

Supplementary materials

Computational Methods Reveal a Series of Cyclic and Linear Lichenysins and Surfactins from the Vietnamese Marine Sediment-Derived *Streptomyces* Strain G222

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Figure S1: A) The molecular networking obtained through the LC-MS/MS analysis of the extract of G222. B) Zoom view of the discriminant cluster of the lipopeptides.

Figure S2: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (1) (m/z 979.6434 $[M + H]^+$).

Figure S3: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (2) (m/z 993.6581 $[M + H]^+$).

Figure S4: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (3) (m/z 1007.6734 $[M + H]^+$).

Figure S5: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (4) (isoform of 3) (m/z 1007.6733 $[M + H]^+$).

Figure S6: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (5) (m/z 1021.6896 $[M + H]^+$).

Figure S7: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (6) (m/z 1035.7063 $[M + H]^+$).

Figure S8: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (7) (isoform of 6) (m/z 1035.7063 $[M + H]^+$).

Figure S9: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (8) (m/z 1063.7366 $[M + H]^+$).

Figure S10: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (9) (m/z 1025.6832 $[M + H]^+$).

Figure S11: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for linear Lichenysin (10) (m/z 1039.6990 $[M + H]^+$).

Figure S12: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for linear Lichenysin (11) (m/z 1053.7147 $[M + H]^+$).

Figure S13: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Surfactin (**12**) (m/z 1008.6618 $[M + H]^+$)

Figure S14: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Surfactin (**13**) (m/z 1022.6767 $[M + H]^+$)

Figure S15: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Surfactin (**14**) (m/z 1036.6906 $[M + H]^+$)

Figure S16: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for linear Surfactin (**15**) (m/z 1054.7003 $[M + H]^+$)

Figure S17: Molecular networks obtained using the Feature-Based Molecular Network workflow on GNPS (<https://gnps.ucsd.edu/ProteoSAFe/status.jsp?task=de168cddecfe40248fa3ba5077705e55>) (accessed on 23 May 2023).

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Figure S24: HSQC-NMR spectrum of Lichenysin (**3-4**) (m/z 1007.6734 and 1007.6733 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S25: HMBC-NMR spectrum of Lichenysin (**3-4**) (m/z 1007.6734 and 1007.6733 $[M + H]^+$) (600 MHz, CD_3OH).

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Figure S27: Structure of Lichenysins (**3-4**) (m/z 1007.6734 and 1007.6733 $[M + H]^+$; respectively).

Figure S28: ^1H -NMR spectrum of Lichenysin (**5a** and **5b**) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD_3OH).

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Figure S30: COSY-NMR spectrum of Lichenysin (**5a** and **5b**) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S31: TOCSY-NMR spectrum of Lichenysin (**5a** and **5b**) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S32: HSQC-NMR spectrum of Lichenysin (**5a** and **5b**) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S33: HMBC-NMR spectrum of Lichenysin (**5a** and **5b**) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S34: ROESY-NMR spectrum of Lichenysin (**5a** and **5b**) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S35: Structure of Lichenysins (**5a** and **5b**) (m/z 1021.6896 $[M + H]^+$).

Figure S36: ^1H -NMR spectrum of Lichenysin (**6-7**) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S37: DEPTQ spectrum of Lichenysin (**6-7**) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S38: COSY-NMR spectrum of Lichenysin (**6-7**) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S39: TOCSY-NMR spectrum of Lichenysin (**6-7**) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S40: HSQC-NMR spectrum of Lichenysin (**6-7**) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S41: HMBC-NMR spectrum of Lichenysin (**6-7**) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S42: ROESY-NMR spectrum of Lichenysin (**6-7**) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD_3OH).

Figure S43: Structure of Lichenysins (**6-7**) (m/z 1035.7063 $[M + H]^+$).

Figure S44: Marfey's analysis in positive ion mode high-resolution ESI mass spectrum

Figure S45: Biofilm-forming activity against *Pseudomonas aeruginosa* MUC-N1.

Figure S46: A) ATP release in *S. aureus* MRSA exhibited by compounds (**3-4**) and (**6-7**) as determined using ATP efflux assay. Squalamine ($100\text{ }\mu\text{g.mL}^{-1}$) was the positive control and water was the negative control. Compounds were tested at a final concentration of $100\text{ }\mu\text{g.mL}^{-1}$, and results are reported as relative luminescence unit. B) Bacterial growth

inhibition exhibited by (3-4) and (6-7) against *S. aureus* MRSA (CF-Marseille) with different concentrations. Positive control was bacteria only and negative control was media only.

Table S1: Product ion spectra data for Lichenysin (1) (m/z 979.6434 [M + H]⁺)

Table S2: Product ion spectra data for Lichenysin (2) (m/z 993.6581 [M + H]⁺)

Table S3: Product ion spectra data for Lichenysin (3) (m/z 1007.6734 [M + H]⁺)

Table S4: Product ion spectra data for Lichenysin (4) (isoform of 3) (m/z 1007.6733 [M + H]⁺)

Table S5: Product ion spectra data for Lichenysin (5) (m/z 1021.6896 [M + H]⁺)

Table S6: Product ion spectra data for Lichenysin (6) (m/z 1035.7063 [M + H]⁺)

Table S7: Product ion spectra data Lichenysin (7) (isoform of 6) (m/z 1035.7063 [M + H]⁺)

Table S8: Product ion spectra data Lichenysin (8) (m/z 1063.7366 [M + H]⁺)

Table S9: Product ion spectra data for Lichenysin (9) (m/z 1025.6832 [M + H]⁺)

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Table S14: Product ion spectra data for Surfactin (14) (m/z 1036.6906 [M + H]⁺)

Table S15: Product ion spectra data for Linear Surfactin (15) (m/z 1054.7003 [M + H]⁺)

Table S16. ¹H, ¹³C, HMBC, and ROESY NMR data of Lichenysin (3-4) (m/z 1007.6734 and 1007.6733 [M + H]⁺) (600 MHz, CD₃OH), isoform *ante-iso* and *iso* (0.8:0.2).

Table S17. ¹H, ¹³C, HMBC, and ROESY NMR data of Lichenysin (5a and 5b) (m/z 1021.6896 [M + H]⁺) (600 MHz, CD₃OH) isoform: *iso* and *n* (0.5:0.5).

Table S18. ¹H, ¹³C, HMBC, and ROESY NMR data of Lichenysin (6-7) (m/z 1035.7063 [M + H]⁺) (600 MHz, CD₃OH) isoform: *ante-iso* and *iso* (0.2:0.8).

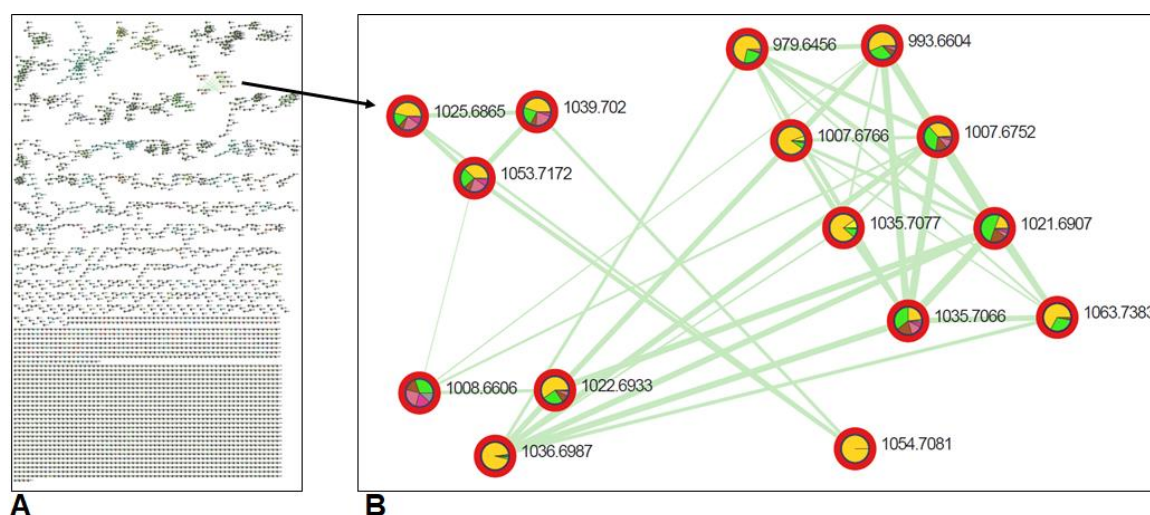


Figure S1. A) The molecular networking obtained through the LC-MS/MS analysis of the extract of G222. B) Zoom view of the discriminant cluster of the lipopeptides.

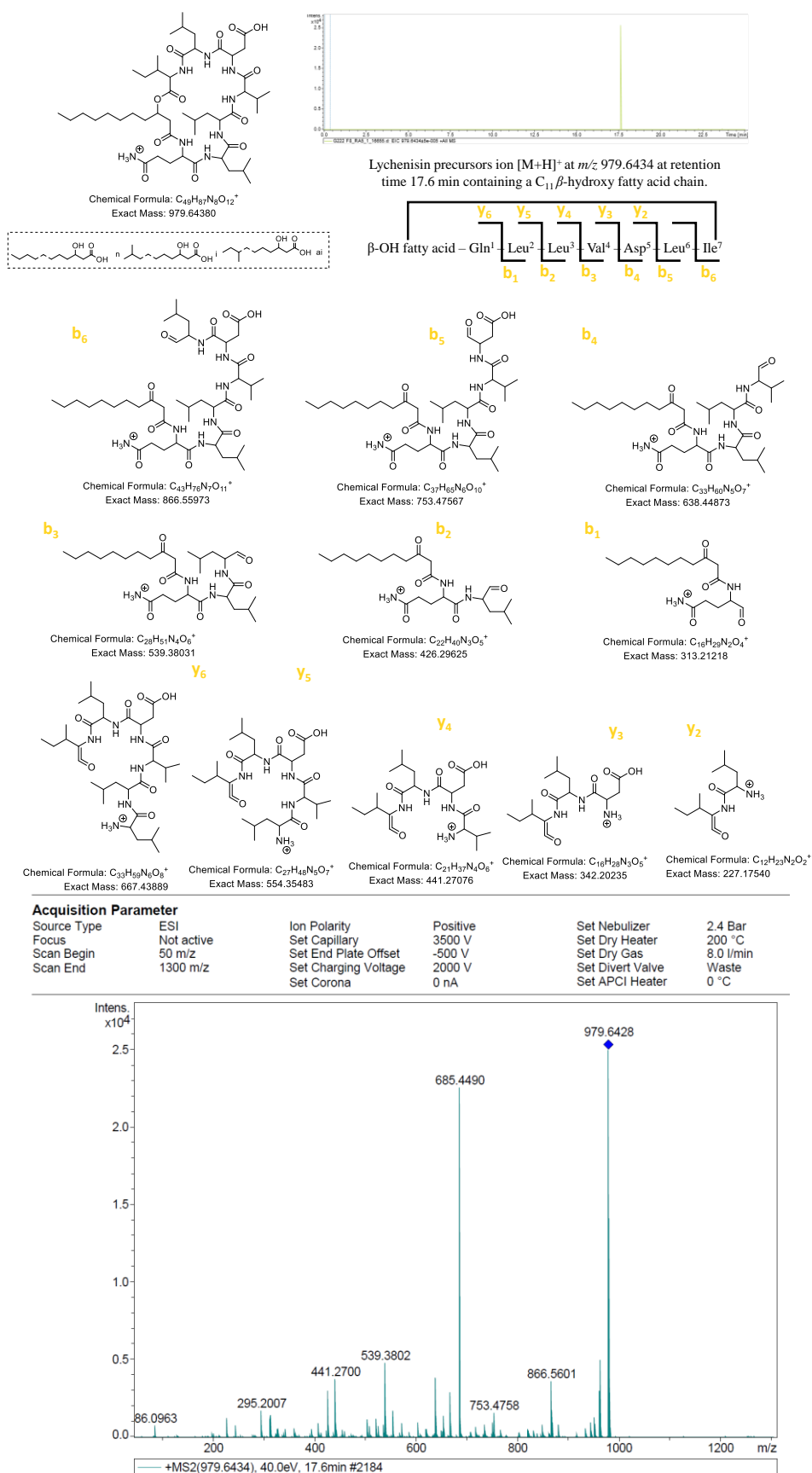


Figure S2: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (**1**) (m/z 979.6501 $[M + H]^+$).

Table S1. Product ion spectra data for Lichenysin (1) (m/z 979.6434 $[M + H]^+$).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	979.6434	0.4	C ₄₉ H ₈₇ N ₈ O ₁₂
b ₆	866.5601	0.4	C ₄₃ H ₇₆ N ₇ O ₁₁
b ₅	753.4758	-0.2	C ₃₇ H ₆₅ N ₆ O ₁₀
b ₄	638.4484	0.5	C ₃₃ H ₆₀ N ₅ O ₇
b ₃	539.3802	0.3	C ₂₈ H ₅₁ N ₄ O ₆
b ₂	426.2960	0.6	C ₂₂ H ₄₀ N ₃ O ₅
b ₁	313.2116	1.8	C ₁₆ H ₂₉ N ₂ O ₄
y ₆	667.4395	-0.9	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3550	-0.3	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2700	1.8	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2022	0.4	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1753	0.7	C ₁₂ H ₂₃ N ₂ O ₂

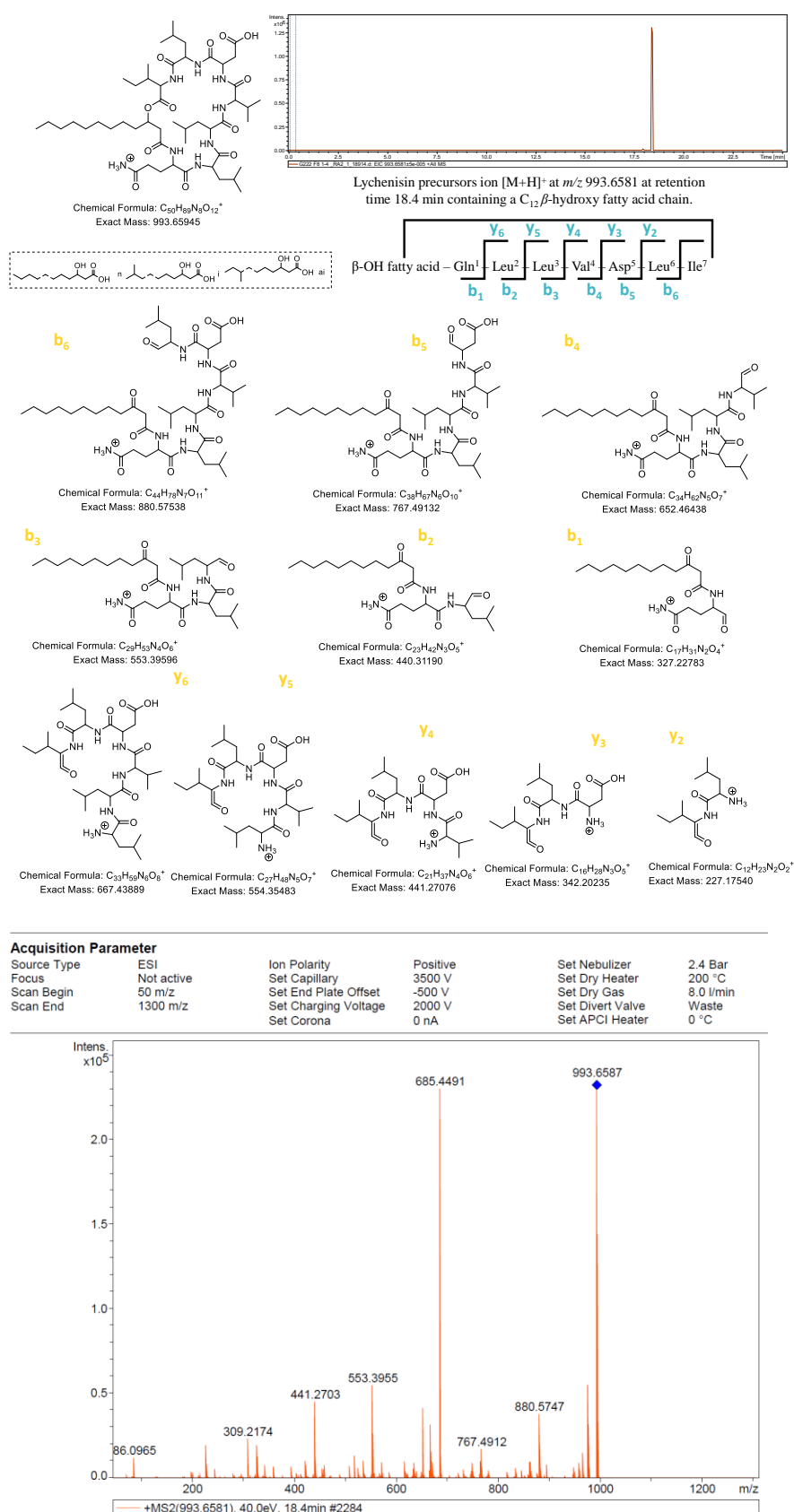


Figure S3: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (2) (m/z 993.6581 $[M+H]^+$)

Table S2. Product ion spectra data for Lichenysin (**2**) (m/z 993.6581 $[M + H]^+$).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	993.6581	-1.7	C ₅₀ H ₈₉ N ₈ O ₁₂
b ₆	880.5754	0.8	C ₄₄ H ₇₈ N ₇ O ₁₁
b ₅	767.4912	0.1	C ₃₈ H ₆₇ N ₆ O ₁₀
b ₄	652.4641	0.4	C ₃₄ H ₆₂ N ₅ O ₇
b ₃	553.3955	0.9	C ₂₉ H ₅₃ N ₄ O ₆
b ₂	440.3115	1.0	C ₂₃ H ₄₂ N ₃ O ₅
b ₁	327.2278	0.5	C ₁₇ H ₃₁ N ₂ O ₄
y ₆	667.4386	0.4	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3545	0.6	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2703	1.0	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2020	1.1	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1754	0.1	C ₁₂ H ₂₃ N ₂ O ₂

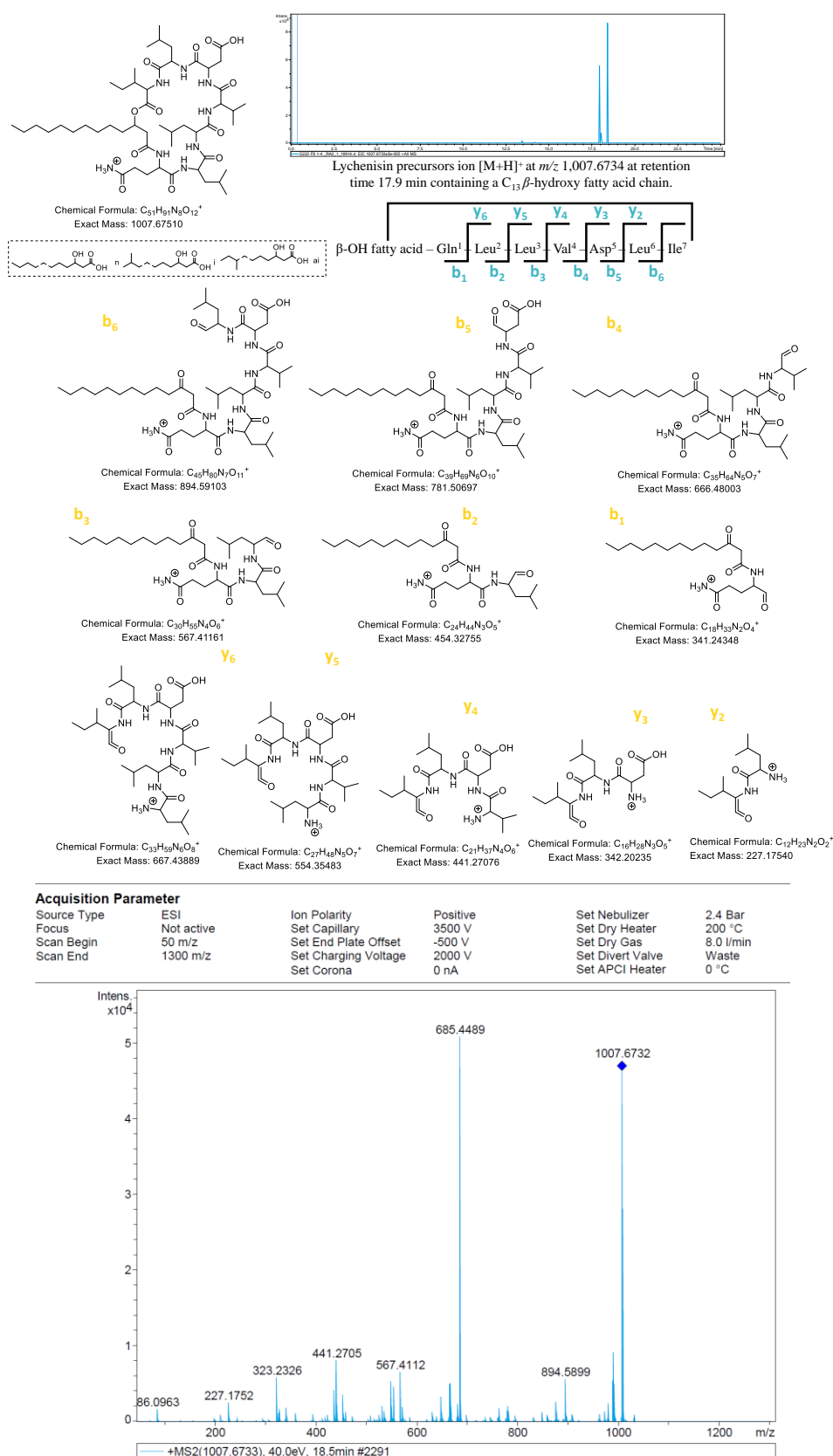


Figure S4: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (3) (m/z 1007.6734 $[M + H]^+$)

Table S3. Product ion spectra data for Lichenysin (**3**) (m/z 1007.6734 $[M + H]^+$).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	1007.6734	-1.7	C ₅₁ H ₉₁ N ₈ O ₁₂
b ₆	894.5896	1.6	C ₄₅ H ₈₀ N ₇ O ₁₁
b ₅	781.5077	-1.0	C ₃₉ H ₆₉ N ₆ O ₁₀
b ₄	666.4787	2.0	C ₃₅ H ₆₄ N ₅ O ₇
b ₃	567.4111	1.0	C ₃₀ H ₅₅ N ₄ O ₆
b ₂	454.3267	1.8	C ₂₄ H ₄₄ N ₃ O ₅
b ₁	341.2429	1.6	C ₁₈ H ₃₃ N ₂ O ₄
y ₆	667.4393	-0.7	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3538	1.8	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2702	1.3	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2026	-0.8	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1755	0.4	C ₁₂ H ₂₃ N ₂ O ₂

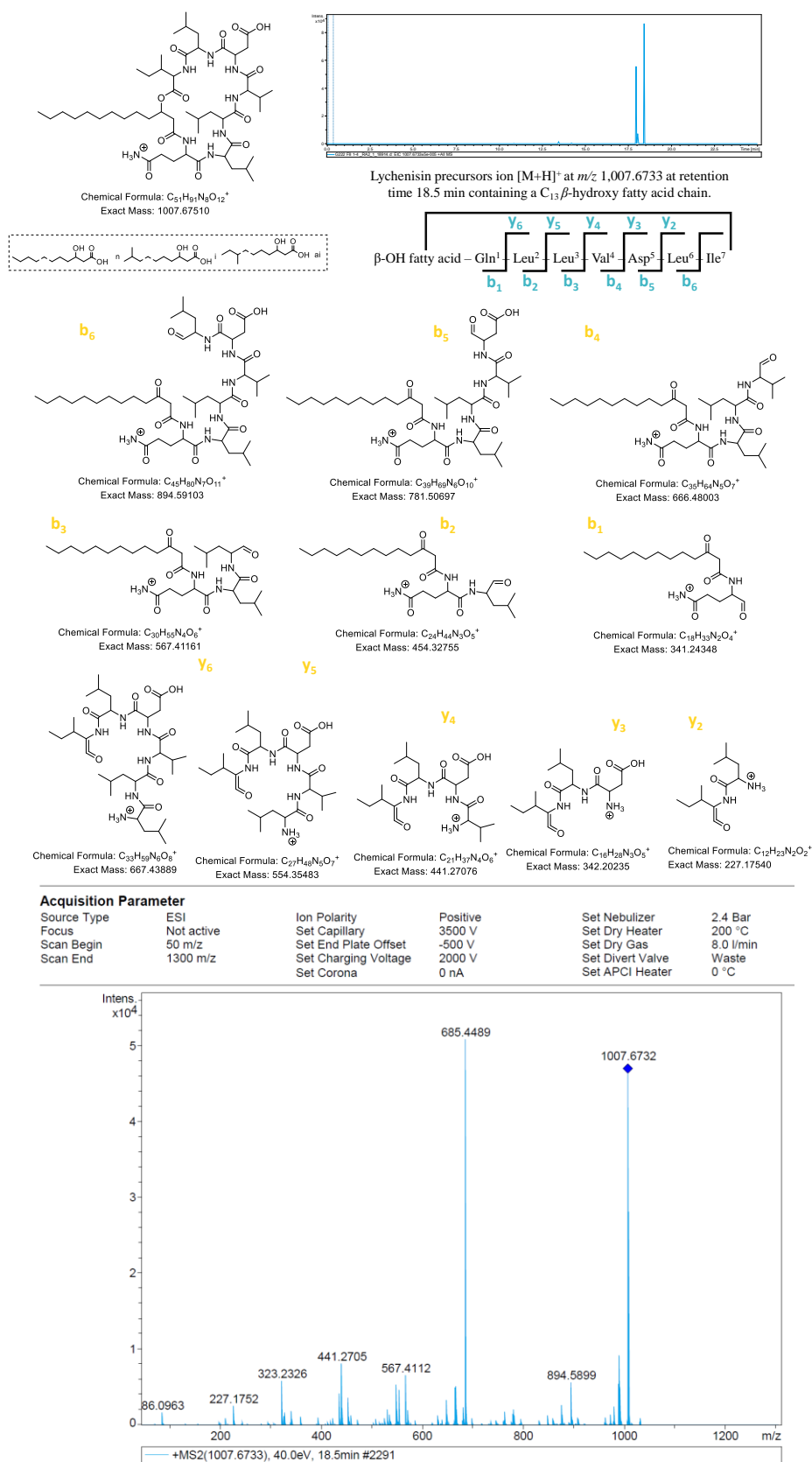


Figure S5: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (4) (isoform of 3) (m/z 1007.6733 $[M + H]^+$)

Table S4: Product ion spectra data for Lichenysin (4) (isoform of 3) (m/z 1007.6733 $[M + H]^+$)

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	1007.6733	-1.8	C ₅₁ H ₉₁ N ₈ O ₁₂
b ₆	894.5899	2.7	C ₄₅ H ₈₀ N ₇ O ₁₁
b ₅	781.5060	2.9	C ₃₉ H ₆₉ N ₆ O ₁₀
b ₄	666.4793	1.1	C ₃₅ H ₆₄ N ₅ O ₇
b ₃	567.4116	0.8	C ₃₀ H ₅₅ N ₄ O ₆
b ₂	454.3265	2.2	C ₂₄ H ₄₄ N ₃ O ₅
b ₁	341.2431	1.2	C ₁₈ H ₃₃ N ₂ O ₄
y ₆	667.4389	0.2	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3540	-0.9	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2705	0.7	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2018	1.6	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1752	0.8	C ₁₂ H ₂₃ N ₂ O ₂

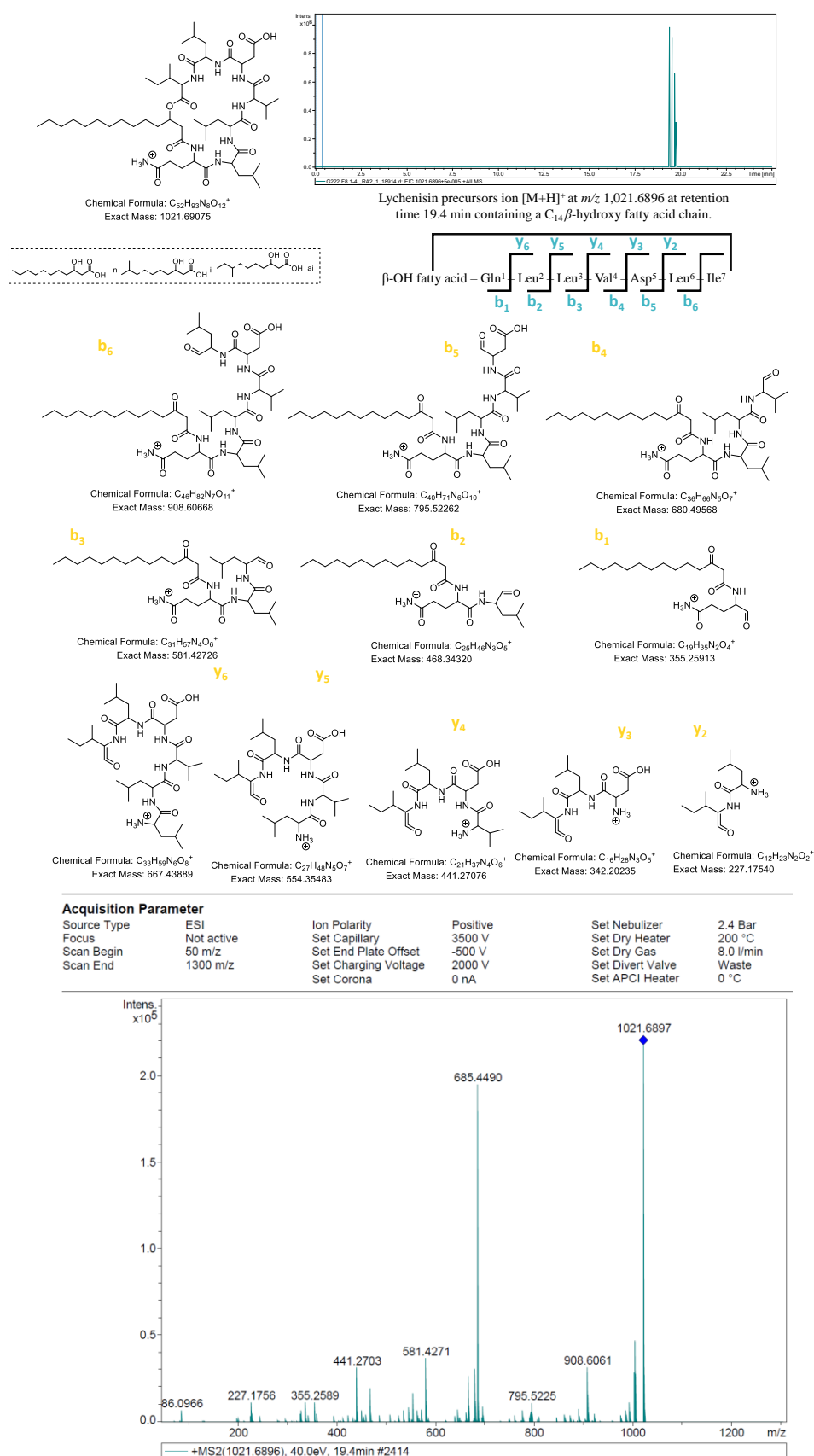


Figure S6: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (5) (m/z 1021.6896 $[M + H]^+$)

Table S5: Product ion spectra data for Lichenysin (5) (m/z 1021.6896 $[M + H]^+$)

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	1021.6896	-0.01	C ₅₂ H ₉₃ N ₈ O ₁₂
b ₆	908.6061	0.6	C ₄₆ H ₈₂ N ₇ O ₁₁
b ₅	795.5225	0.2	C ₄₀ H ₇₁ N ₆ O ₁₀
b ₄	680.4950	1.0	C ₃₆ H ₆₆ N ₅ O ₇
b ₃	581.4271	0.3	C ₃₁ H ₅₇ N ₄ O ₆
b ₂	468.3427	1.0	C ₂₅ H ₄₆ N ₃ O ₅
b ₁	355.2589	0.6	C ₁₉ H ₃₅ N ₂ O ₄
y ₆	667.4388	0.2	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3547	0.2	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2703	1.1	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2022	0.5	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1756	-0.8	C ₁₂ H ₂₃ N ₂ O ₂

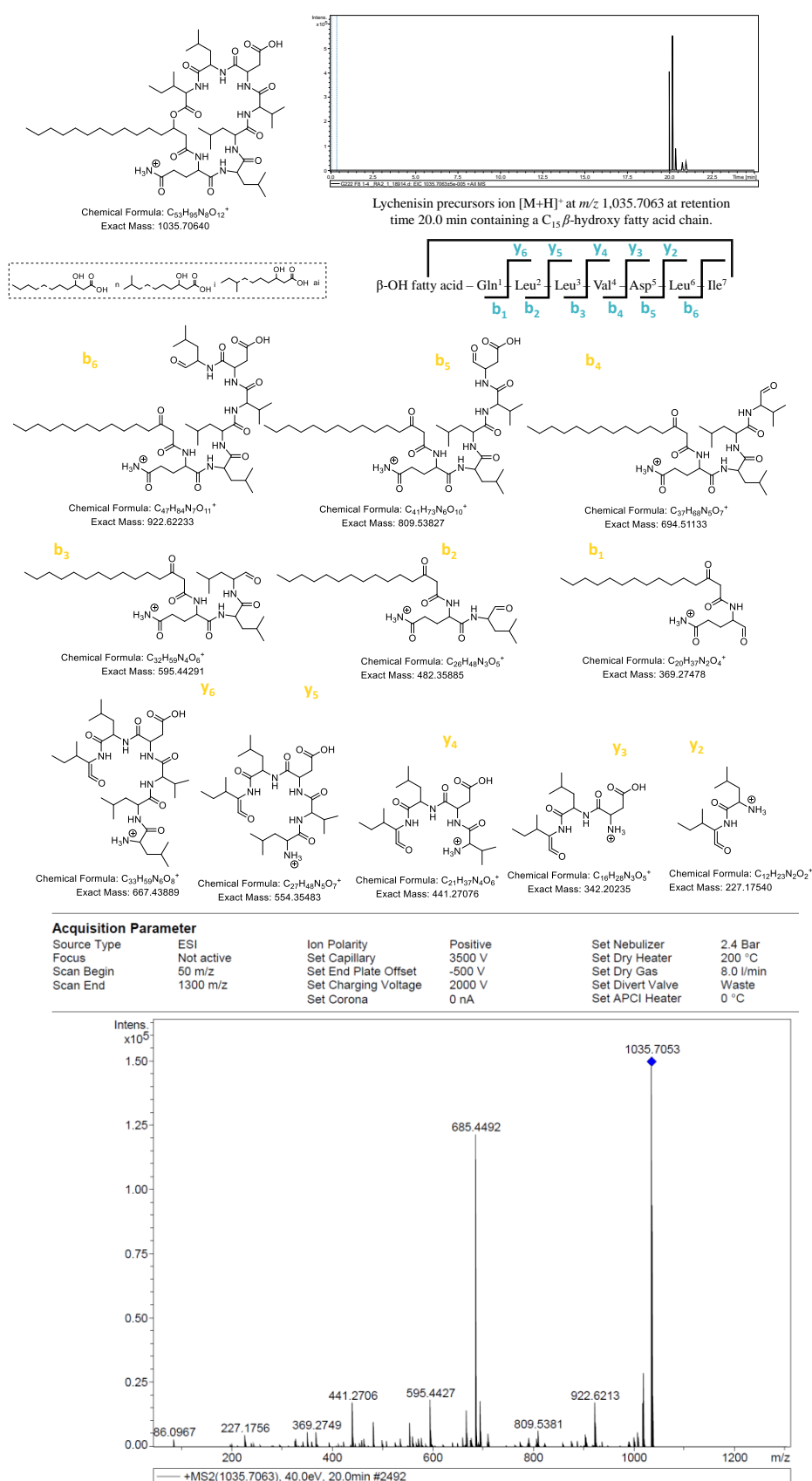


Figure S7: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (6) (m/z 1035.7063 $[M + H]^+$)

Table S6. Product ion spectra data for Lichenysin (**6**) (m/z 1035.7063 [M + H]⁺).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	1035.7063	-0.01	C ₅₃ H ₉₅ N ₈ O ₁₂
b ₆	922.6213	1.1	C ₄₇ H ₈₄ N ₇ O ₁₁
b ₅	809.5381	0.2	C ₄₁ H ₇₃ N ₆ O ₁₀
b ₄	694.5113	0.4	C ₃₇ H ₆₈ N ₅ O ₇
b ₃	595.4427	0.3	C ₃₂ H ₅₉ N ₄ O ₆
b ₂	482.3585	0.8	C ₂₆ H ₄₈ N ₃ O ₅
b ₁	369.2749	-0.2	C ₂₀ H ₃₇ N ₂ O ₄
y ₆	667.4391	-0.3	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3545	0.5	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2706	0.4	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2026	-0.6	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1756	-0.7	C ₁₂ H ₂₃ N ₂ O ₂

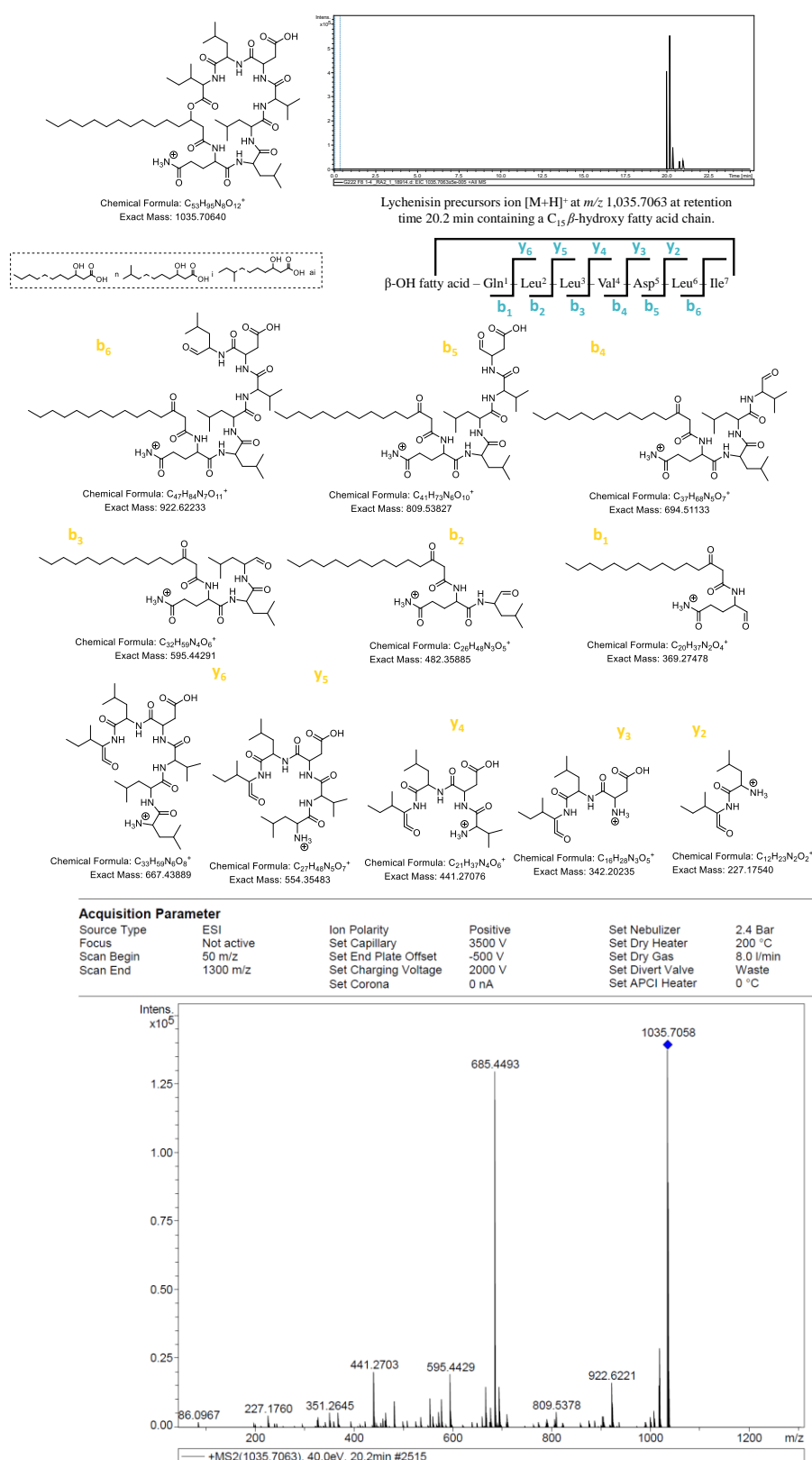


Figure S8: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (7) (isoform of 6) (m/z 1035.7063 $[M + H]^+$)

Table S7. Product ion spectra data Lichenysin (7) (isoform of 6) (m/z 1035.7063 $[M + H]^+$).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	1035.7063	-0.01	C ₅₃ H ₉₅ N ₈ O ₁₂
b ₆	922.6221	0.2	C ₄₇ H ₈₄ N ₇ O ₁₁
b ₅	809.5378	0.6	C ₄₁ H ₇₃ N ₆ O ₁₀
b ₄	694.5109	0.6	C ₃₇ H ₆₈ N ₅ O ₇
b ₃	595.4429	0.0	C ₃₂ H ₅₉ N ₄ O ₆
b ₂	482.3589	0.0	C ₂₆ H ₄₈ N ₃ O ₅
b ₁	369.2748	0.0	C ₂₀ H ₃₇ N ₂ O ₄
y ₆	667.4388	0.1	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3552	-0.6	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2703	1.0	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2025	-0.5	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1760	-2.5	C ₁₂ H ₂₃ N ₂ O ₂

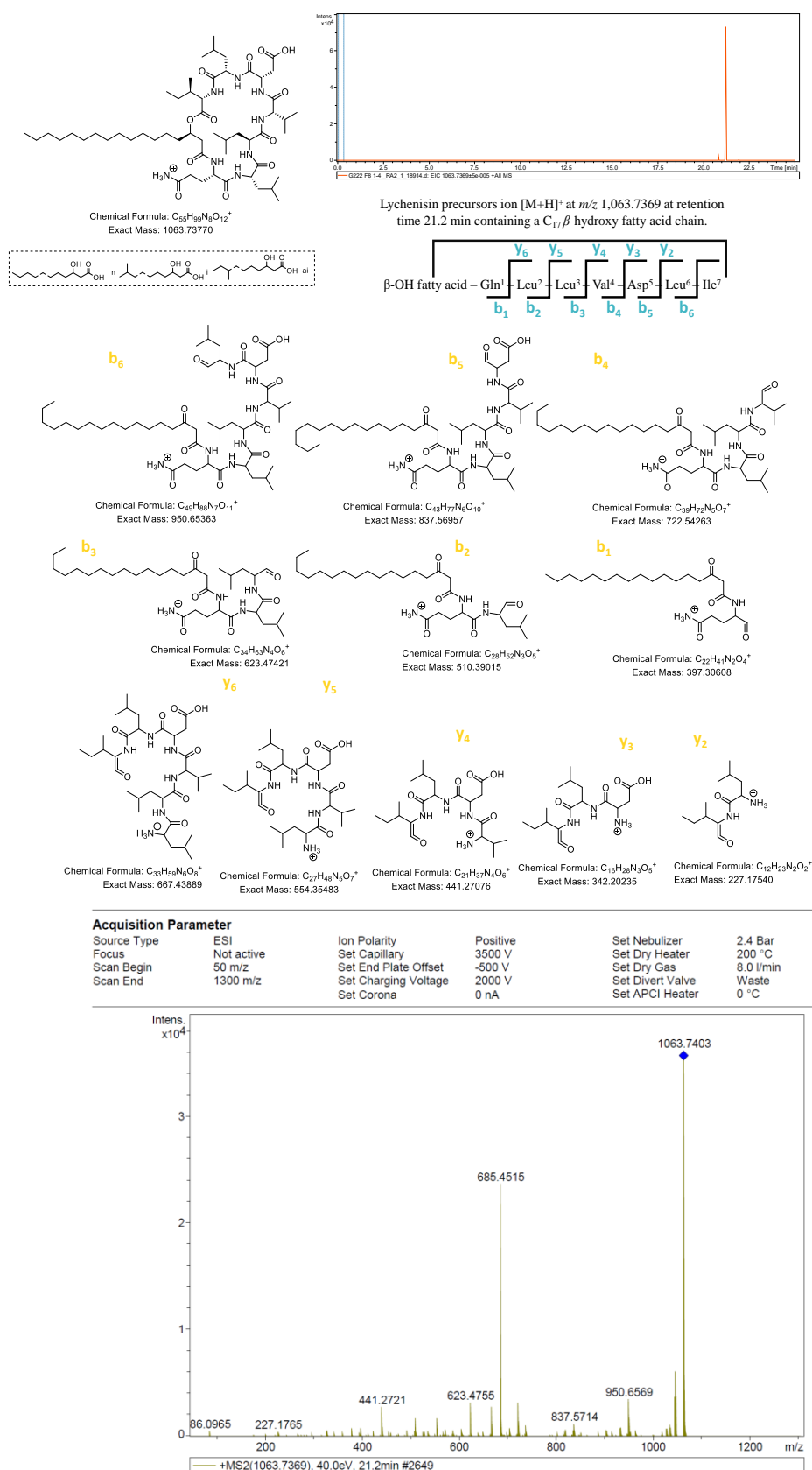


Figure S9: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Lichenysin (8) (m/z 1063.7366 $[M + H]^+$)

Table S8. Product ion spectra data Lichenysin (8) (m/z 1063.7366 $[M + H]^+$).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	1063.7366	-1.0	C ₅₅ H ₉₉ N ₈ O ₁₂
b ₆	950.6569	-3.4	C ₄₉ H ₈₈ N ₇ O ₁₁
b ₅	837.5714	-0.6	C ₄₃ H ₇₇ N ₆ O ₁₀
b ₄	722.5448	-3.1	C ₃₉ H ₇₂ N ₅ O ₇
b ₃	623.4755	-2.1	C ₃₄ H ₆₃ N ₄ O ₆
b ₂	510.3915	-2.7	C ₂₈ H ₅₂ N ₃ O ₅
b ₁	397.3070	-2.4	C ₂₂ H ₄₁ N ₂ O ₄
y ₆	667.4414	2.2	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3564	-2.8	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2721	-2.1	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2044	-6.0	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1765	-4.8	C ₁₂ H ₂₃ N ₂ O ₂

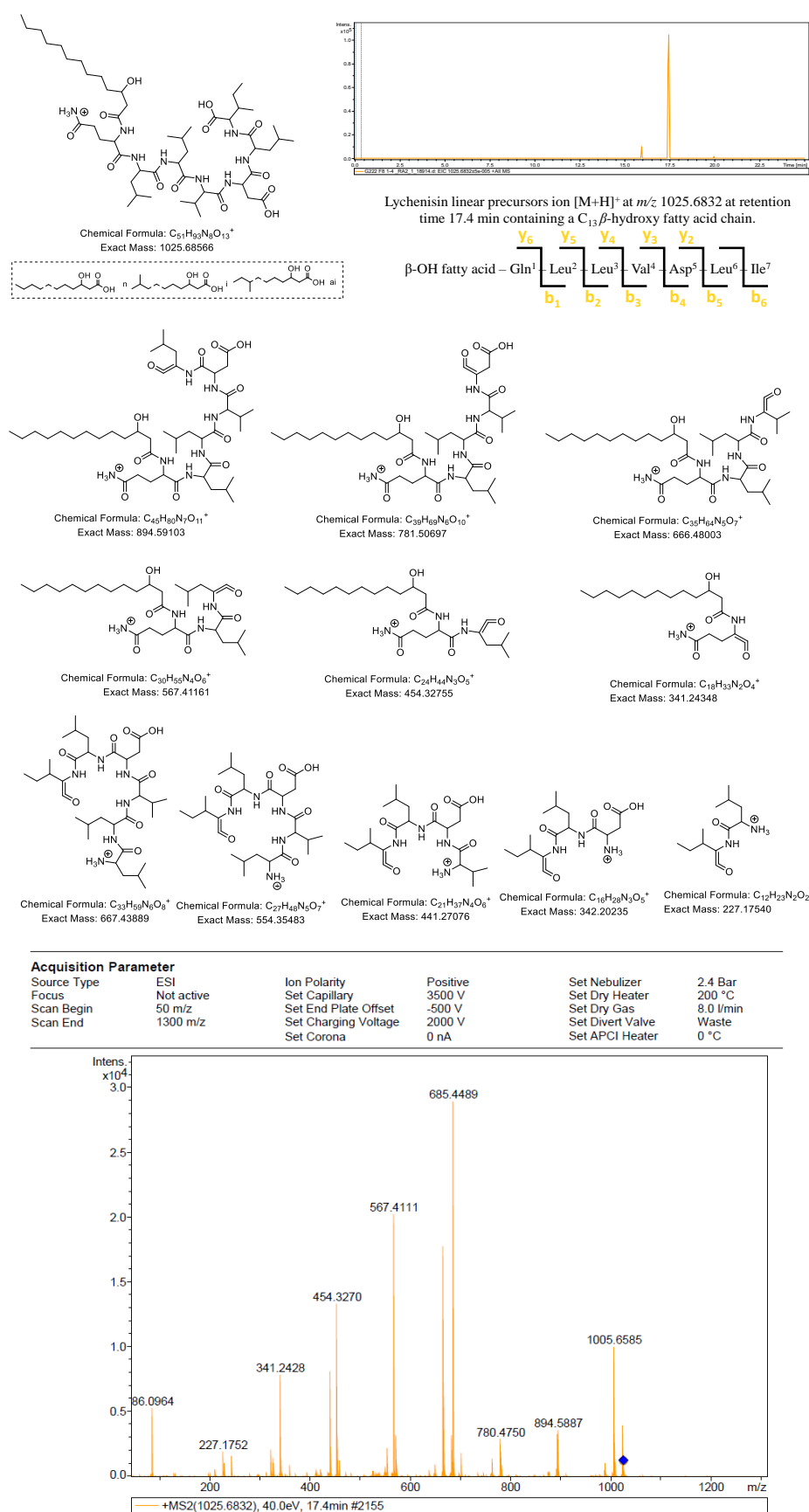


Figure S10. Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for linear Lichenysin (9) (m/z 1025.6832 $[M + H]^+$).

Table S9. Product ion spectra data for Lichenysin (**9**) (m/z 1025.6832 $[M + H]^+$).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	1025.6832	-2.4	C ₅₁ H ₉₃ N ₈ O ₁₃
b ₆	894.5887	2.6	C ₄₅ H ₈₀ N ₇ O ₁₁
b ₅	781.5056	1.7	C ₃₉ H ₆₉ N ₆ O ₁₀
b ₄	666.4797	0.5	C ₃₅ H ₆₄ N ₅ O ₇
b ₃	567.4111	0.9	C ₃₀ H ₅₅ N ₄ O ₆
b ₂	454.3270	1.2	C ₂₄ H ₄₄ N ₃ O ₅
b ₁	341.2428	2.1	C ₁₈ H ₃₃ N ₂ O ₄
y ₆	667.4831	5.1	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3541	1.4	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2701	1.5	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2459	12.0	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1752	0.7	C ₁₂ H ₂₃ N ₂ O ₂

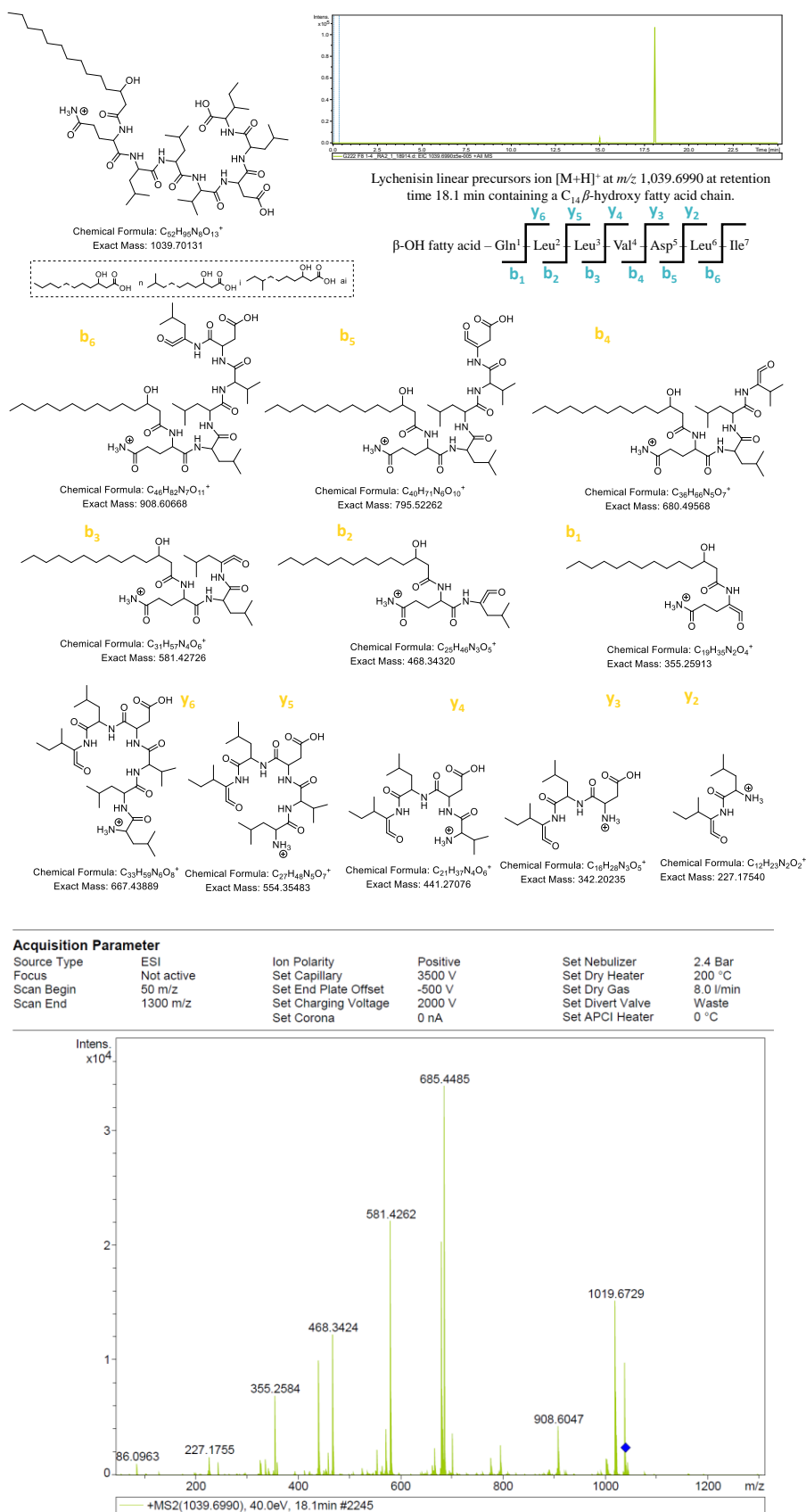


Figure S11: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for linear Lichenysin (10) (m/z 1039.6990 $[M+H]^+$)

Table S10. Product ion spectra data for Linear Lichenysin (**10**) (m/z 1039.6990 $[M + H]^+$).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	1039.6990	-2.22	C ₅₂ H ₉₅ N ₈ O ₁₃
b ₆	908.6047	2.1	C ₄₆ H ₈₂ N ₇ O ₁₁
b ₅	795.5204	2.8	C ₄₀ H ₇₁ N ₆ O ₁₀
b ₄	680.4947	1.4	C ₃₆ H ₆₆ N ₅ O ₇
b ₃	581.4262	1.8	C ₃₁ H ₅₇ N ₄ O ₆
b ₂	468.3424	1.8	C ₂₅ H ₄₆ N ₃ O ₅
b ₁	355.2584	2.5	C ₁₈ H ₃₃ N ₂ O ₄
y ₆	667.4383	0.8	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3537	2.0	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2698	2.2	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2024	-0.1	C ₁₉ H ₃₅ N ₅ O ₃
y ₂	227.1755	-0.3	C ₁₂ H ₂₃ N ₂ O ₂

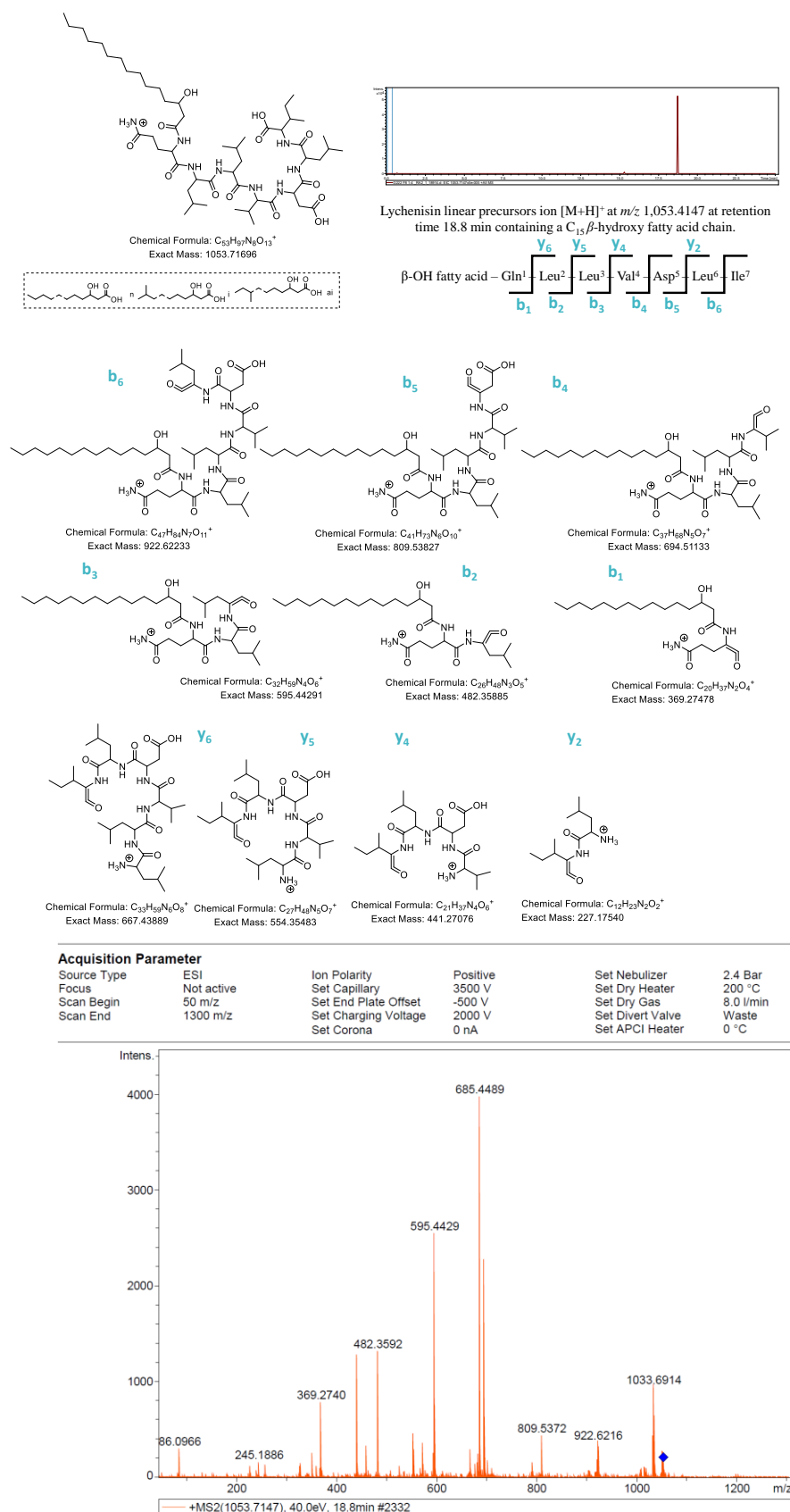


Figure S12 Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for linear Lichenysin (11) (m/z 1053.7147 $[M + H]^+$)

Table S11. Product ion spectra data for Linear Lichenysin (11) (m/z 1053.7147 $[M + H]^+$).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Gln ¹ – Leu ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Ile ⁷	1053.7147	-2.1	C ₅₃ H ₉₇ N ₈ O ₁₃
b ₆	922.6216	0.8	C ₄₇ H ₈₄ N ₇ O ₁₁
b ₅	809.5372	1.3	C ₄₁ H ₇₃ N ₆ O ₁₀
b ₄	694.5120	-0.9	C ₃₇ H ₆₈ N ₅ O ₇
b ₃	595.4429	0.0	C ₃₂ H ₅₉ N ₄ O ₆
b ₂	482.3592	-0.7	C ₂₆ H ₄₈ N ₃ O ₅
b ₁	369.2740	2.0	C ₂₀ H ₃₇ N ₂ O ₄
y ₆	667.4389	0.9	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3551	-0.5	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2698	2.5	C ₂₁ H ₃₇ N ₄ O ₆
y ₂	227.1755	0.5	C ₁₂ H ₂₃ N ₂ O ₂

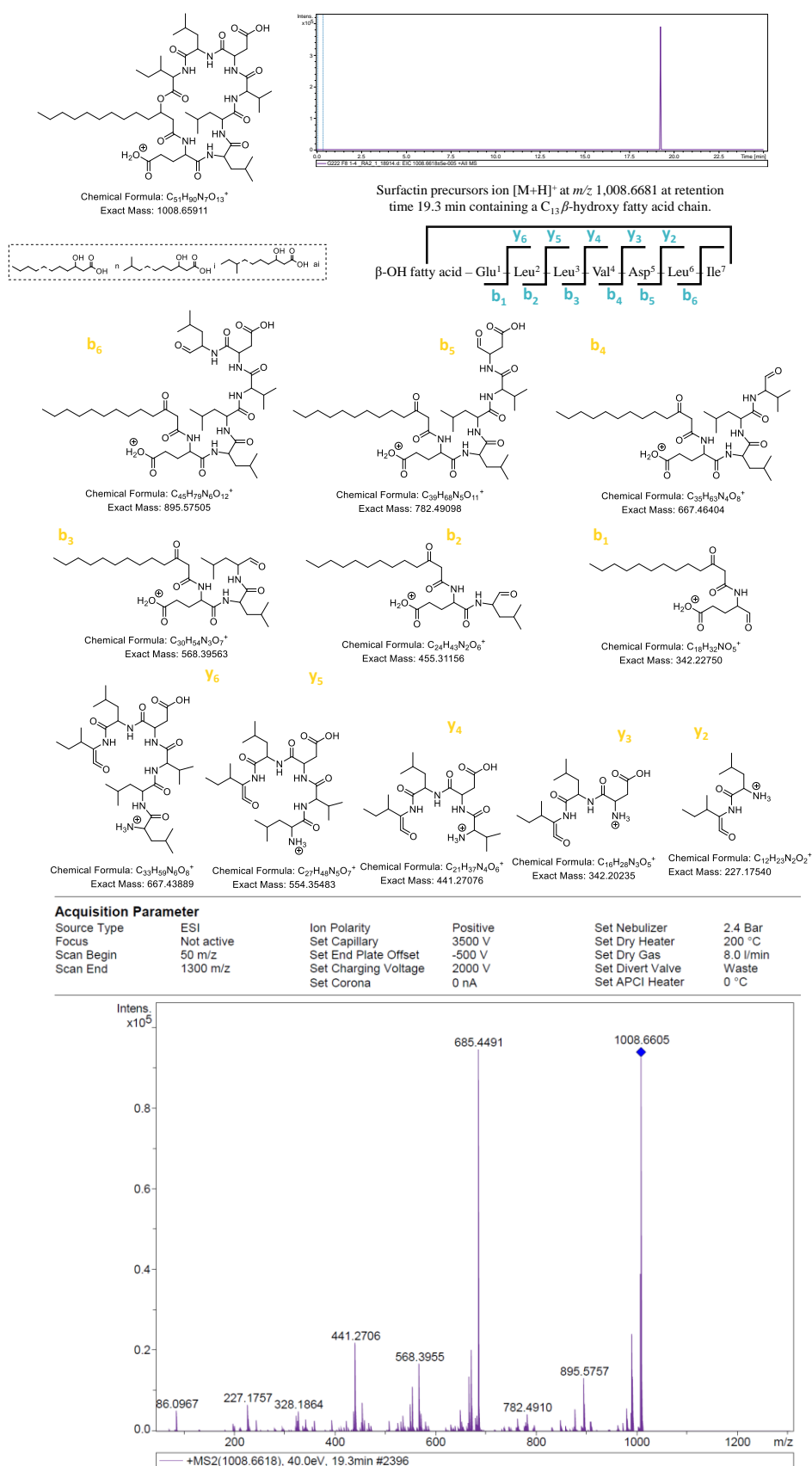


Figure S13: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Surfactin (12) (m/z 1008.6618 $[M + H]^+$)

Table S12. Product ion spectra data for Surfactin (**12**) (m/z 1008.6618 $[M + H]^+$).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Glu ¹ – Leu/Ile ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Leu/Ile ⁷	1008.6618	2.7	C ₅₁ H ₉₀ N ₇ O ₁₃
b ₆	895.5757	0.8	C ₄₅ H ₇₉ N ₆ O ₁₂
b ₅	782.4910	0.0	C ₃₉ H ₆₈ N ₅ O ₁₁
b ₄	667.4605	0.6	C ₃₅ H ₆₃ N ₄ O ₈
b ₃	568.3955	0.2	C ₃₀ H ₅₄ N ₃ O ₇
b ₂	455.3114	0.4	C ₂₄ H ₄₃ N ₂ O ₆
b ₁	342.2277	-0.6	C ₁₈ H ₃₃ N ₁ O ₅
y ₆	667.4412	-3.4	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3546	0.3	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2706	0.3	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2023	0.1	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.0655	-1.4	C ₁₂ H ₂₃ N ₂ O ₂

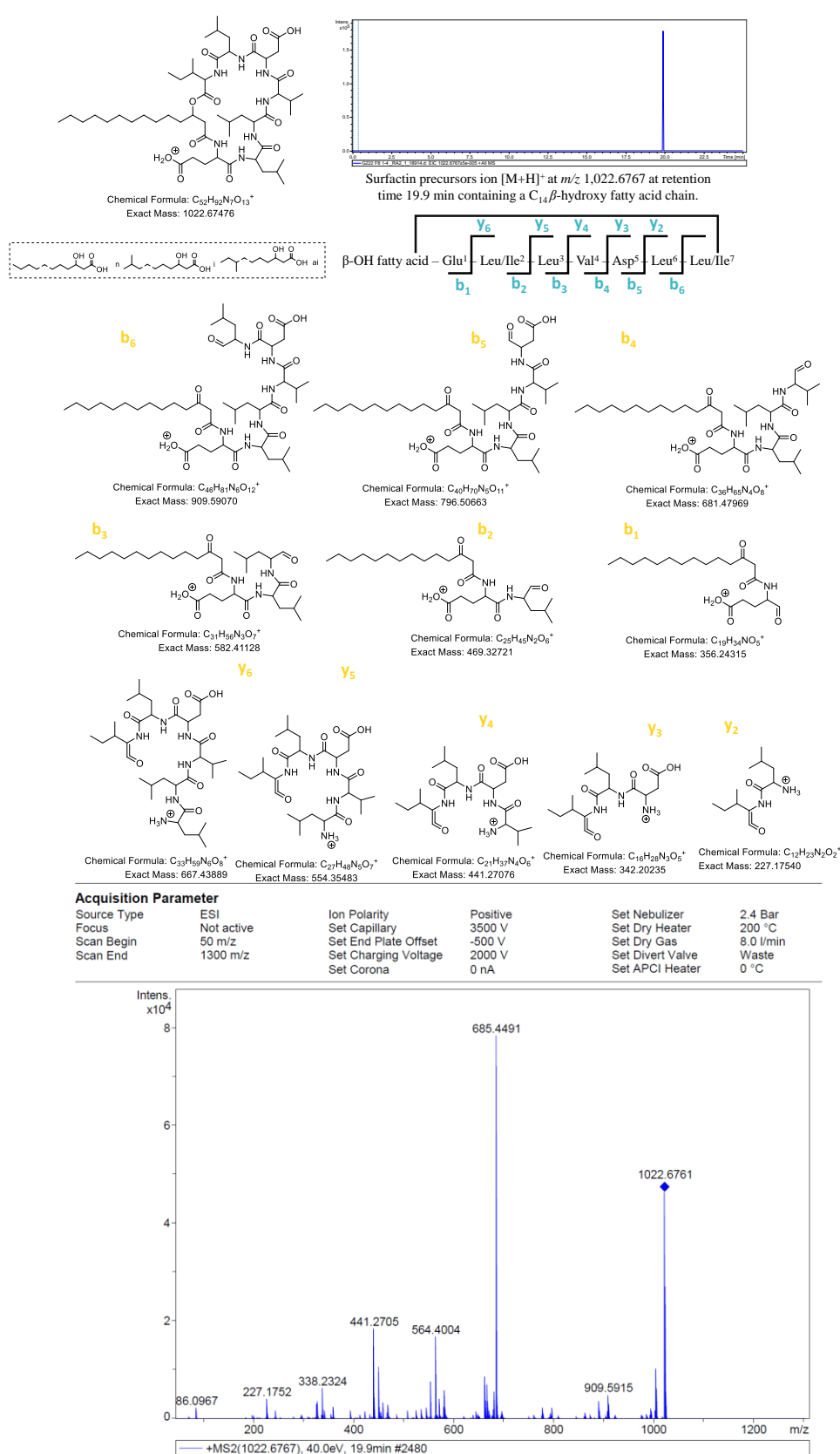


Figure S14: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Surfactin (13) (m/z 1022.6767 $[M + H]^+$)

Table S13. Product ion spectra data for Surfactin (13) (m/z 1022.6767 [M + H]⁺).

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Glu ¹ – Leu/Ile ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Leu/Ile ⁷	1022.6767	1.9	C ₅₂ H ₉₂ N ₇ O ₁₃
b ₆	909.5915	-0.9	C ₄₆ H ₈₁ N ₆ O ₁₂
b ₅	796.5079	-1.6	C ₄₀ H ₇₀ N ₅ O ₁₁
b ₄	681.4800	1.5	C ₃₆ H ₆₅ N ₄ O ₈
b ₃	582.4119	-1.1	C ₃₁ H ₅₆ N ₃ O ₇
b ₂	469.3273	-0.1	C ₂₅ H ₄₅ N ₂ O ₆
b ₁	356.2429	0.6	C ₁₉ H ₃₄ NO ₅
y ₆	667.4383	0.9	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3546	0.4	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2705	0.6	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2024	-0.1	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1752	0.8	C ₁₂ H ₂₃ N ₂ O ₂

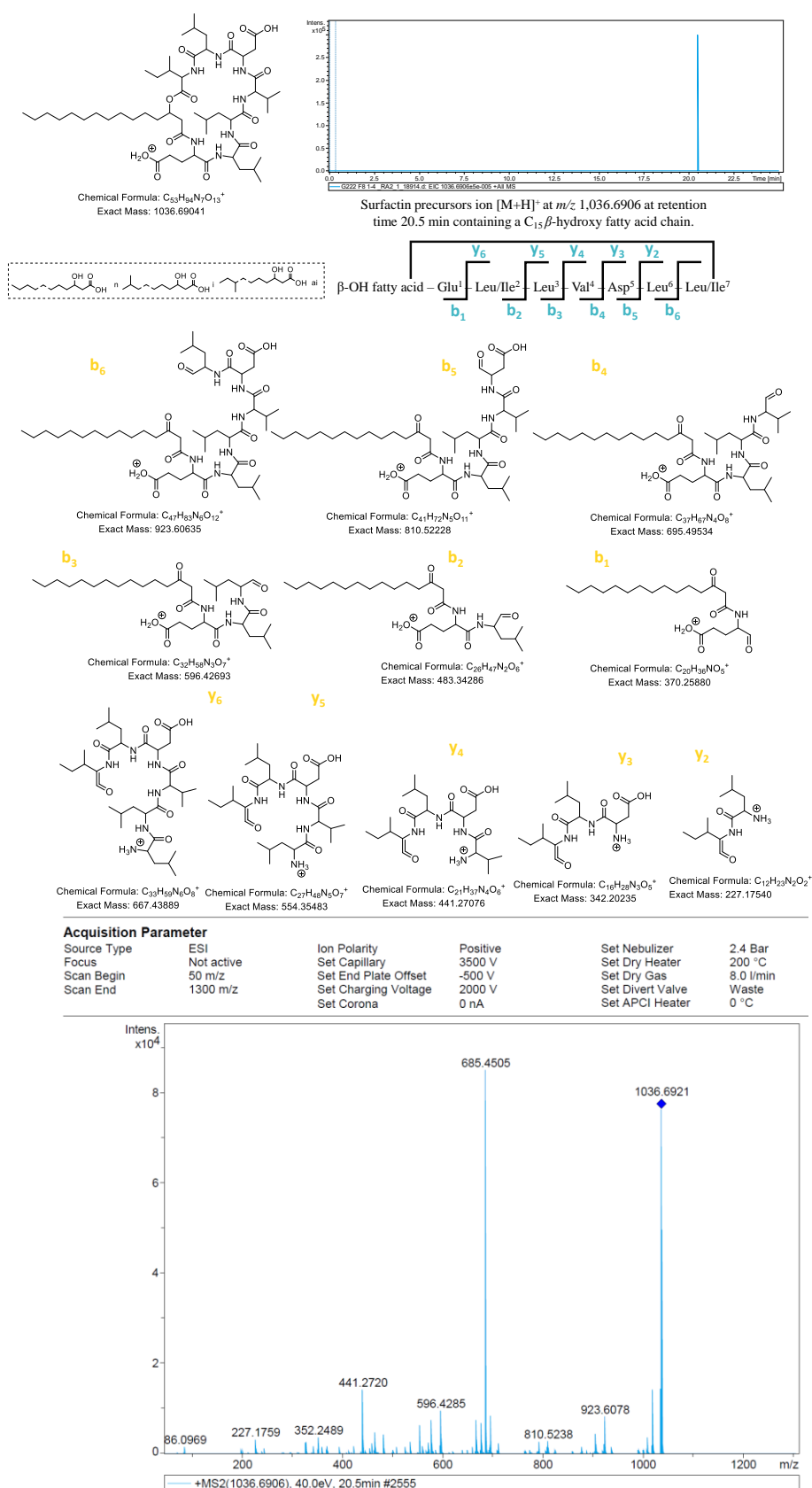


Figure S15: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for Surfactin (14) (m/z 1036.6906 $[M + H]^+$)

Table S14: Product ion spectra data for Surfactin (**14**) (m/z 1036.6906 $[M + H]^+$)

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Glu ¹ – Leu/Ile ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Leu/Ile ⁷	1036.6906	0.2	C ₅₃ H ₉₄ N ₇ O ₁₃
b ₆	923.6078	-1.6	C ₄₇ H ₈₃ N ₆ O ₁₂
b ₅	810.5238	-1.8	C ₄₁ H ₇₂ N ₅ O ₁₁
b ₄	695.4970	-2.4	C ₃₇ H ₆₇ N ₄ O ₈
b ₃	596.4285	-2.6	C ₃₂ H ₅₈ N ₃ O ₇
b ₂	483.3440	0.5	C ₂₆ H ₄₇ N ₂ O ₆
b ₁	370.2599	4.2	C ₂₀ H ₃₆ N ₁ O ₅
y ₆	667.4401	-1.8	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3548	-2.1	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2720	-2.9	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2033	-2.8	C ₁₆ H ₃₂ N ₅ O ₃
y ₂	227.1759	-2.1	C ₁₂ H ₂₃ N ₂ O ₂

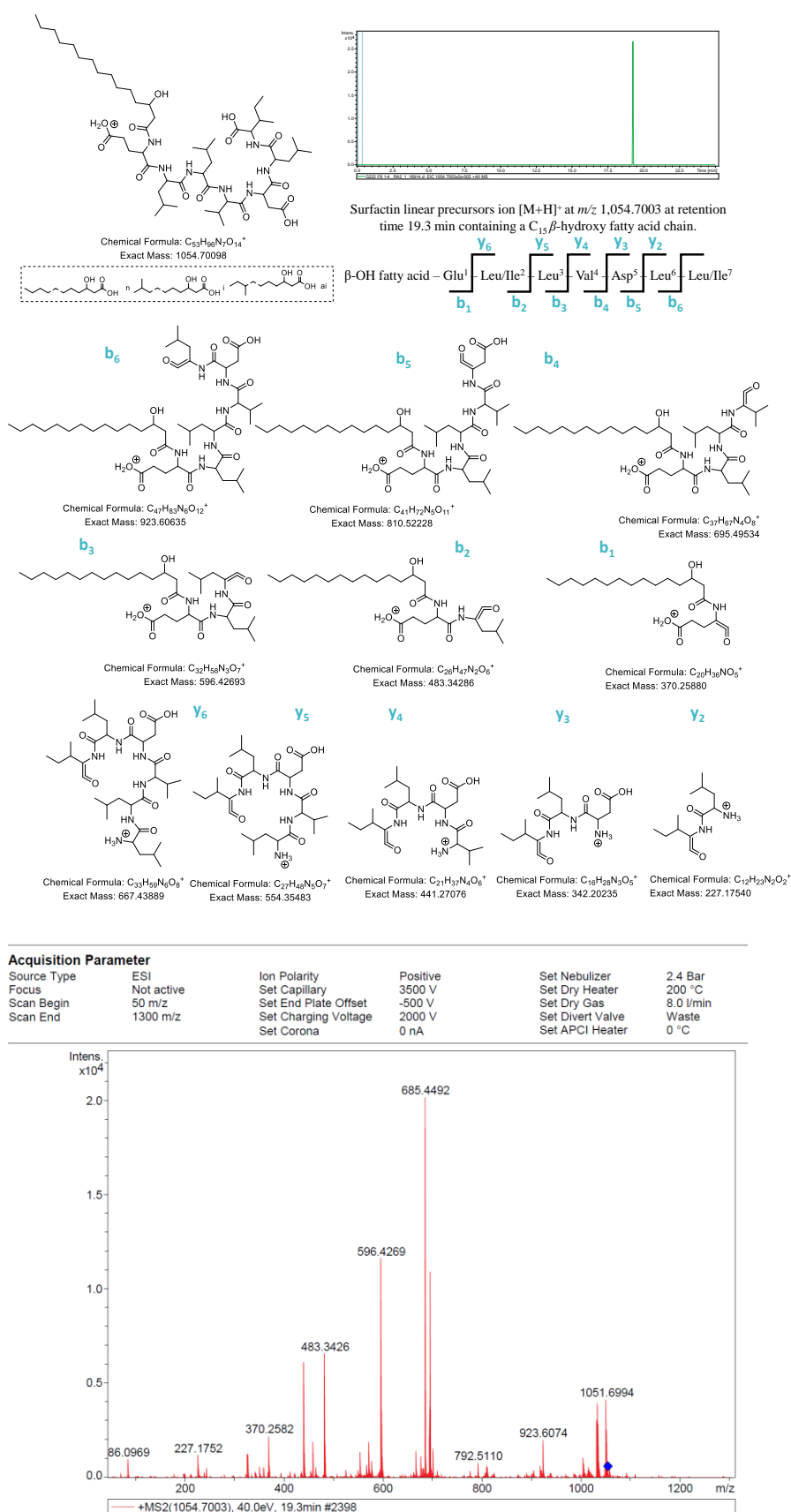


Figure S16: Fragmentations pattern and positive ion mode high-resolution ESI MS/MS spectrum for linear Surfactin (15) (m/z 1054.7003 $[M + H]^+$)

Table S15: Product ion spectra data for Linear Surfactin (**15**) (m/z 1054.7003 $[M + H]^+$)

Product ion Assignment	(m/z)	Error, pm	Molecular Formula
β -OH fatty acid – Glu ¹ – Leu/Ile ² – Leu ³ – Val ⁴ – Asp ⁵ – Leu ⁶ – Leu/Ile ⁷	1054.7003	0.6	C ₅₃ H ₉₆ N ₇ O ₁₄
b ₆	923.6074	-1.2	C ₄₇ H ₈₃ N ₆ O ₁₂
b ₅	810.5224	-0.1	C ₄₁ H ₇₂ N ₅ O ₁₁
b ₄	695.4949	2.5	C ₃₇ H ₆₇ N ₄ O ₈
b ₃	596.4269	0.0	C ₃₂ H ₅₈ N ₃ O ₇
b ₂	483.3426	0.6	C ₂₆ H ₄₇ N ₂ O ₆
b ₁	370.2582	1.7	C ₂₀ H ₃₆ N ₁ O ₅
y ₆	667.4322	0.4	C ₃₃ H ₅₉ N ₈ O ₆
y ₅	554.3551	-0.6	C ₂₇ H ₄₈ N ₅ O ₇
y ₄	441.2708	0.0	C ₂₁ H ₃₇ N ₄ O ₆
y ₃	342.2024	-0.1	C ₁₆ H ₂₈ N ₃ O ₅
y ₂	227.1752	1.0	C ₁₂ H ₂₃ N ₂ O ₂

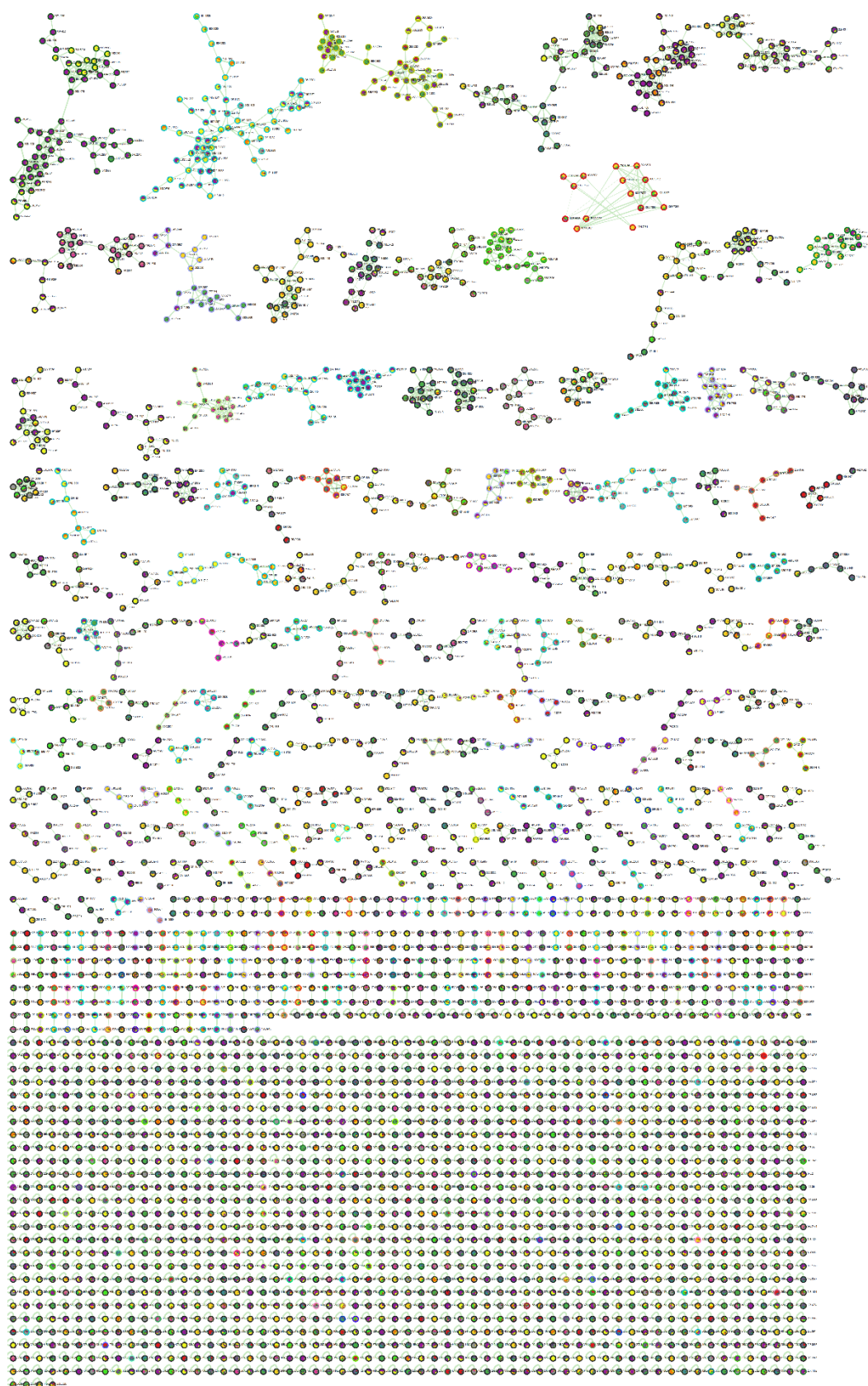


Figure S17: Molecular networks obtained using the Feature-Based Molecular Network workflow on GNPS (<https://gnps.ucsd.edu/ProteoSAFe/status.jsp?task=de168cddecfe40248fa3ba5077705e55>) (accessed on 23 May 2023).

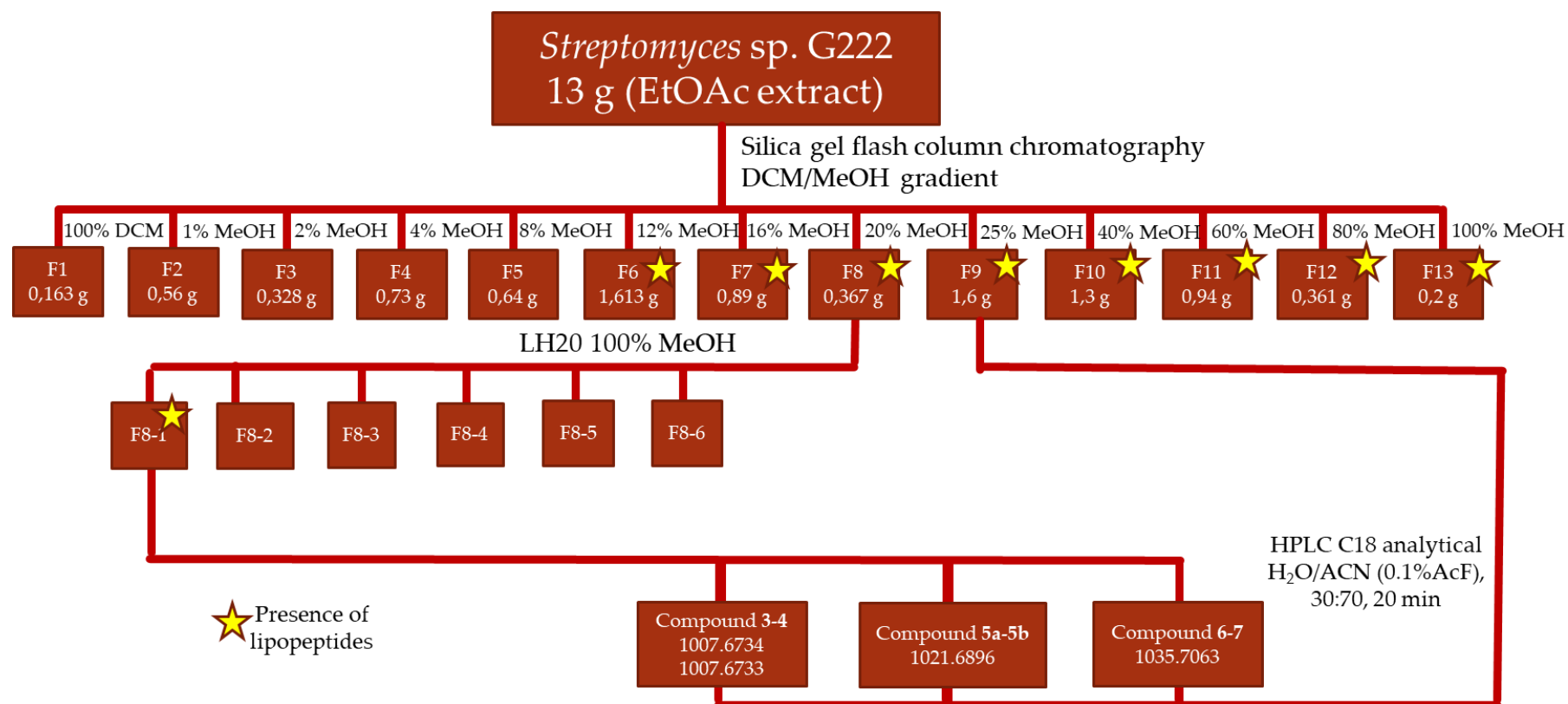


Figure S18: Chemical fractionation schema of *Streptomyces* sp. G222 crude extract.

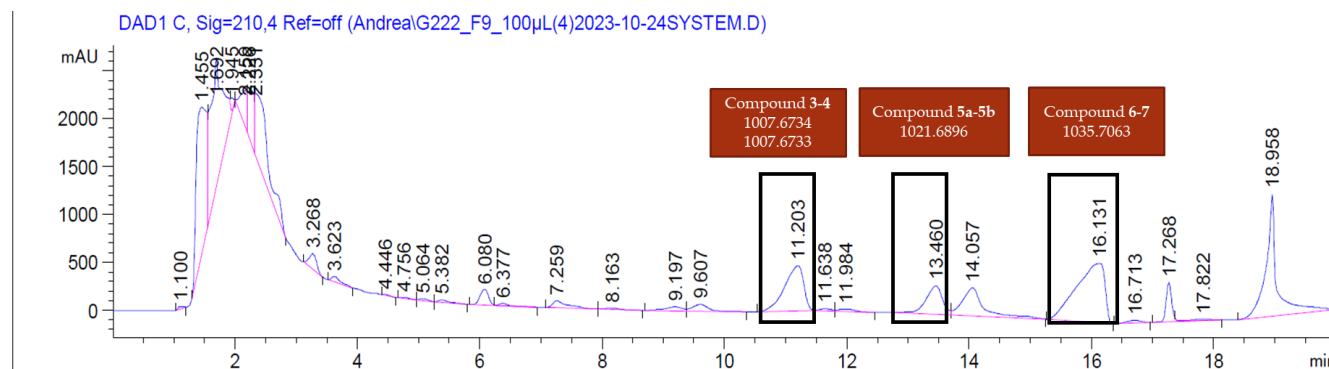


Figure S19: HPLC profile of the subfraction F9 at 210 nm.

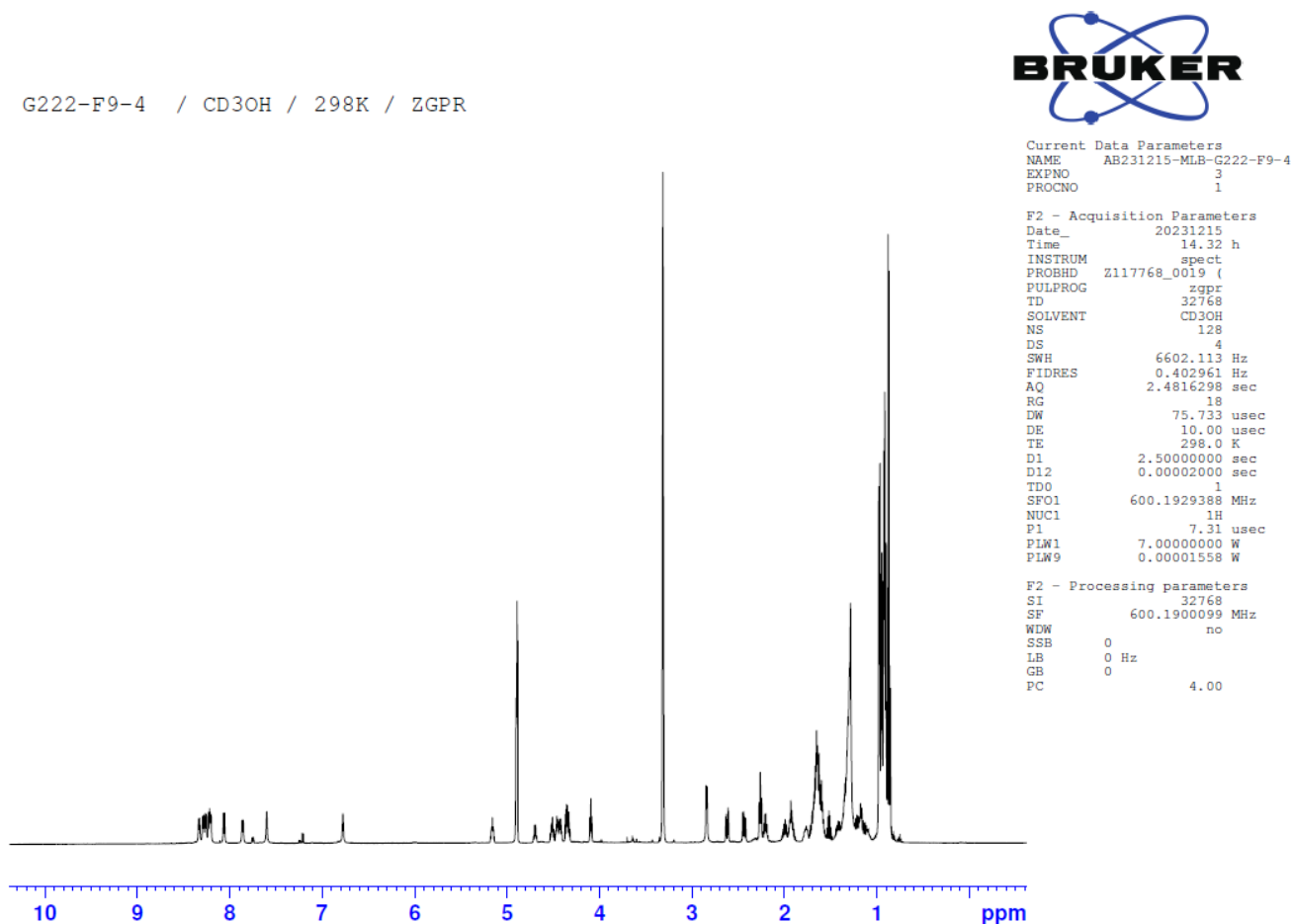


Figure S20: ^1H -NMR spectrum of Lichenysin (3-4) (m/z 1007.6734 and 1007.6733 $[\text{M} + \text{H}]^+$) (600 MHz, CD_3OH).

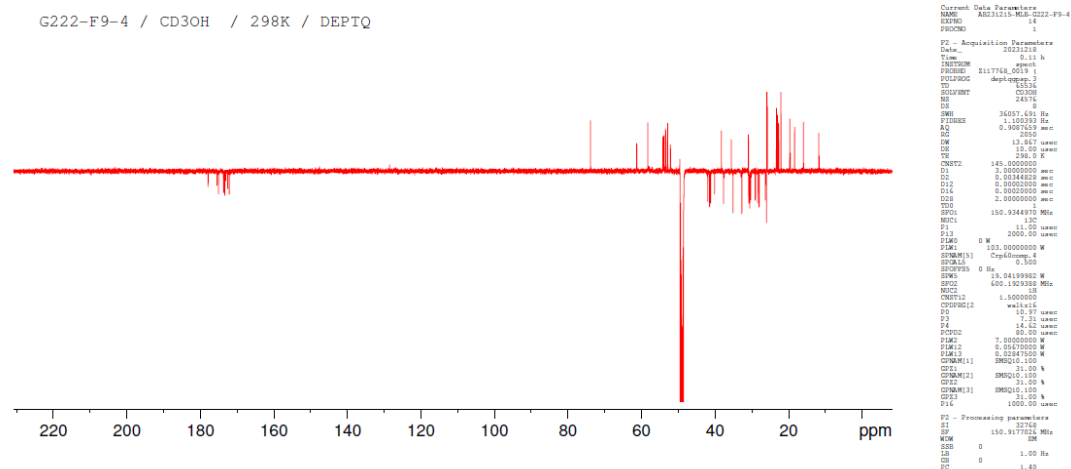


Figure S21: DEPTQ spectrum of Lichenysin (3-4) (m/z 1007.6734 and 1007.6733 $[M + H]^+$) (600 MHz, CD_3OH).

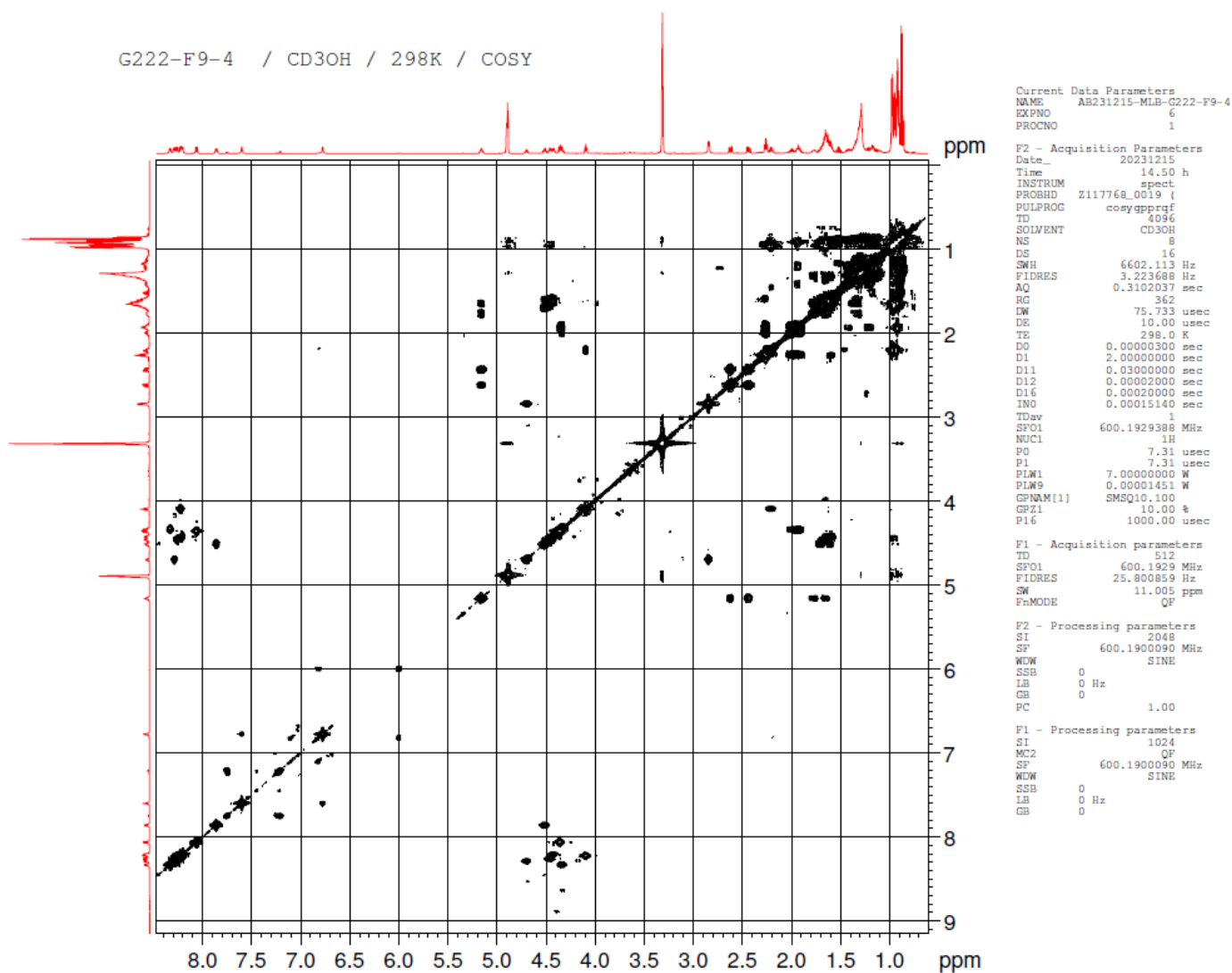


Figure S22: COSY-NMR spectrum of Lichenysin (3-4) (m/z 1007.6734 and 1007.6733 $[M + H]^+$) (600 MHz, CD_3OH).

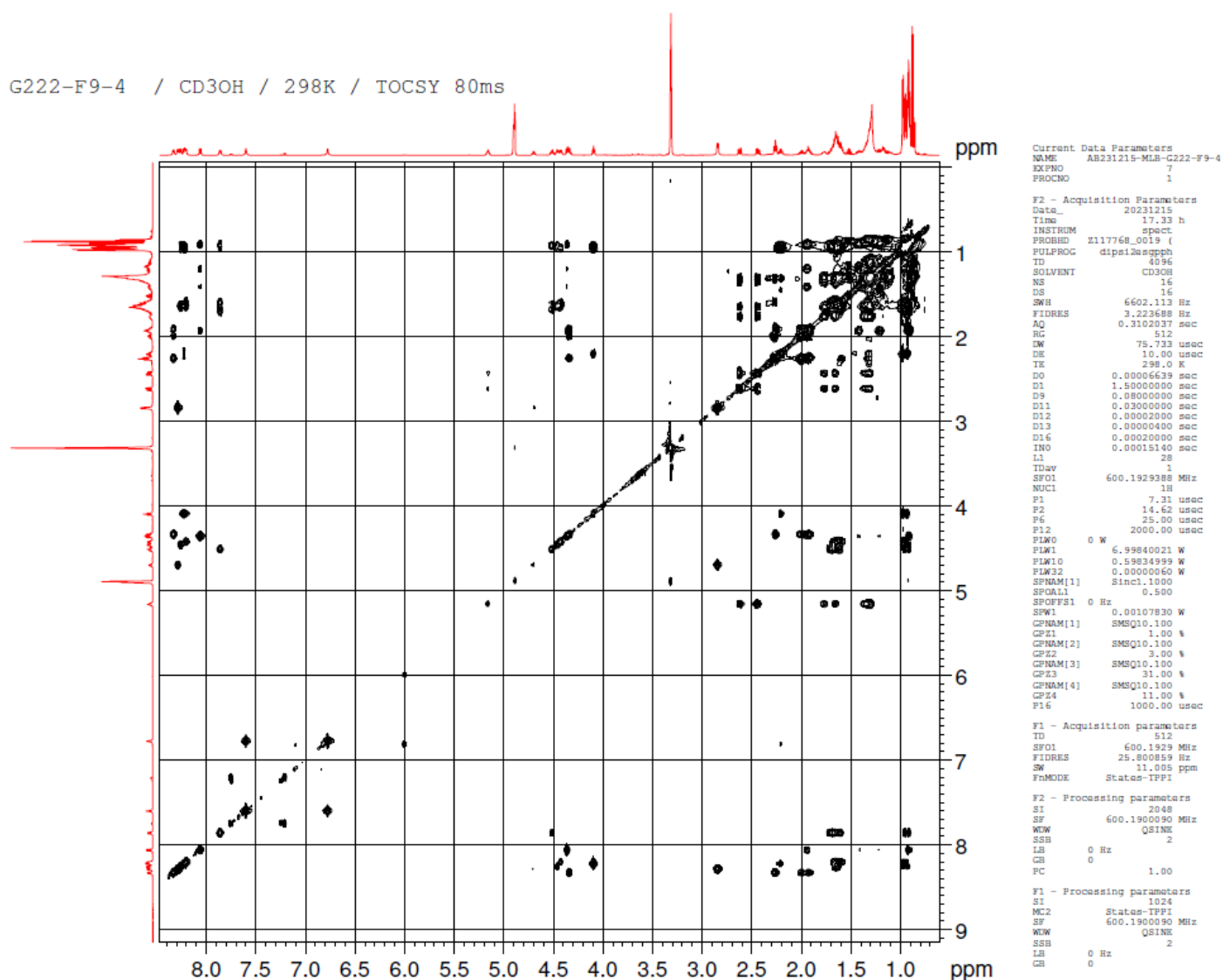


Figure S23: TOCSY-NMR spectrum of Lichenysin (3-4) (m/z 1007.6734 and 1007.6733 $[M + H]^+$) (600 MHz, CD_3OH).

G222-F9-4 / CD3OH / 298K / HSQCEDETGPSP.3

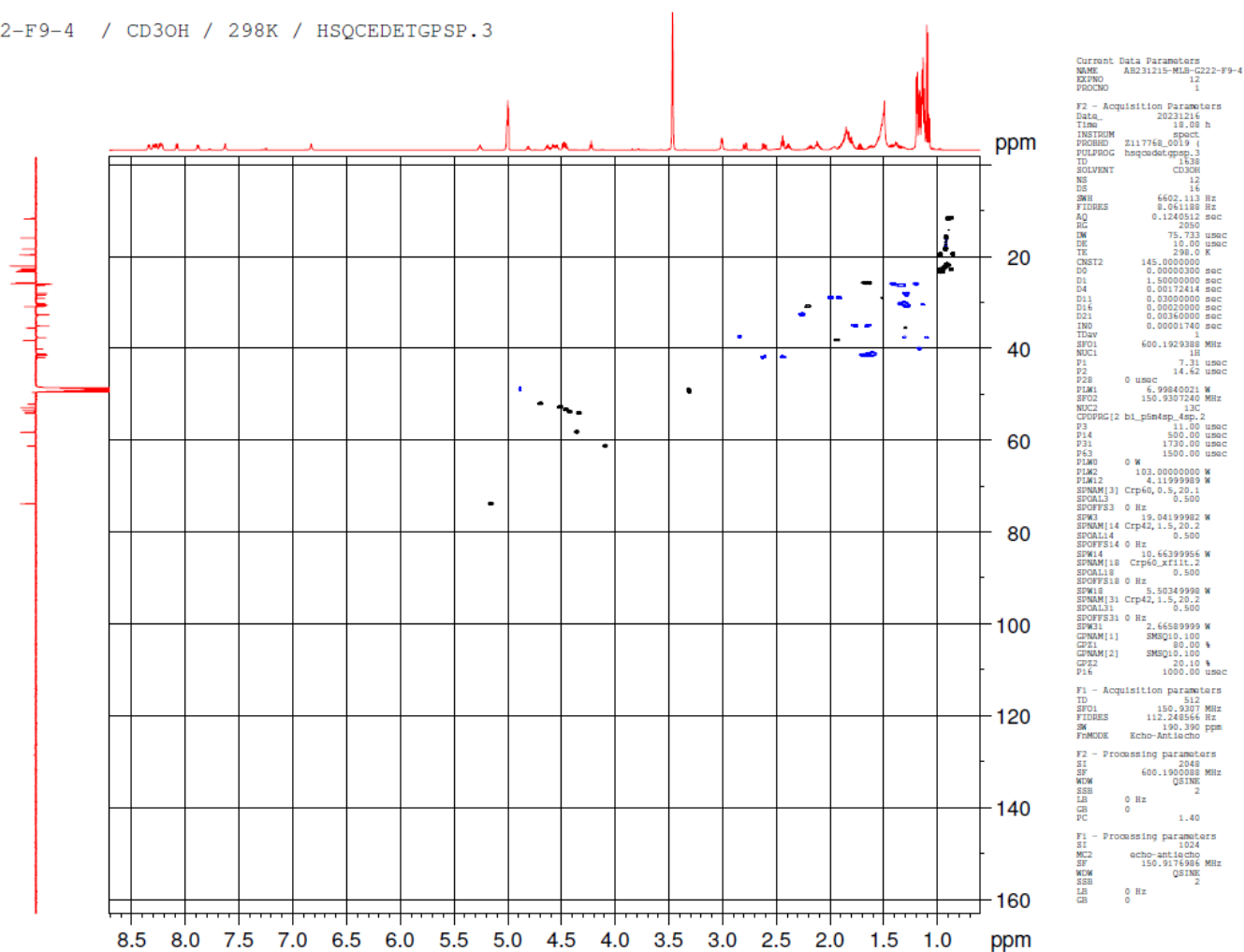


Figure S24: HSQC-NMR spectrum of Lichenysin (3-4) (m/z 1007.6734 and 1007.6733 $[M + H]^+$) (600 MHz, CD_3OH)

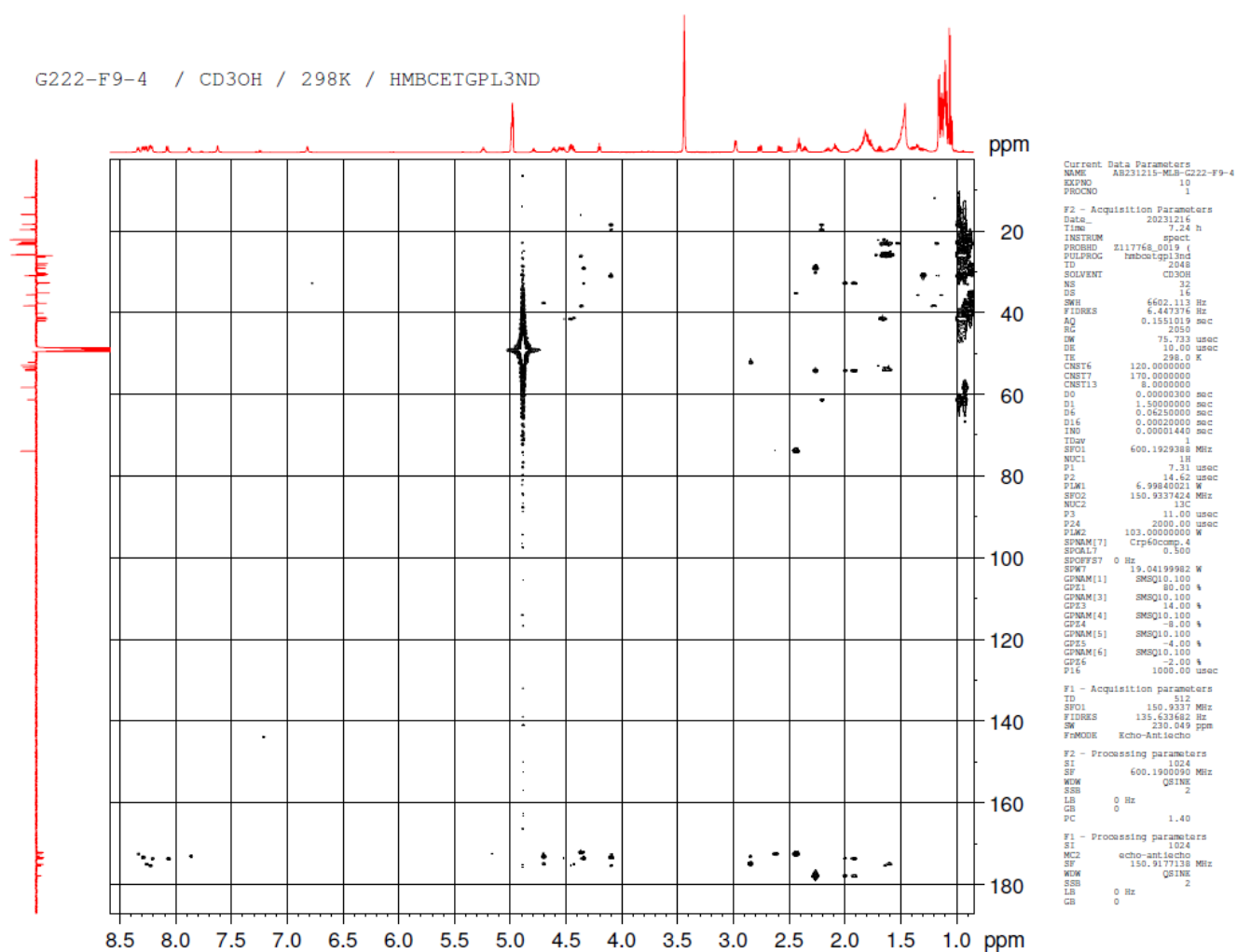


Figure S25: HMBC-NMR spectrum of Lichenysin (3-4) (m/z 1007.6734 and 1007.6733 $[M + H]^+$) (600 MHz, CD_3OH).

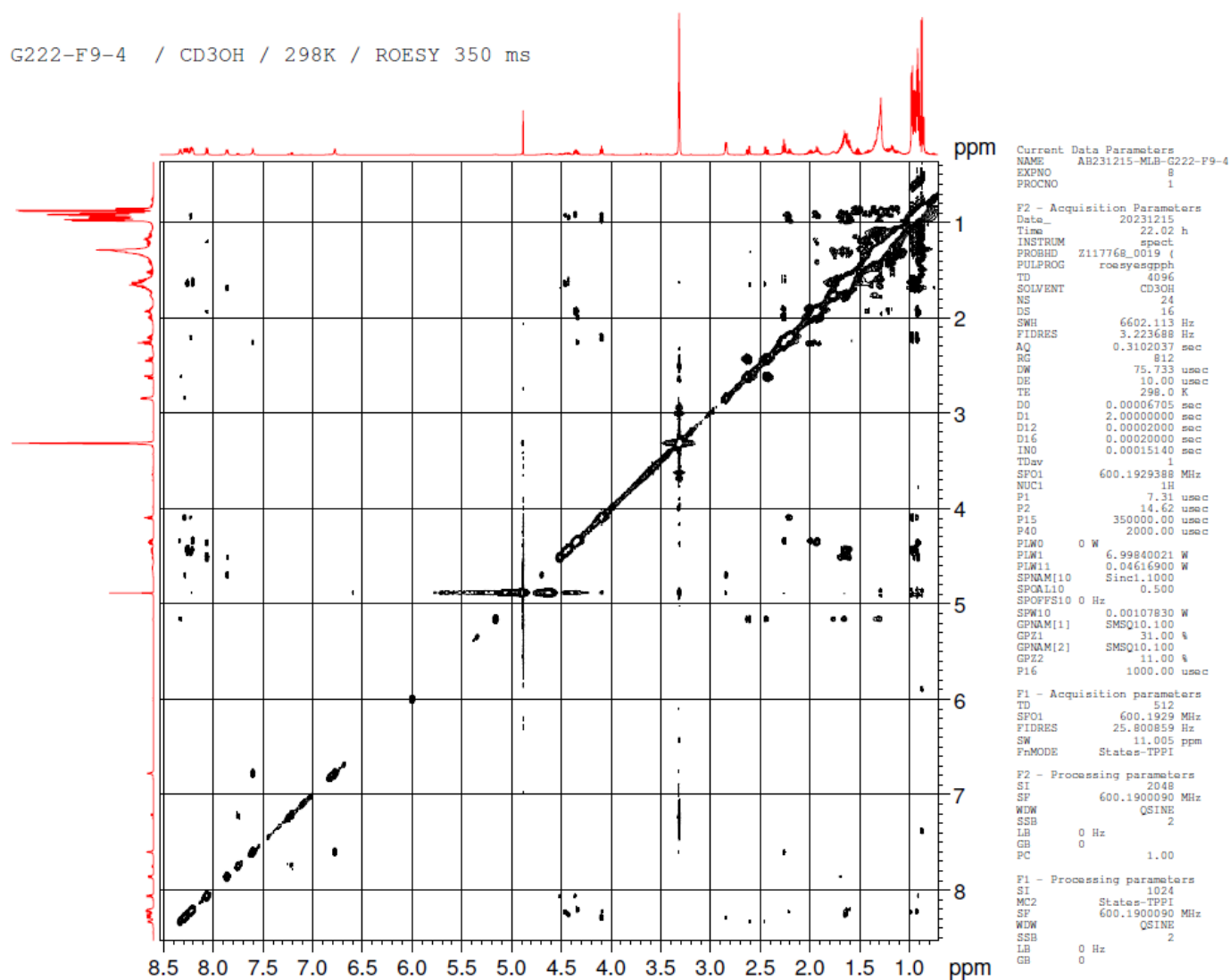


Figure S26: ROESY-NMR spectrum of Lichenysin (3-4) (m/z 1007.6734 and 1007.6733 $[M + H]^+$) (600 MHz, CD₃OH).

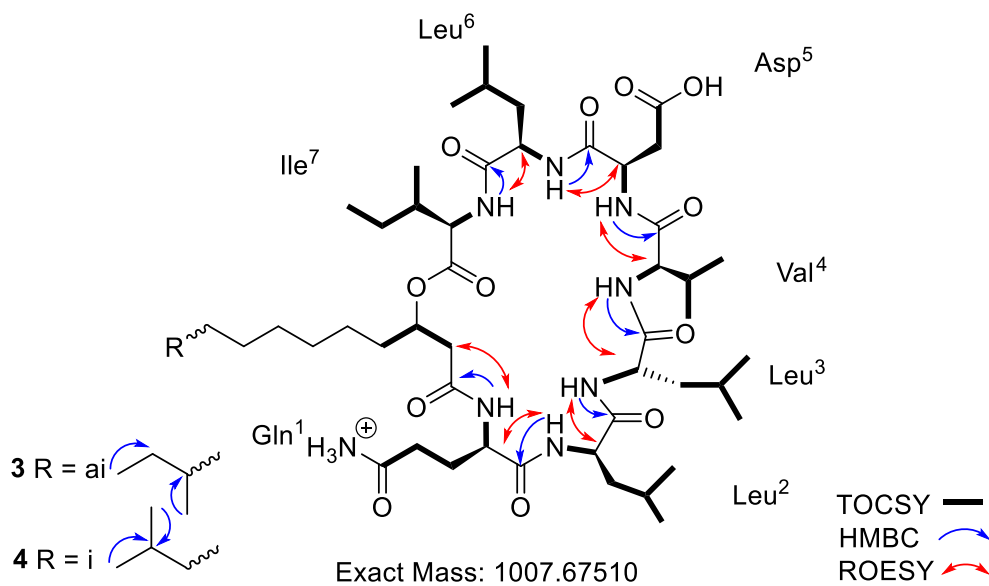


Figure S27: Structure of Lichenysins (3-4) (m/z 1007.6734 and 1007.6733 $[M + H]^+$; respectively).

Table S16. ^1H , ^{13}C , HMBC, and ROESY NMR data of Lichenysin (3-4) (m/z 1007.6734 and 1007.6733 $[\text{M} + \text{H}]^+$) (600 MHz, CD_3OH), isoform *ante-iso* and *iso* (0.8:0.2).

Unit	Pos.	δ_{C} , Mult.	δ_{H} (J in Hz)	HMBC	ROESY ^a
L-Gln ¹	NH	-	8.33, d (7.46)	Gln ¹ - α , FA-CO,	FA-2-a, FA-2-b, FA-3
	C=O	173.62, C	-	-	-
	α -C	54.15, CH	4.33, ddd (7.46, 9.93, 14.13)	Gln ¹ -CO, Gln ¹ - β , Gln ¹ - γ	Leu ² -NH
	β -C	29.05, CH ₂	1.99, m 1.91, m	Gln ¹ -CO, Gln ¹ - α , Gln ¹ - γ , Gln ¹ - δ -C=O	Leu ² -NH
	γ -C	32.75, CH ₂	2.26, t (7.46)	Gln ¹ - α , Gln ¹ - β , Gln ¹ - δ -C=O	Leu ² -NH
	δ -C=O	177.87, CO ²	-	-	-
	ϵ -NH ₂	-	7.61, brs 6.77, brs	Gln ¹ - δ -C=O, Gln ¹ - γ	-
L-Leu ²	NH	-	8.20, dd (7.57)	Gln ¹ -CO	Gln ¹ - α , Gln ¹ - β , Gln ¹ - γ
	C=O	175.09, C	-	-	-
	α -C	53.90, CH	4.42, ddd (5.93, 7.57, 14.06)	Leu ² -CO, Leu ² - β , Leu ² - γ	Leu ³ -NH
	β -C	41.17, CH ₂	1.64, m 1.61, m	Leu ² -CO, Leu ² - α , Leu ² - γ	Leu ³ -NH
	γ -C	25.87, CH	1.64, m	Leu ² - α , Leu ² - β , Leu ² - δ , Leu ² - δ'	Leu ³ -NH
	δ -C	22.98, CH ₃	0.97, d (4.13)	Leu ² - β , Leu ² - γ , Leu ² - δ'	-
	δ' -C	23.28, CH ₃	0.95, d (3.79)	Leu ² - β , Leu ² - γ , Leu ² - δ	-
D-Leu ³	NH	-	8.26, d (7.18)	Leu ² -CO, Leu ³ - α	Leu ² - α , Leu ² - β , Leu ² - γ
	C=O	175.52, C	-	-	-
	α -C	53.44, CH	4.45, ddd (4.31, 7.18, 12.09)	Leu ³ -CO, Leu ³ - β , Leu ³ - γ	Val ⁴ -NH
	β -C	41.44, CH ₂	1.61, m 1.64, m	Leu ³ -CO, Leu ³ - α , Leu ³ - γ	Val ⁴ -NH
	γ -C	25.70, CH	1.64, m	Leu ³ - α , Leu ³ - β	Val ⁴ -NH
	δ -C	22.07, CH ₃	0.91, d (4.31)	Leu ³ - β , Leu ³ - γ , Leu ³ - δ'	-
	δ' -C	23.26, CH ₃	0.97, d (4.31)	Leu ³ - β , Leu ³ - γ , Leu ³ - δ	-
L-Val ⁴	NH	-	8.22, d (7.32)	Val ⁴ - α , Val ⁴ - β , Leu ³ -CO	Leu ³ - α , Leu ³ - β , Leu ³ - γ
	C=O	173.35 C	-	-	-
	α -C	61.26, CH	4.09, dd (5.54, 7.32)	Val ⁴ -CO, Val ⁴ - β , Val ⁴ - γ , Val ⁴ - γ'	Asp ⁵ -NH
	β -C	30.89, CH	2.20, ddd (5.54, 7.32, 13.92)	Val ⁴ - α , Val ⁴ - γ , Val ⁴ - γ'	Asp ⁵ -NH
	γ -C	19.62, CH ₃	0.97, d (5.54)	Val ⁴ - β , Val ⁴ - γ'	Asp ⁵ -NH
	γ' -C	18.31, CH ₃	0.92, d (5.54)	Val ⁴ - β , Val ⁴ - γ	Asp ⁵ -NH
L-Asp ⁵	NH	-	8.28, d (7.46)	Val ⁴ -CO, Asp ⁵ - α	Val ⁴ - α , Val ⁴ - β , Val ⁴ - γ , Val ⁴ - γ' , Leu ⁶ -NH
	C=O	173.11, C	-	-	-
	α -C	52.11, CH	4.70, dd (7.46, 13.76)	Asp ⁵ -CO, Asp ⁵ - β , Asp ⁵ - γ -C=O	Leu ⁶ -NH
	β -C	37.64, CH ₂	2.84, d (7.46)	Asp ⁵ -CO, Asp ⁵ - α , Asp ⁵ - γ -C=O	Leu ⁶ -NH
	γ -C=O	175.07, CO ²	-	-	-
D-Leu ⁶	NH	-	7.85, d (8.32)	Asp ⁵ -CO	Asp ⁵ - α , Asp ⁵ -NH, Ile ⁶ -NH
	C=O	173.60, C	-	-	-
	α -C	52.91, CH	4.51, ddd (5.21, 8.32, 14.15)	Leu ⁶ -CO, Leu ⁶ - β , Leu ⁶ -CO, Leu ⁶ - α , Leu ⁶ - γ	Ile ⁷ -NH
	β -C	41.49, CH ₂	1.70, m 1.60, m	Leu ⁶ - α , Leu ⁶ - β	Ile ⁷ -NH
	γ -C	25.86, CH	1.64, m	Leu ⁶ - α , Leu ⁶ - β	Ile ⁷ -NH
	δ -C	22.07, CH ₃	0.91, d (1.68)	Leu ⁶ - β , Leu ⁶ - δ	-
	δ' -C	22.65, CH ₃	0.94, d (1.89)	Leu ⁶ - β , Leu ⁶ - δ'	-

Unit	Pos.	δ_C , Mult.	δ_H (J in Hz)	HMBC	ROESY ^a
L-Ile ⁷	NH	-	8.07, d (8.47)	Leu ⁶ -CO	Leu ⁶ -NH, Leu ⁶ - α , Leu ⁶ - β , Leu ⁶ - γ
	C=O	172.13, C	-	-	-
	α -C	58.30, CH	4.36, dd (5.29, 8.47)	Ile ⁷ -CO, Ile ⁷ - β , Ile ⁷ - γ , Ile ⁷ - δ	-
	β -C	38.24, CH	1.93, q (8.47)	Ile ⁷ - β , Ile ⁷ - γ , Ile ⁷ - δ , Ile ⁷ - δ'	-
	γ -C	25.99, CH ₂ a. b	1.41, m 1.20, m	Ile ⁷ - β , Ile ⁷ - δ , Ile ⁷ - δ'	-
	δ -C	15.93, CH ₃	0.92, d (3.66)	Ile ⁷ - α , Ile ⁷ - β , Ile ⁷ - γ	-
	δ' -C	11.77, CH ₃	0.89, t (3.12)	Ile ⁷ - β , Ile ⁷ - γ	-
Fatty acid	C=O	172.55, C	-	-	-
	2	41.96, CH ₂ a. b	2.62, dd (5.02, 12.97) 2.43, dd (7.46, 12.97)	FA-3, FA-4, FA-CO	Gln ¹ -NH
	3	73.83, CH	5.15 ddd (5.02, 7.46, 12.97)	FA-4, FA-CO	Gln ¹ -NH
	4	35.14, CH ₂ a. b	1.77, m 1.65, m	FA-2-a, FA-2-b, FA-3 FA-3, FA-5-9	- -
	ai	5	26.32, CH ₂ [*]	1.35 ^b , m	FA-5-9, FA-10, FA-11
ai	6	27.98, CH ₂ [*]	1.29 ^b , m	FA-5-9, FA-10, FA-11	-
ai	7	30.24, CH ₂ [*]	1.29 ^b , m	FA-5-9, FA-10, FA-11	-
ai	8	30.53, CH ₂ [*]	1.29 ^b , m	FA-5-9, FA-10, FA-11	-
ai	9	37.67, CH ₂ a. b	1.09, m 1.29, m	FA-5-9, FA-10, FA-11	-
ai	10	35.54, CH	1.29 ^b , m	FA-11, FA-5-9	-
ai	11	30.53, CH ₂ a. b	1.29 ^b , m 1.13 ^b , m	FA-10, FA-12, FA-13	-
ai	12	19.50, CH ₃	0.85, t (4.54)	FA-10	-
ai	13	11.63, CH ₃	0.86, d (5.32)	FA-9, FA-10, FA-11	-
i	5	26.33, CH ₂ [*]	1.35 ^b , m	FA-5-9, FA-10, FA-11	-
i	6	28.36, CH ₂ [*]	1.29 ^b , m	FA-5-9, FA-10, FA-11	-
i	7	30.28, CH ₂ [*]	1.29 ^b , m	FA-5-9, FA-10, FA-11	-
i	8	30.78, CH ₂ [*]	1.29 ^b , m	FA-5-9, FA-10, FA-11	-
i	9	26.32, CH ₂	1.29 ^b , m	FA-10, FA-12, FA-13	-
i	10	40.13, CH ₂	1.17 ^b , m	FA-11, FA-12, FA-13	-
i	11	29.02, CH	1.51, ept (6.65)	FA-10, FA-12, FA-13	-
i	12	22.91, CH ₃	0.87, d (6.65)	FA-9, FA-11, FA-13	-
i	13	22.91, CH ₃	0.87, d (6.65)	FA-9, FA-11, FA-12	-

^a Sequential ROEs ^b Overlapped signals prevent determination of constant couplings * May be interchanged.

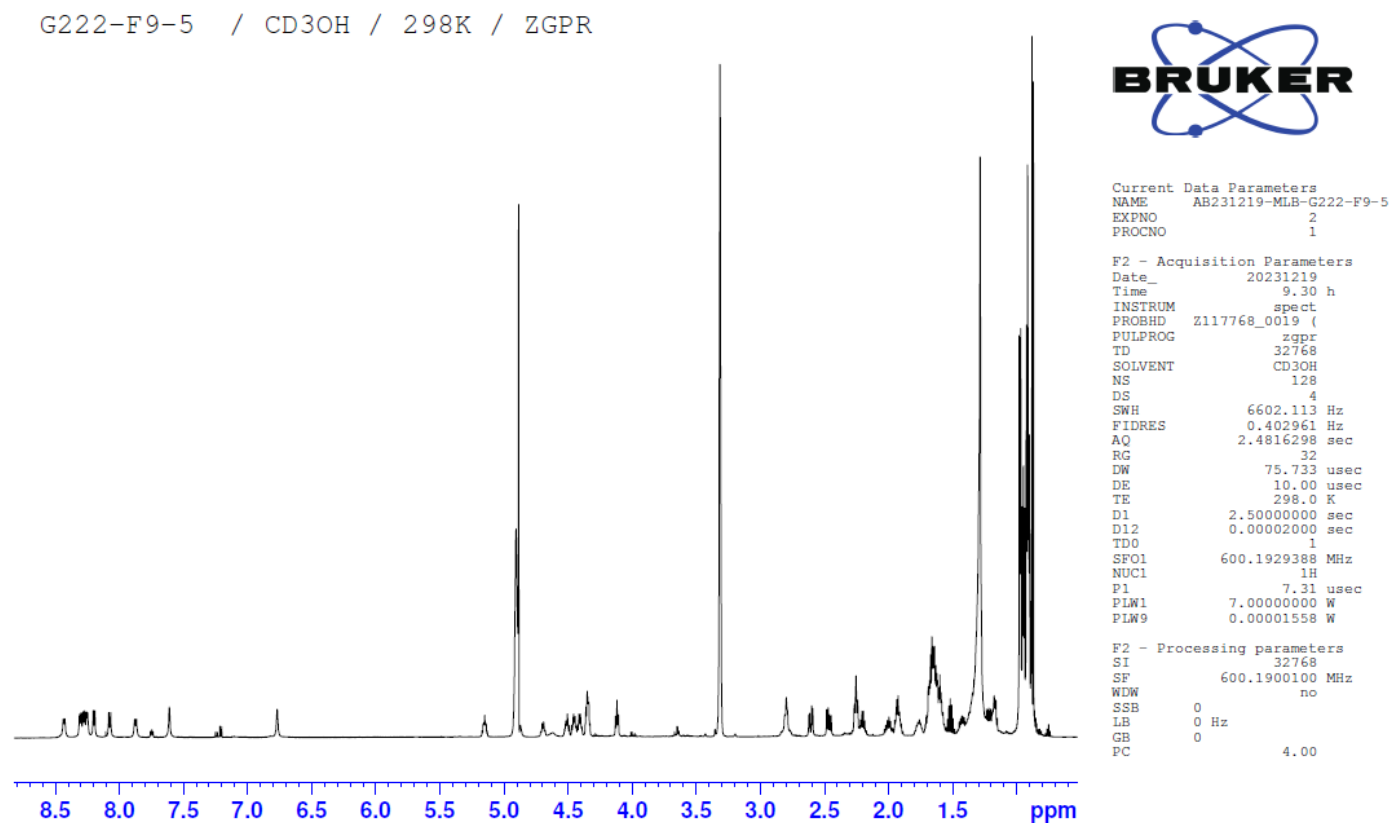


Figure S28: ^1H -NMR spectrum of Lichenysin (**5a** and **5b**) (m/z 1021.6896 $[\text{M} + \text{H}]^+$) (600 MHz, CD_3OH).

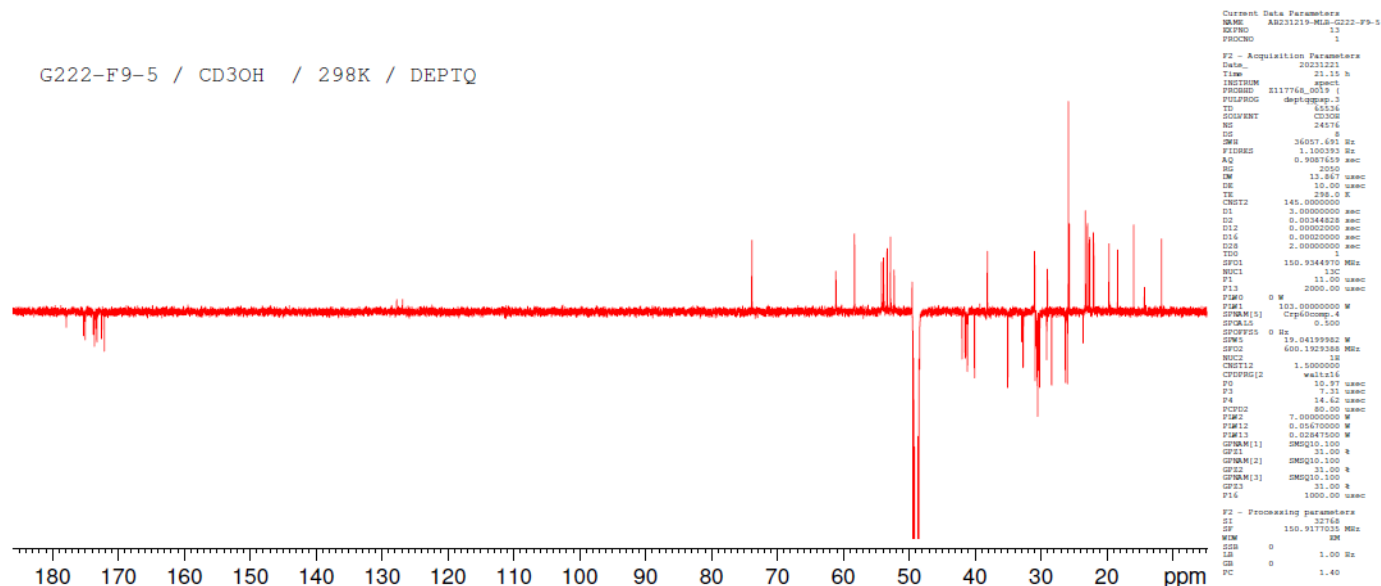


Figure S29: DEPTQ spectrum of Lichenysin (**5a** and **5b**) (m/z 1021.6896 $[\text{M} + \text{H}]^+$) (600 MHz, CD_3OH).

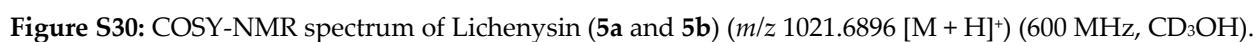


Figure S30: COSY-NMR spectrum of Lichenysin (**5a** and **5b**) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD_3OH).

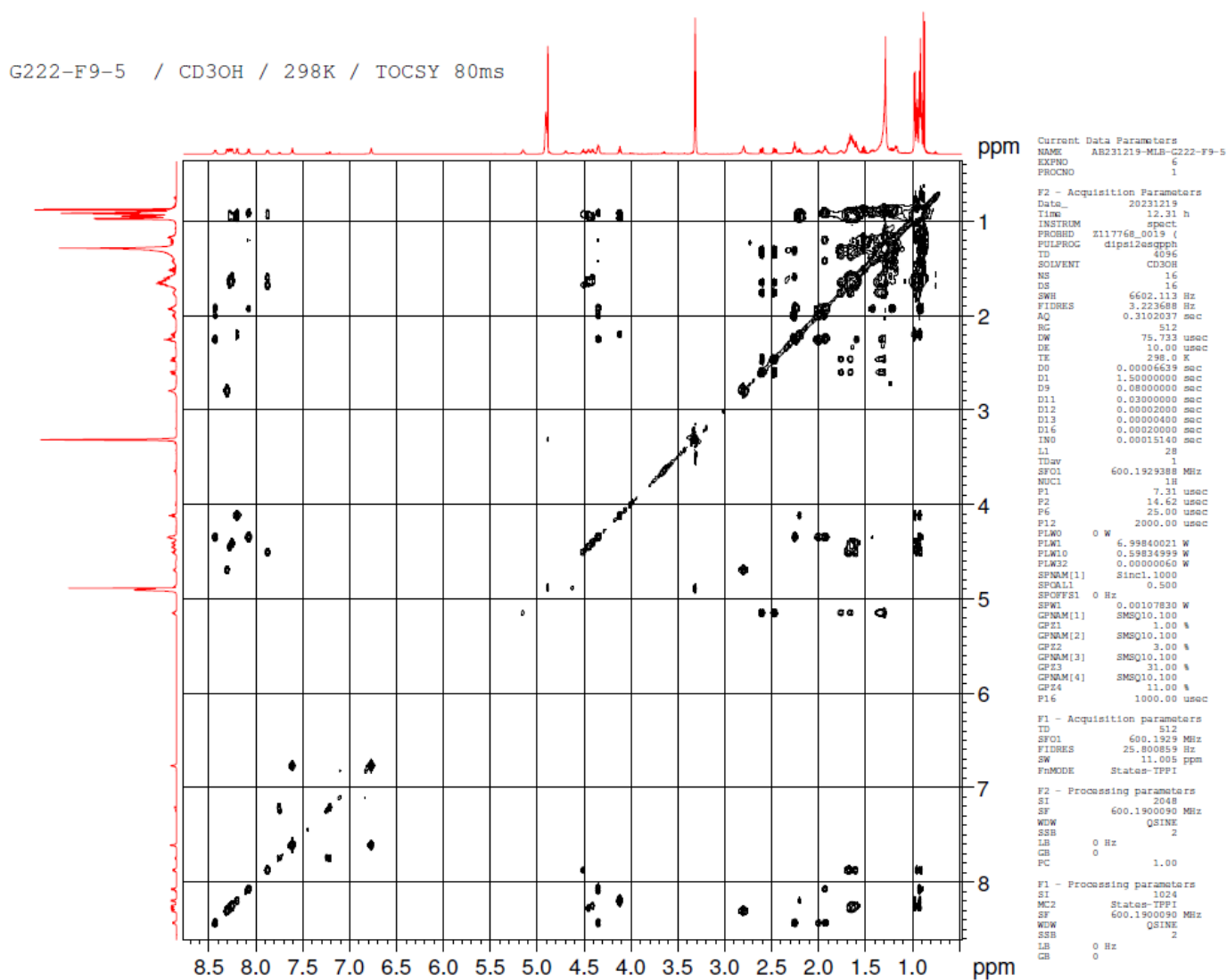


Figure S31: TOCSY-NMR spectrum of Lichenysin (5a and 5b) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD₃OH).

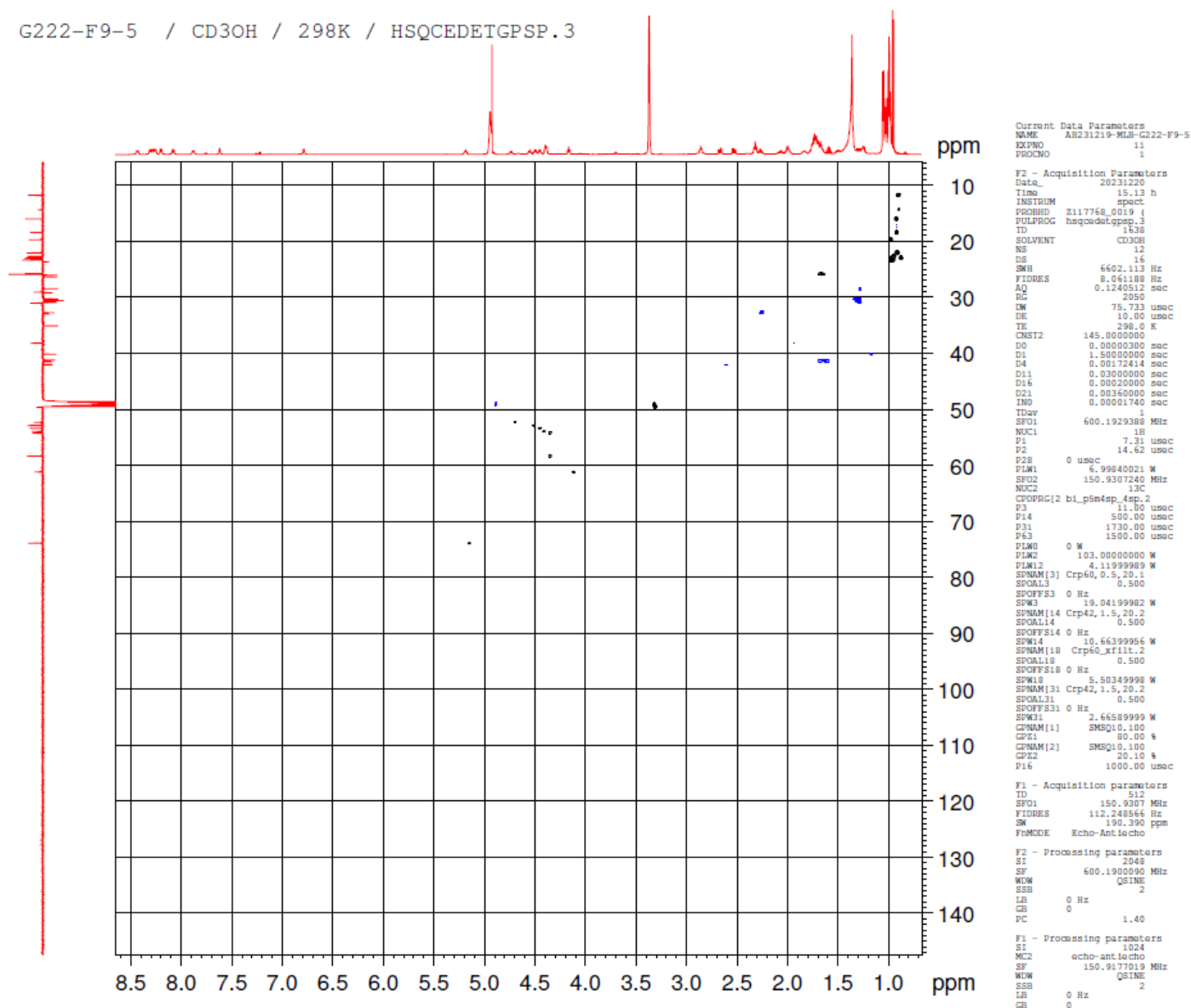


Figure S32: HSQC-NMR spectrum of Lichenysin (5a and 5b) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD₃OH).

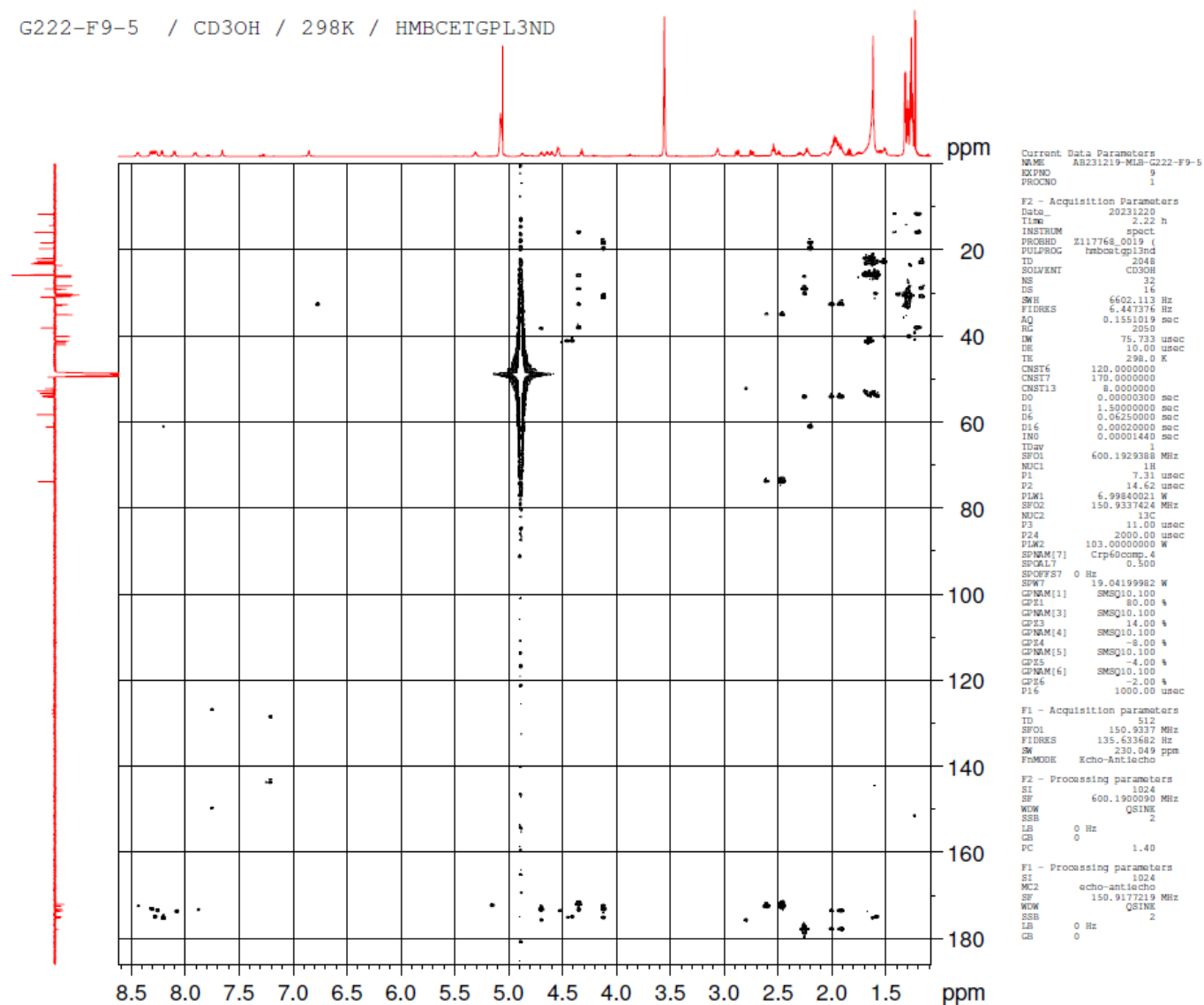


Figure S33: HMBC-NMR spectrum of Lichenysin (5a and 5b) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD₃OH).

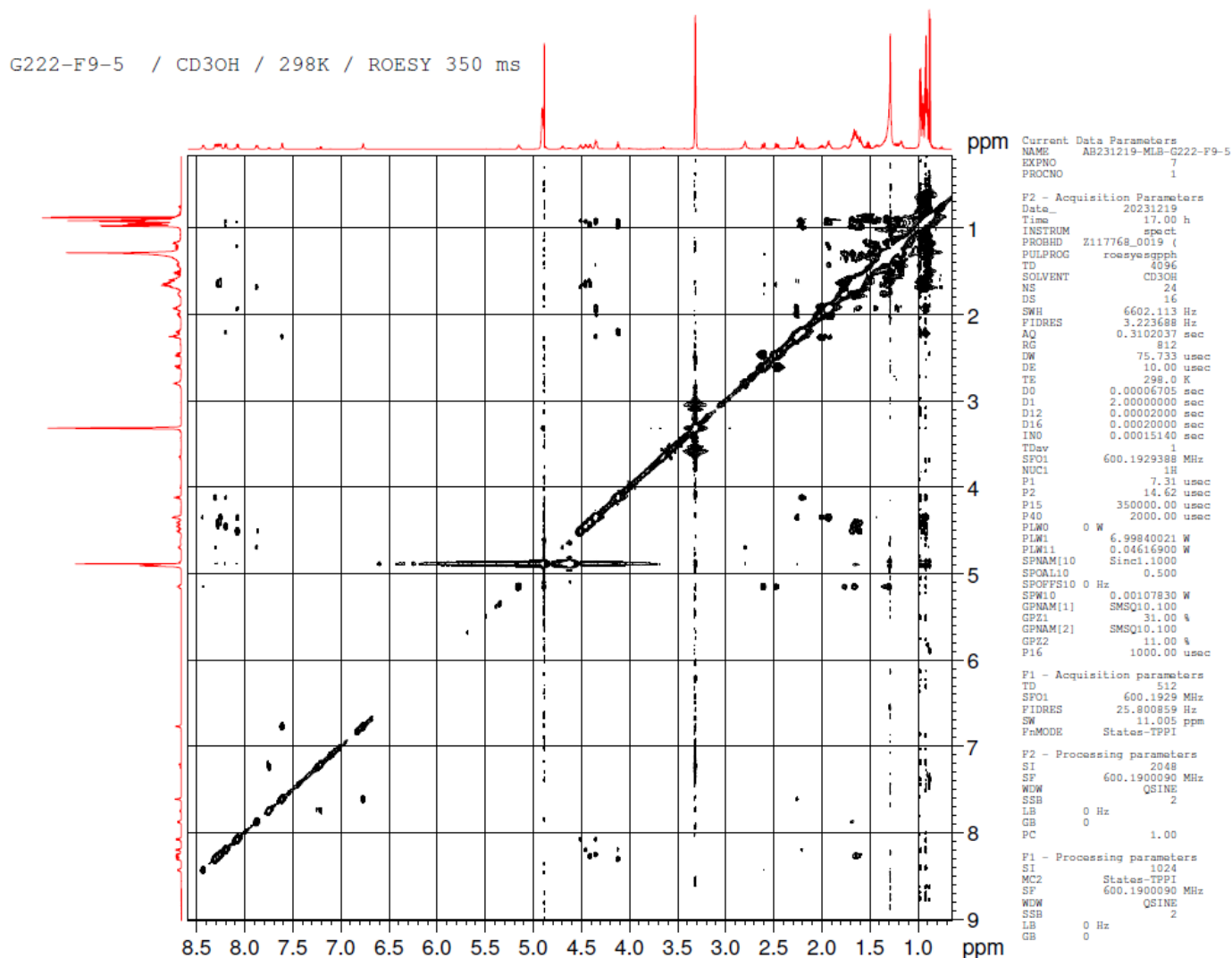


Figure S34: ROESY-NMR spectrum of Lichenysin (5a and 5b) (m/z 1021.6896 $[M + H]^+$) (600 MHz, CD₃OH).

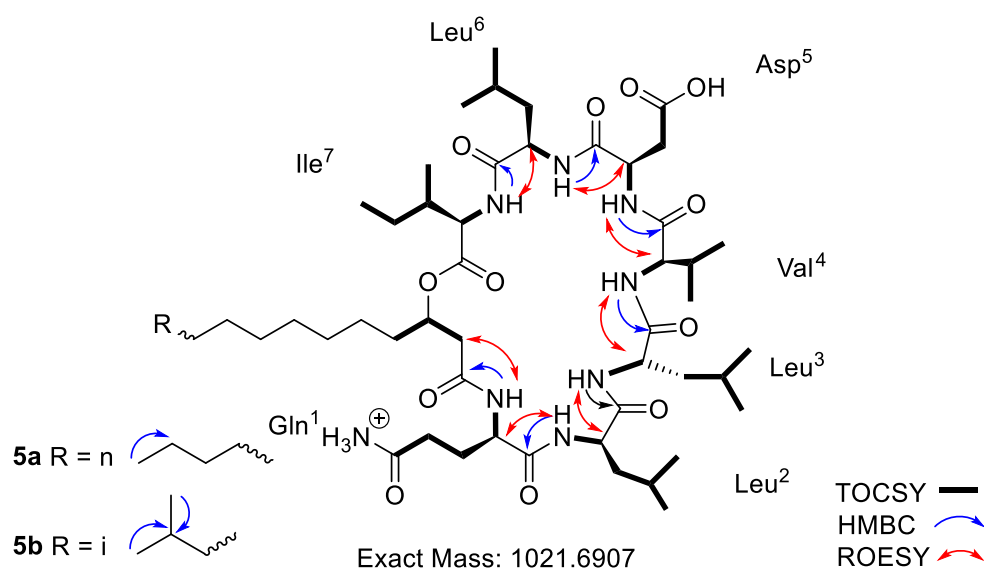


Figure S35: Structure of Lichenysins (5a and 5b) (m/z 1021.6896 $[M + H]^+$).

Table S17. ^1H , ^{13}C , HMBC, and ROESY NMR data of Lichenysin (**5a** and **5b**) (m/z 1021.6896 $[\text{M} + \text{H}]^+$) (600 MHz, CD_3OH) isoform: *iso* and *n* (0.5:0.5).

Unit	Pos.	δ_{C} , Mult.	δ_{H} (J in Hz)	HMBC	ROESY ^a
L-Gln ¹	NH	-	8.43, d (7.72)	FA-CO	FA-2-a, FA-2-b, FA-3
	C=O	173.63, C	-	-	-
	α -C	54.19, CH	4.34, ddd (4.72, 7.72, 10.13)	Gln ¹ -CO, FA-CO	Leu ² -NH
	β -C	29.13, CH ₂ a. b.	1.98, m 1.91, m	Gln ¹ -CO, Gln ¹ - α , Gln ¹ - γ , Gln ¹ - δ -C=O	Leu ² -NH
	γ -C	32.72, CH ₂	2.25, t (7.72)	Gln ¹ - α , Gln ¹ - β , Gln ¹ - δ -C=O	Leu ² -NH
	δ -C=O	177.89, CO ²	-	-	-
	ϵ -NH ₂	- a. b.	7.61, brs 6.80 brs	Gln ¹ - δ -C=O, Gln ¹ - γ	-
L-Leu ²	NH	-	8.24, d (5.82)	Leu ² - α , Leu ² - β , Gln ¹ - CO	Gln ¹ - α , Gln ¹ - β , Gln ¹ - γ
	C=O	175.01, C	-	-	-
	α -C	53.90, CH	4.41, ddd (2.26, 5.82, 7.62)	Leu ² -CO, Leu ² - β , Leu ² - γ , Gln ¹ -CO	Leu ³ -NH
	β -C	41.18, CH ₂ a. b.	1.63 m 1.59 m	Leu ² -CO, Leu ² - α , Leu ² - γ	Leu ³ -NH
	γ -C	25.87, CH	1.64, m	Leu ² - α , Leu ² - β , Leu ² - δ , Leu ² - δ'	Leu ³ -NH
	δ -C	23.27, CH ₃	0.95, d (2.26)	Leu ² - γ , Leu ² - δ'	-
	δ' -C	23.27, CH ₃	0.97, d (2.26)	Leu ² - γ , Leu ² - δ	-
D-Leu ³	NH	-	8.27, d (7.31)	Leu ² -CO, Leu ³ - α	Leu ² - α , Leu ² - β , Leu ² - γ
	C=O	175.23, C	-	-	-
	α -C	53.36, CH	4.45, ddd (5.08, 7.31, 11.85)	Leu ³ -CO, Leu ³ - β , Leu ³ - γ	Val ⁴ -NH
	β -C	41.24, CH ₂ a. b.	1.63, m 1.61, m	Leu ³ -CO, Leu ³ - α , Leu ³ - γ	Val ⁴ -NH
	γ -C	25.72, CH	1.64, m	Leu ³ - α , Leu ³ - β	Val ⁴ -NH
	δ -C	23.00, CH ₃	0.97, d (1.69)	Leu ³ - γ , Leu ³ - δ'	-
	δ' -C	22.05, CH ₃	0.91, d (1.69)	Leu ³ - γ , Leu ³ - δ	-
L-Val ⁴	NH	-	8.19, d (7.72)	Val ⁴ - α , Leu ³ -CO	Leu ³ - α , Leu ³ - β , Leu ³ - γ
	C=O	173.28, C	-	-	-
	α -C	61.17, CH	4.14, dd (7.72, 13.83)	Val ⁴ -CO, Val ⁴ - β , Val ⁴ - γ , Val ⁴ - γ'	Asp ⁵ -NH
	β -C	31.03, CH	2.20, ddd (3.85, 6.93, 11.80)	Val ⁴ - α , Val ⁴ - γ , Val ⁴ - γ'	Asp ⁵ -NH
	γ -C	19.69, CH ₃	0.97, d (1.89)	Val ⁴ - α , Val ⁴ - β , Val ⁴ - γ'	Asp ⁵ -NH
	γ' -C	18.41, CH ₃	0.92, d (1.89)	Val ⁴ - α , Val ⁴ - β , Val ⁴ - γ	Asp ⁵ -NH
L-Asp ⁵	NH	-	8.31, d (7.53)	Val ⁴ -CO	Val ⁴ - α , Val ⁴ - β , Val ⁴ - γ , Val ⁴ - γ' , Leu ⁶ -NH
	C=O	173.41, C	-	-	-
	α -C	52.30, CH	4.69, dd (7.53, 13.75)	Asp ⁵ -CO, Asp ⁵ - β , Asp ⁵ - γ -C=O	Leu ⁶ -NH

	β -C	38.31, CH ₂	2.81, dd (7.53, 13.75)	Asp ⁵ -CO, Asp ⁵ - α , Asp ⁵ - γ -C=O	Leu ⁶ -NH
	γ -C=O	175.82, CO ²	-	-	-
D-Leu ⁶	NH	-	7.88, d (8.69)	Asp ⁵ -CO	Asp ⁵ - α , Asp ⁵ -NH, Ile ⁶ -NH
	C=O	173.80, C	-	-	-
	α -C	52.89, CH	4.51 ddd (5.12, 8.69, 13.32)	Leu ⁶ -CO, Leu ⁶ - β ,	Ile ⁷ -NH
	β -C	41.50, CH ₂ ^{a.} ^b	1.68, m 1.61, m	Leu ⁶ -CO, Leu ⁶ - α , Leu ⁶ - γ	Ile ⁷ -NH
	γ -C	25.86, CH	1.64, m	Leu ⁶ - α , Leu ⁶ - β	Ile ⁷ -NH
	δ -C	21.98, CH ₃	0.91, d (1.68)	Leu ⁶ - β , Leu ³ - δ	-
	δ' -C	22.65, CH ₃	0.94, d (1.89)	Leu ⁶ - β , Leu ⁶ - δ'	-
L-Ile ⁷	NH	-	8.07, d (8.53)	Leu ⁶ -CO	Leu ⁶ -NH, Leu ⁶ - α , Leu ⁶ - β , Leu ⁶ - γ
	C=O	172.13, C	-	-	-
	α -C	58.35, CH	4.35, ddd (4.31, 8.53, 11.47)	Ile ⁷ -CO, Ile ⁷ - β , Ile ⁷ - γ , Ile ⁷ - δ	-
	β -C	38.13, CH	1.92, d (8.53)	Ile ⁷ -CO	-
	γ -C	25.99, CH ₂ ^{a.} ^b	1.43, m 1.20, m	Ile ⁷ - β , Ile ⁷ - δ , Ile ⁷ - δ'	-
	δ -C	15.96, CH ₃	0.92, d (4.31)	Ile ⁷ - α , Ile ⁷ - β	-
	δ' -C	11.72, CH ₃	0.90, t (2.33)	Ile ⁷ - β	-
Fatty acid	C=O	172.55, C	-	-	-
	2	41.99, CH ₂ ^{a.} ^b	2.61, dd 2.46, dd	FA-3, FA-4, FA-CO	Gln ¹ -NH
	3	73.83, CH	5.15 ddd (5.24, 6.89, 12.81)	FA-4, FA-CO	Gln ¹ -NH
	4	35.11, CH ₂ ^{a.} ^b	1.75, m 1.65, m	FA-2-a, FA-2-b, FA-3 FA-5-9, FA-10, FA-11	- -
	5	30.22, CH ₂	1.35 ^b , m	FA-5-9, FA-10, FA-11	-
	6	30.34, CH ₂	1.29 ^b , m	FA-5-9, FA-10, FA-11	-
	7	30.52, CH ₂	1.29 ^b , m	FA-5-9, FA-10, FA-11	-
	8	30.54, CH ₂	1.29 ^b , m	FA-5-9, FA-10, FA-11	-
	9	30.60, CH ₂	1.29 ^b , m	FA-5-9, FA-10, FA-11	-
i	10	26.35, CH ₂	1.29 ^b , m	FA-11, FA-5-9	-
i	11	40.02, CH ₂	1.16, m	FA-12	-
i	12	29.06, CH	1.51, m	FA-11	-
i	13	22.92, CH ₃	0.87, d (6.60)	FA-11, FA-12, FA-14	-
i	14	22.92, CH ₃	0.87, d (6.60)	FA-11, FA-12, FA-13	-
n	10	30.65, CH ₂	1.29 ^b , m	FA-11, FA-5-9	-
n	11	30.90, CH ₂	1.29 ^b , m	FA-12	-
n	12	29.06, CH ₂	1.29 ^b , m	FA-11	-
n	13	23.59, CH ₂	1.29 ^b , m	FA-11, FA-12, FA-14	-
n	14	14.32, CH ₃	0.90, t (6.60)	FA-11, FA-12, FA-13	-

^aSequential ROEs. ^bOverlapped signals prevent determination of constant couplings.

G222-F9-7 / CD3OH / 298K / ZGPR



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PROCNO 1

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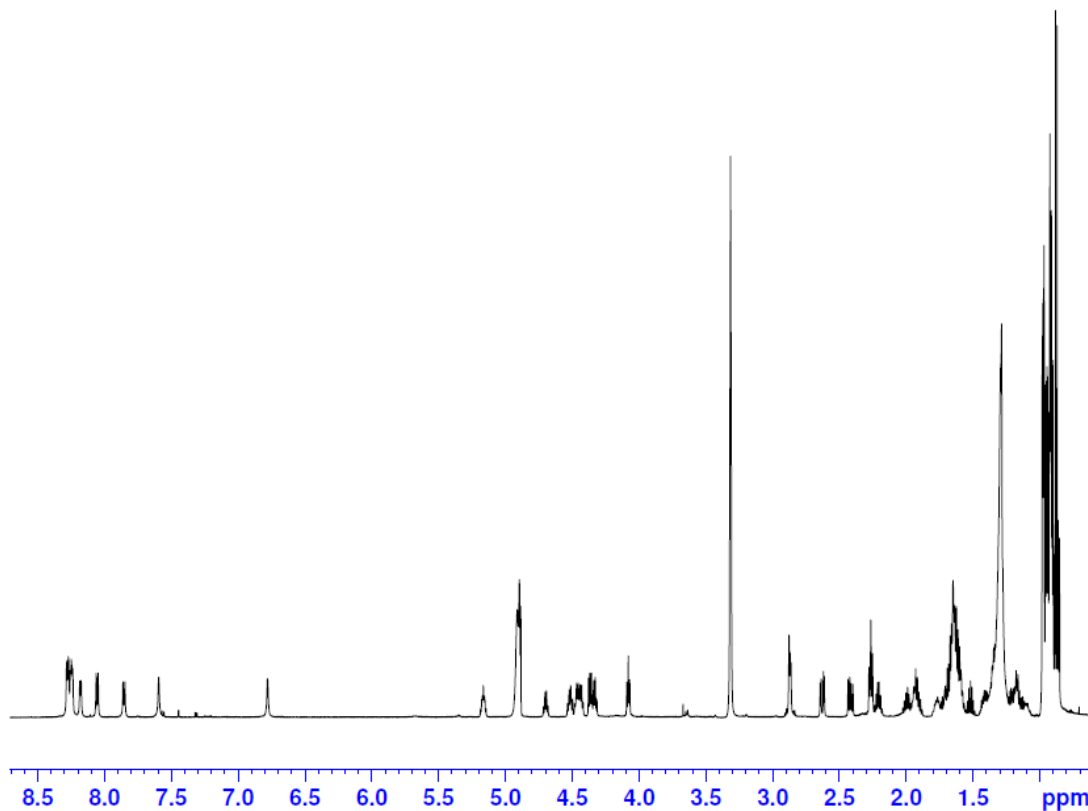


Figure S36: ^1H -NMR spectrum of Lichenysin (6-7) (m/z 1035.7063 $[\text{M} + \text{H}]^+$) (600 MHz, CD_3OH).

G222-F9-7 / CD3OH / 298K / DEPTQ

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PROCNO 1

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SFOFFS 0 Hz
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CHN112 1.50000000
CHN112 CPDPRG2 wait16
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P2 7.34 usec
P3 14.18 usec
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CP22 31.00 %
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CP23 31.00 %
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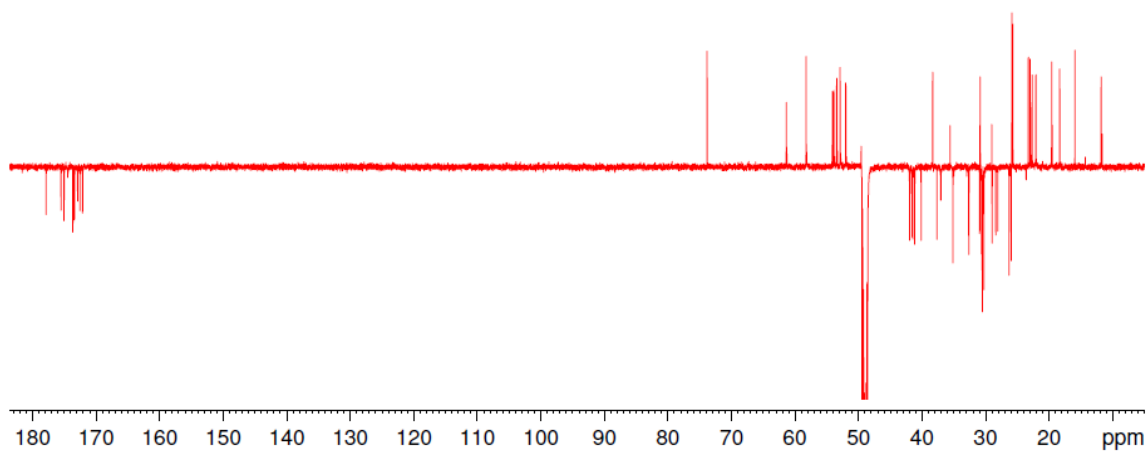


Figure S37: DEPTQ spectrum of Lichenysin (6-7) (m/z 1035.7063 $[\text{M} + \text{H}]^+$) (600 MHz, CD_3OH).

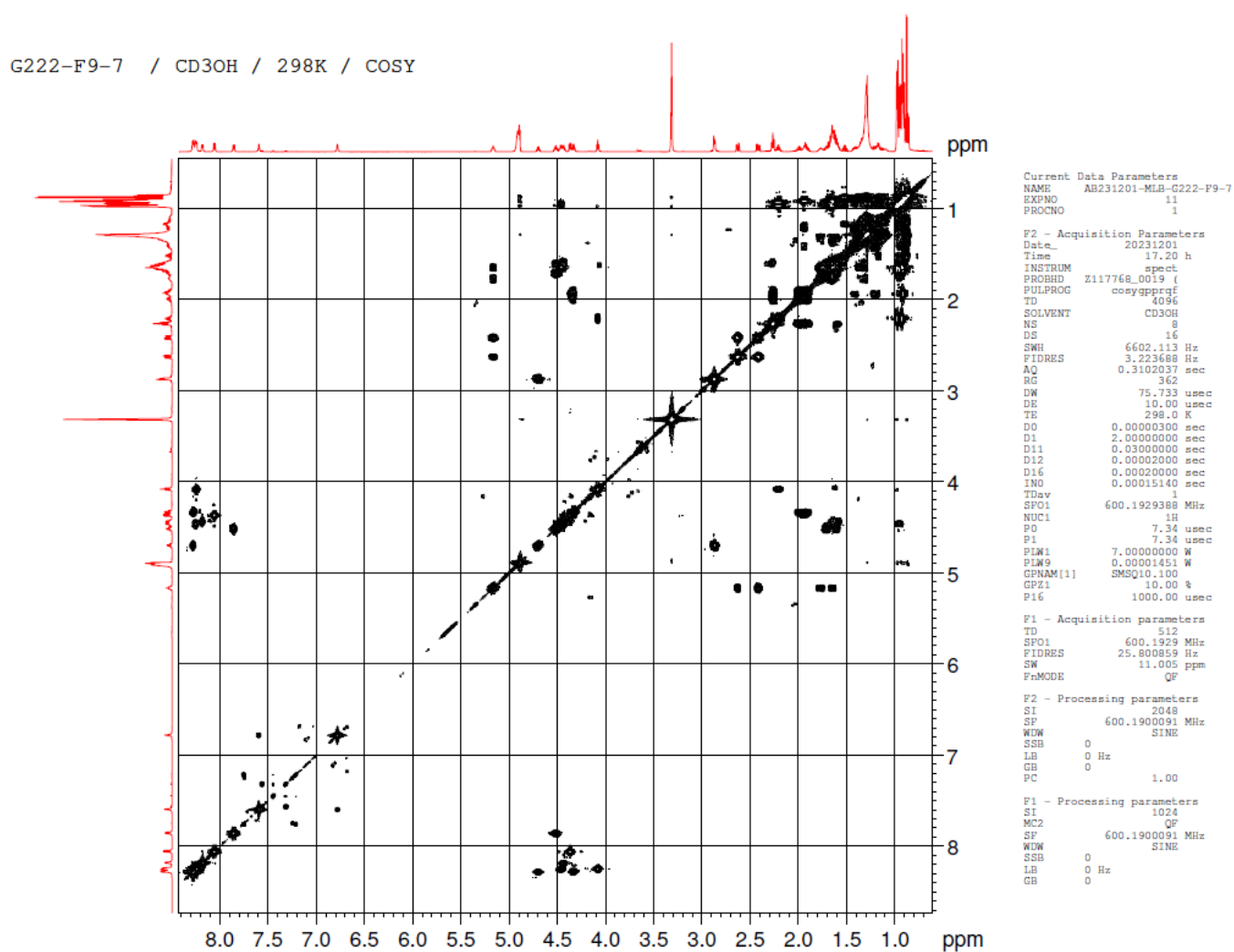


Figure S38: COSY-NMR spectrum of Lichenysin (6-7) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD_3OH).

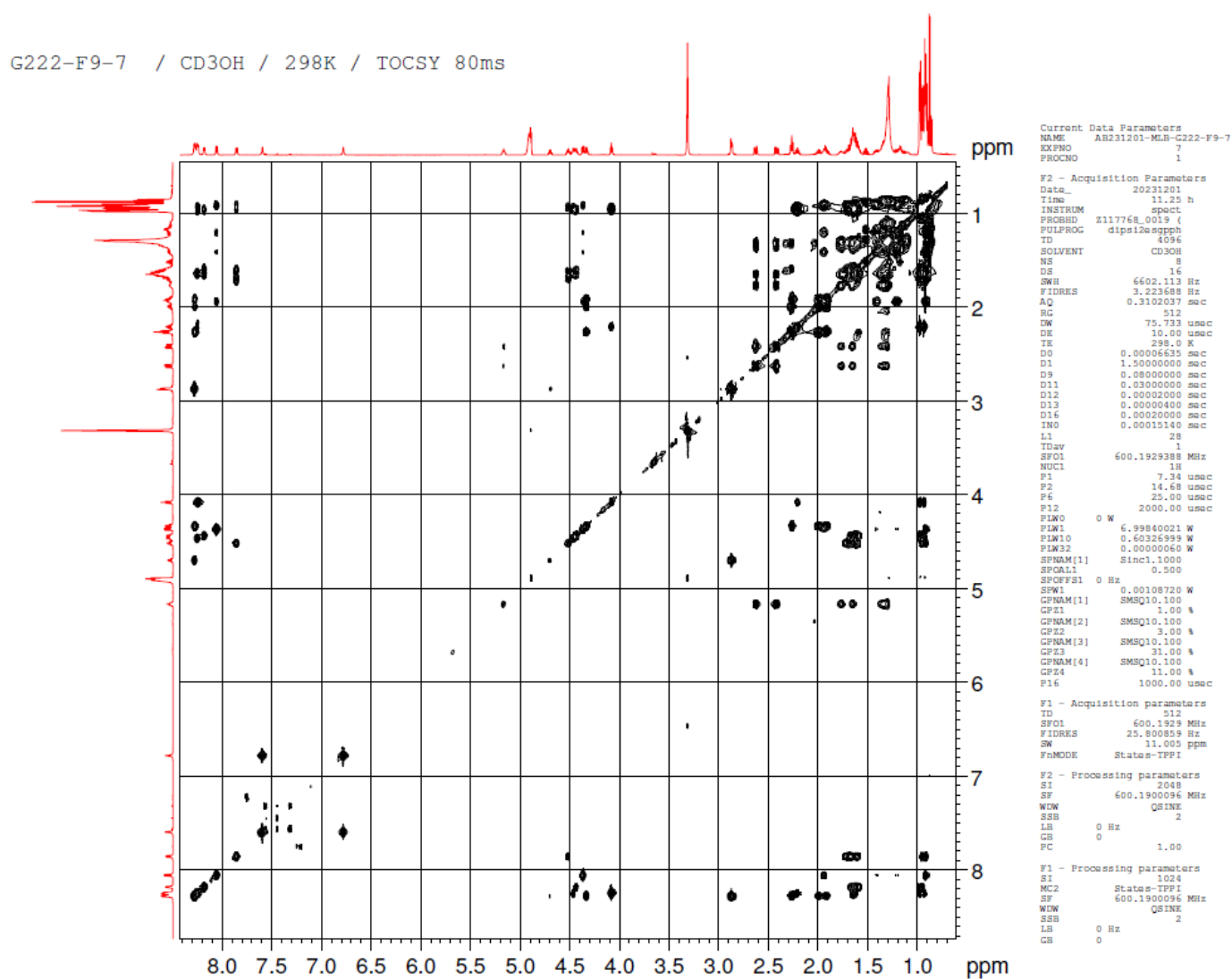


Figure S39: TOCSY-NMR spectrum of Lichenysin (6-7) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD₃OH).

G222-F9-7 / CD3OH / 298K / HSQCEDETGPSP.3

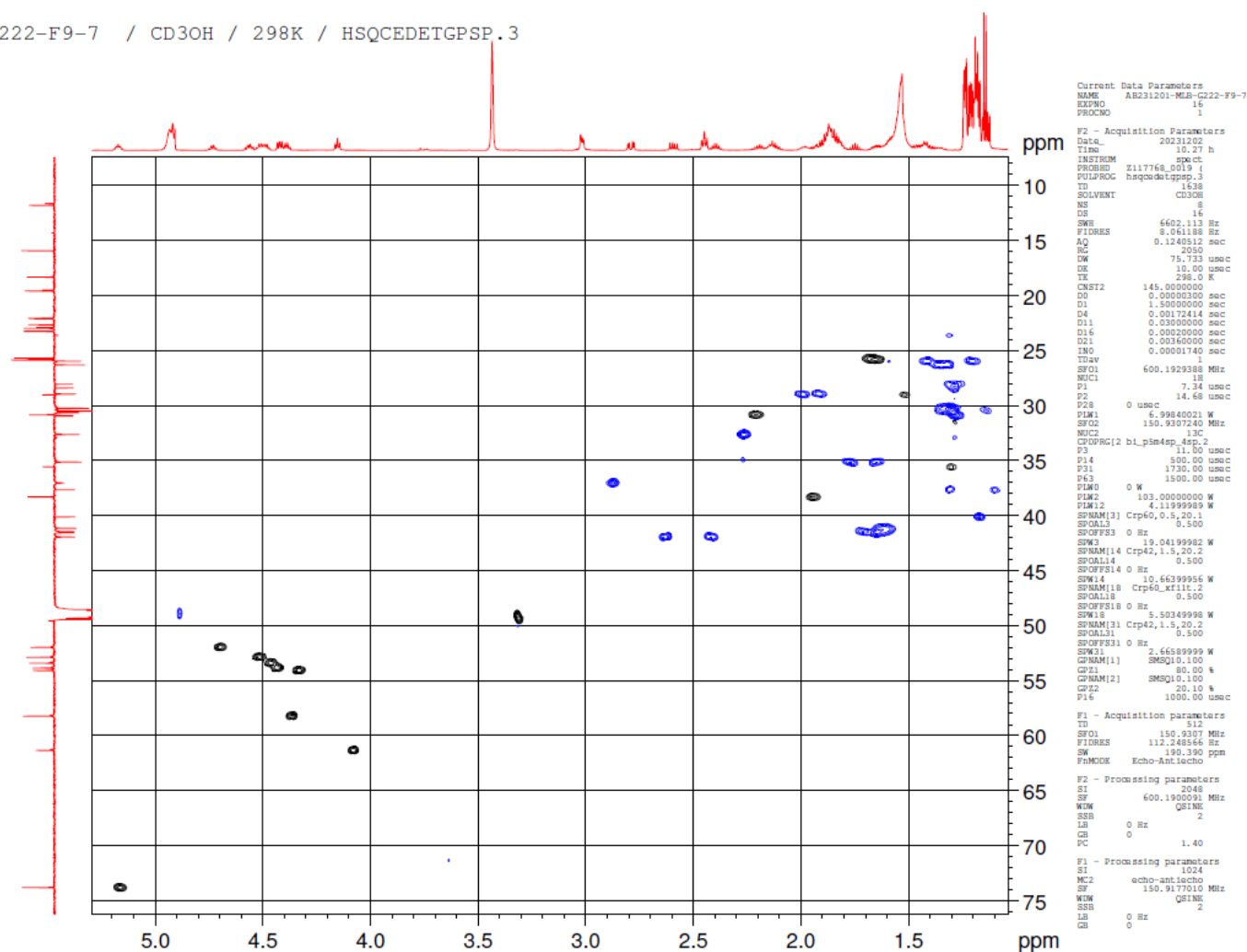


Figure S40: HSQC-NMR spectrum of Lichenysin (6-7) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD_3OH).

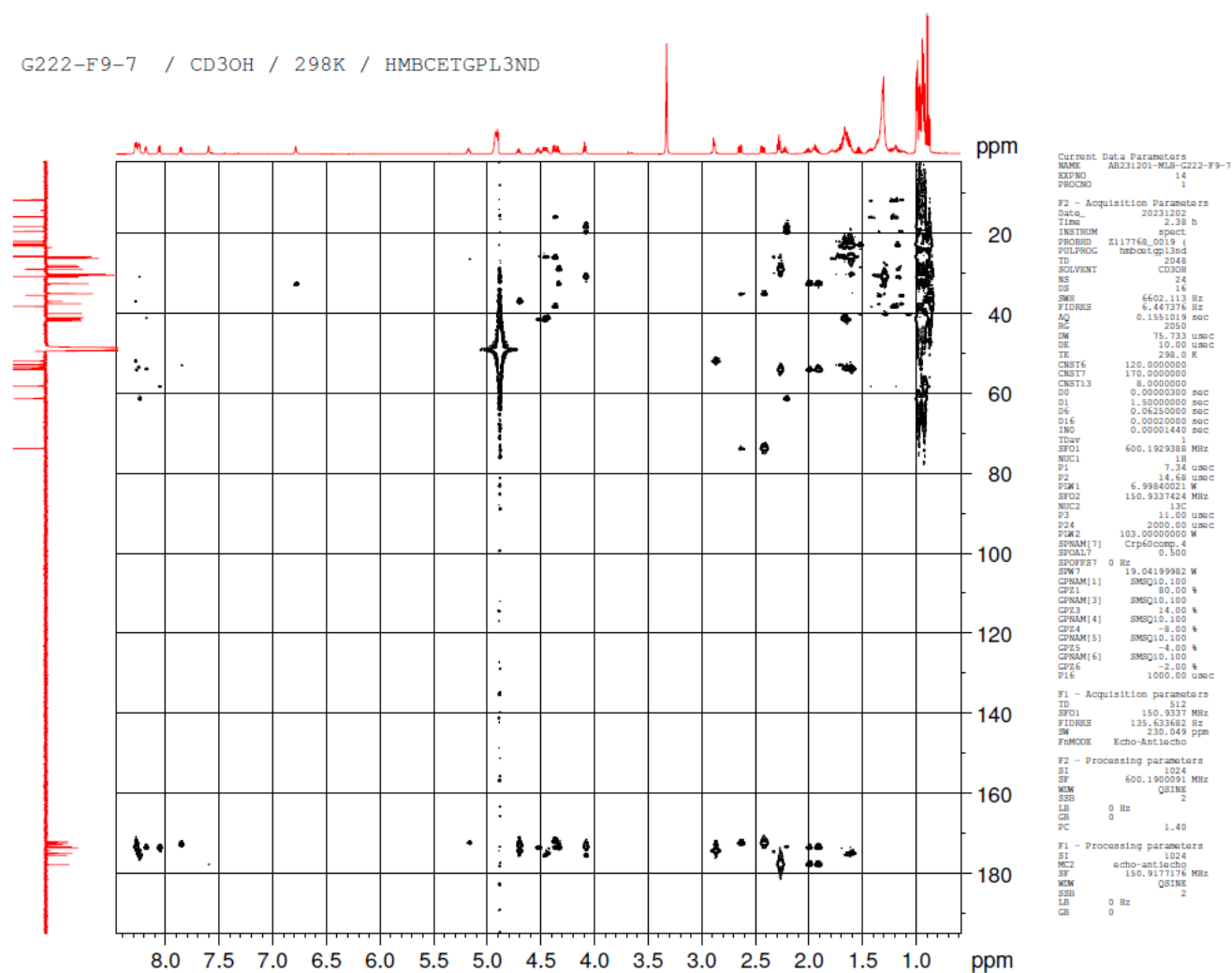


Figure S41: HMBC-NMR spectrum of Lichenysin (6-7) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD_3OH).

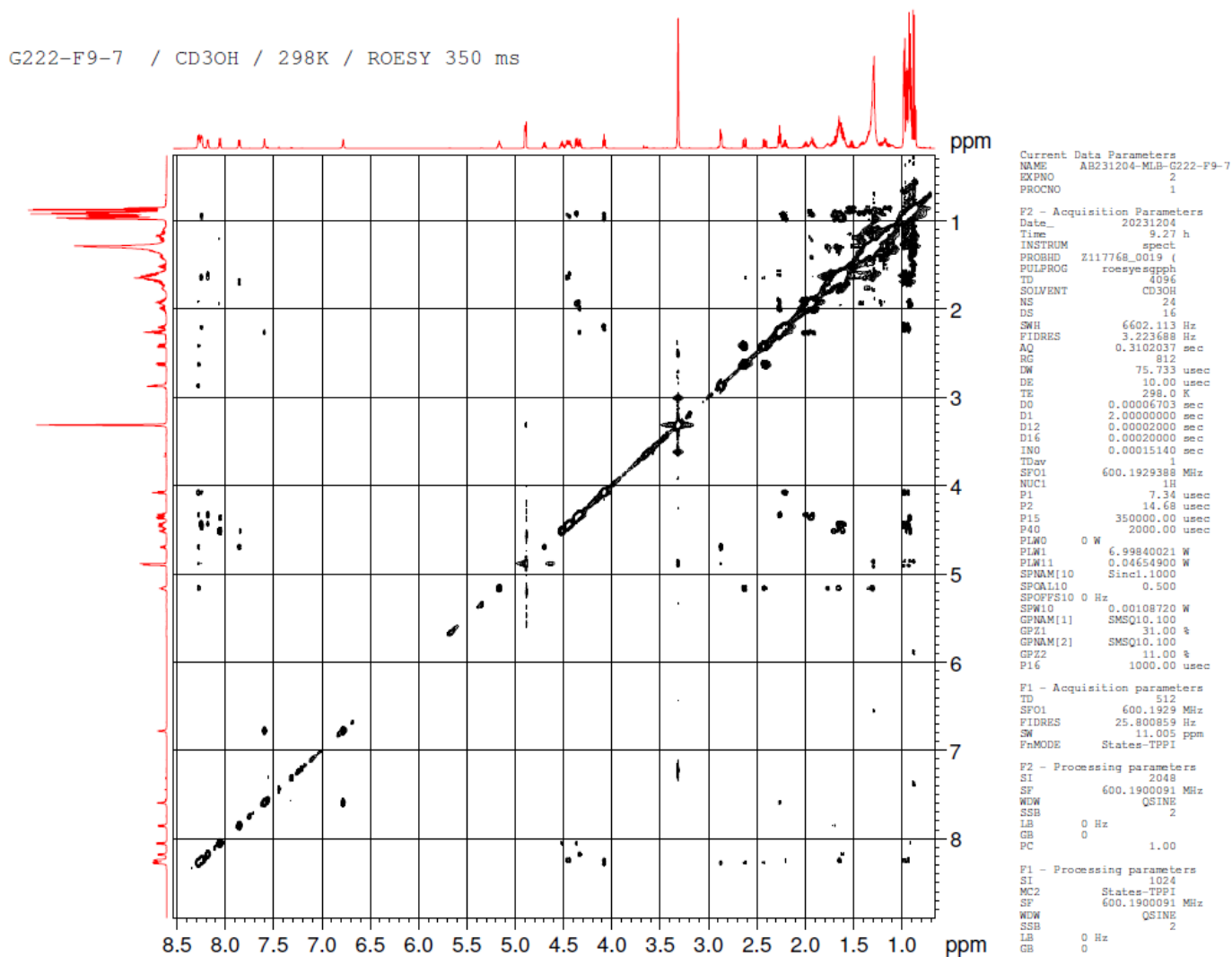


Figure S42: ROESY-NMR spectrum of Lichenysin (6-7) (m/z 1035.7063 $[M + H]^+$) (600 MHz, CD₃OH).

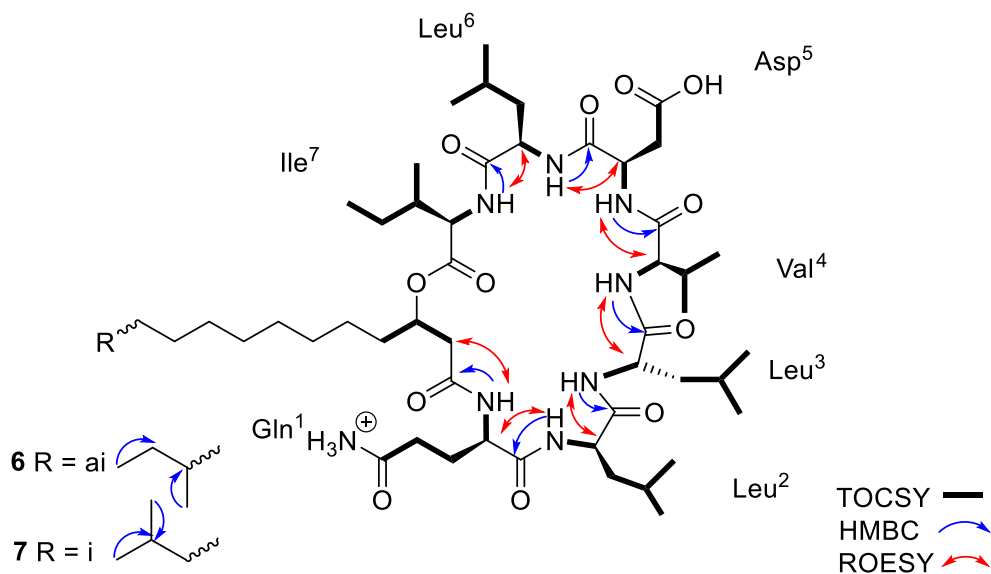


Figure S43: Structure of Lichenysins (6-7) (m/z 1035.7063 $[M + H]^+$).

Table S18. ^1H , ^{13}C , HMBC, and ROESY NMR data of Lichenysin (6-7) (m/z 1035.7064 [$\text{M} + \text{H}$] $^+$) (600 MHz, CD_3OH) isoform: *ante-iso* and *iso* (0.2:0.8).

Unit	Pos.	δ_{C} , Mult.	δ_{H} (J in Hz)	HMBC	ROESY ^a
L-Gln ¹	NH	-	8.26, d (7.56)	Gln ¹ - α , FA-CO,	FA-2-a, FA-2-b, FA-3
	C=O	173.62, C	-	-	-
	α -C	54.12, CH	4.33, ddd (7.56, 11.13, 13.23)	Gln ¹ -CO, Gln ¹ - β , Gln ¹ - γ	Leu ² -NH
	β -C	28.96, CH ₂ a. b.	1.98, m 1.91, m	Gln ¹ -CO, Gln ¹ - α , Gln ¹ - γ , Gln ¹ - δ -C=O	Leu ² -NH
	γ -C	32.63, CH ₂	2.26, t (7.56)	Gln ¹ - α , Gln ¹ - β , Gln ¹ - δ -C=O	Leu ² -NH
	δ -C=O	177.89, CO ²	-	-	-
	ε -NH ₂	- a. b.	7.59, brs 6.77 brs	Gln ¹ - δ -C=O, Gln ¹ - γ	-
L-Leu ²	NH	-	8.17, d (6.35)	Gln ¹ -CO	Gln ¹ - α , Gln ¹ - β , Gln ¹ - γ
	C=O	175.09, C	-	-	-
	α -C	53.90, CH	4.43, ddd (5.09, 6.35, 14.25)	Leu ² -CO, Leu ² - β , Leu ² - γ	Leu ³ -NH
	β -C	41.17, CH ₂ a. b.	1.64, m 1.61, m	Leu ² -CO, Leu ² - α , Leu ² - γ	Leu ³ -NH
	γ -C	25.87, CH	1.64, m	Leu ² - α , Leu ² - β , Leu ² - δ , Leu ² - δ'	Leu ³ -NH
	δ -C	22.98, CH ₃	0.97, d (4.88)	Leu ² - β , Leu ² - γ , Leu ² - δ'	-
	δ' -C	23.28, CH ₃	0.95, d (4.88)	Leu ² - β , Leu ² - γ , Leu ² - δ	-
D-Leu ³	NH	-	8.25, d (6.63)	Leu ² -CO, Leu ³ - α	Leu ² - α , Leu ² - β , Leu ² - γ
	C=O	175.52, C	-	-	-
	α -C	53.44, CH	4.45, ddd (5.94, 6.63, 14.25)	Leu ³ -CO, Leu ³ - β , Leu ³ - γ	Val ⁴ -NH
	β -C	41.56, CH ₂ a. b.	1.61, m 1.64, m	Leu ³ -CO, Leu ³ - α , Leu ³ - γ	Val ⁴ -NH
	γ -C	25.70, CH	1.64, m	Leu ³ - α , Leu ³ - β	Val ⁴ -NH
	δ -C	22.07, CH ₃	0.91, d (3.95)	Leu ³ - β , Leu ³ - γ , Leu ³ - δ'	-
	δ' -C	23.26, CH ₃	0.97, d (4.88)	Leu ³ - β , Leu ³ - γ , Leu ³ - δ	-
L-Val ⁴	NH	-	8.24, d (6.70)	Val ⁴ - α , Val ⁴ - β , Leu ³ -CO	Leu ³ - α , Leu ³ - β , Leu ³ - γ
	C=O	173.37 C	-	-	-
	α -C	61.35, CH	4.09, dd (5.26, 6.70)	Val ⁴ -CO, Val ⁴ - β , Val ⁴ - γ , Val ⁴ - γ'	Asp ⁵ -NH
	β -C	30.93, CH	2.20, ddd (5.26, 6.70, 13.20)	Val ⁴ - α , Val ⁴ - γ , Val ⁴ - γ'	Asp ⁵ -NH
	γ -C	19.62, CH ₃	0.97, d (4.88)	Val ⁴ - β , Val ⁴ - γ'	Asp ⁵ -NH
	γ' -C	18.31, CH ₃	0.92, d (3.95)	Val ⁴ - β , Val ⁴ - γ	Asp ⁵ -NH
L-Asp ⁵	NH	-	8.28, d (7.43)	Val ⁴ -CO, Asp ⁵ - α	Val ⁴ - α , Val ⁴ - β , Val ⁴ - γ , Val ⁴ - γ' , Leu ⁶ -NH
	C=O	172.99, C	-	-	-
	α -C	52.01, CH	4.70, dd (7.43, 12.55)	Asp ⁵ -CO, Asp ⁵ - β , Asp ⁵ - γ -C=O	Leu ⁶ -NH
	β -C	37.10, CH ₂	2.84, dd (3.19, 7.43)	Asp ⁵ -CO, Asp ⁵ - α , Asp ⁵ - γ -C=O	Leu ⁶ -NH
	γ -C=O	174.49, CO ²	-	-	-
D-Leu ⁶	NH	-	7.85, d (8.63)	Asp ⁵ -CO	Asp ⁵ - α , Asp ⁵ -NH, Ile ⁶ -NH
	C=O	173.60, C	-	-	-
	α -C	52.91, CH	4.51, ddd (5.77, 8.63, 14.59)	Leu ⁶ -CO, Leu ⁶ - β ,	Ile ⁷ -NH
	β -C	41.45, CH ₂ a. b.	1.70, m 1.60, m	Leu ⁶ -CO, Leu ⁶ - α , Leu ⁶ - γ	Ile ⁷ -NH
	γ -C	25.86, CH	1.64, m	Leu ⁶ - α , Leu ⁶ - β	Ile ⁷ -NH
	δ -C	22.07, CH ₃	0.91, d (3.95)	Leu ⁶ - β , Leu ³ - δ	-
	δ' -C	22.65, CH ₃	0.94, d (3.58)	Leu ⁶ - β , Leu ⁶ - δ'	-

Unit	Pos.	δ_C , Mult.	δ_H (J in Hz)	HMBC	ROESY ^a
L-Ile ⁷	NH	-	8.05, d (8.27)	Leu ⁶ -CO	Leu ⁶ -NH, Leu ⁶ - α , Leu ⁶ - β , Leu ⁶ - γ
	C=O	172.12, C	-	-	-
	α -C	58.21, CH	4.36, dd (5.60, 8.27)	Ile ⁷ -CO, Ile ⁷ - β , Ile ⁷ - γ , Ile ⁷ - δ	-
	β -C	38.37, CH	1.93, q (8.27)	Ile ⁷ - β , Ile ⁷ - γ , Ile ⁷ - δ , Ile ⁷ - δ'	-
	γ -C	25.99, CH ₂ a. b	1.41, m 1.20, m	Ile ⁷ - β , Ile ⁷ - δ , Ile ⁷ - δ'	-
	δ -C	15.96, CH ₃	0.92, d (4.31)	Ile ⁷ - α , Ile ⁷ - β , Ile ⁷ - γ	-
	δ' -C	11.79, CH ₃	0.89, t (2.33)	Ile ⁷ - β , Ile ⁷ - γ	-
Fatty acid	C=O	172.55, C	-	-	-
	2	41.96, CH ₂ a. b	2.62, dd (4.58, 12.21) 2.43, dd (7.46, 14.31)	FA-3, FA-4, FA-CO	Gln ¹ -NH
	3	73.83, CH	5.15 ddd (4.58, 7.46, 12.21)	FA-4, FA-CO	Gln ¹ -NH
	4	35.14, CH ₂ a. b	1.77, m 1.65, m	FA-2-a, FA-2-b, FA-3 FA-3, FA-5-11	- -
ai	5	26.32, CH ₂ *	1.35 ^b , m	FA-5-11, FA-12, FA-13	-
ai	6	28.07, CH ₂ *	1.29 ^b , m	FA-5-11, FA-12, FA-13	-
ai	7	30.24, CH ₂ *	1.29 ^b , m	FA-5-11, FA-12, FA-13	-
ai	8	30.51, CH ₂ *	1.29 ^b , m	FA-5-11, FA-12, FA-13	-
ai	9	30.52, CH ₂ *	1.29 ^b , m	FA-5-11, FA-12, FA-13	-
ai	10	30.65, CH ₂ *	1.29 ^b , m	FA-5-11, FA-12, FA-13	-
ai	11	37.67, CH ₂ a. b	1.09, m 1.29, m	FA-5-11, FA-12, FA-13	-
ai	12	35.54, CH ₂	1.29 ^b , m	FA-13, FA-5-11	-
ai	13	30.53, CH a. b	1.29 ^b , m 1.13, m	FA-12, FA-14, FA-15	-
ai	14	19.50, CH ₃	0.85, d (4.84)	FA-12	-
ai	15	11.63, CH ₃	0.86, t (6.15)	FA-11, FA-12, FA-13	-
i	5	26.33, CH ₂ *	1.35 ^b , m	FA-5-11, FA-12, FA-13	-
i	6	28.42, CH ₂ *	1.29 ^b , m	FA-5-11, FA-12, FA-13	-
i	7	30.40, CH ₂ *	1.29 ^b , m	FA-5-11, FA-12, FA-13	-
i	8	30.51, CH ₂ *	1.29 ^b , m	FA-5-11, FA-12, FA-13	-
i	9	30.53, CH ₂	1.29 ^b , m	FA-5-11, FA-12, FA-13	-
i	10	30.65, CH ₂ *	1.29 ^b , m	FA-5-11, FA-12, FA-13	-
i	11	26.32, CH ₂	1.29 ^b , m	FA-12, FA-14, FA-15	-
i	12	40.13, CH ₂	1.17, m	FA-11, FA-14, FA-15	-
i	13	29.02, CH ₂	1.51, m	FA-12, FA-14, FA-15	-
i	14	22.91, CH ₃	0.87, d (6.55)	FA-11, FA-13, FA-15	-
i	15	22.91, CH ₃	0.87, d (6.55)	FA-11, FA-13, FA-14	-

^a Sequential ROEs ^b Overlapped signals prevent determination of constant couplings * May be interchanged.

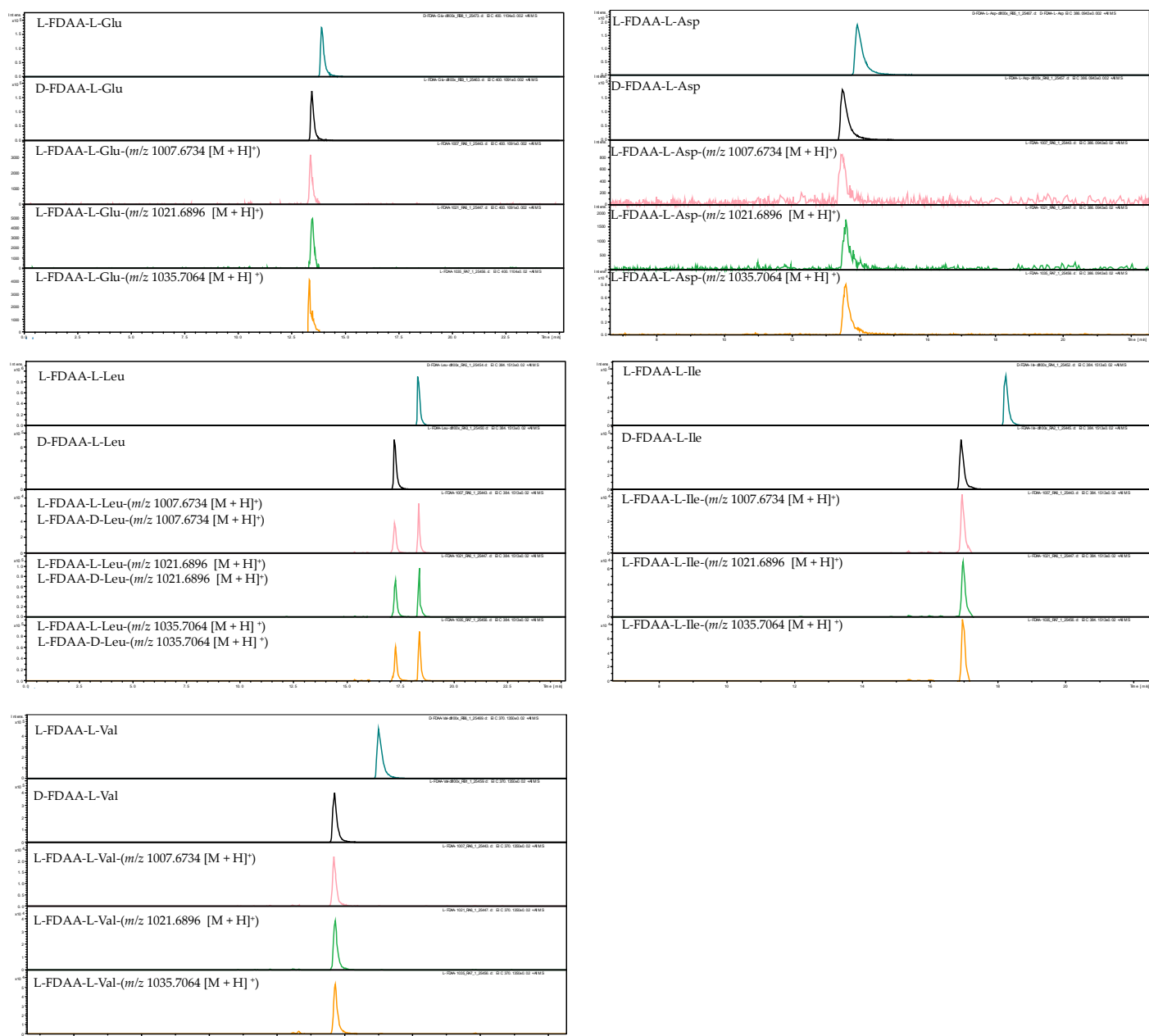


Figure S44: Marfey's analysis in positive ion mode high-resolution ESI mass spectrum

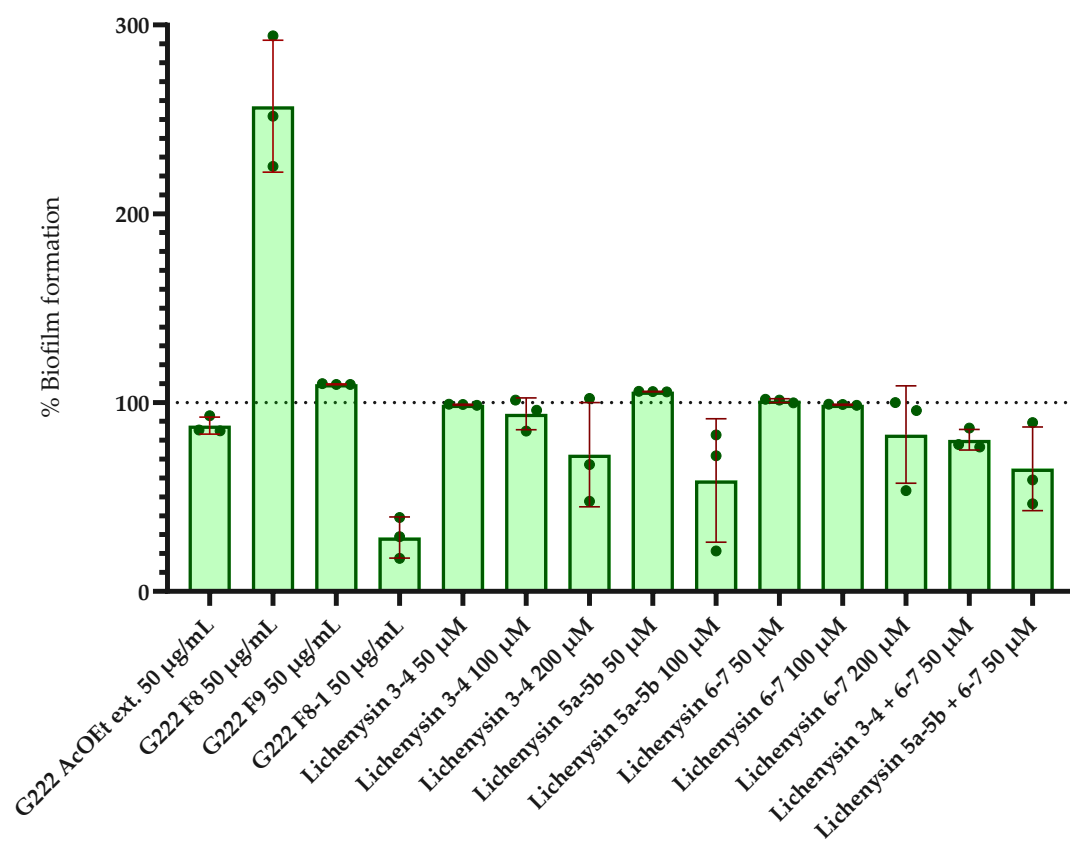


Figure S45: Biofilm-forming activity against *Pseudomonas aeruginosa* MUC-N1 (n = 3).

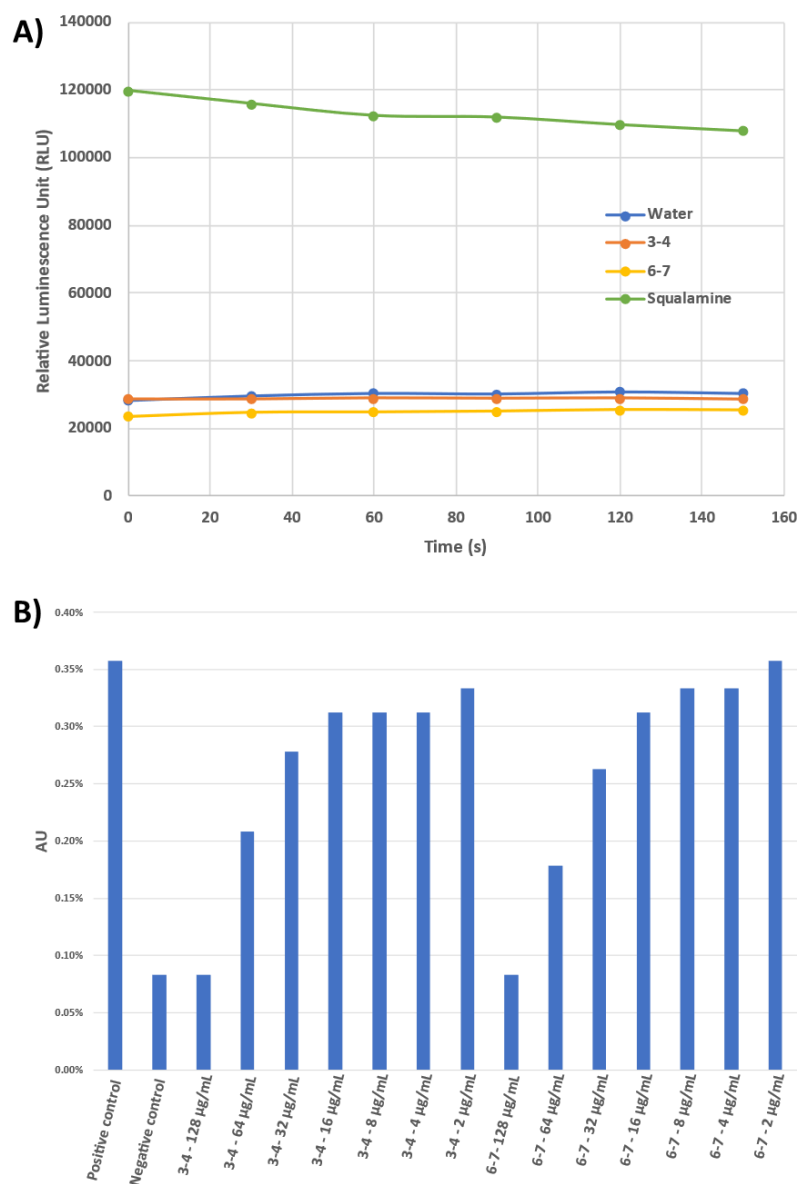


Figure S46: A) ATP release in *S. aureus* MRSA exhibited by compounds (3-4) and (6-7) as determined using ATP efflux assay. Squalamine ($100 \mu\text{g.mL}^{-1}$) was the positive control and water was the negative control. Compounds were tested at a final concentration of $100 \mu\text{g.mL}^{-1}$, and results are reported as relative luminescence unit. B) Bacterial growth inhibition exhibited by (3-4) and (6-7) against *S. aureus* MRSA (CF-Marseille) with different concentrations. Positive control was bacteria only and negative control was media only.