

Supplementary material for the manuscript, ‘Prevalence and Preferred Niche of Small Eukaryotes with Mixotrophic Potentials in the Global Ocean’, accepted for publication in the *Microorganisms* journal by Dong et al.

Supplementary Table S1. Species and cellular 18S rRNA gene copy numbers used in this study to retrieve gene-cell abundance correction factors for small eukaryotes.

Taxa	Group	Reference	Method	Biovolume (μm^3)	Gene Copies per Cell
<i>Ostreococcus tauri</i>		Gong et al. 2019	bioinformatics	0.5	3.4
<i>Trebouxia</i> sp.		Gong et al. 2019	bioinformatics	1767.1	78.1
<i>Bathycoccus prasinos</i>		Zhu et al. 2005	qPCR	3.8	5.2
<i>Emiliana huxleyi</i>		Zhu et al. 2005	qPCR	64.3	2.5
<i>Mesopedinella arctica</i>		Zhu et al. 2005	qPCR	252.8	7.4
<i>Micromonas pusilla</i>		Zhu et al. 2005	qPCR	4.0	3.0
<i>Nannochloropsis salina</i>		Zhu et al. 2005	qPCR	4.0	1.0
<i>Ostreococcus tauri</i>		Zhu et al. 2005	qPCR	0.3	2.5
<i>Pelagomonas calceolata</i>		Zhu et al. 2005	qPCR	7.7	1.3
<i>Rhodomonas salina</i>	other	Zhu et al. 2005	qPCR	217.0	28.8
<i>Tetraselmis</i> sp.	eukaryotes	Zhu et al. 2005	qPCR	208.7	7.6
<i>Chrysochromulina</i> sp.		Li et al., 2022	qPCR	20.6	2.0
<i>Chrysochromulina</i> sp.		Li et al., 2022	qPCR	41.6	2.0
ChrysoH		Li et al., 2022	qPCR	5.6	1.0
DictyX		Li et al., 2022	qPCR	78.0	4.5
<i>Florenciella Parvula</i>		Li et al., 2022	qPCR	26.5	1.2
Hap2		Li et al., 2022	qPCR	47.7	3.0
<i>Rhizochromulina</i>		Li et al., 2022	qPCR	14.1	1.2
<i>Triparma</i>		Li et al., 2022	qPCR	8.2	0.8
<i>Florenciella</i> sp.		Li et al., 2022	qPCR	31.1	2.0
<i>Symbiodinium kawagutii</i>		Gong et al. 2019	bioinformatics	288.7	160.5
<i>Symbiodinium minutum</i>	dinoflagellate	Gong et al. 2019	bioinformatics	220.9	116.0
<i>Amphidinium carterae</i>		Zhu et al. 2005	qPCR	432.0	83.7
<i>Prorocentrum nux</i>		Zhu et al. 2005	qPCR	79.9	22.1
<i>Extubocellulus spinifer</i>		Godhe et al. 2008	qPCR	150.0	19.0
<i>Phaeodactylum triconutum</i>		Gong et al. 2019	bioinformatics	101.6	4.2
<i>Thalassiosira oceaniaca</i>	diatom	Gong et al. 2019	bioinformatics	179.5	17.2
<i>Nitzschia closterium</i>		Zhu et al. 2005	qPCR	640.0	28.0
<i>Thalassiosira weissflogii</i>		Zhu et al. 2005	qPCR	471.0	14.8

Supplementary Table S2. Stepwise regression analysis revealed significant variables contributing to the prevalence of mixotrophs in both surface and chlorophyll maximal layer (SUR and CML) samples, and surface (SUR) samples alone. Significance of P values and coefficient for 8 selected predictors were given in each row.

SUR and CML			SUR		
Predictor [†]	P [‡]	Coefficient	Predictor [†]	P [‡]	Coefficient
PAR	**	0.1	CML	***	0.15
NO ₃ ⁻ and NO ₂ ⁻	***	-0.08	<i>Prochlorococcus</i>	**	-0.82
Density	***	0.80	NO ₃ ⁻ &NO ₂ ⁻	***	-0.15
PIC	**	207.5	PIC	*	218.3
CML	***	0.10	PO ₄ ³⁻	*	0.28
Hbac	**	1.25	Density	*	0.78
<i>Prochlorococcus</i>	*	-0.53	PAR	**	0.12
/	/	/	NPP	.	0.08

[†]Abbreviations: PAR, photosynthetically available radiation; PIC, particulate inorganic carbon; Hbac, heterotrophic bacteria; NPP, net primary production

[‡]Significant codes: '***' p<0.001, '**' p<0.01, '*' p<0.05, '.' p<0.1

Supplementary figures

Figure S1. Redundancy analysis for environmental impact on the distribution of trophic groups in surface oceans, derived from 18S rRNA gene corrected cell abundances. Only 17 *Tara Oceans* lineages passed Escoufier selectin (0.9 threshold) were shown, and adjusted R^2 of the analysis is of 0.54. Environmental variables selected were denoted in cyan-blue arrows and text. Abbreviations: PAR, photosynthetically active radiations; CML, chlorophyll maximum layer; MLD, mixed layer depth; POC, particulate organic carbon; PIC, particulate inorganic carbon; Pro, *Prochlorococcus*.

Figure S2. Linear regression between $TI_{A/H}$ (all in cell abundances) and resource variables retrieved from the SUR (104 samples) and SUR and CML dataset (165 samples). Different significance levels were marked with different number of red stars (one star if $0.001 < p < 0.05$ and two stars if $p < 0.001$), and shaded bands are pointwise 95% confidence interval on the fitted values (the line). Abbreviations: PAR, photosynthetically active radiations; N: inorganic nutrients of nitrate (NO_3^-) and nitrite (NO_2^-); Hbac: heterotrophic bacteria; CML, chlorophyll maximal layer.

Figure S3. Linear regression between TI_g (all in cell abundances) and single environmental variables that have shown significant correlations, retrieved from all 165 SUR and CML samples. Different significance levels were marked with different number of red stars (one star if $0.001 < p < 0.05$ and two stars if $p < 0.001$), and shaded bands are pointwise 95% confidence interval on the fitted values (the line). Abbreviations: CML, chlorophyll maximal layer; Chl *a*, total chlorophyll *a*, Pro, *Prochlorococcus*.

Figure S1

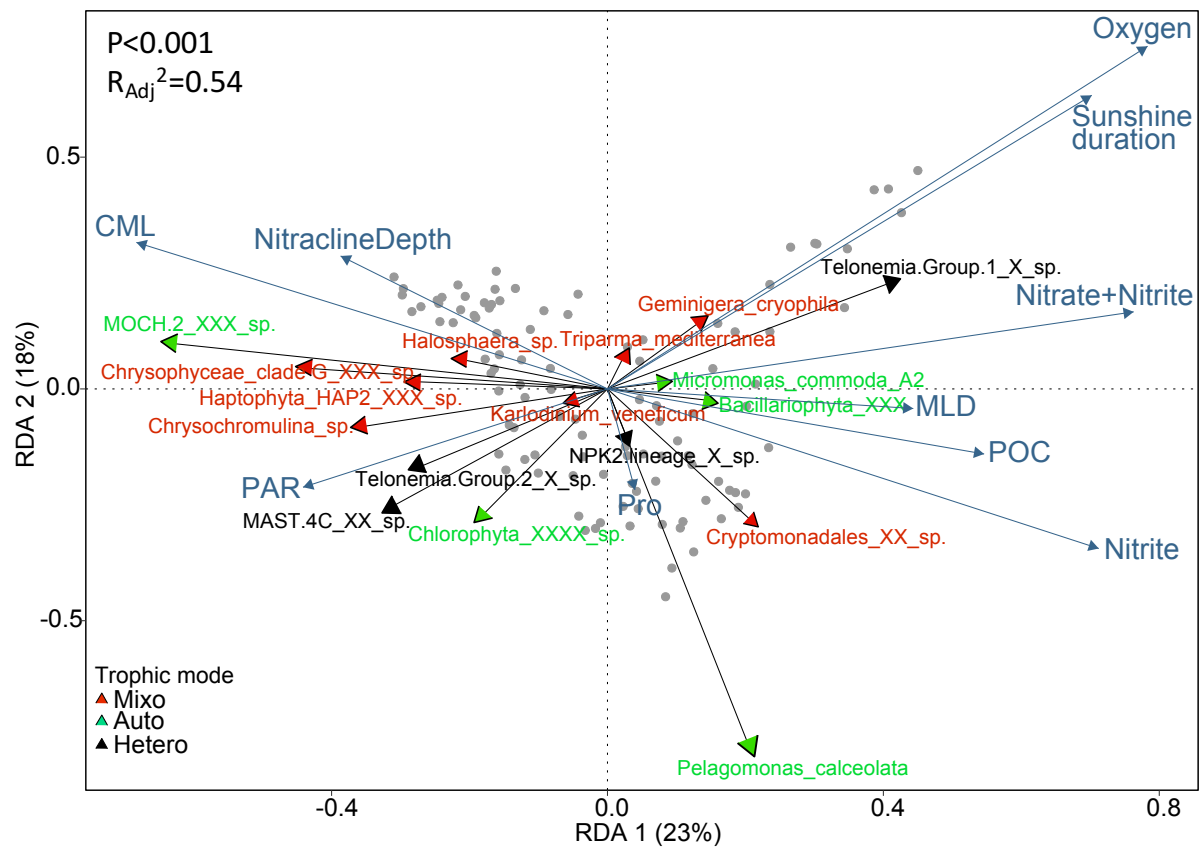


Figure S2

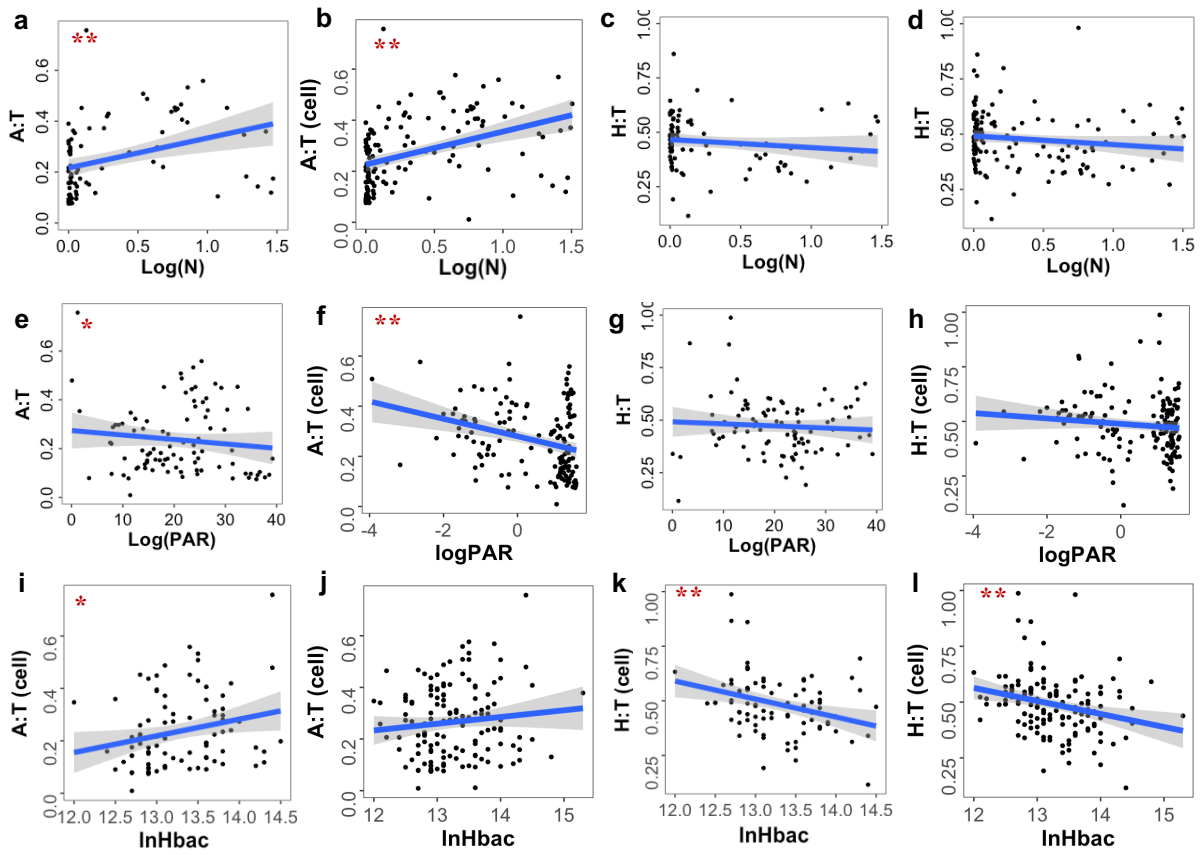
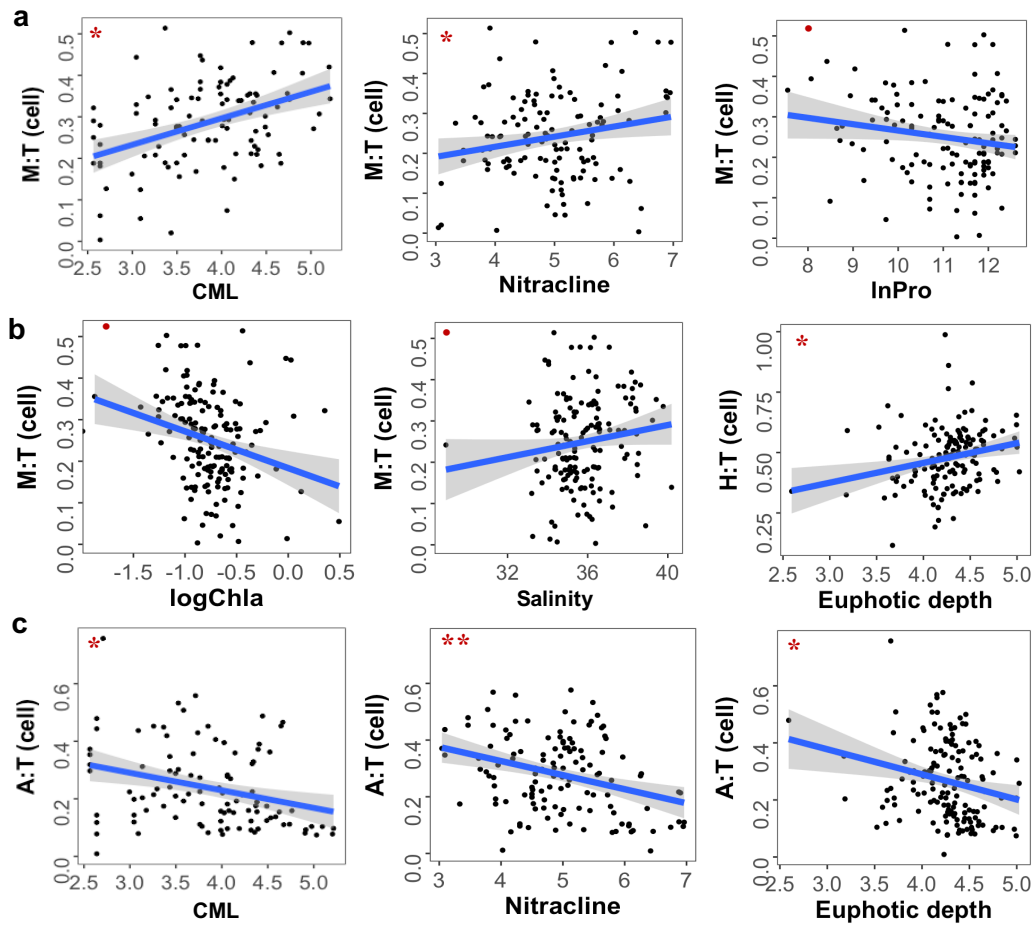


Figure S3



References

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