

Regioselective one-pot synthesis, biological activity and molecular docking studies of novel conjugates *N*-(*p*-aryltriazolyl)-1,5-benzodiazepin-2-ones as potent antibacterial and antifungal agents

Asma NSIRA¹, Hasan MTIRAOUI¹, Sami CHNITI¹, Hanan Al-GHULIKAH^{2,*}, Rafik GHARBI³ and Moncef MSADDEK¹

¹ Laboratory of Heterocyclic Chemistry Natural Products and Reactivity/CHPNR, Department of Chemistry, Faculty of Science of Monastir, University of Monastir, 5000 Monastir, Tunisia

² Department of Chemistry, College of Sciences, Princess Nourah bint Abdulrahman University, P.O. Box 84428, Riyadh 11671, Saudi Arabia

³ Laboratory of applied chemistry and environment, Department of Chemistry, Faculty of Science of Monastir, University of Monastir, 5000 Monastir, Tunisia

* Correspondence: haalghulikah@pnu.edu.sa; Tel.: +966-11823-6011

Table of contents:

I. The detailed description of NMR 4b.....	S2-S5
II. The detailed description of NMR 6b.....	S6-S15
II. Spectra of HRMS of 4b and 4g.....	S16-S17
II.2 Copies of ¹H and ¹³C NMR	S18-S29

✚ Thus, the formation of the triazole unit in **4b** chosen as an example was confirmed by the disappearance in the ¹H NMR spectra of a triplet (t, H-3", 2.35 ppm) corresponding to the acetylenic proton in front of the appearance of the characteristic singlet (s, H-5", 8,05 ppm) which is directly attributed to the proton of the triazole ring.

Moreover the deshielding alkynes functions characteristic signals at 4.30 ppm (d, 2H, CH2-1', *J* = 15.6 Hz), to the signals corresponding to the methylene alpha to the triazole ring at 4.96 ppm (d, 1H, H-1a", *J* = 14.7 Hz) and 5.27 ppm (d, 1H, H-1b", *J* = 15 Hz), as can be seen when compared with their displacements in the starting molecules. Note also the presence of a singlet at 3.89 ppm directly attributed to the methoxy group and signals in the aromatic region at (δH 6,98-8,16 ppm, 12H) corresponding to the aromatic protons of both diazepine and the phenyl ring of the azide used.

Additional supports for our assignments arose from the HMBC spectrum which revealed significant long range correlation between the triazolic proton (s, H-5'', 8.01 ppm) and both quaternary carbons C-4'' and C-1''' at 144.6 ppm and 154.2 ppm respectively. Also, the proton H-3''' (7.62 ppm) correlates with carbons OCH₃, C-2''' et C-1''' appearing at 55,6 ppm, 122,0 ppm and 154,2 ppm respectively. Thus came supporting the obtention of the proposed molecular structures.

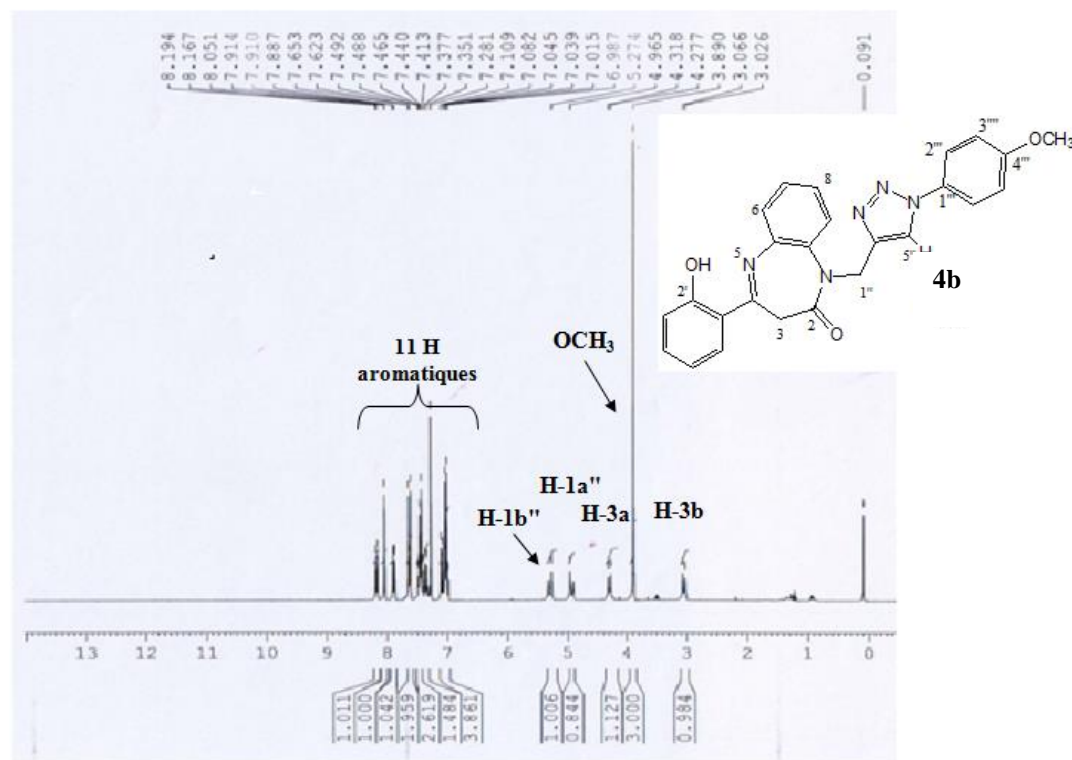


Figure S1 . NMR spectra ¹H (300 MHz, CDCl₃) of compound **4b**

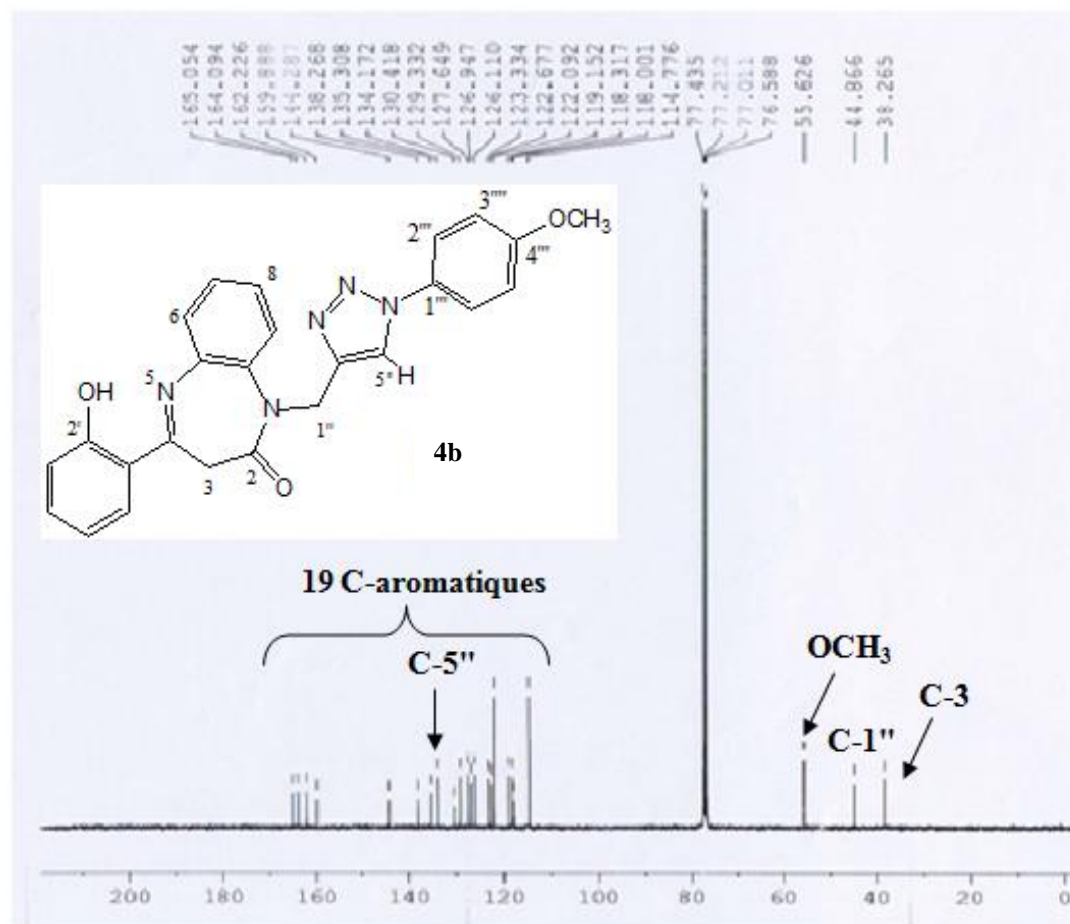


Figure S2 . NMR spectra ^{13}C (75,47 MHz, CDCl_3) of compound **4b**

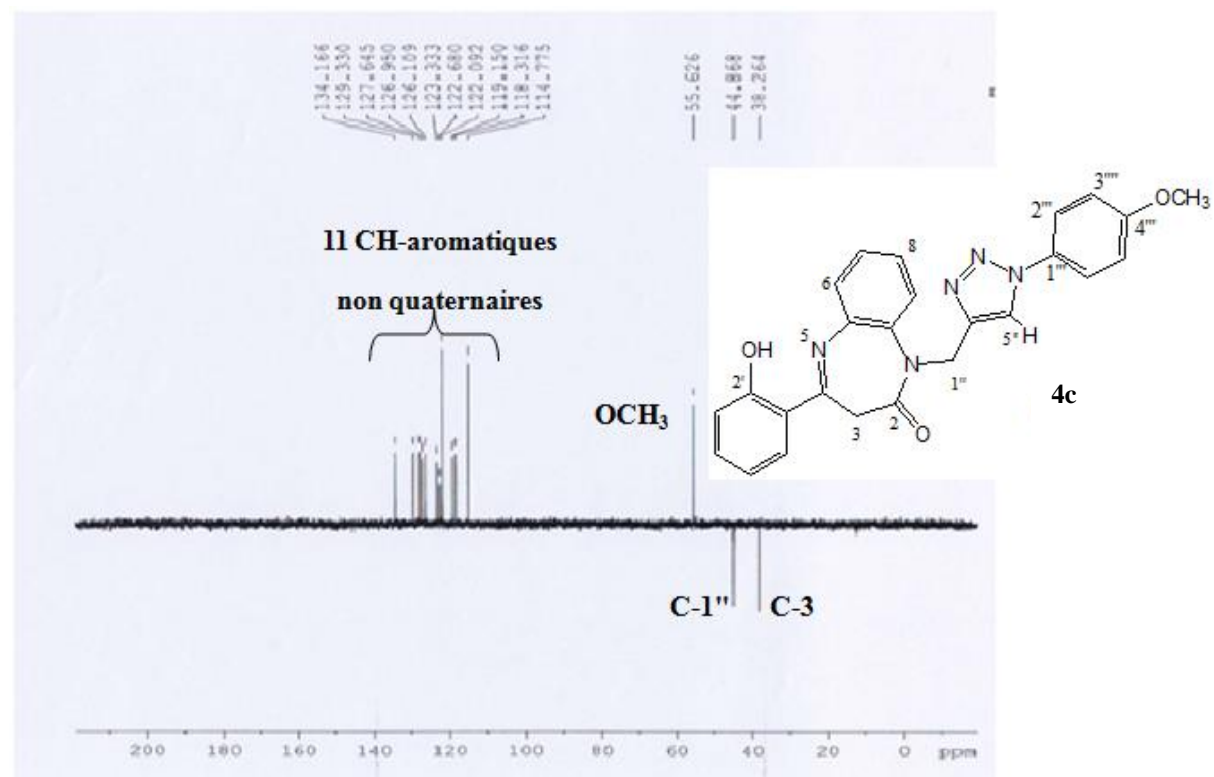


Figure S3 . DEPT 135 of compound **4b**

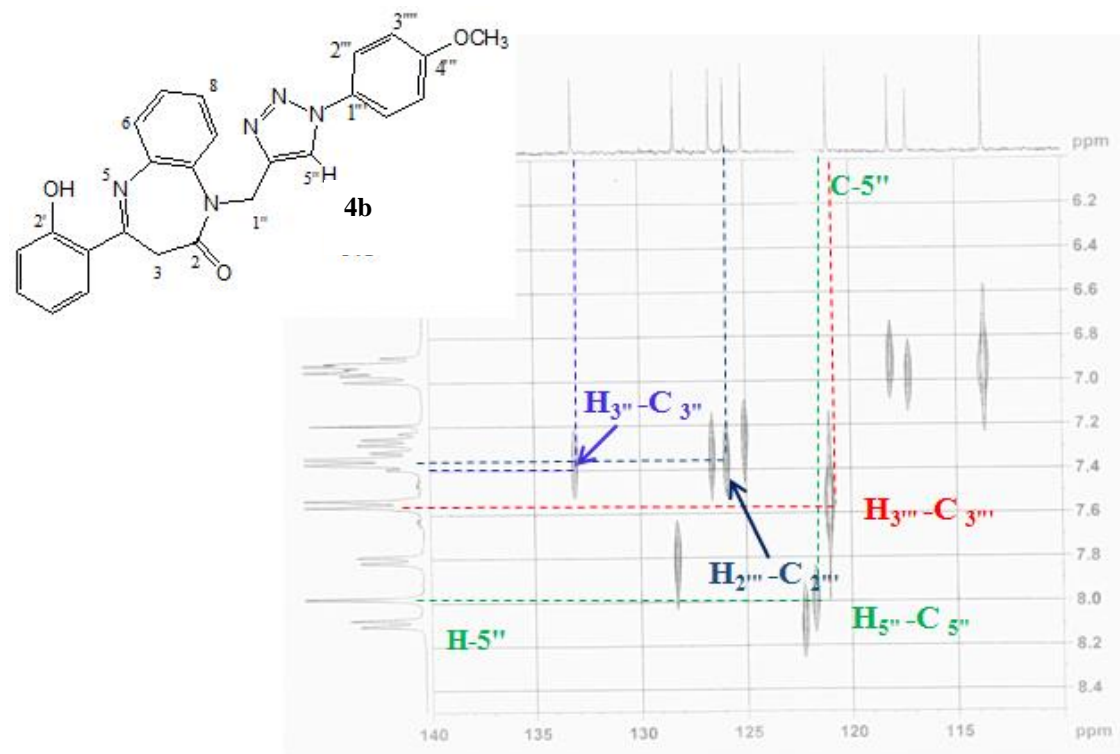


Figure S4 . CHcorr spectra of compound 4b

- The ^1H NMR spectrum of the *N*-galactosyl-triazoles **6b** showed chemical shifts of H-1''' (anomeric proton) at 5.40 ppm (doublet, $J = 4.8$ Hz) downfield compared to the precursor azide at 5.56 ppm. Except these characteristic signals taken from the ^1H NMR spectrum of compound **6b**, ^{13}C NMR (75.4 MHz in CDCl_3) was used and more particularly the DEPT 135 experiment. In addition to the observed peaks for the starting compound, four peaks appeared between 22.6 and 25.9 ppm corresponding to the methyl of the galactopyranosyl part. Moreover, the peaks at $\delta = 66.8(\text{C-2}''')$, $67.1(\text{C-3}''')$, $70.2(\text{C-4}''')$, $70.7(\text{C-5}''')$, $96.2(\text{C-1}''')$, $109.9(\text{C-isop})$, $109.9(\text{C-isop})$ ppm are assigned to the introduced galactosyl moiety. The peak at 50.56 ppm is attributed to the C-6''' carbon. A signal at 126.9 ppm is related to carbon C-5'' of the triazole ring.

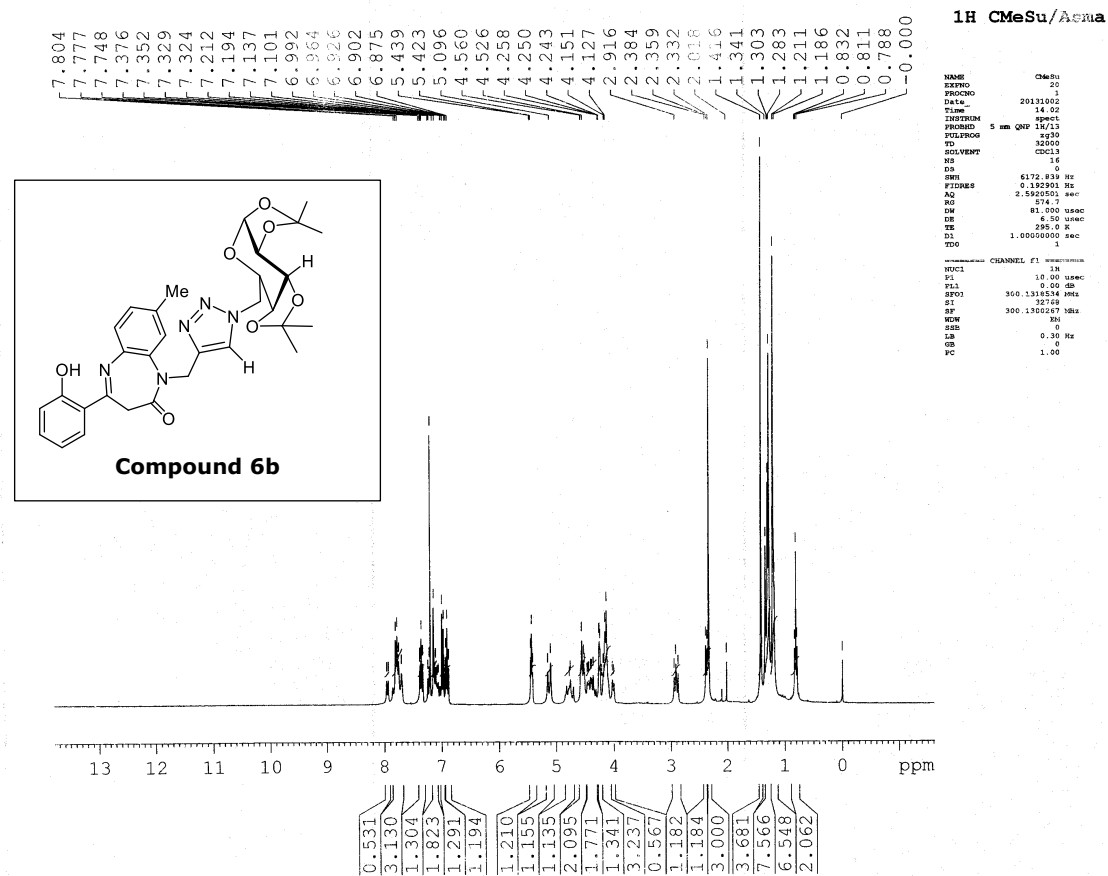


Figure S5. NMR spectra ¹H (300 MHz, CDCl₃) of compound **6b**

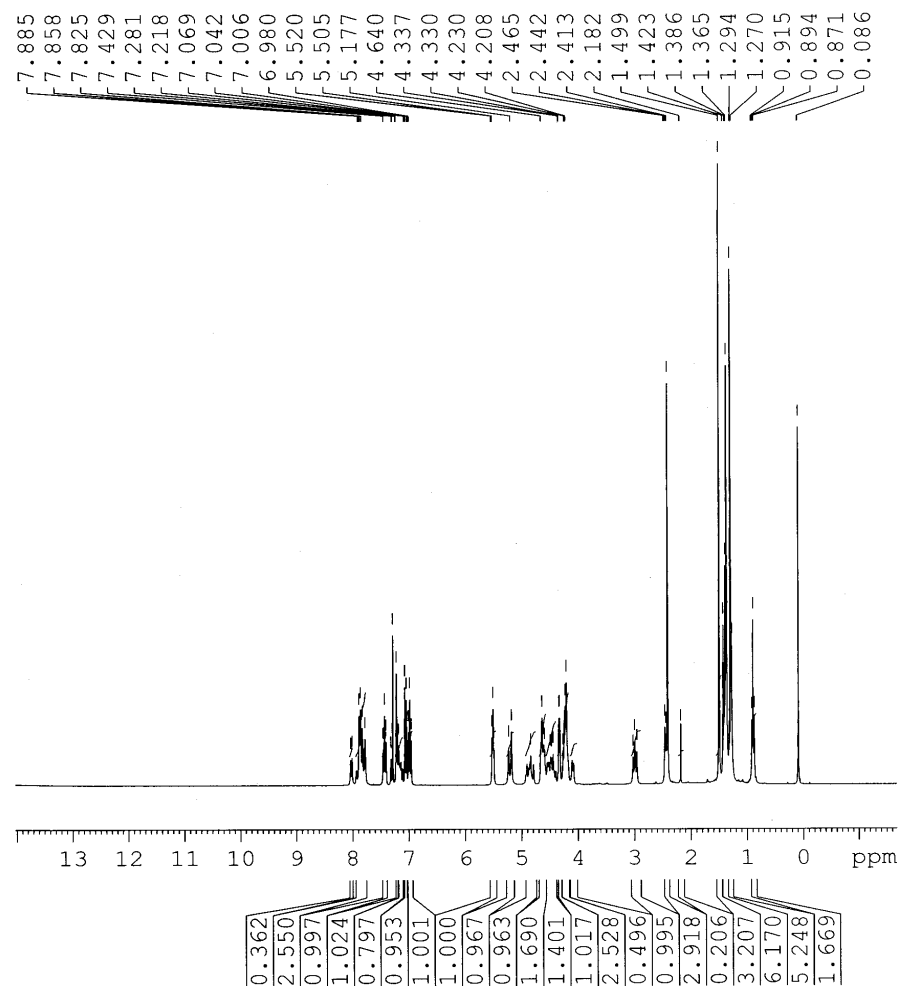
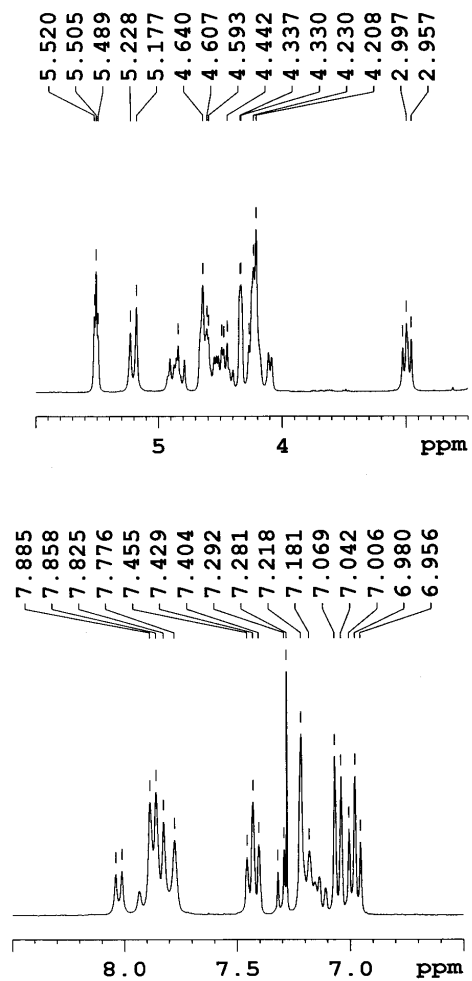


Figure S6. NMR spectra ^1H (300 MHz, CDCl_3) of compound **6b**

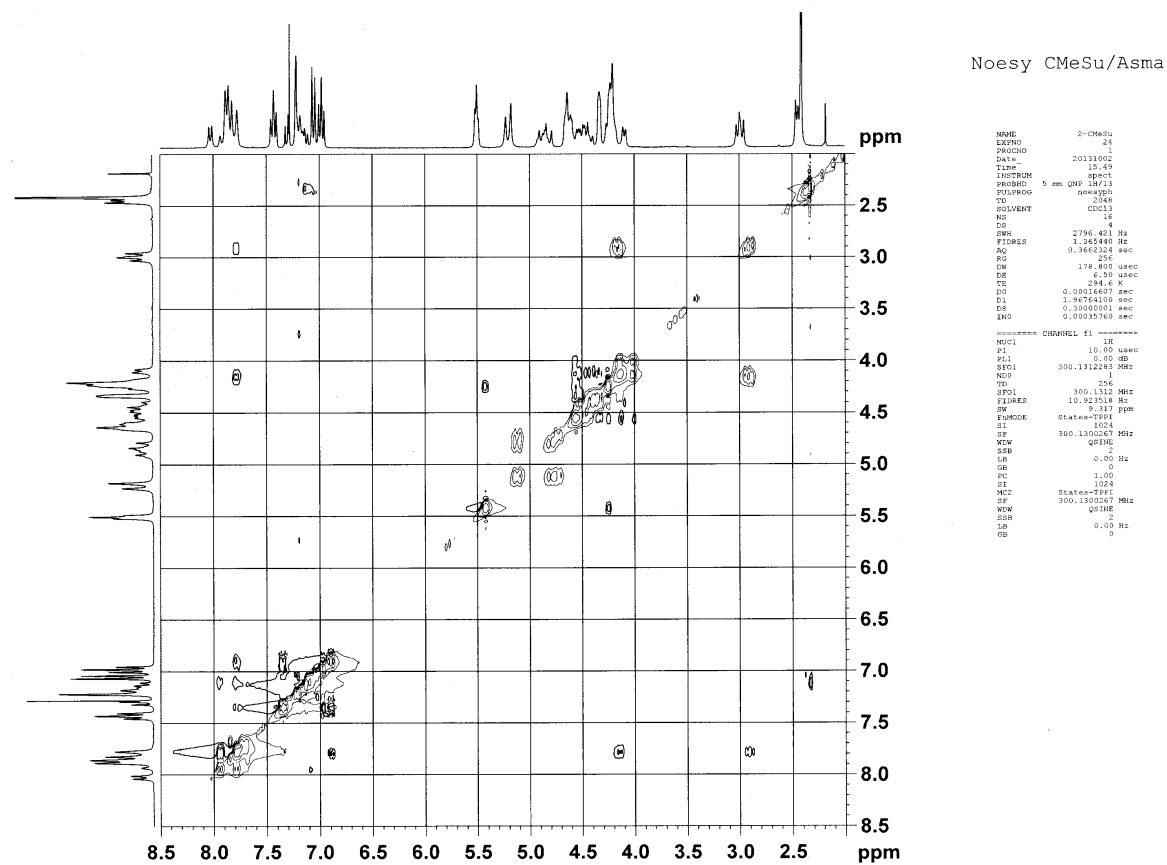


Figure S7. NOESY Spectra of compound **6b**

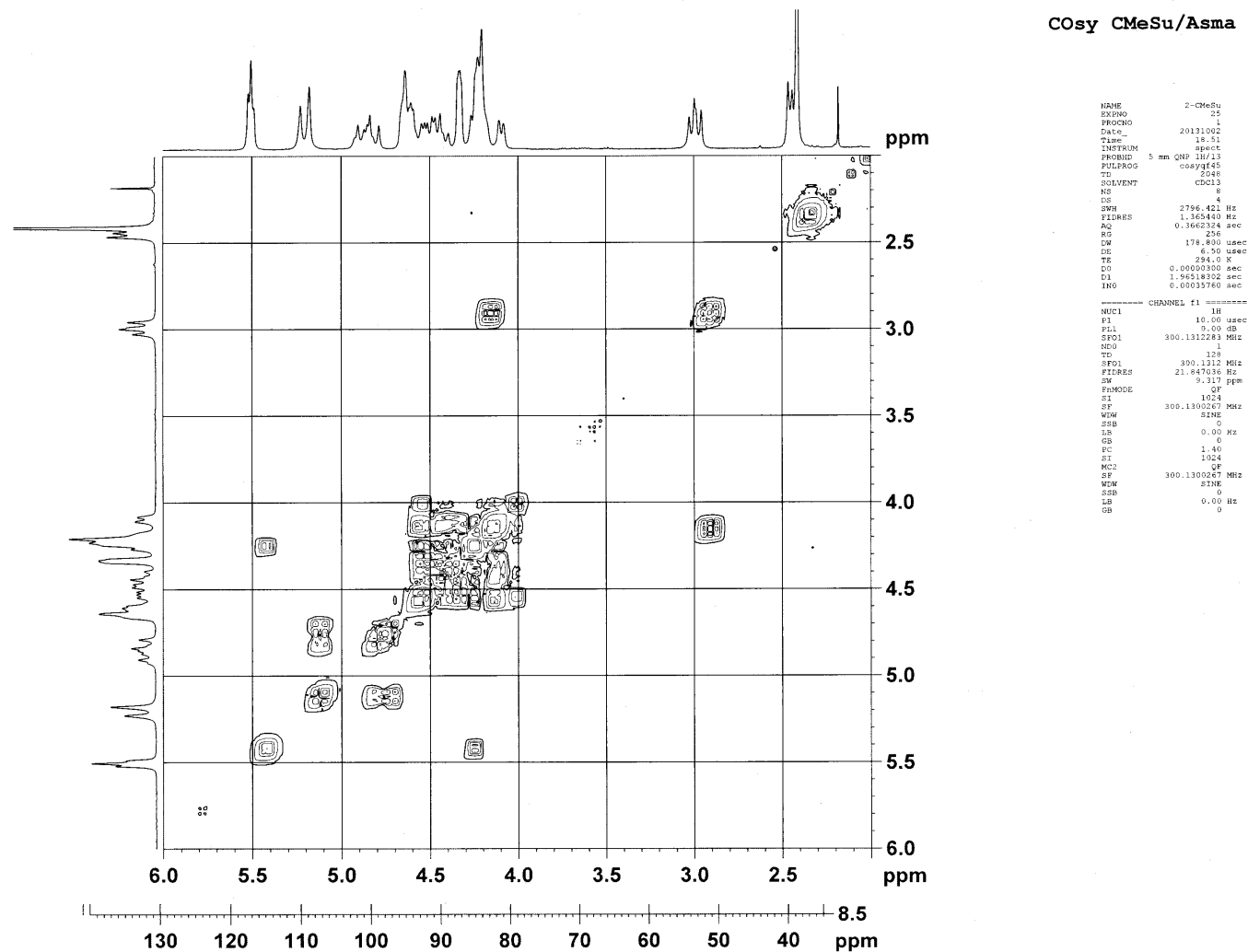


Figure S8 . COSY ^1H - ^1H spectra of compound **6b**

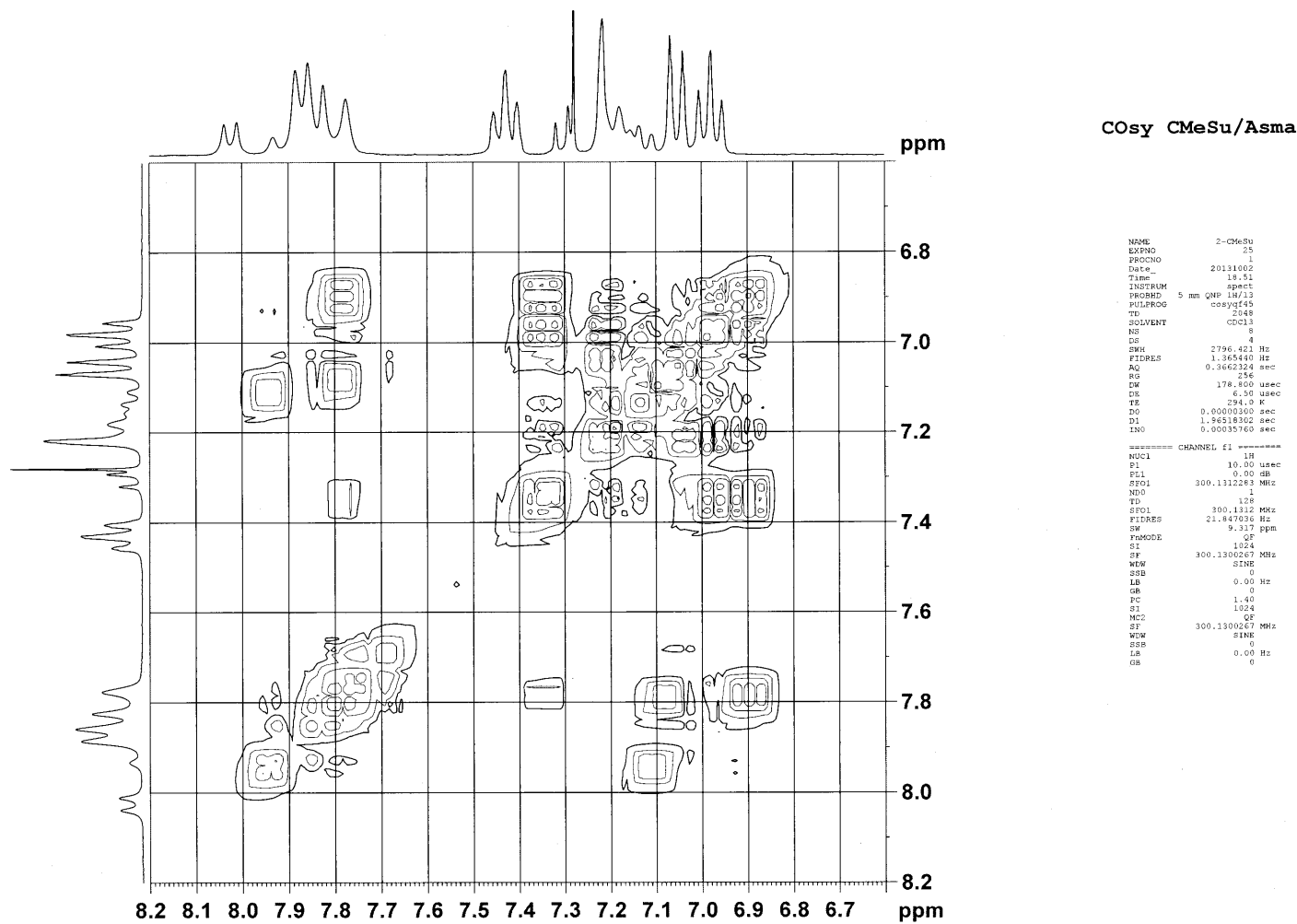


Figure S9 . COSY ^1H - ^1H spectra of compound **6b**

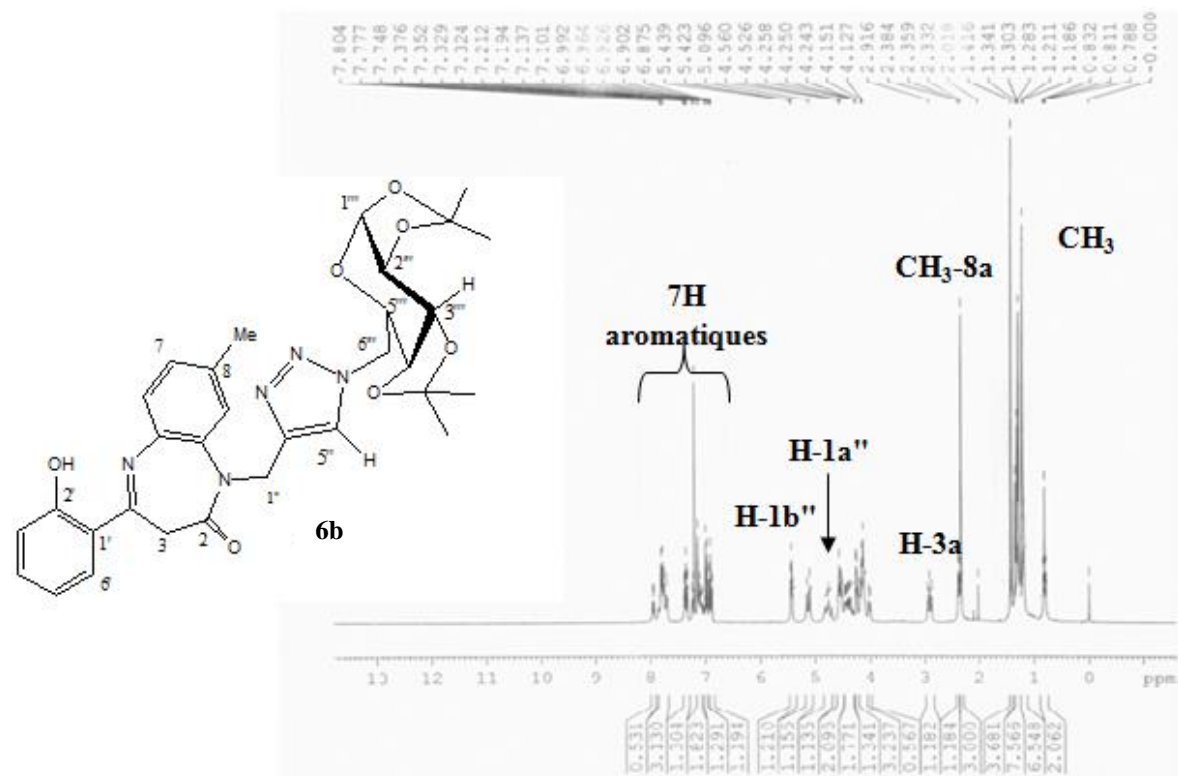


Figure S10. NMR spectra ^1H (300 MHz, CDCl_3) of compound **6b**

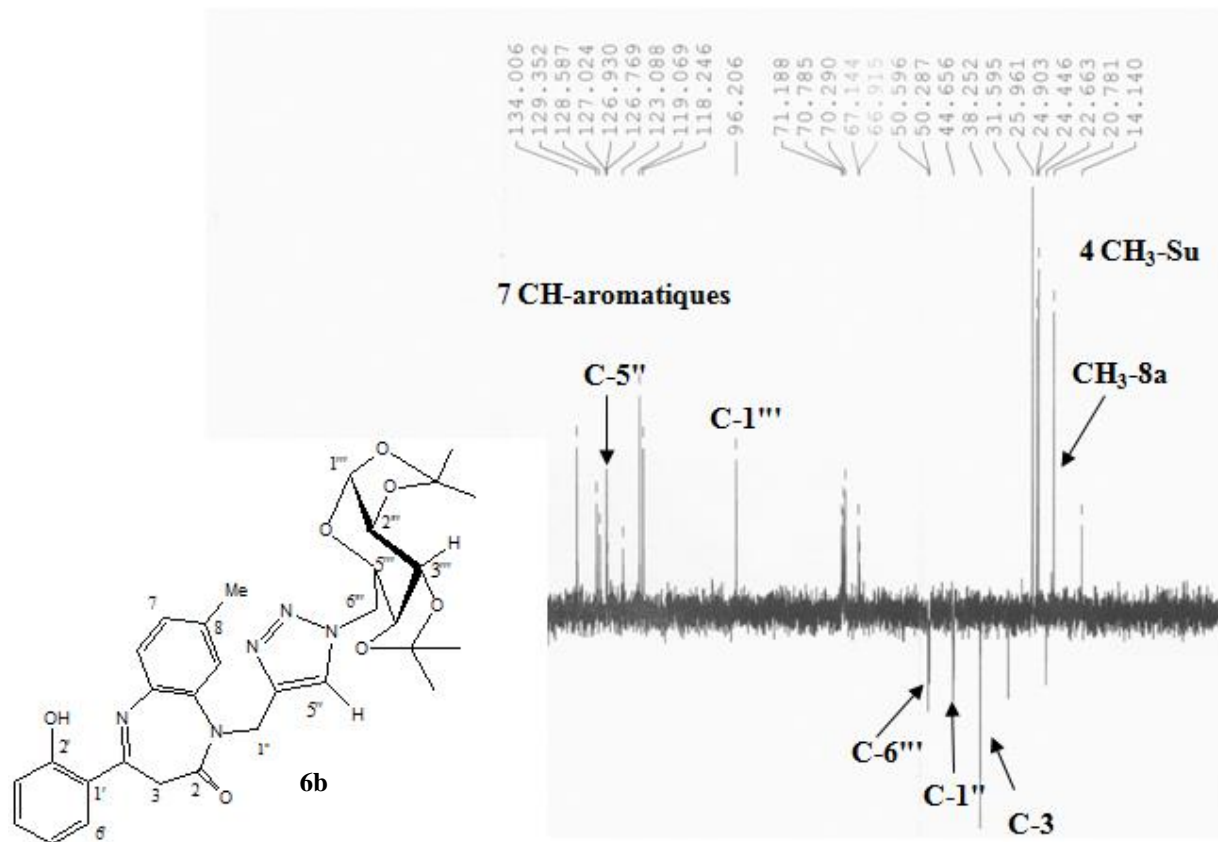


Figure S11. DEPT 135 (75,47 MHz, CDCl₃) of compound **6b**

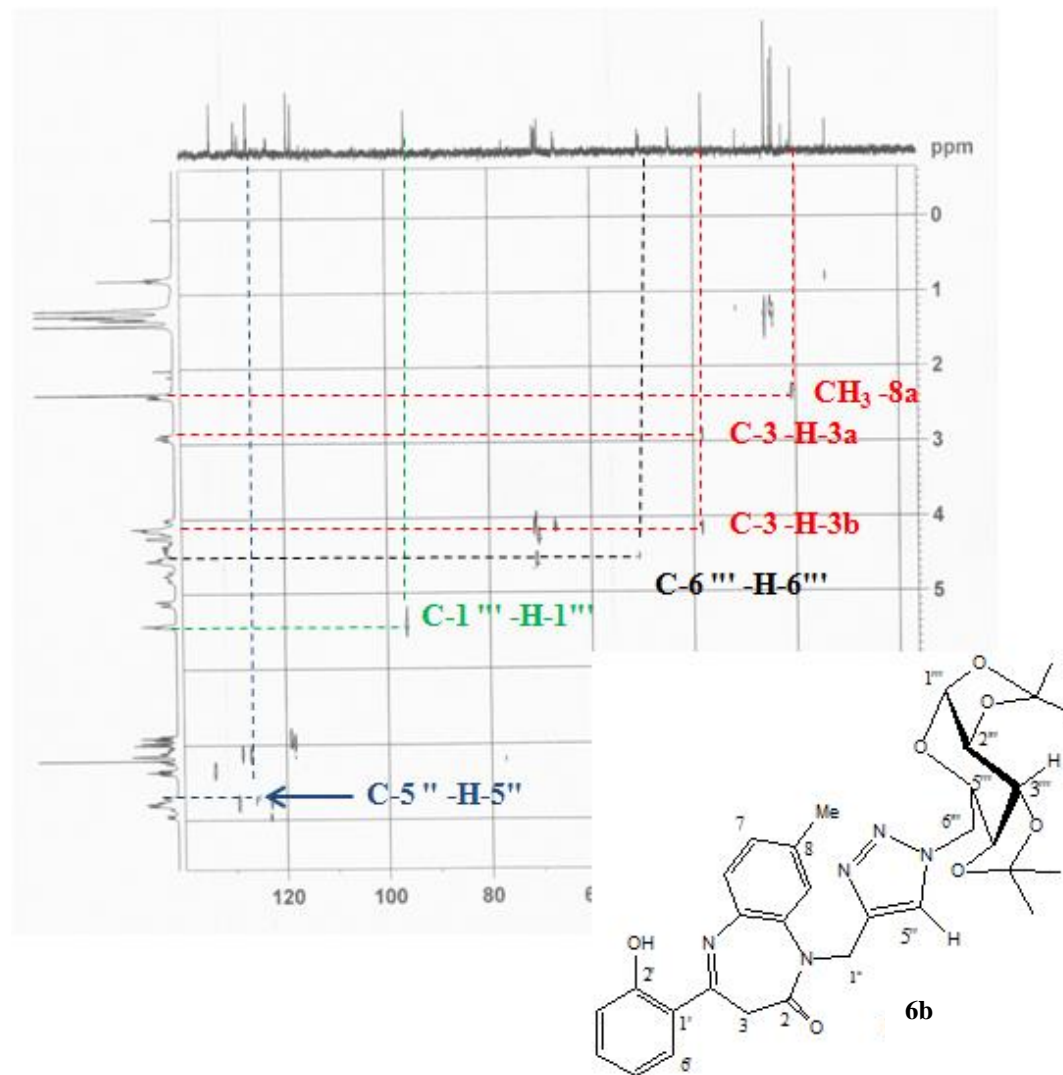


Figure S12 . CHcorr Spectra of compound **6b**

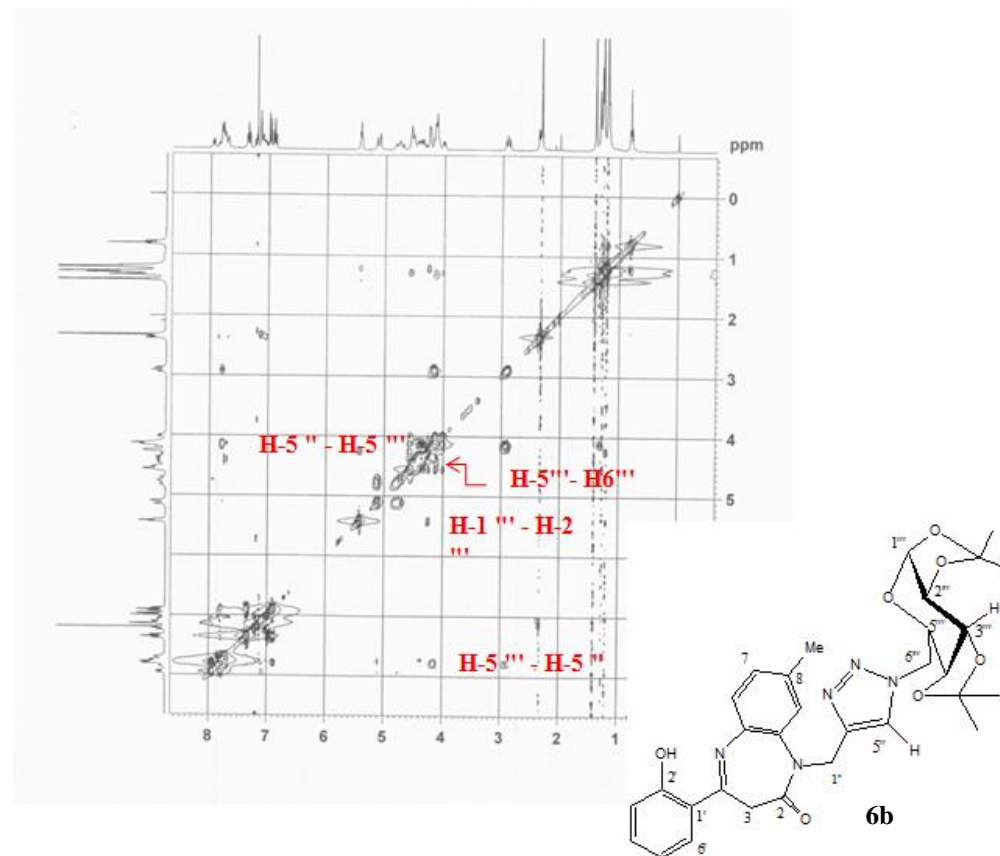


Figure S14. NOESY Spectra of compound **6b**

ANALYSE HRMS

Laboratoire : C-TAC
 Demandeur : Rafik Gharbi
 Nom échantillon : CI015
 Nom fichier : infusion_RAFIK_2198
 Ionisation / polarité / mode : ESI +

Date d'acquisition : 18-07-2017

Compound 4b

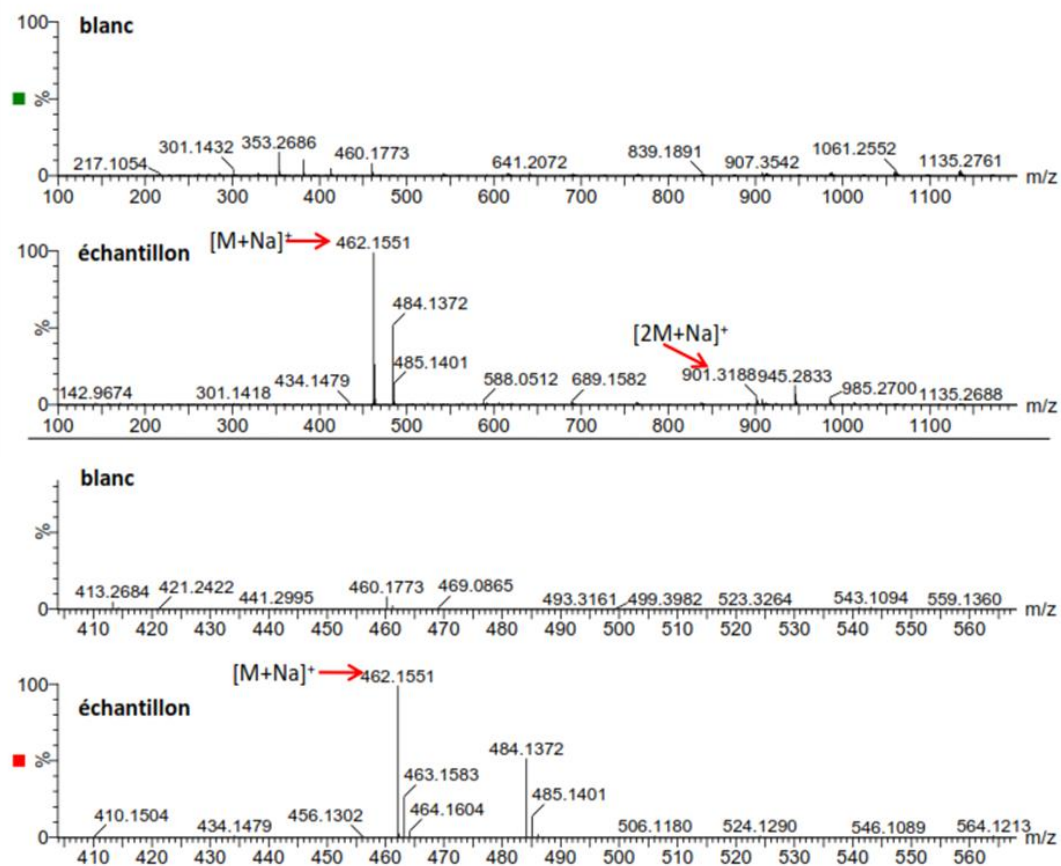


Figure S15. HRMS Spectra of compound 4b

ANALYSE HRMS

Laboratoire : C-TAC

Date d'acquisition : 18-07-2017

Demandeur : Rafik Gharbi

Compound 4g

Nom échantillon : CI021

Nom fichier : infusion_RAFIK_2202

Ionisation / polarité / mode : ESI +

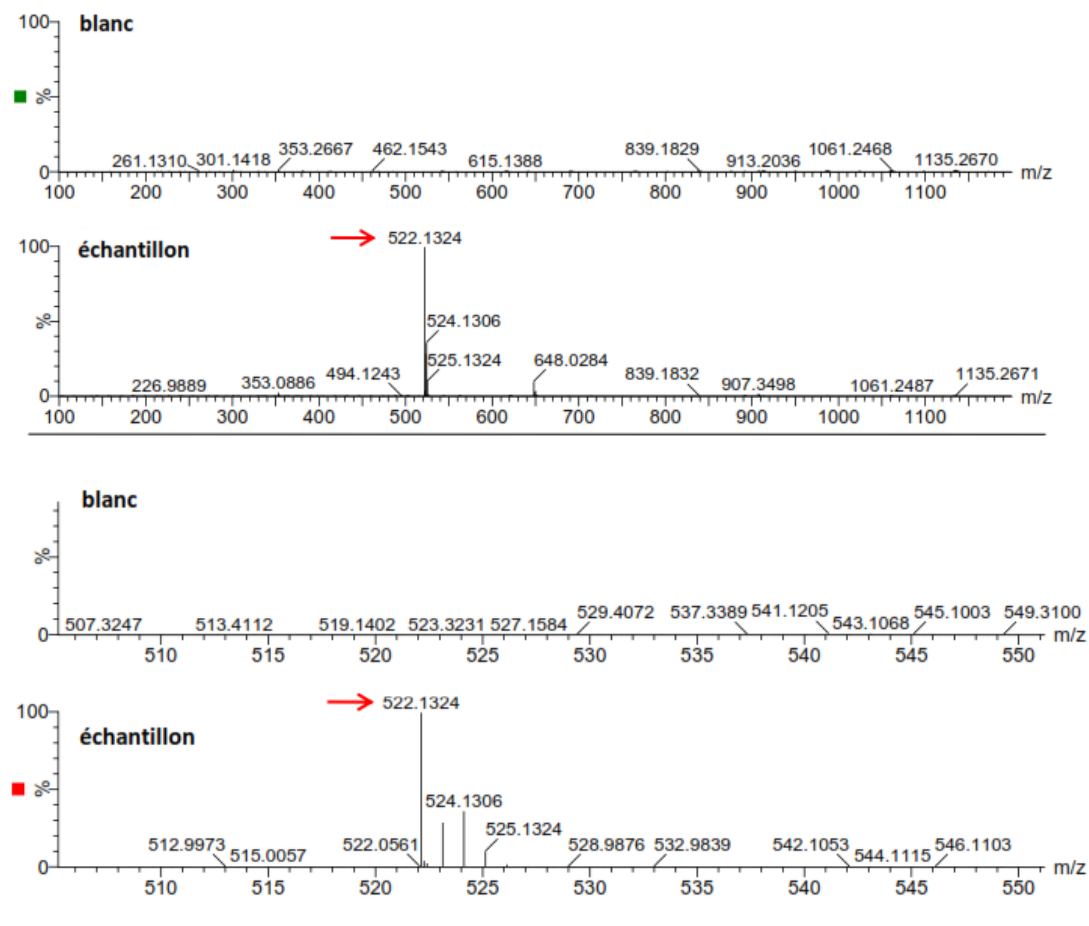


Figure S16. HRMS Spectra of compound 4g

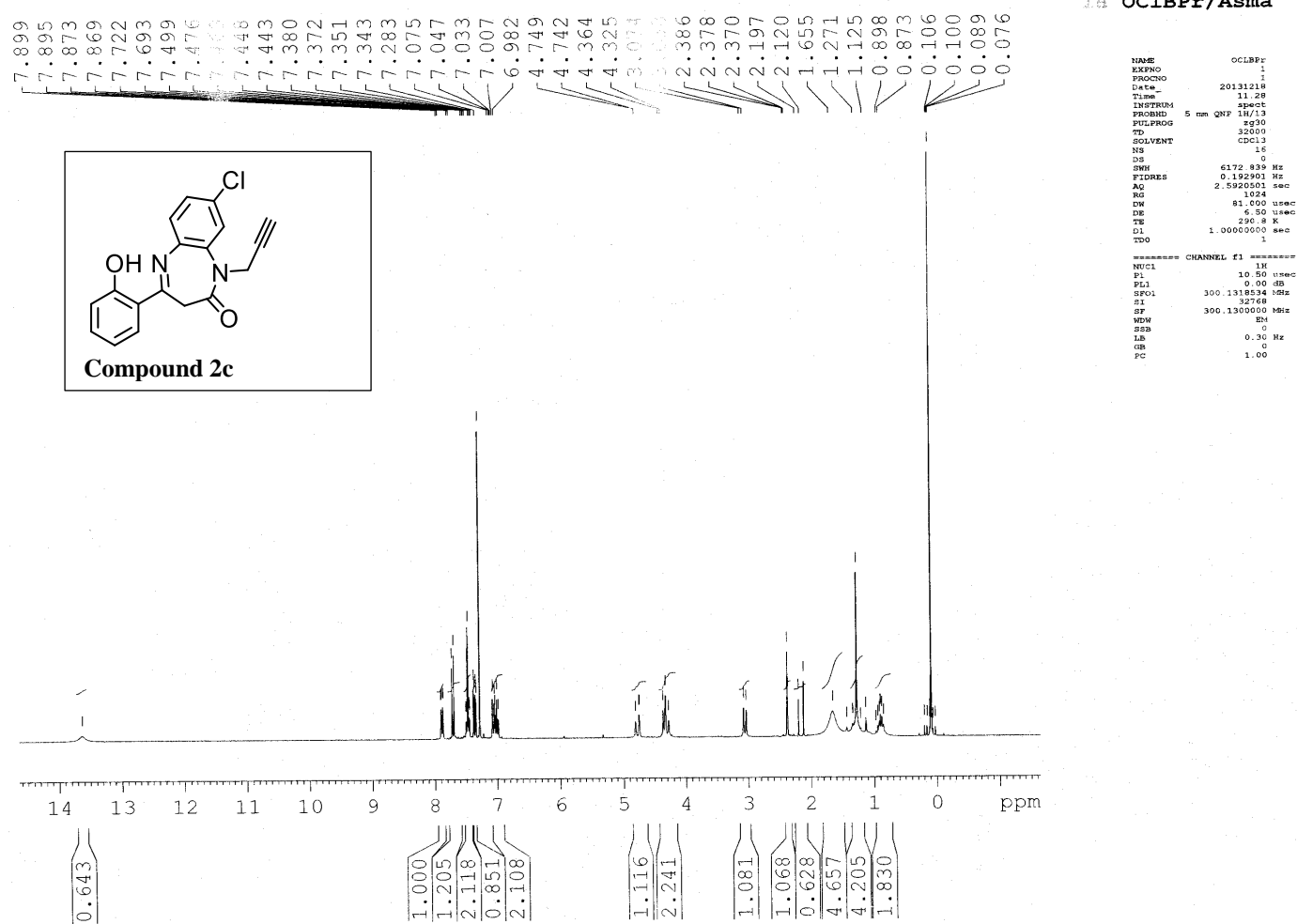


Figure S17. NMR spectra ^1H (300 MHz, CDCl_3) of compound **2c**

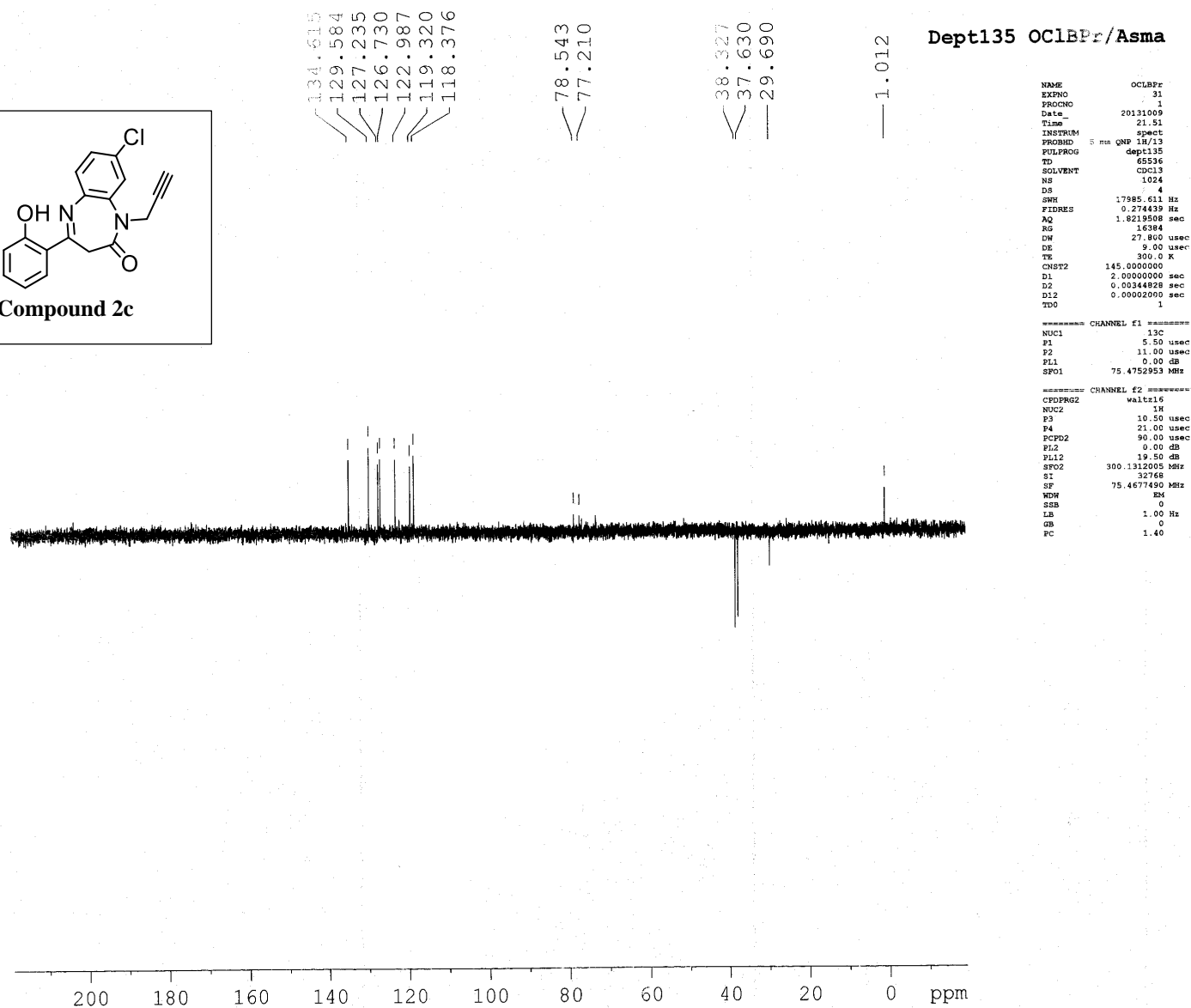
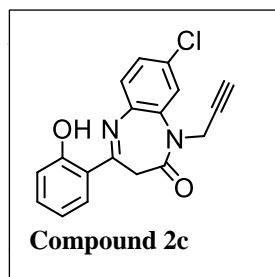


Figure S18 . DEPT 135 of compound 2c

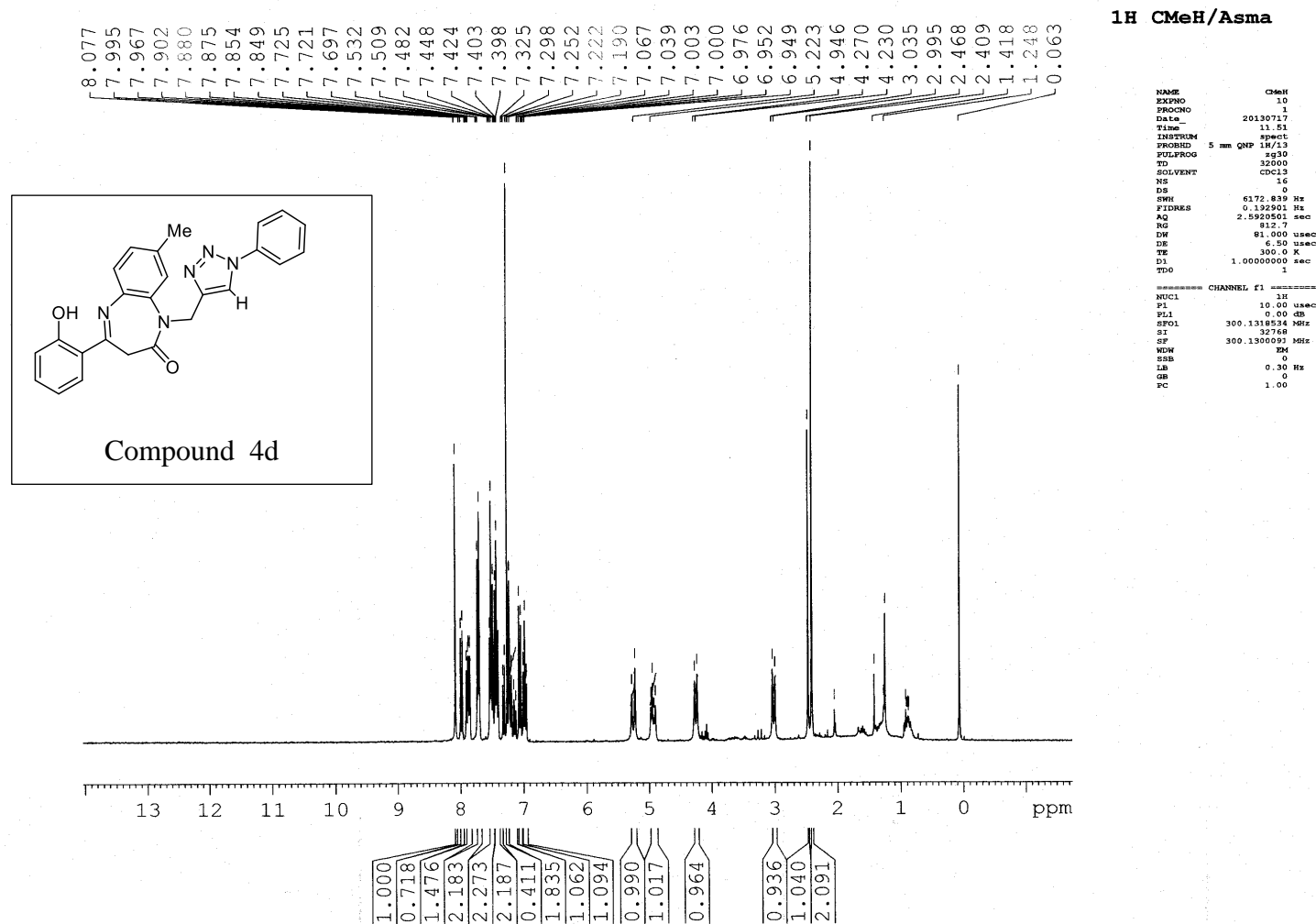


Figure S19. NMR spectra ¹H (300 MHz, CDCl₃) of compound **4d**

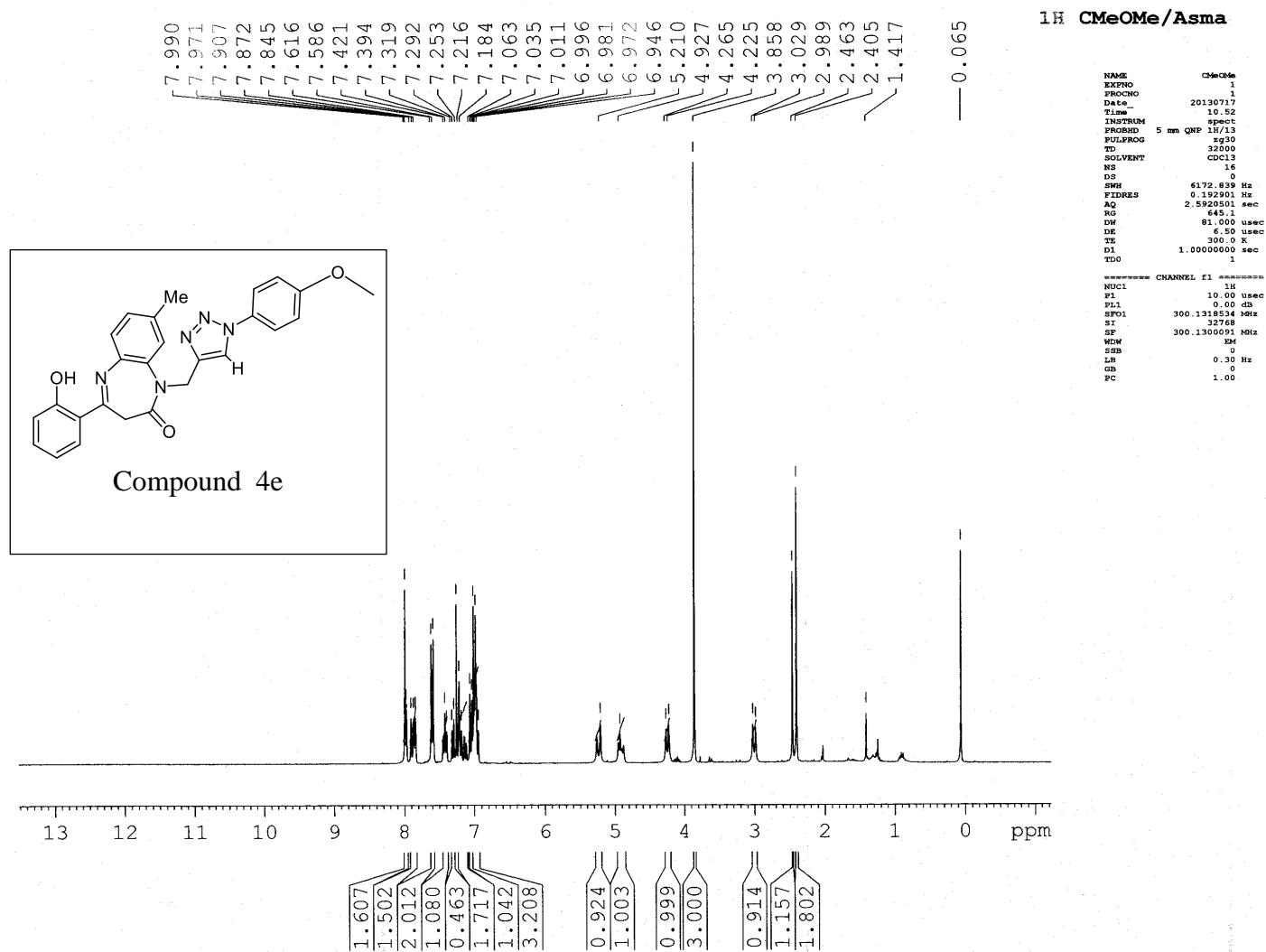


Figure S21. NMR spectra ¹H (300 MHz, CDCl₃) of compound **4e**

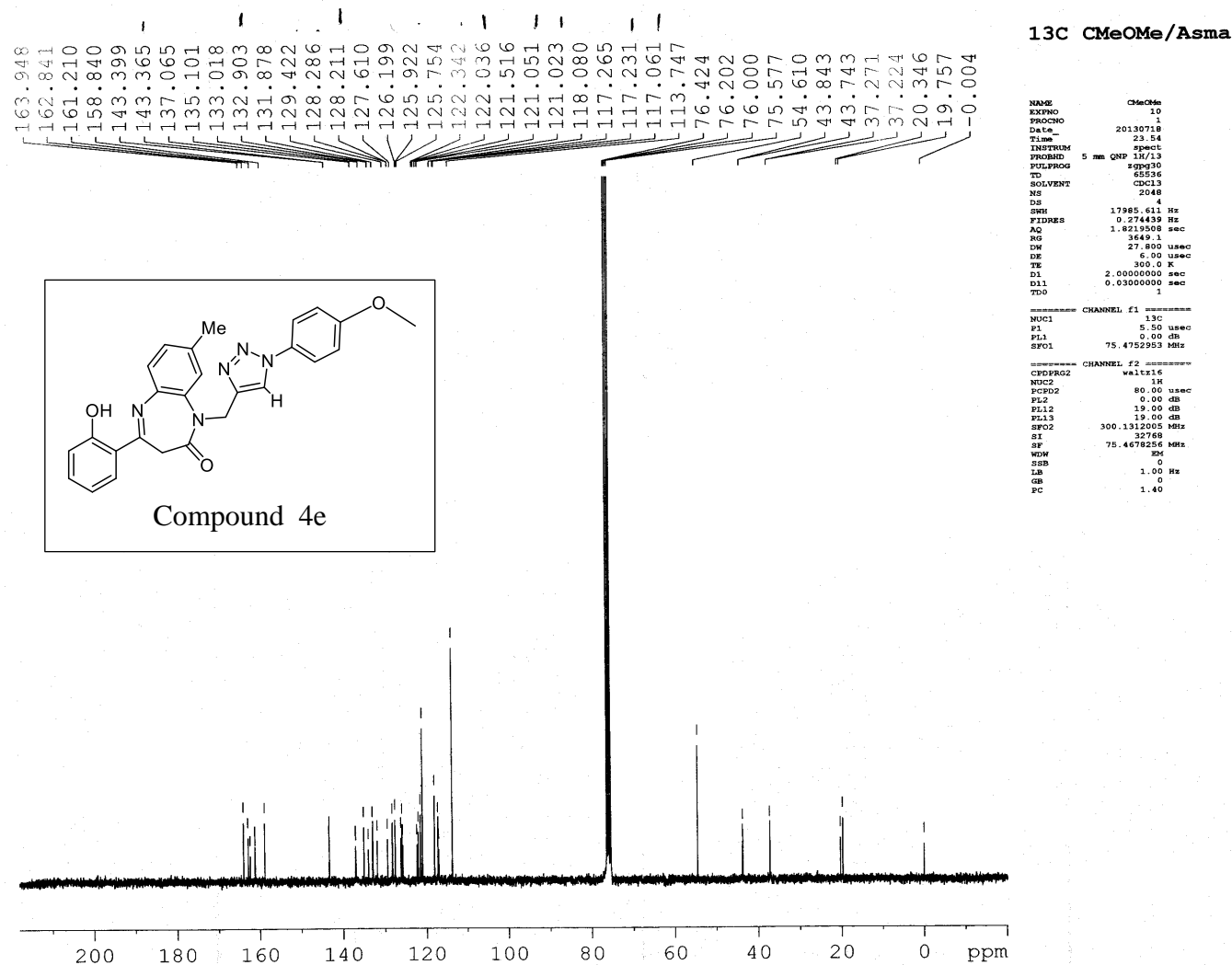


Figure S22. NMR spectra ^{13}C (75,47 MHz, CDCl_3) of compound **4e**

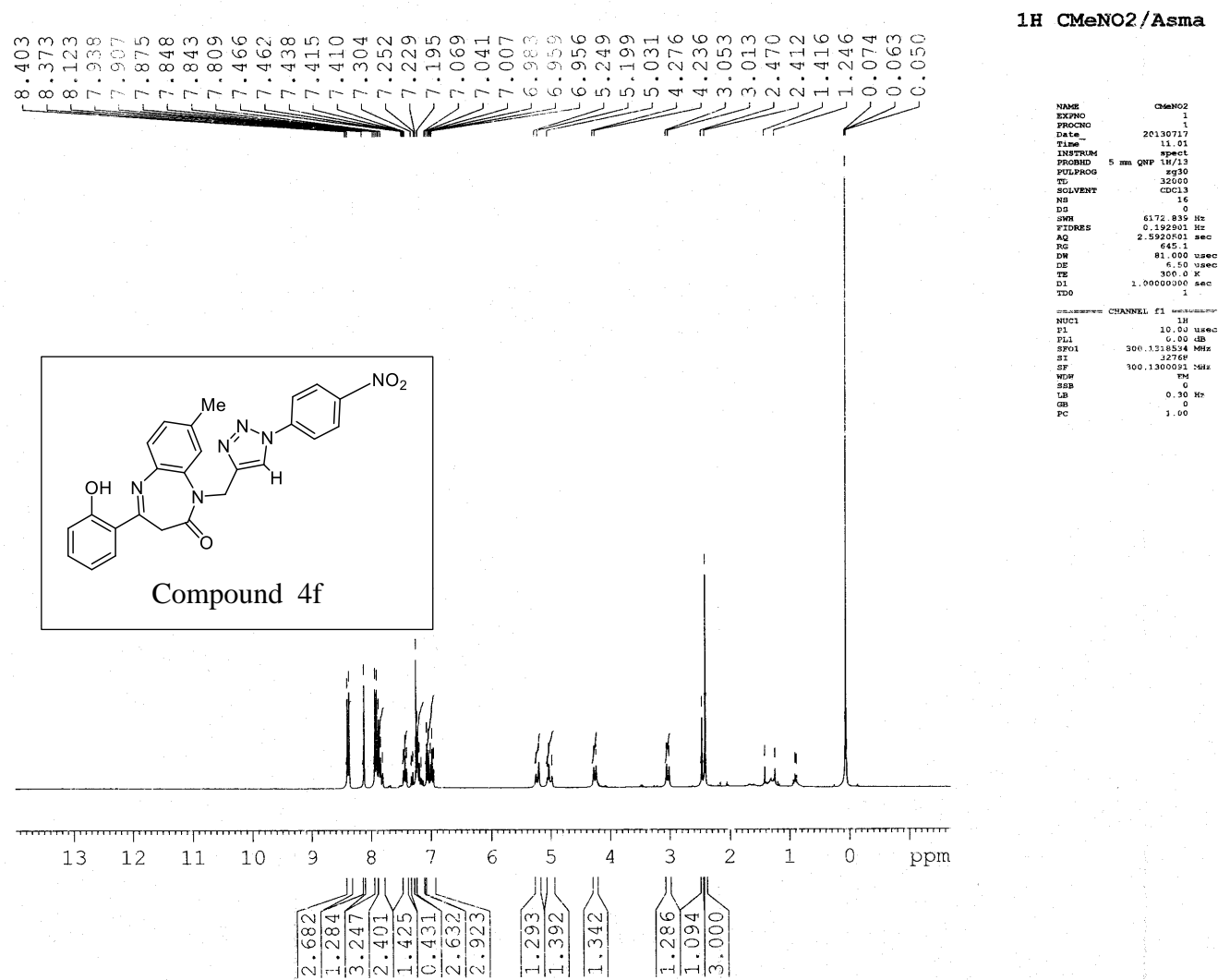


Figure S23. NMR spectra ¹H (300 MHz, CDCl₃) of compound **4f**

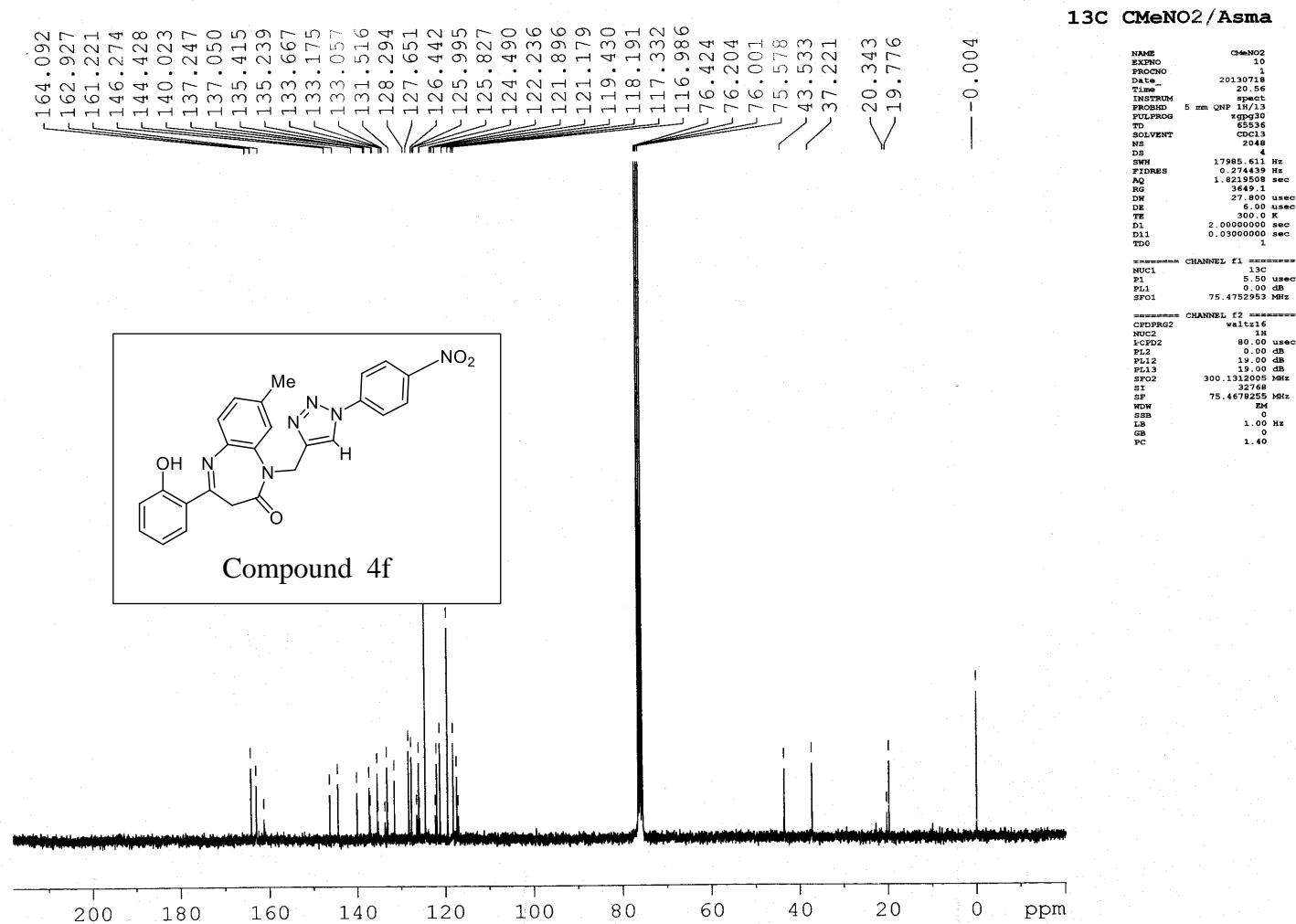


Figure S24. NMR spectra ¹³C (75,47 MHz, CDCl₃) of compound **4f**

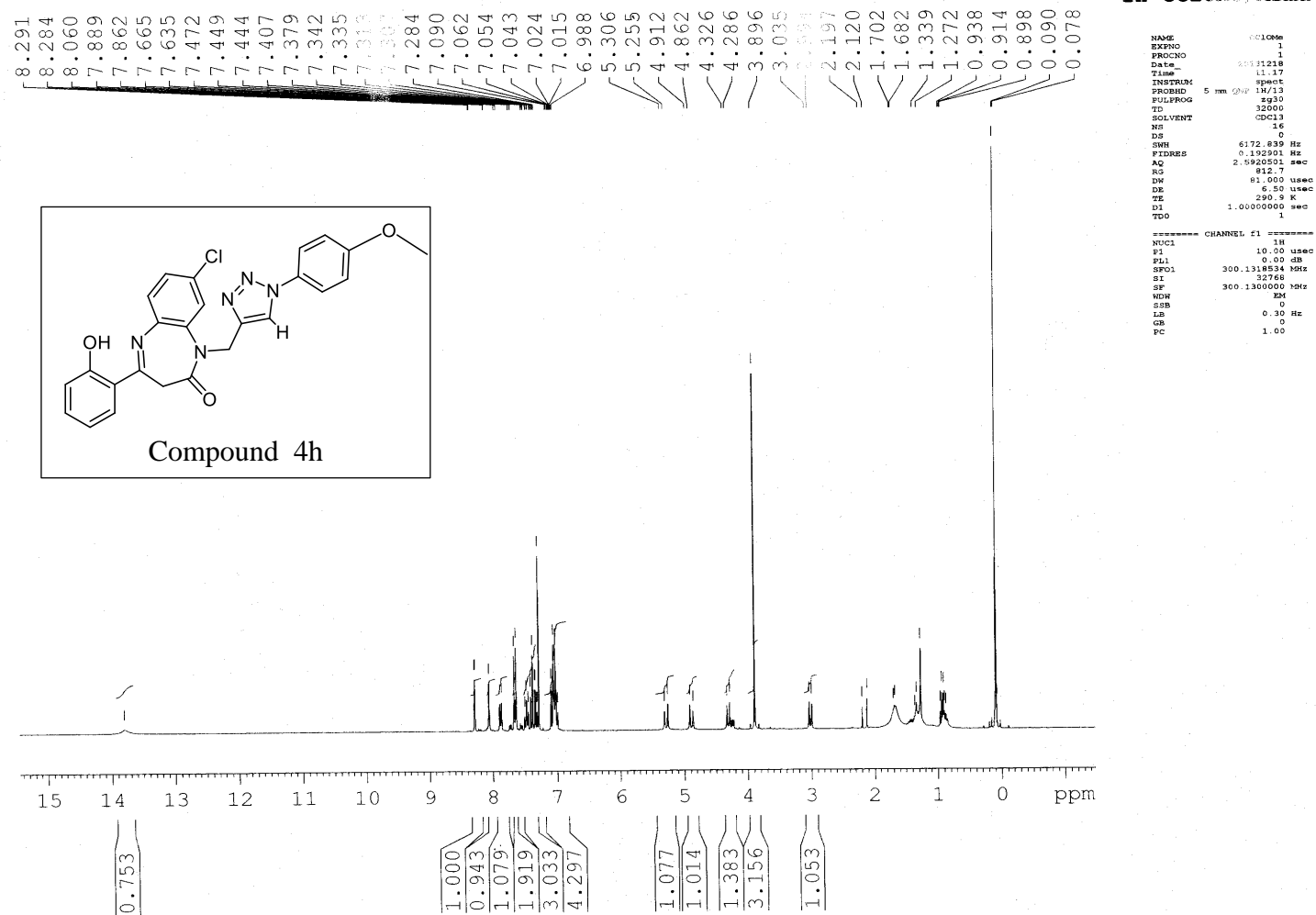


Figure S25. NMR spectra ^1H (300 MHz, CDCl_3) of compound **4h**

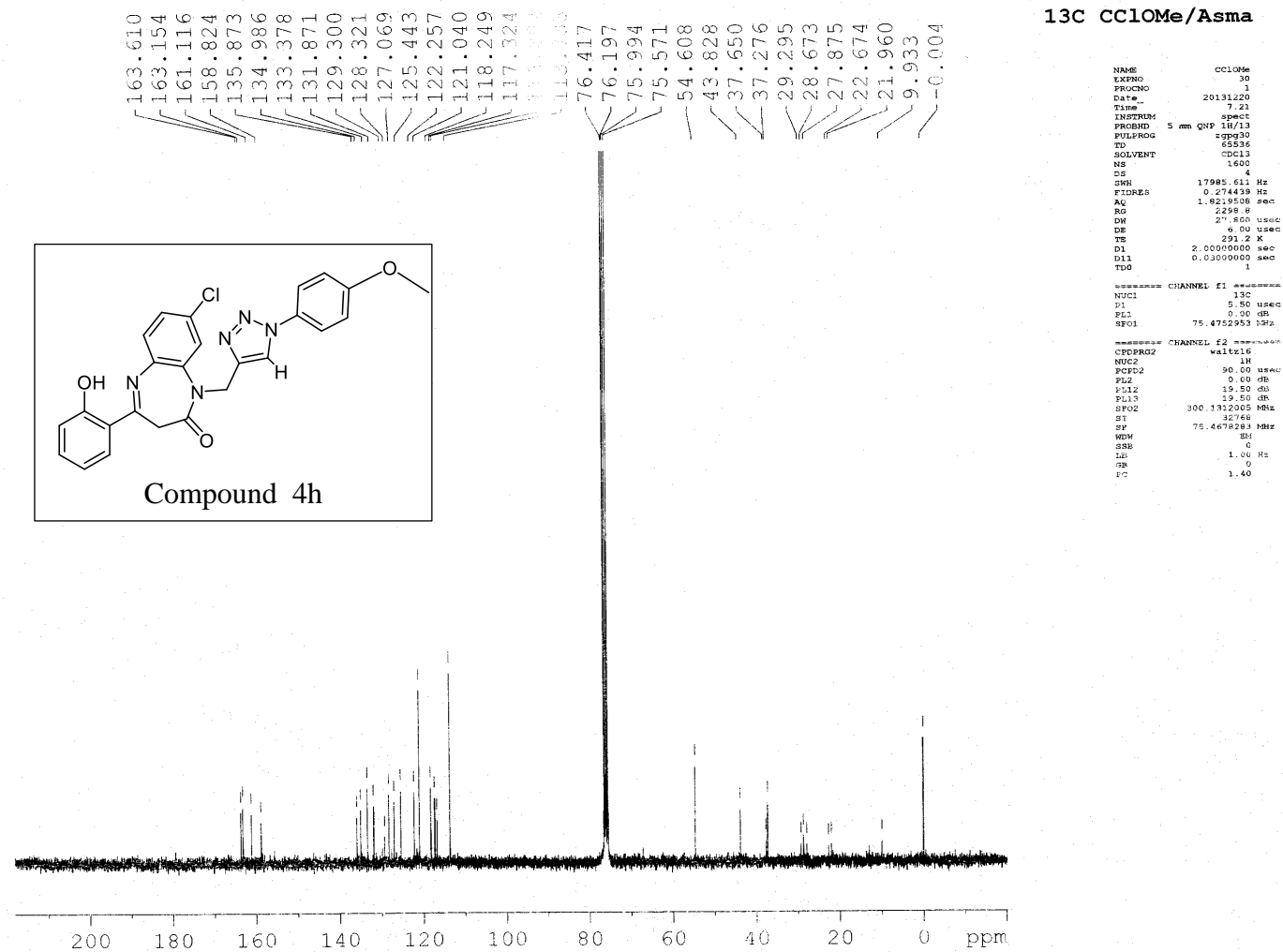


Figure S26. NMR spectra ¹³C (75,47 MHz, CDCl₃) of compound **4h**

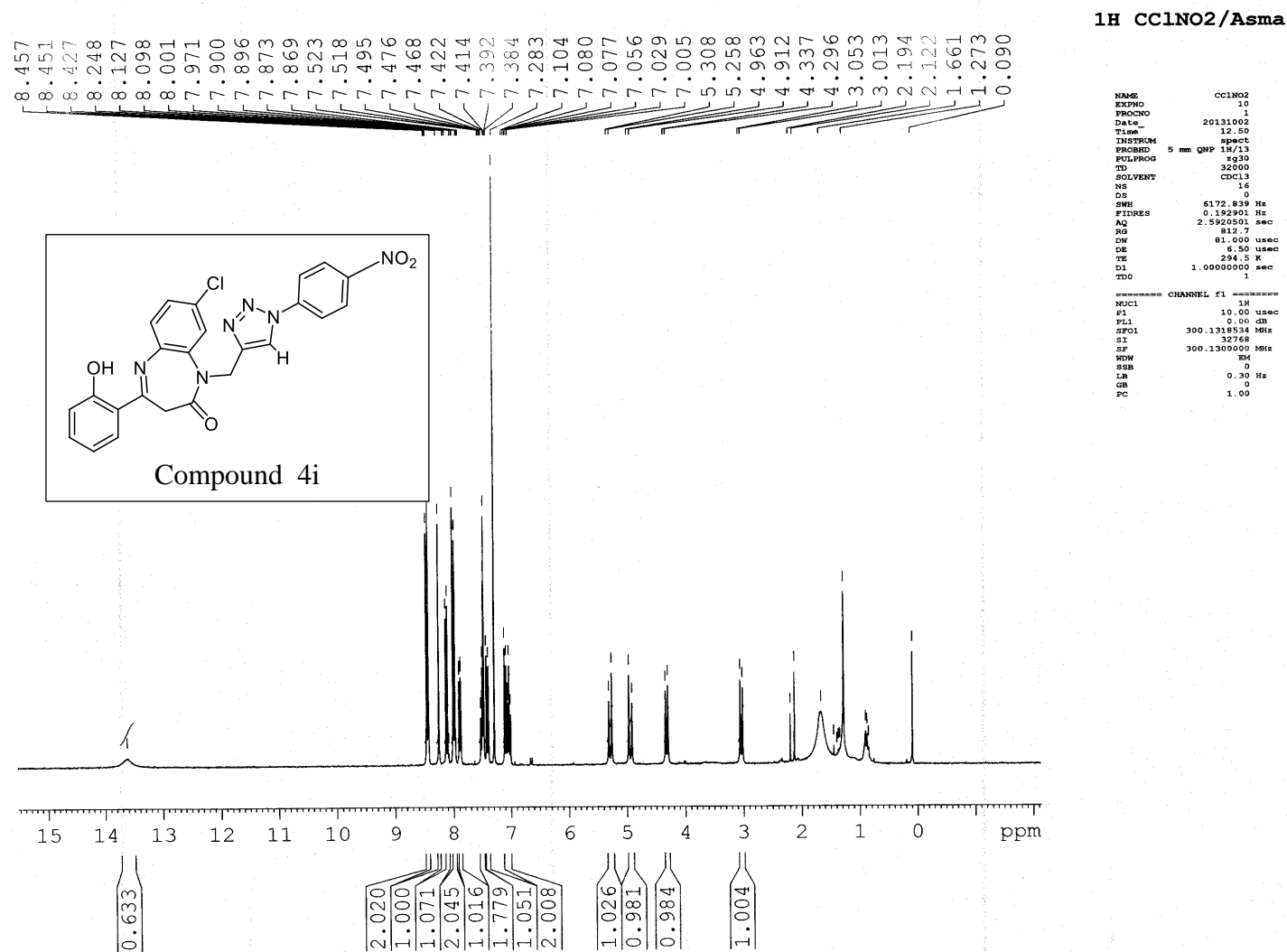


Figure S27. NMR spectra ^1H (300 MHz, CDCl_3) of compound **4i**

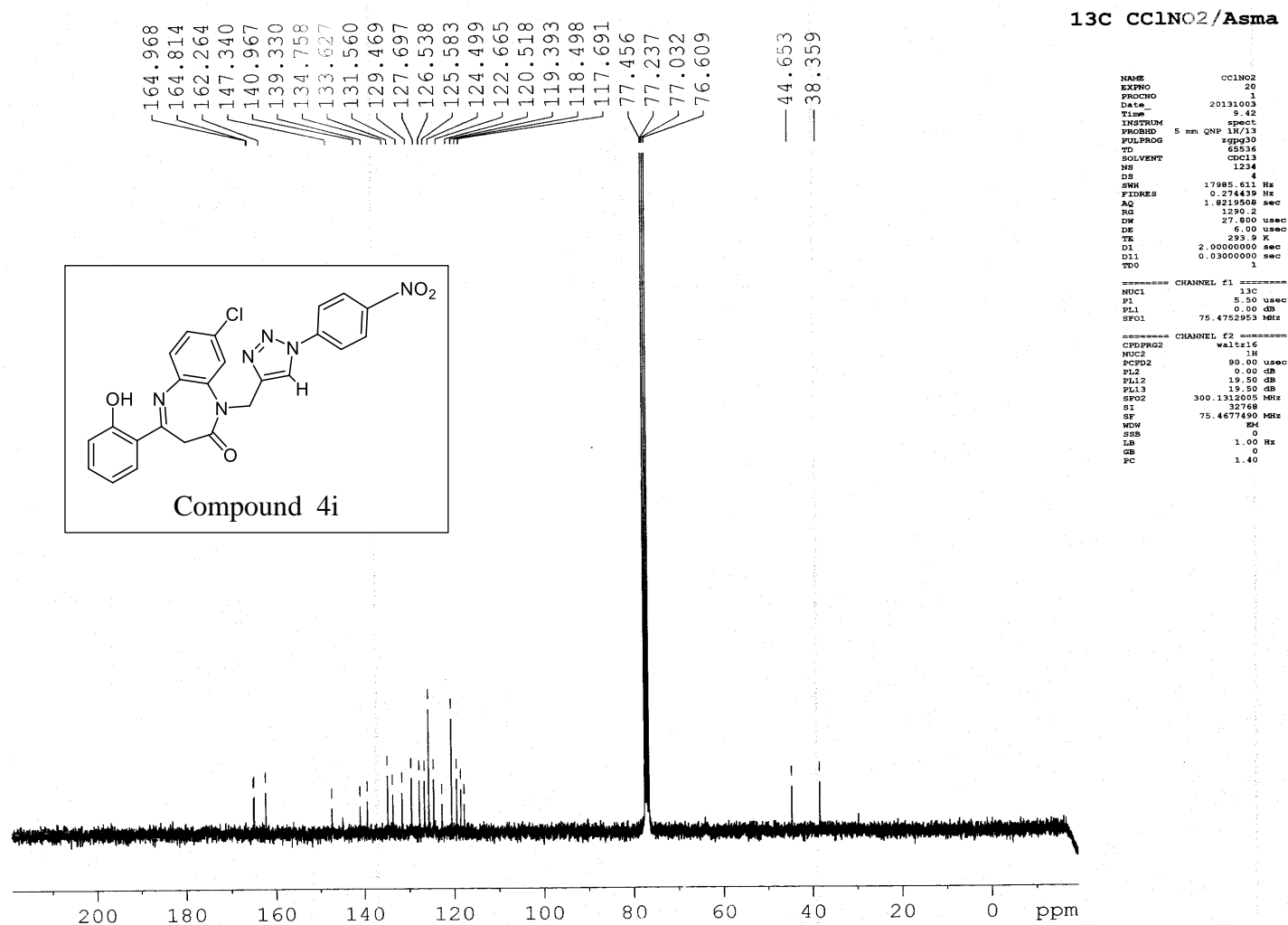


Figure S28. NMR spectra ¹³C (75,47 MHz, CDCl₃) of compound **4i**