± 2.5
[^{177}Lu]Lu-(R)-DOTAGA-rhCCK-16	7.63 ± 0.13	14.0 ± 1.3	23.2 ± 2.7	32.2 ± 2.1	135 ± 9

* Internalisation values are depicted as % of the reference [^{177}Lu]Lu-DOTA-PP-F11N

Table S3. Total cell uptake (37 °C, RPMI 1640, 5 mM L-Gln, 5 mL non-essential amino acids (100×), 10% FCS, 0.25 pmol/well) determined as percent (%) of the applied activity of [¹⁷⁷Lu]Lu-(R)-DOTAGA-rhCCK-9 and -16 as well as the references [¹⁷⁷Lu]Lu-DOTA-PP-F11N and [¹⁷⁷Lu]Lu-CP04 using AR42J cells (3.0 × 10⁵ cells/well) at different time points (1, 2, 4 and 6 h). Data are corrected for non-specific binding (10 μmol/well, [^{nat}Lu]Lu-DOTA-PP-F11N).

Peptide	Total cell uptake (%)			
	1 h	2 h	4 h	6 h
[¹⁷⁷ Lu]Lu-DOTA-PP-F11N	7.03 ± 1.45	10.6 ± 1.5	18.5 ± 2.5	23.5 ± 1.4
[¹⁷⁷ Lu]Lu-CP04	6.00 ± 0.42	10.0 ± 0.4	16.5 ± 0.9	20.4 ± 1.7
[¹⁷⁷ Lu]Lu-(R)-DOTAGA-rhCCK-9	2.25 ± 0.23	3.46 ± 0.56	8.31 ± 0.59	13.4 ± 0.7
[¹⁷⁷ Lu]Lu-(R)-DOTAGA-rhCCK-16	8.37 ± 0.07	14.4 ± 1.3	23.6 ± 2.8	32.8 ± 2.3

Table S4. Biodistribution data of [¹⁷⁷Lu]Lu-DOTA-PP-F11N, [¹⁷⁷Lu]Lu-(R)-DOTAGA-rhCCK-9 and [¹⁷⁷Lu]Lu-(R)-DOTAGA-rhCCK-16 in selected organs at 24 h p.i. in AR42J tumour-bearing CB17-SCID mice (100 pmol each). Data are expressed as %ID/g, mean ± SD.

Organ	[¹⁷⁷ Lu]Lu-DOTA-PP-F11N (n = 4)	[¹⁷⁷ Lu]Lu-(R)-DOTAGA-rhCCK-9 (n = 4)	[¹⁷⁷ Lu]Lu-(R)-DOTAGA-rhCCK-16 (n = 4)	[¹⁷⁷ Lu]Lu-(R)-DOTAGA-rhCCK-16 competition studies (n = 2)
Blood	0.00 ± 0.00	0.02 ± 0.00	0.01 ± 0.00	0.05 ± 0.00
Heart	0.02 ± 0.01	0.09 ± 0.03	0.11 ± 0.01	0.13 ± 0.01
Lung	0.03 ± 0.02	0.10 ± 0.05	0.09 ± 0.02	2.59 ± 1.27
Liver	0.67 ± 0.64	1.07 ± 0.54	1.18 ± 0.27	2.31 ± 0.66
Spleen	0.07 ± 0.03	0.70 ± 0.69	0.62 ± 0.03	1.73 ± 0.01
Pancreas	0.05 ± 0.01	0.31 ± 0.20	0.39 ± 0.12	0.20 ± 0.01
Stomach	0.36 ± 0.05	1.80 ± 0.37	3.51 ± 0.50	0.15 ± 0.04
Intestine	0.04 ± 0.02	0.26 ± 0.04	0.20 ± 0.04	0.16 ± 0.03
Kidney	3.08 ± 0.51	84.4 ± 22.7	85.5 ± 11.3	105 ± 21
Adrenal	0.03 ± 0.03	0.45 ± 0.30	1.49 ± 1.50	0.40 ± 0.11
Muscle	0.00 ± 0.00	0.03 ± 0.01	0.05 ± 0.01	0.06 ± 0.04
Bone	0.03 ± 0.01	0.82 ± 0.42	0.17 ± 0.02	0.24 ± 0.04
Tumour	1.88 ± 0.82	6.40 ± 1.48	15.70 ± 3.27	0.91 ± 0.04

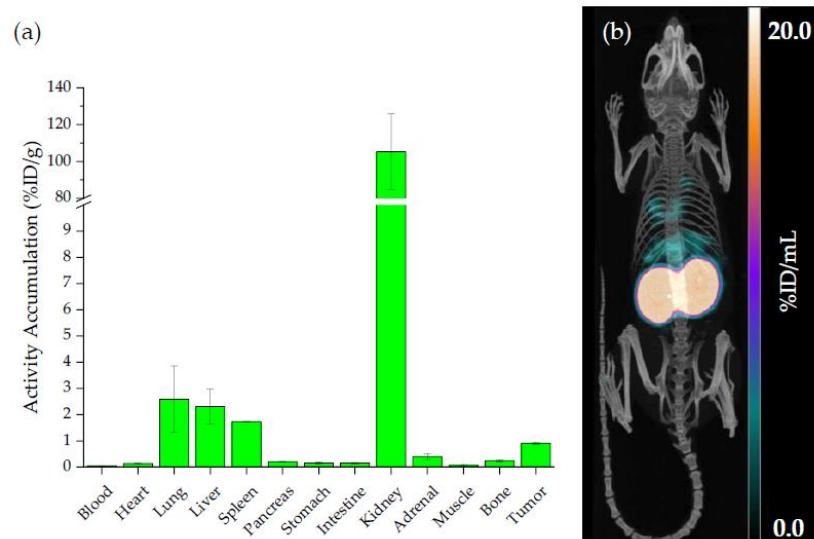


Figure S4. (a) Biodistribution of [^{177}Lu]Lu-DOTA-rhCCK-16 (100 pmol) co-injected with [$^{\text{nat}}\text{Lu}$]Lu-DOTA-MGS5 (40 nmol) in selected organs (%ID/g) at 24 h p.i. in AR42J tumour-bearing CB17-SCID mice. Data is expressed as mean \pm SD ($n = 2$). (b) Representative $\mu\text{SPECT/CT}$ images of [^{177}Lu]Lu-DOTA-rhCCK-16 co-injected with [$^{\text{nat}}\text{Lu}$]Lu-DOTA-MGS5 (40 nmol) at 24 h p.i. in AR42J tumour-bearing CB17-SCID mice.