

SUPPLEMENTARY INFORMATION

[¹⁸F]Fluoride activation and ¹⁸F-labelling in hydrous conditions – towards a microfluidic synthesis of PET radiopharmaceuticals

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Figure S1. Picture of iMiDEV™ microfluidic cassette

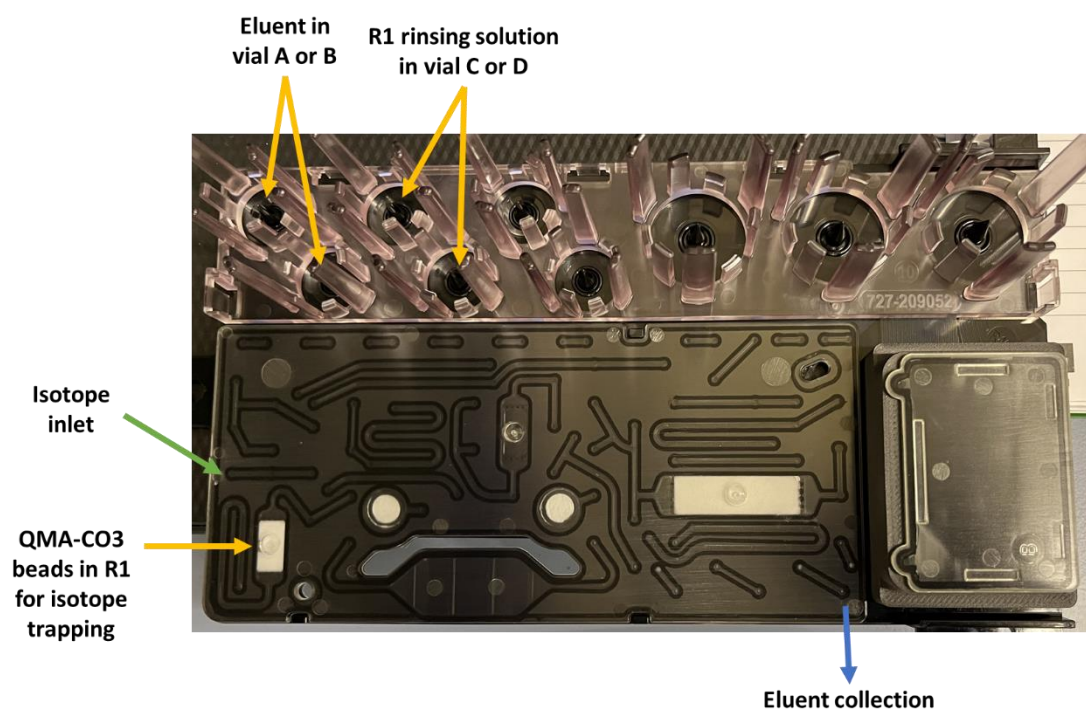


Figure S2. Cassette architecture with liquid and gas pathways marked in green, orange and yellow. A) Trapping ^{18}F fluoride on QMA- CO_3 (Waters®) beads in R1; B) rinsing the QMA- CO_3 beads with MeCN from C (green) or alternatively from D (orange); C) elution ^{18}F fluoride from R1 with eluent in A (green) or alternatively in B (orange).

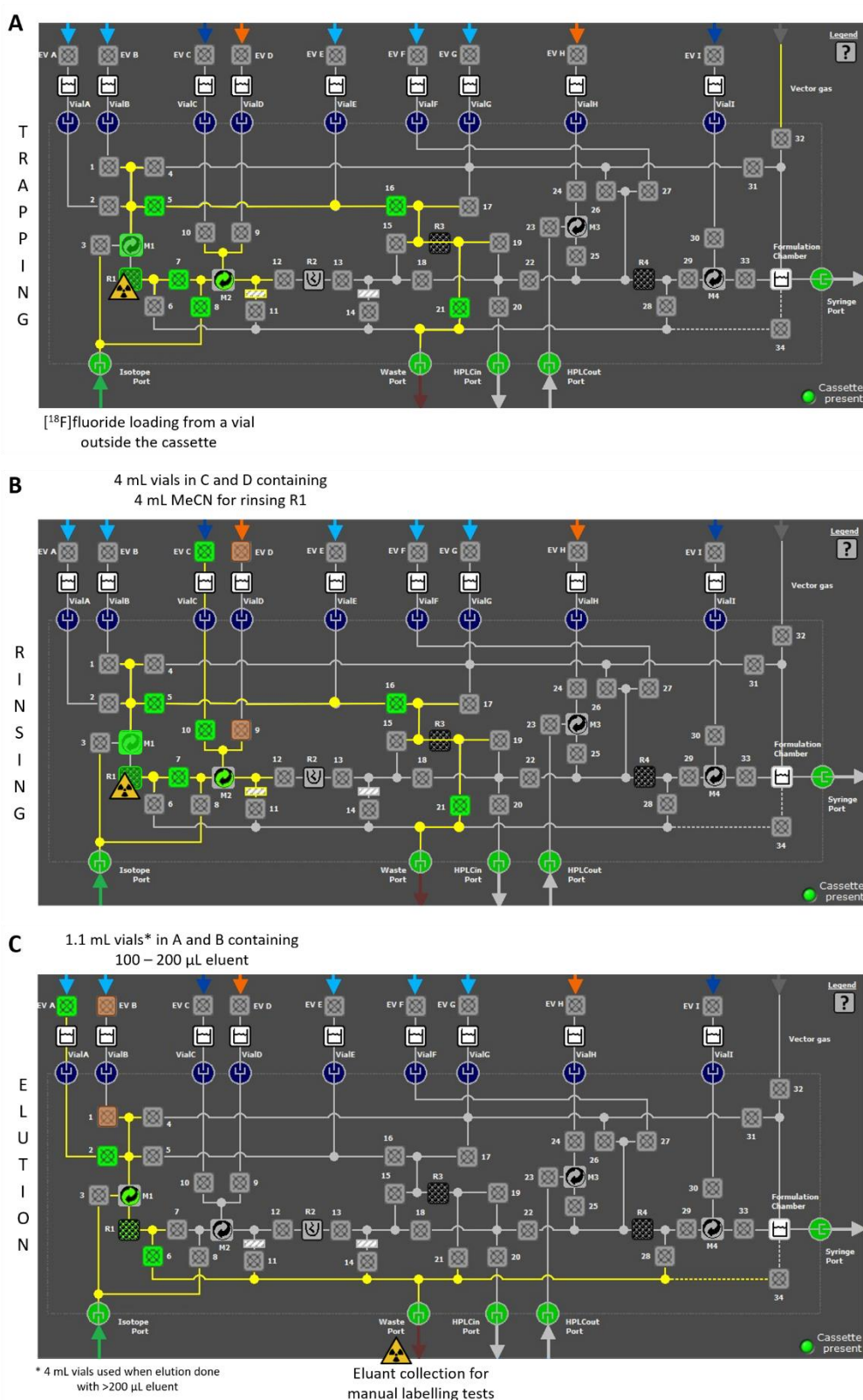


Table S1. Results of the azeotropic drying free nucleophilic ^{18}F -fluorination (n = 3).

Entry	Product	Precursor quantity	Activating agent (150 μL)	Precursor Solvent (150 μL)	Labelling conditions	RCC, %
1	^{18}F FTAG	5 mg (10.4 μmol)	3% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% CH_3CN	95 $^\circ\text{C}$ 5 min	82.2 \pm 5.8
2	^{18}F FTAG	5 mg (10.4 μmol)	5% H_2O TBAB _{40%} in CH_3CN	100% CH_3CN	95 $^\circ\text{C}$ 10 min	89.9 \pm 2.4
3	^{18}F F-Me-OTs	7 mg (19.6 μmol)	3% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	9/1 $\text{CH}_3\text{CN}/\text{H}_2\text{O}$	120 $^\circ\text{C}$ 10 min	63.9 \pm 4.3
4	^{18}F F-Me-OTs	7 mg (19.6 μmol)	3% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	8/2 $\text{CH}_3\text{CN}/\text{H}_2\text{O}$	120 $^\circ\text{C}$ 10 min	49.2 \pm 17.8
5	^{18}F F-Me-OTs	7 mg (19.6 μmol)	5% H_2O TBAB _{40%} in CH_3CN	8/2 $\text{CH}_3\text{CN}/\text{H}_2\text{O}$	120 $^\circ\text{C}$ 10 min	49.7 \pm 0.9
6	^{18}F DPA-714 (OTs prec)	1.5 mg (2.7 μmol)	2% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	52.5 \pm 10.1
7	^{18}F DPA-714 (OTs prec)	1.5 mg (2.7 μmol)	5% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	41.2 \pm 9.0
8	^{18}F DPA-714 (OTs prec)	1.2 mg (2.2 μmol)	5% H_2O TBAB _{40%} in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	66.0 \pm 10%
9	^{18}F Fallypride (OTs prec)	1.0 mg (1.9 μmol)	2% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	41.6 \pm 9.8
10	^{18}F Fallypride (OTs prec)	1.0 mg (1.9 μmol)	5% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	18.7 \pm 1.4 (n=2)
11	^{18}F LBT-999 (Cl prec)	1.5 mg (4.3 μmol)	2% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	39.5 \pm 1.7
12	^{18}F LBT-999 (Cl prec)	1.5 mg (4.3 μmol)	5% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	29.8 \pm 1.4
13	^{18}F FPyNHS (DABCO prec)	2.4 mg (4.9 μmol)	2% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	40 $^\circ\text{C}$ 10 min	16.4 \pm 3.2
14	^{18}F FPyNHS (DABCO prec)	2.5 mg (5.2 μmol)	5% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	40 $^\circ\text{C}$ 10 min	5.0 \pm 3.4
15	^{18}F FPyOBn (DABCO prec)	1.3 mg (2.7 μmol)	2% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	80 $^\circ\text{C}$ 10 min	97.9 \pm 0.2 (n=2)
16	^{18}F FPyOBn (DABCO prec)	1.2 mg (2.5 μmol)	5% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	80 $^\circ\text{C}$ 10 min	95.7 \pm 0.6 (n=2)
17	^{18}F FPyZIDE (Me_3N^+ prec)	1.3 mg (2.8 μmol)	2% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	60.9 \pm 14.1
18	^{18}F FPyZIDE (Me_3N^+ prec)	1.4 mg (3.0 μmol)	5% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	27.1 \pm 2.6
19	^{18}F FPyZIDE (NO_2 prec)	1.2 mg (4.0 μmol)	2% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	12.6 \pm 2.4
20	^{18}F FPyZIDE (NO_2 prec)	1.2 mg (4.0 μmol)	5% H_2O $\text{K}_{222}/\text{K}_2\text{CO}_3$ in CH_3CN	100% DMSO	130 $^\circ\text{C}$ 10 min	1.4 \pm 0.9

Details of radio-HPLC analyses:

HPLCs were performed on:

[HPLC A]: Alliance e2695 system equipped with a 2998 PDA detector and Herm LB500 equipped with NaI detector (Berthold) controlled by the Empower Software (Waters), Zorbax SB-C18 5 μ m 4.6 \times 250 mm analytical column (Agilent).

[HPLC B]: Alliance e2695 system equipped with a 2998 PDA detector and Herm LB500 equipped with NaI detector (Berthold) controlled by the Empower Software (Waters), Symmetry-M[®] C-18 5 μ m 150 \times 4.6 mm analytical column (Waters).

[HPLC C]: Alliance 2690 system equipped with a 996 PDA detector (Waters) and a LB509 radioactivity detector (Berthold) controlled by the Empower Software (Waters), Symmetry-M[®] C-18 5 μ m 50 \times 4.6 mm analytical column (Waters).

Elutions were performed using the following solvents:

S1: water containing 0.1% TFA

S2: acetonitrile containing 0.1% TFA

S3: water containing Low-UV PIC[®] B7 reagent (20 mL for 1000 mL) (Waters)

S4: water:acetonitrile 30:70 (v:v) containing Low-UV PIC[®] B7 reagent (20 mL for 1000 mL) (Waters)

Solvent mixtures, flow rates, λ detection and retention times of radiotracers are summarized in Table S2.

Table S2: HPLC conditions.

Radiotracer	HPLC system	Eluant mixture (v:v)	Flow rate (mL/min)	λ (nm)	Radio-HPLC Rt (min)
[¹⁸ F]F-Me-OTs	A	S1:S2 (55:45)	1	254	9.6
[¹⁸ F]Tosyl fluoride	A	S1:S2 (55:45)	1	254	13.3
[¹⁸ F]DPA-714	B	S1:S2 (55:45)	1	263	3.9
[¹⁸ F]DPA-714	C	S3:S4 (40:60)	2	254	2.1
[¹⁸ F]Fallypride	C	S3:S4 (60:40)	2.5	220	1.6
[¹⁸ F]LBT-999	C	S3:S4 (55:45)	2	220	2.2
[¹⁸ F]FPyNHS	C	S3:S4 (75:25)	2	260	1.8
[¹⁸ F]FPyZIDE	C	S3:S4 (60:40)	2	267	2.6
[¹⁸ F]FPyOBn	C	S3:S4 (35:65)	2	254	2.7

Table S3. Comparison of the mass of anion exchange beads between different suppliers

Mass\Supplier	Waters Sep-Pak Accell Plus QMA Carbonate Light Cartridge	Eichrom QMA-S-BC, (QMA-CO ₃)	Maxi-Clean S-Pure QMA-CO ₃
Mass declared by supplier	130 mg	125 mg	125 mg
Mass measured (n = 3)	108 \pm 1 mg	118.7 \pm 0.4 mg	54.4 \pm 1.9 mg

Table S4. Results of the elution efficiencies (EE) obtained in manual [^{18}F]fluoride fractionated elution tests using kryptofix-based method ($n = 3$) and 3 different QMA- CO_3 types of beads (25 mg/cartridge)

Entry	Volume of eluent ($\text{K}_{222}/\text{K}_2\text{CO}_3/\text{MeCN}$, 3% H_2O)	EE average (Waters)	EE average (Eichrom)	EE average (S*Pure)
1	0.1 mL	2.5 \pm 1.2%	7.3 \pm 6.8%	0.3 \pm 0.3%
2	0.2 mL	34.4 \pm 2.6%	41.8 \pm 6.3%	4.7 \pm 3.6%
3	0.3 mL	67.5 \pm 4.3%	73.4 \pm 9.4%	20.1 \pm 10.7%
4	0.4 mL	86.3 \pm 3.1%	90.0 \pm 5.3%	43.1 \pm 12.7%
5	0.5 mL	94.1 \pm 1.4%	96.4 \pm 2.9%	68.3 \pm 7.6%

Table S5. Results of the elution efficiencies (EE) obtained in manual [^{18}F]fluoride fractionated elution tests using tetrabutylammonium-based method ($n = 3$) and 3 different QMA- CO_3 types of beads (25 mg/cartridge)

Entry	Volume of eluent (TBAB 5%)	EE average (Waters)	EE average (Eichrom)	EE average (S*Pure)
1	0.1 mL	0.6 \pm 0.5%	0.1 \pm 0.1%	0.1 \pm 0.2%
2	0.2 mL	16.1 \pm 4.7%	10.9 \pm 4.6%	3.7 \pm 2.6%
3	0.3 mL	40.0 \pm 5.4%	36.9 \pm 9.8%	14.0 \pm 3.5%
4	0.4 mL	60.5 \pm 7.5%	55.5 \pm 9.8%	30.0 \pm 3.6%
5	0.5 mL	73.8 \pm 3.4%	70.8 \pm 7.4%	47.0 \pm 2.3%

Table S6. Results of the [^{18}F]fluoride EE using kryptofix-based elution method ($n \geq 3$) obtained in microfluidic cassette using reactor R1 filled with approximately 25 mg of QMA- CO_3 beads from Waters®.

Entry	Volume of eluent ($\text{K}_{222}/\text{K}_2\text{CO}_3/\text{CH}_3\text{CN}$ eluent)	EE with 1% H_2O	EE with 2% H_2O	EE with 3% H_2O	EE with 5% H_2O
1	0.1 mL	51.4 \pm 14.6%	43.3 \pm 26.6	46.7 \pm 11.2%	55.9 \pm 26.8%
2	0.15 mL	69.4 \pm 5.4%	82.2 \pm 9.4%	90.6 \pm 2.5%	94.1 \pm 1.9%
3	0.2 mL	70.0 \pm 7.4%	86.9 \pm 2.9%	92.5 \pm 3.9%	93.1 \pm 4.5%
4	1 mL	90.8 \pm 1.2%	97.6 \pm 0.9%	98.2 \pm 0.6%	98.7 \pm 0.9%