

# Supplementary materials

## **Antibacterial Activities and Life Cycle Stages of *Asparagopsis armata*: Implications of the Metabolome and Microbiome**

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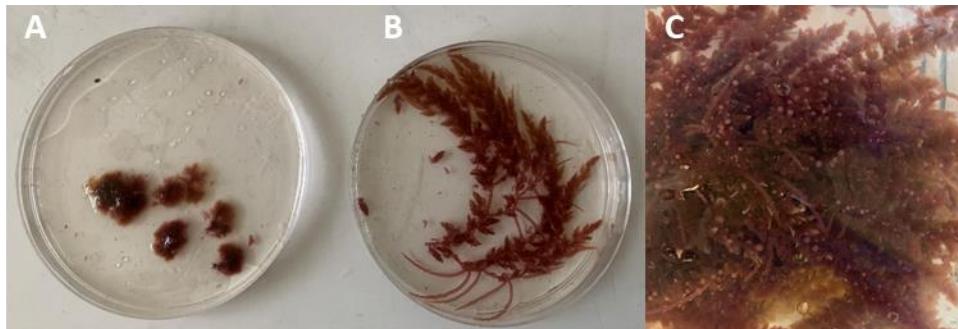
## Table of contents

Supplementary methods .....	4
Supplementary method 1: Pictures of the stages of <i>A. armata</i> life cycle collected in Banyuls-sur-Mer with the tetrasporophyte (A), the gametophyte without cystocarps (B) and the gametophyte with developed cystocarps (C).....	4
Supplementary method 2: Details of data pre-processing for the metabolomics study.....	4
Supplementary method 3: Composition of bacterial media. The quantities (in g) correspond to the preparation of 1 L of medium.....	5
Supplementary figures.....	6
Figure S1: Hierarchical clustering analysis of the metabolome of three stages of the life cycle of <i>A. armata</i> analysed in UHPLC-ESI(-)-HRMS/MS (distance measure: Euclidean, clustering algorithm: Ward).....	6
Figure S2: Chromatogram of a quality control sample analysed with the UHPLC-ESI(-)-HRMS spectrometer and displaying the ion at $m/z$ 572.50134 [M-H] <sup>-</sup> (most intense isotopic mass) of the molecule C <sub>5</sub> H <sub>2</sub> Br <sub>6</sub> O <sub>2</sub> with the highest intensity on the chromatogram.....	7
Figure S3: Chromatogram of the most active fractions (flash-chromatography fractionation) analysed by UHPLC-ESI(-)-HRMS/MS .....	8
Figure S4: ESI(-)-MS/MS spectrum of C <sub>5</sub> H <sub>2</sub> Br <sub>6</sub> O <sub>2</sub> (compound 1) and the possible fragmentation pattern. ....	9
Figure S5: ESI(-)-MS/MS spectrum of C <sub>5</sub> H <sub>2</sub> Br <sub>5</sub> ClO <sub>2</sub> (compound 2) and the possible fragmentation pattern.....	9
Figure S6: ESI(-)-MS/MS spectrum of C <sub>5</sub> H <sub>3</sub> Br <sub>4</sub> ClO <sub>2</sub> (compound 5) and the possible fragmentation pattern.....	10
Figure S7: ESI(-)-MS/MS spectrum of C <sub>4</sub> H <sub>3</sub> Br <sub>3</sub> O <sub>3</sub> (compound 6) and the possible fragmentation pattern. ....	10
Figure S8: ESI(-)-MS/MS spectrum of C <sub>5</sub> H <sub>3</sub> Br <sub>5</sub> O <sub>2</sub> (compound 7) and the possible fragmentation pattern. ....	11
Figure S9: Chromatogram of the most active fractions of the second fractionation (HPLC (Waters 1525) coupled to a UV detector (Waters 2487)) analysed by UHPLC-ESI(-)-HRMS/MS .....	12

Figure S10: EIC of ion with a $m/z$ 572.5021 (most abundant isotopic mass) corresponding to a monoisotopic $m/z$ of 566.5085 in active fractions, including 40 % H <sub>2</sub> O-MeOH (A), 30 % H <sub>2</sub> O-MeOH (B), F9 (C) and F10 (D).....	13
Figure S11: Rarefaction curves of 16S rRNA gene sequences for the gametophyte with developed cystocarps (A), the gametophyte (B) and the tetrasporophyte (C) samples. ....	14
Figure S12: Relative abundance of bacterial families associated with the three <i>A. armata</i> stages. Codes “C” represents samples of the gametophyte with developed cystocarps, “G” the gametophyte samples and “T” the tetrasporophyte samples. ..	15
Figure S13: KO pathways of the most predicted abundant functions associated with GC (A), G (B) and T (C).....	16
Figure S14: Scores plot of the multiblock PLS-DA analysis (DIABLO) of <i>A. armata</i> gametophyte ( <i>G</i> ), with developed cystocarps ( <i>GC</i> ), and tetrasporophyte ( <i>T</i> ) phases. ....	17
Figure S15: Correlations between the first dimension (A) and the second dimension (B) of each dataset (Chem= Chemistry; Metabarc= Metabarcoding) for the two PLS models. Codes “C” represents samples of the gametophyte with developed cystocarps, “G” the gametophyte samples and “T” the tetrasporophyte samples. ....	18
Figure S16: Heatmap of significant ASV and metabolites of the three phases of <i>A. armata</i> (“C” represents samples of the gametophyte with developed cystocarps, “G” the gametophyte samples and “T” the tetrasporophyte samples).....	19

## **Supplementary methods**

**Supplementary method 1: Pictures of the stages of *A. armata* life cycle collected in Banyuls-sur-Mer with the tetrasporophyte (A), the gametophyte without cystocarps (B) and the gametophyte with developed cystocarps (C)**



**Supplementary method 2: Details of data pre-processing for the metabolomics study**

Data acquisitions were performed using Xcalibur 4.1.31.9 (Thermo Fisher Scientific). Raw data were converted to mzML files with MSconvert (version 3.0, from Proteowizard library). mzML files were uploaded and processed using the Galaxy web platform (version 3.3) (Giacomoni et al., 2015; Guitton et al., 2017). The workflow used for data pre-processing and used parameters are published on the Galaxy Workflow4Metabolomics platform at: [https://workflow4metabolomics.usegalaxy.fr/u/christelle\\_parchemin/w/workflowparcheminalgae](https://workflow4metabolomics.usegalaxy.fr/u/christelle_parchemin/w/workflowparcheminalgae). Briefly, the preprocessing consisted in a chromatographic peak detection (Galaxy Version 3.12+galaxy0) using the CentWave method with a minimum and maximum peak width of 5 and 60s, and 4 successive scans with an intensity above 500 000 as limit for consideration of region of interest. The chromatographic peak detection was followed by a peak grouping using the PeakDensity method step, a loess/non-linear “PeakGroups” retention time adjustment (degree of smoothing: 0.8), a peak filling and “CAMERA” peak annotation. A matrix of features with peak intensity, m/z value and retention time was generated. A clean-up based on p-Values and t-Stat outputs generated by the “CAMERA” step was performed in order to eliminate all features that are significantly detected in blanks. Then, an “inter/intra-batch” signal correction was applied using the “Batch correction” function with a “loess” regression model (span = 0.8) (Van Der Kloet et al., 2009). This step was followed by a second clean-up according to feature's CV in pool QC injections (all features with area relative standard deviation upper than 30% through pool QC injections were eliminated from the

dataset) (Thévenot et al., 2015). Finally, redundancies due to isotopes were manually eliminated (only monoisotopic mass was kept). For identification, most probable molecular formula was determined using using Sirius (v4.9.15 (Dürkop et al., 2019))), MetLin database, characteristics isotopic clusters, MS/MS spectra and comparison with literature.

### **Supplementary method 3: Composition of bacterial media. The quantities (in g) correspond to the preparation of 1 L of medium**

Code	Strain	Medium	Culture temperature (°C)	Incubation time (h)	Susceptible to antibiotic
Ea	<i>Edwardsiella anguillarum</i> (DSMZ-27202)	Luria Broth <sup>a</sup>	27	24	Kanamycine
Lg	<i>Lactococcus garvieae</i> (CIP102507T)	Brain Heart Infusion <sup>b</sup>	37	24	Erythromycine
Tm	<i>Tenacibaculum maritimum</i> (CIP103528T)	Marine Broth <sup>c</sup>	27	24	Ampicilline
Va	<i>Vibrio anquillarum</i> (CIP 63.36T)	Marine Broth <sup>c</sup>	27	24	Kanamycine
Vh	<i>Vibrio harveyi</i> (CIP103192T)	Marine Broth <sup>c</sup>	27	24	Kanamycine
Yr	<i>Yersina ruckeri</i> (CIP82.80T)	Marine Broth <sup>c</sup>	27	24	Amoxicilline

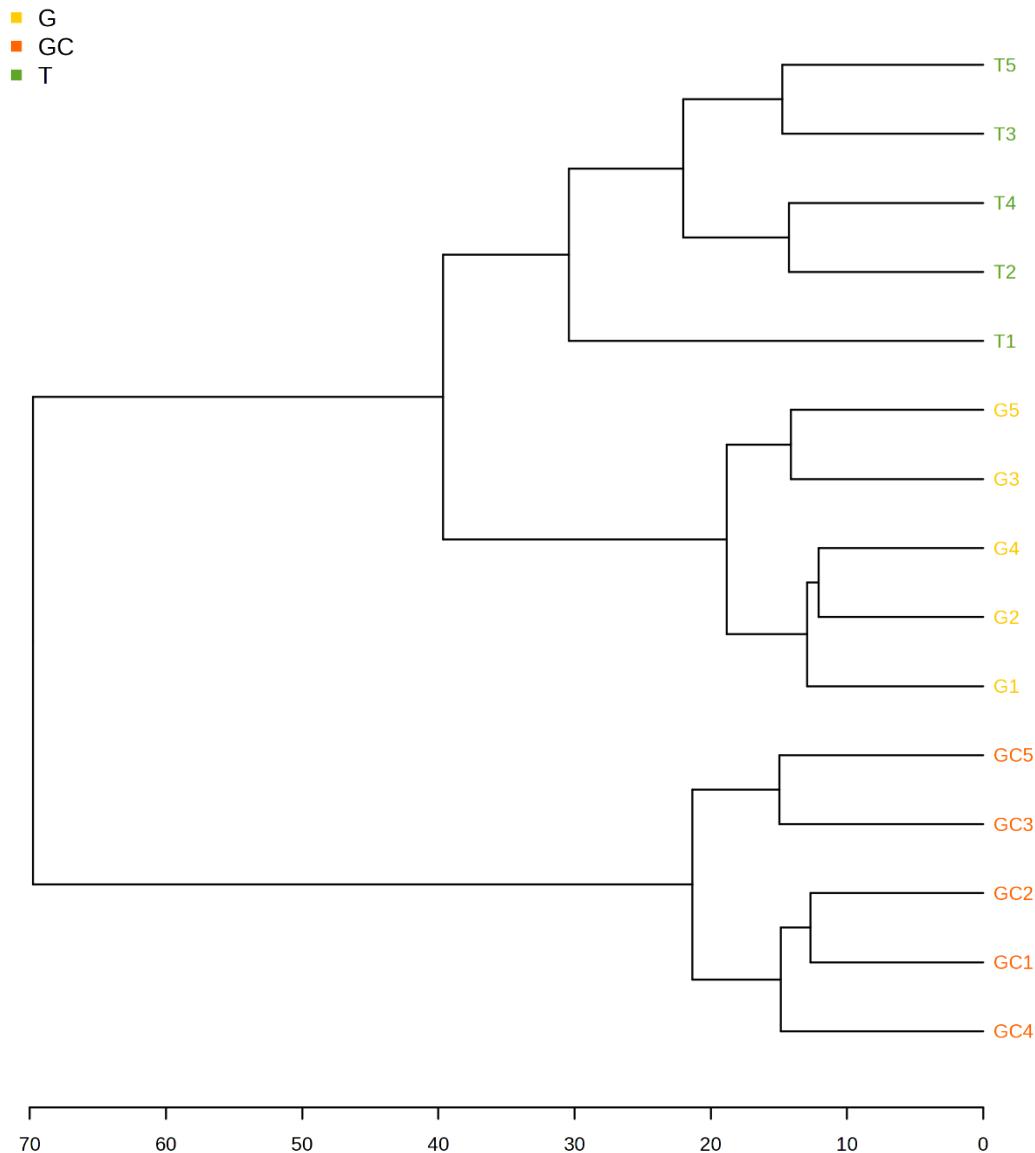
<sup>a</sup>Luria Broth = 5 g of yeast extract (Yeast Extract 70161-500G (Fluka Analytical, Sigma-Aldrich®, Saint-Louis USA)), 10g of peptone (Peptone from casein, pancreatic digest 70169-500G (Fluka Analytical, Sigma-Aldrich®, Saint-Louis USA)), 10g of sea salts (Sea salts S9883-500G (Sigma Life Science, Sigma-Aldrich®, Saint-Louis USA)) and 15 g of bacteriological agar (Bacteriological agar A5306-250G (Sigma-Aldrich®, Saint-Louis USA)).

<sup>b</sup>Brain Heart Infusion Agar = 52 g for 1 L of medium (Brain Heart Infusion Agar 70138-500G (Sigma-Aldrich®, Saint-Louis USA))

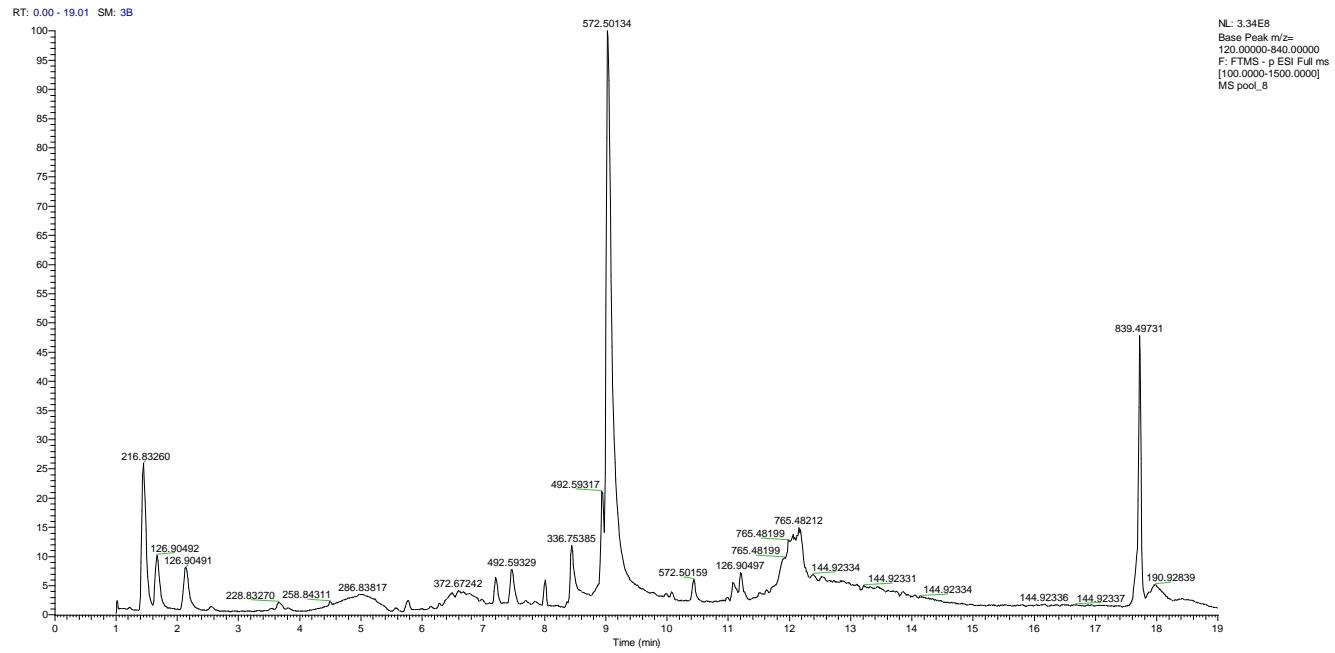
<sup>c</sup>Marine Broth = 37.4 g for 1 L of medium (Difco™ Marine Broth 2216 (Becton, Dickinson and Company, Sparks, USA))

## Supplementary figures

**Figure S1: Hierarchical clustering analysis of the metabolome of three stages of the life cycle of *A. armata* analysed in UHPLC-ESI(-)-HRMS/MS (distance measure: Euclidean, clustering algorithm: Ward)**

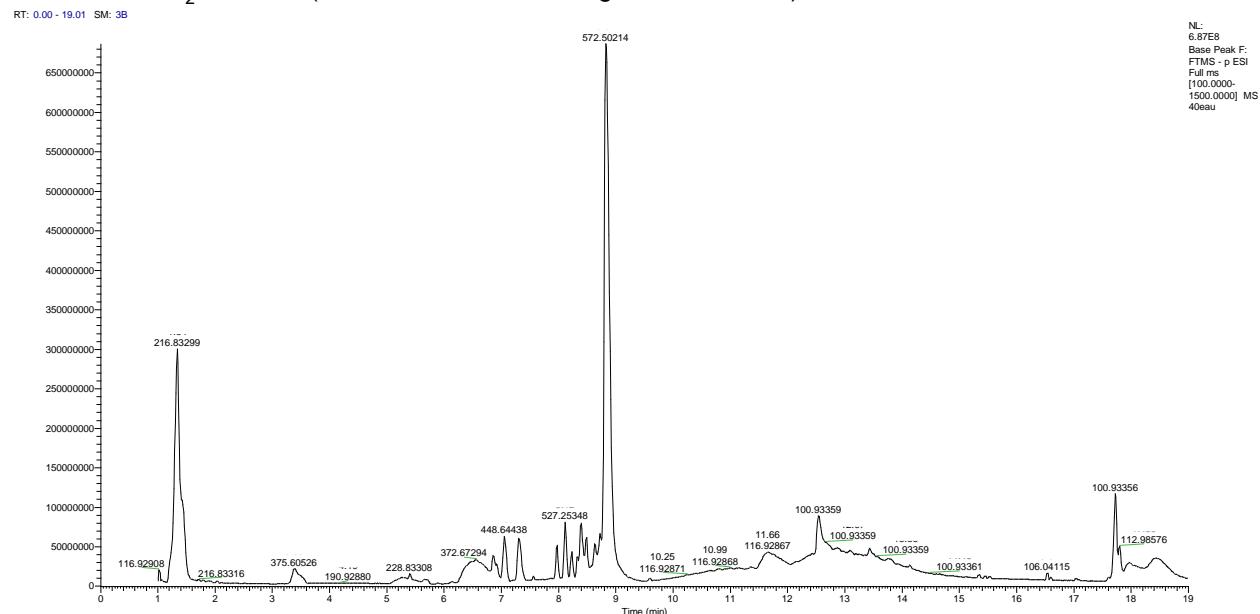


**Figure S2: Chromatogram of a quality control sample analysed with the UHPLC-ESI(-)-HRMS spectrometer and displaying the ion at  $m/z$  572.50134 [M-H] $^-$  (most intense isotopic mass) of the molecule C<sub>5</sub>H<sub>2</sub>Br<sub>6</sub>O<sub>2</sub> with the highest intensity on the chromatogram.**

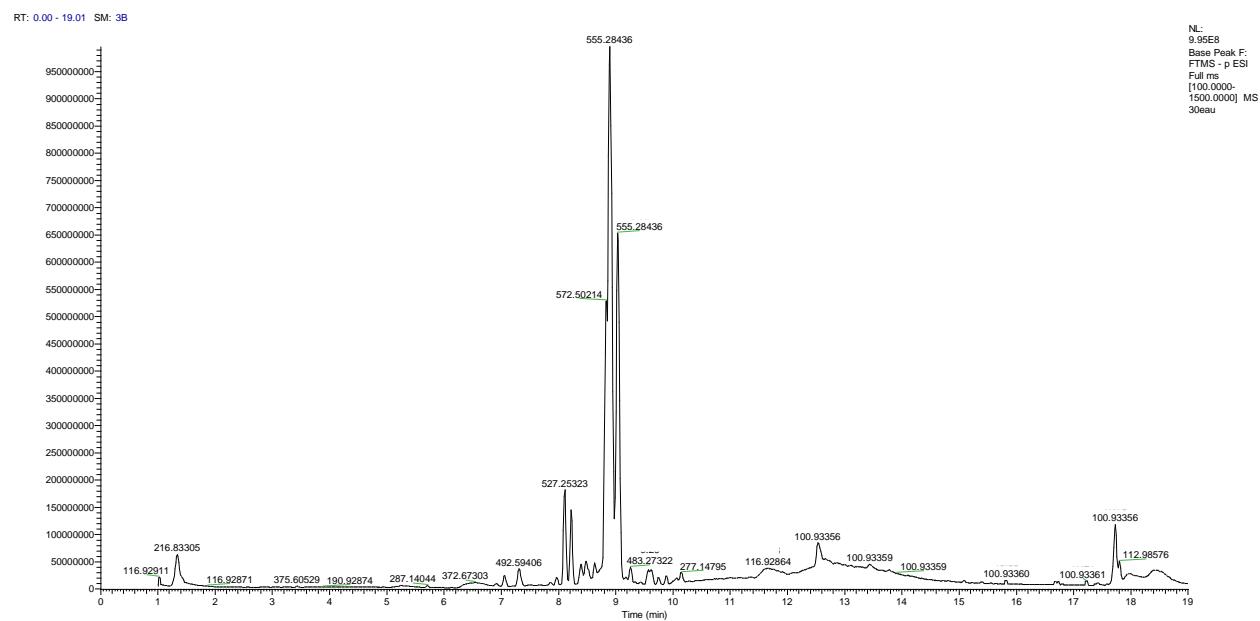


**Figure S3: Chromatogram of the most active fractions (flash-chromatography fractionation) analysed by UHPLC-ESI(-)-HRMS/MS**

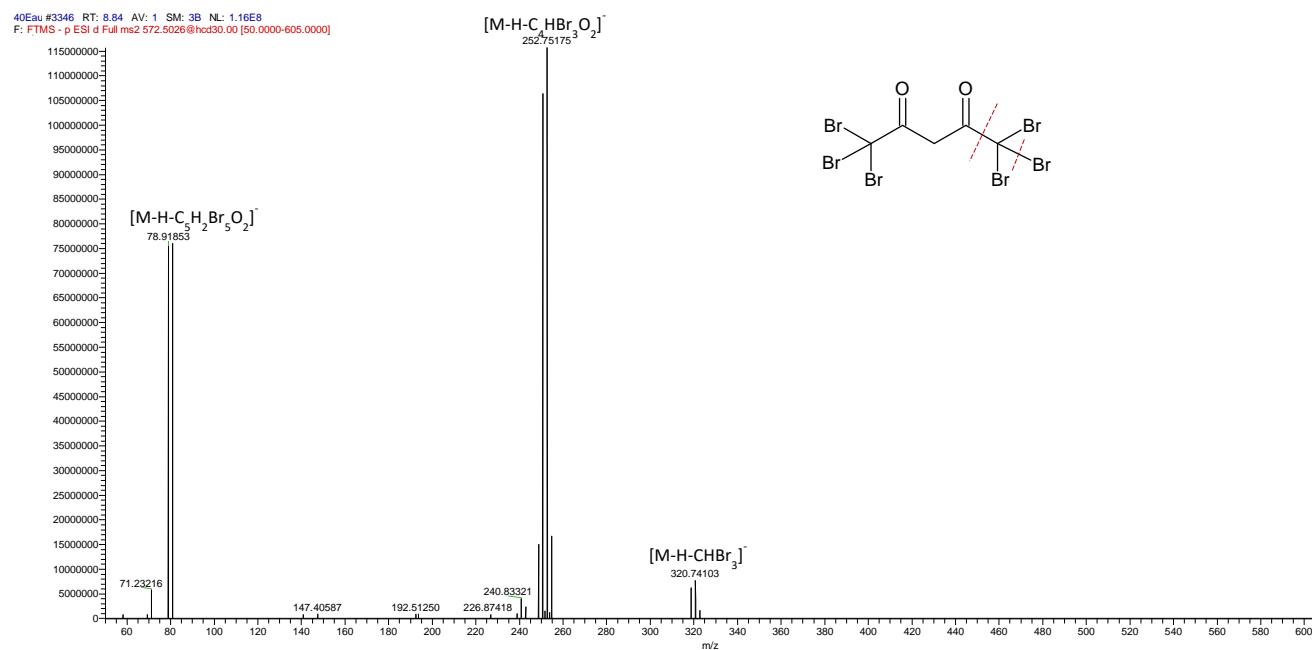
**A. 40 % H<sub>2</sub>O-MeOH (98.9±0.4 % of bacterial growth inhibition)**



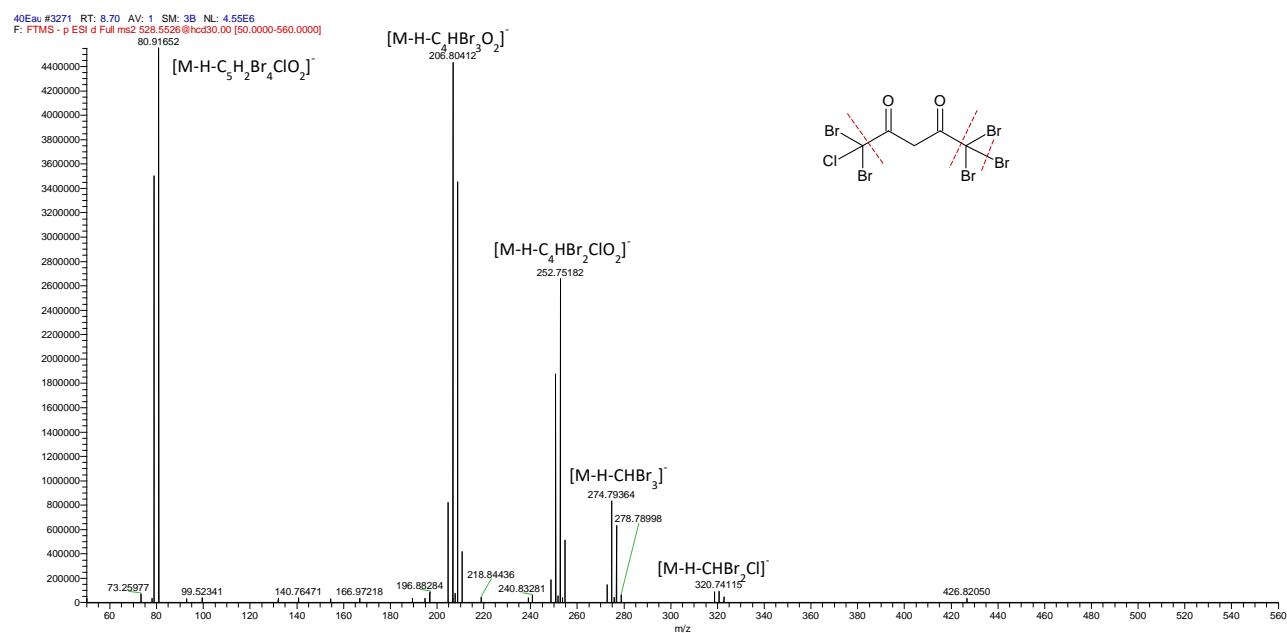
**B. 30 % H<sub>2</sub>O-MeOH (97.6±0.8 % of bacterial growth inhibition)**



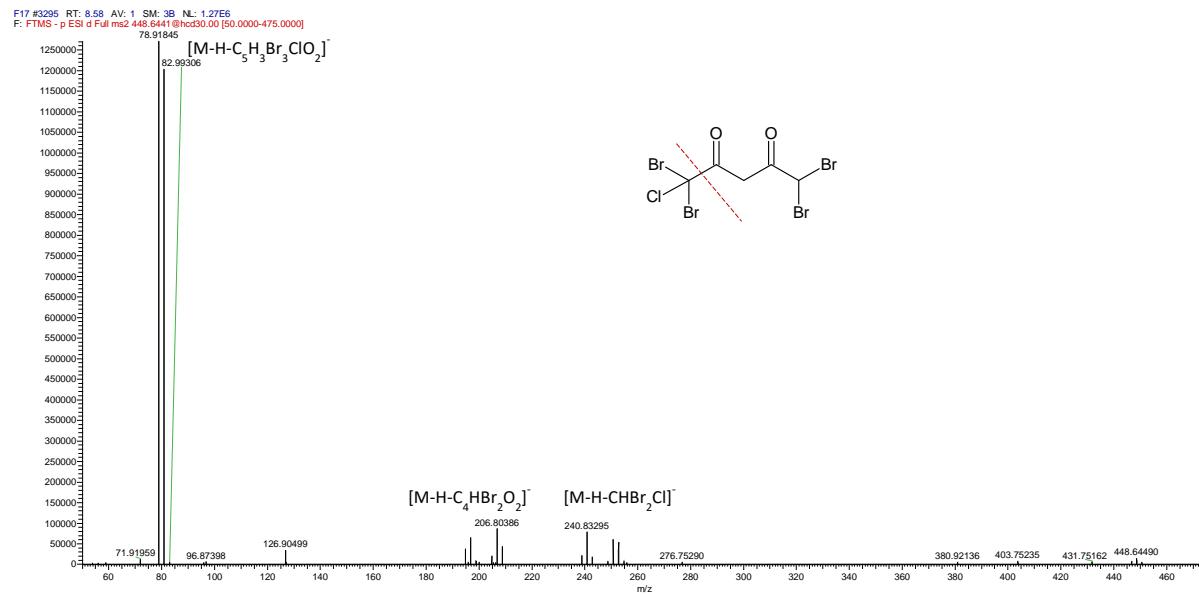
**Figure S4:** ESI(-)-MS/MS spectrum of C<sub>5</sub>H<sub>2</sub>Br<sub>6</sub>O<sub>2</sub> (compound 1) and the possible fragmentation pattern.



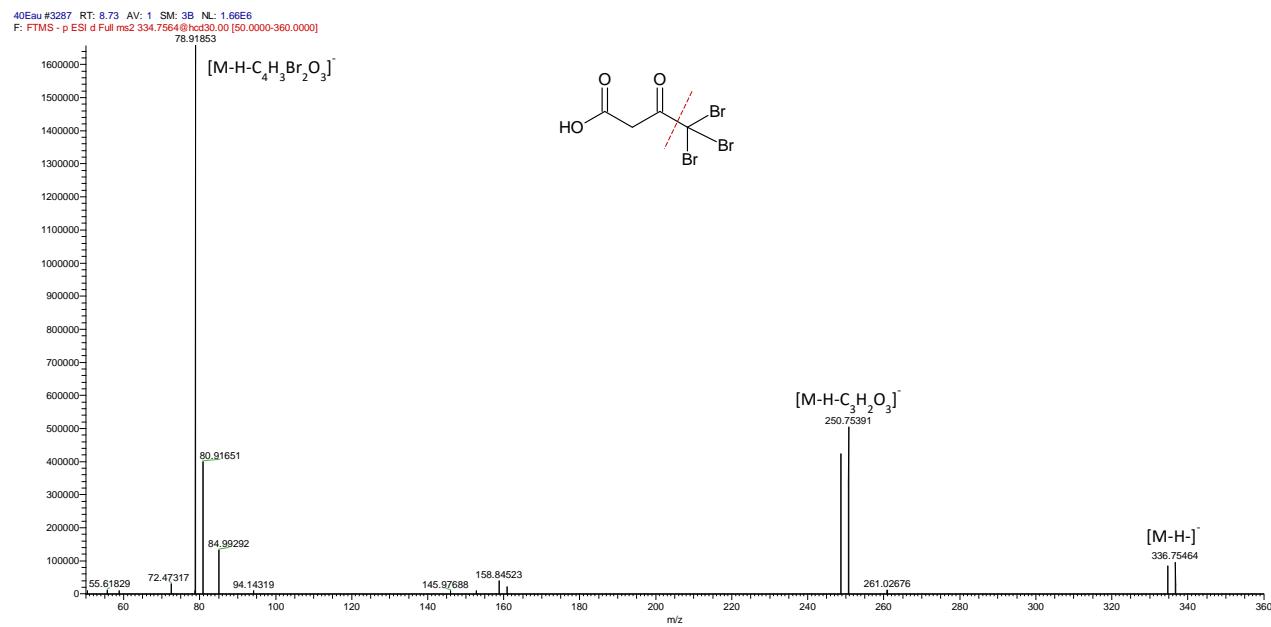
**Figure S5:** ESI(-)-MS/MS spectrum of C<sub>5</sub>H<sub>2</sub>Br<sub>5</sub>ClO<sub>2</sub> (compound 2) and the possible fragmentation pattern.



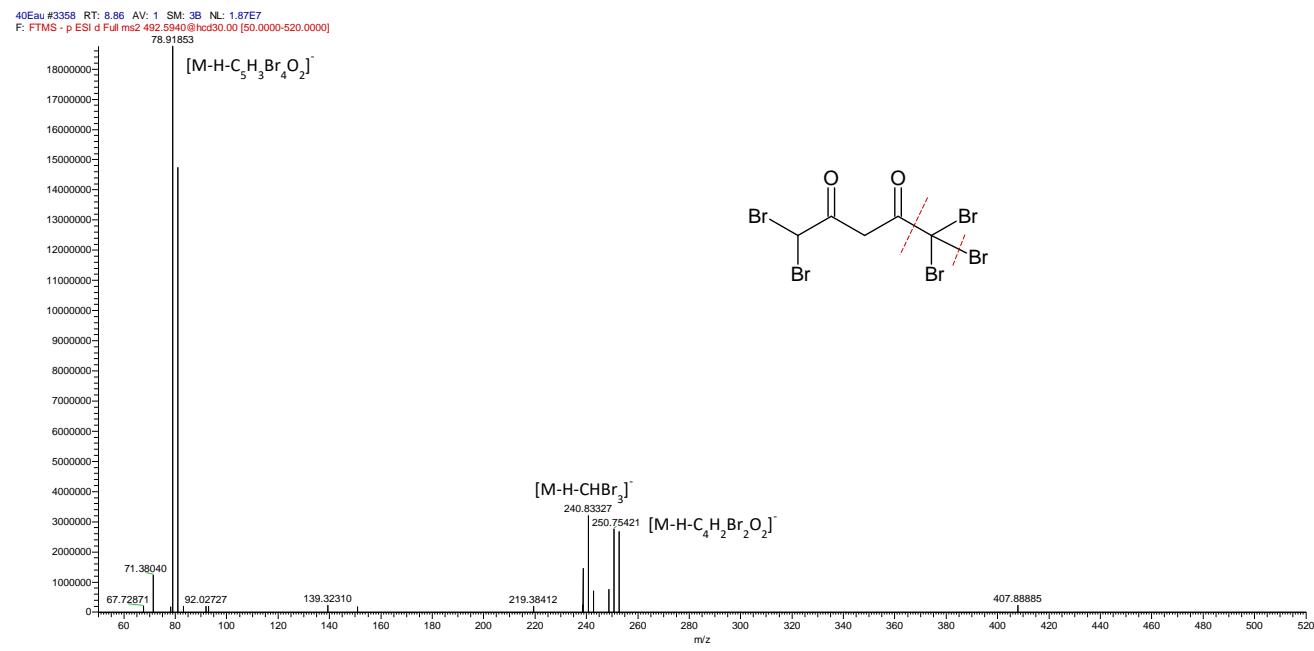
**Figure S6: ESI(-)-MS/MS spectrum of C<sub>5</sub>H<sub>3</sub>Br<sub>4</sub>ClO<sub>2</sub> (compound 5) and the possible fragmentation pattern.**



**Figure S7: ESI(-)-MS/MS spectrum of C<sub>4</sub>H<sub>3</sub>Br<sub>3</sub>O<sub>3</sub> (compound 6) and the possible fragmentation pattern.**

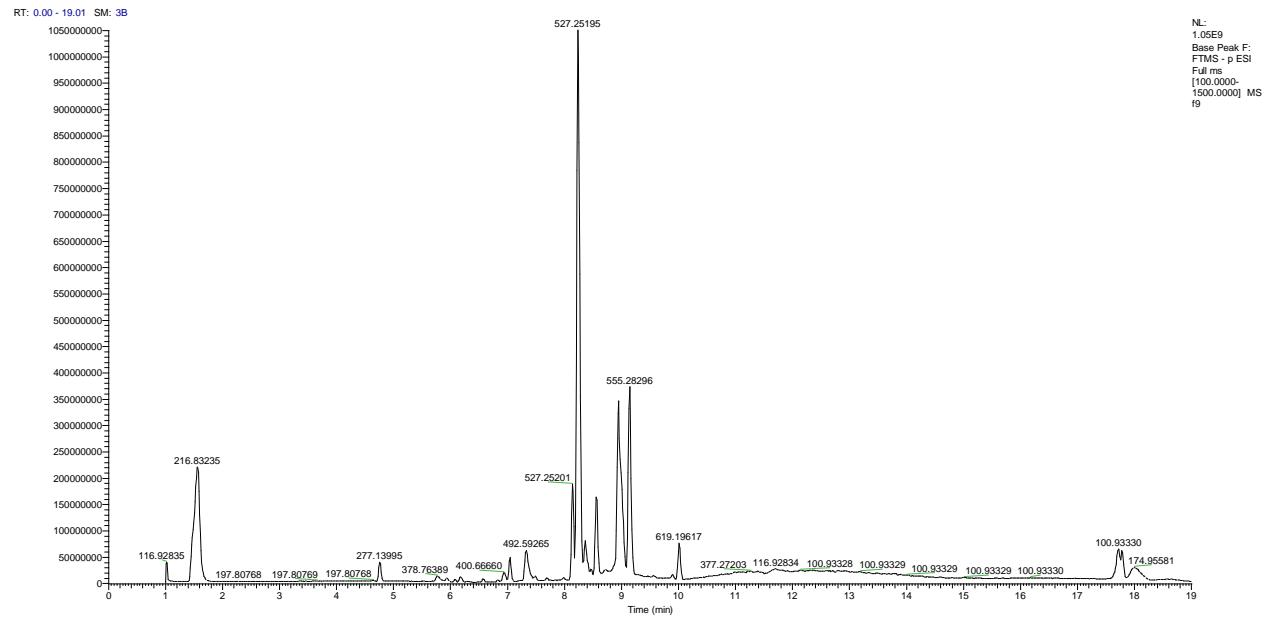


**Figure S8: ESI(-)-MS/MS spectrum of C<sub>5</sub>H<sub>3</sub>Br<sub>5</sub>O<sub>2</sub> (compound 7) and the possible fragmentation pattern.**

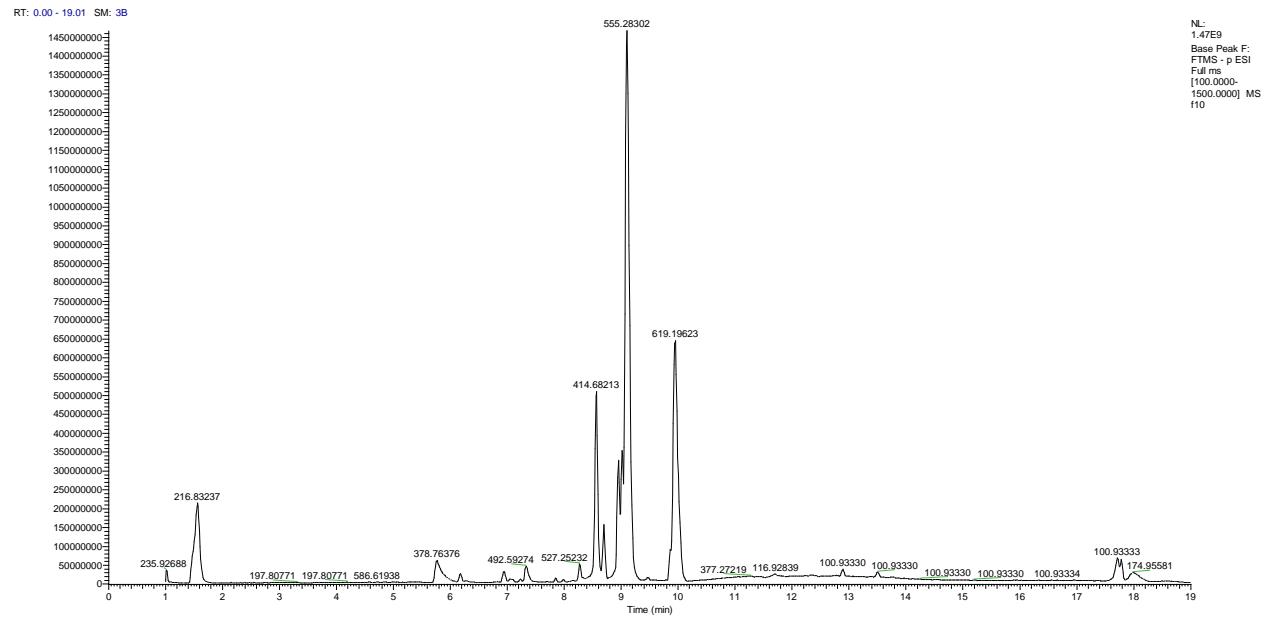


**Figure S9: Chromatogram of the most active fractions of the second fractionation (HPLC (Waters 1525) coupled to a UV detector (Waters 2487)) analysed by UHPLC-ESI(-)-HRMS/MS**

**A. F9 (99±0.0 % of bacterial growth inhibition)**

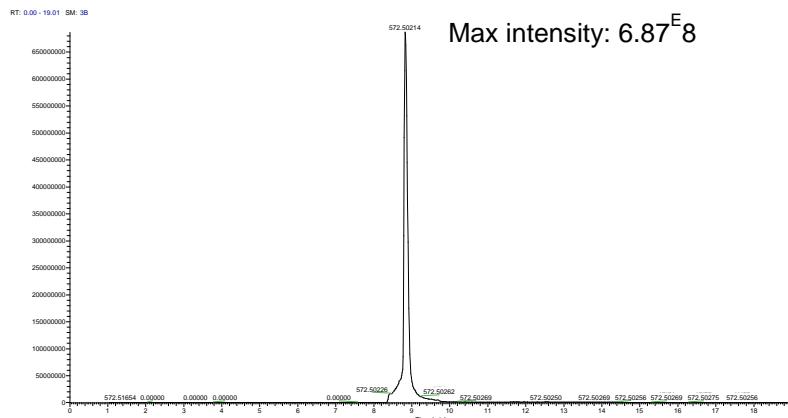


**B. F10 (93.7±0.3 % of bacterial growth inhibition)**

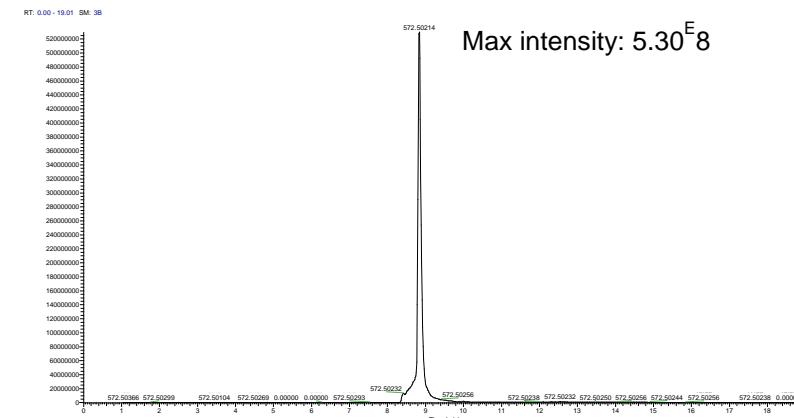


**Figure S10: EIC of ion with a  $m/z$  572.5021 (most abundant isotopic mass) corresponding to a monoisotopic  $m/z$  of 566.5085 in active fractions, including 40 % H<sub>2</sub>O-MeOH (A), 30 % H<sub>2</sub>O-MeOH (B), F9 (C) and F10 (D).**

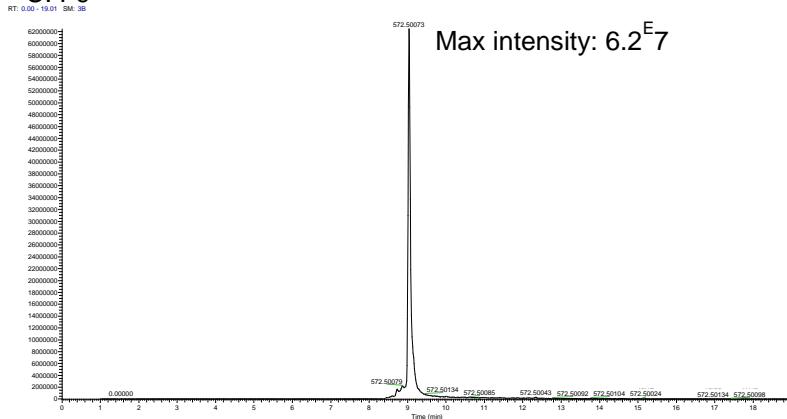
A. 40% H<sub>2</sub>O-MeOH



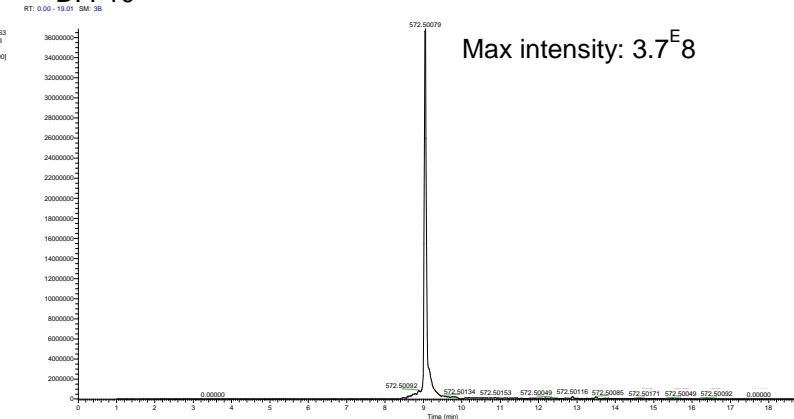
B. 30% H<sub>2</sub>O-MeOH



C. F9

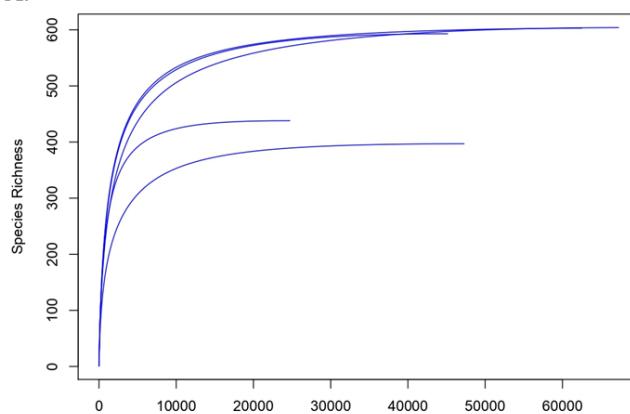


D. F10

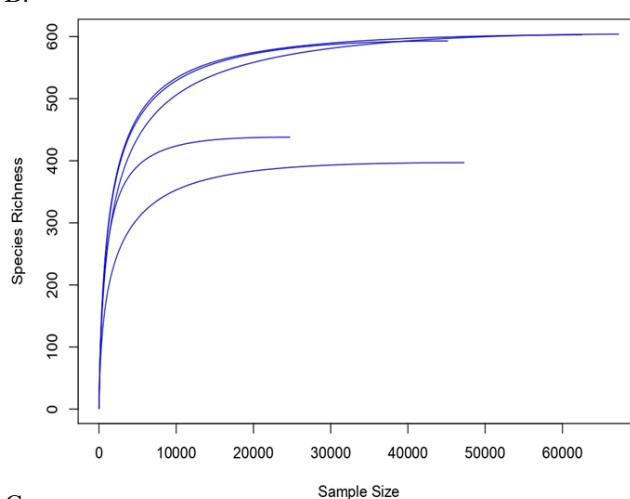


**Figure S11: Rarefaction curves of 16S rRNA gene sequences for the gametophyte with developed cystocarps (A), the gametophyte (B) and the tetrasporophyte (C) samples.**

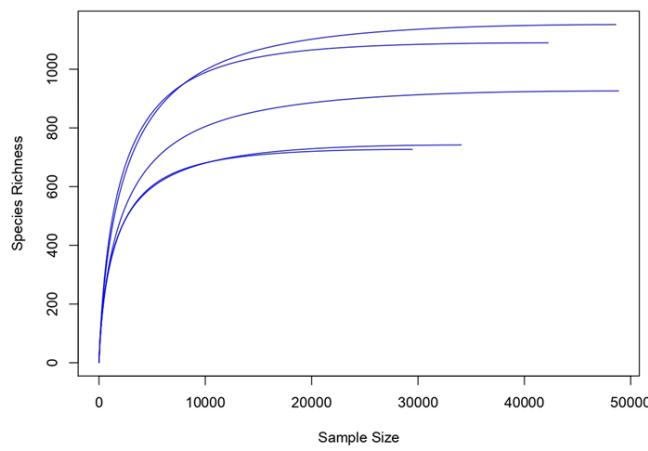
A.



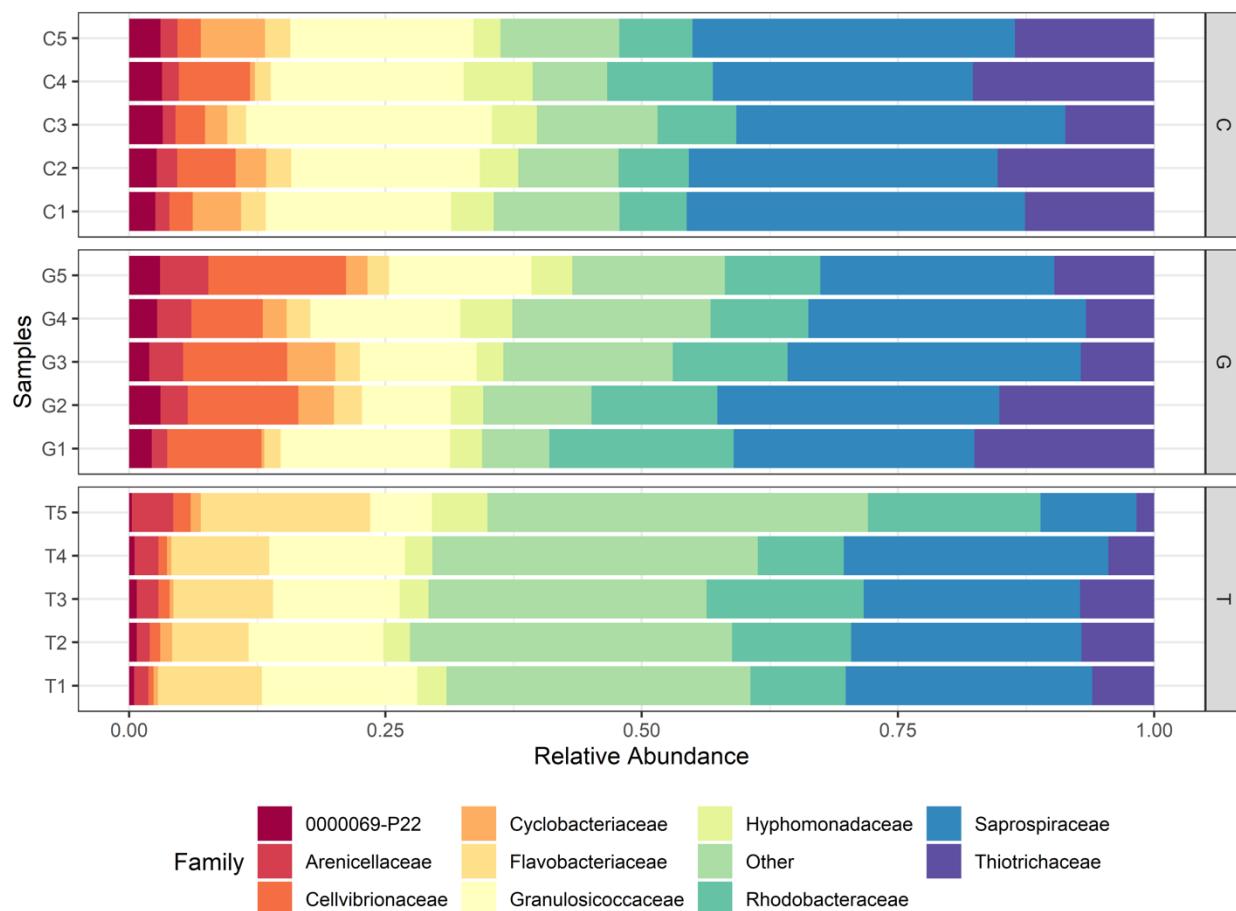
B.



C.



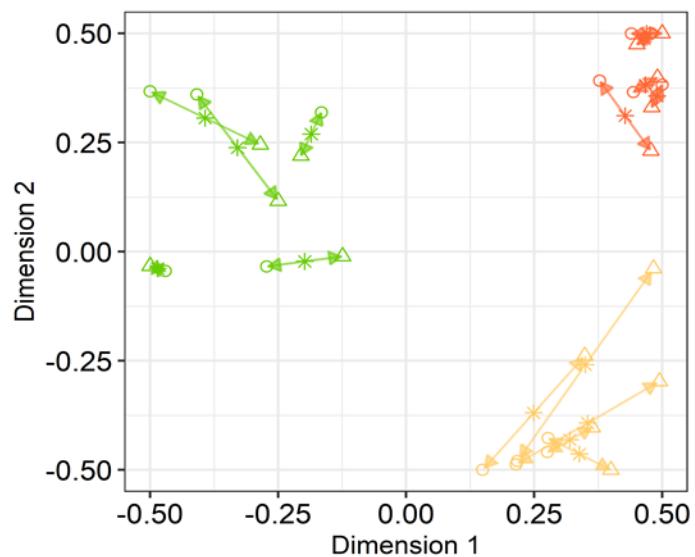
**Figure S12: Relative abundance of bacterial families associated with the three *A. armata* stages. Codes “C” represents samples of the gametophyte with developed cystocarps, “G” the gametophyte samples and “T” the tetrasporophyte samples.**



**Figure S13: KO pathways of the most predicted abundant functions associated with GC (A), G (B) and T (C).**

A. GC	B. G
<b>Metabolism</b>	<b>Metabolism</b>
Global and overview maps 01100 Metabolic pathways (4)	Global and overview maps 01100 Metabolic pathways (25)
Carbohydrate metabolism 00040 Pentose and glucuronate interconversions (1)	01110 Biosynthesis of secondary metabolites (12)
00050 Butanate metabolism (1)	01120 Microbial metabolism in diverse environments (15)
Energy metabolism 00920 Sulfur metabolism (1)	01200 Carbon metabolism (7)
Nucleotide metabolism 00230 Purine metabolism (1)	01210 2-Oxocarboxylic acid metabolism (1)
Amino acid metabolism 00260 Glycine, serine and threonine metabolism (1)	01212 Fatty acid metabolism (8)
Environmental Information Processing	01230 Biosynthesis of amino acids (2)
Membrane transport 02010 ABC transporters (5)	01240 Biosynthesis of cofactors (1)
Signal transduction 02020 Two-component system (2)	Carbohydrate metabolism 00010 Glycolysis / Gluconeogenesis (2)
<b>Cellular Processes</b>	00053 Ascorbate and aldarate metabolism (1)
Cell growth and death 04113 Meiosis - yeast (1)	00060 Pyruvate metabolism (5)
Cellular community - prokaryotes 02024 Quorum sensing (1)	00630 Glyoxylate and dicarboxylate metabolism (5)
02025 Biofilm formation - Pseudomonas aeruginosa (1)	00640 Propanoate metabolism (5)
02026 Biofilm formation - Escherichia coli (1)	00650 Butanoate metabolism (6)
<b>Organismal Systems</b>	00660 CS-5-hydroxy dibasic acid metabolism (1)
Aging 04213 Longevity regulating pathway - multiple species (1)	Energy metabolism 00720 Carbon fixation pathways in prokaryotes (3)
<b>Human Diseases</b>	00680 Methane metabolism (1)
Drug resistance: antimicrobial 01503 Cationic antimicrobial peptide (CAMP) resistance (1)	00910 Nitrogen metabolism (1)
<b>C. T</b>	00920 Sulfur metabolism (1)
<b>Metabolism</b>	Lipid metabolism 00061 Fatty acid biosynthesis (3)
Global and overview maps 01100 Metabolic pathways (6)	00071 Fatty acid degradation (8)
01110 Biosynthesis of secondary metabolites (5)	00120 Primary bile acid biosynthesis (1)
01120 Microbial metabolism in diverse environments (3)	00561 Glycerolipid metabolism (1)
01200 Carbon metabolism (3)	00592 alpha-Linolenic acid metabolism (1)
01212 Fatty acid metabolism (1)	Amino acid metabolism 00250 Alanine, aspartate and glutamate metabolism (2)
01230 Biosynthesis of amino acids (1)	00260 Glycine, serine and threonine metabolism (1)
01250 Biosynthesis of nucleotide sugars (1)	00280 Valine, leucine and isoleucine degradation (11)
01240 Biosynthesis of cofactors (2)	00290 Valine, leucine and isoleucine biosynthesis (1)
Carbohydrate metabolism 00010 Glycolysis / Gluconeogenesis (2)	00310 Lysine degradation (5)
00020 Citrate cycle (TCA cycle) (2)	00220 Arginine biosynthesis (2)
00052 Galactose metabolism (1)	00330 Arginine and proline metabolism (1)
00520 Amino sugar and nucleotide sugar metabolism (1)	00340 Histidine metabolism (1)
00620 Pyruvate metabolism (2)	00350 Tyrosine metabolism (1)
00630 Glyoxylate and dicarboxylate metabolism (1)	00360 Phenylalanine metabolism (2)
00640 Propanoate metabolism (1)	00380 Tryptophan metabolism (4)
00562 Inositol phosphate metabolism (1)	Metabolism of cofactors and vitamins 00410 beta-Alanine metabolism (3)
Energy metabolism 00061 Fatty acid metabolism (1)	00480 Glutathione metabolism (1)
Lipid metabolism 00061 Fatty acid biosynthesis (1)	Metabolism of terpenoids and polyketides 00760 Nicotinate and nicotinamide metabolism (1)
Amino acid metabolism 00260 Glycine, serine and threonine metabolism (2)	00770 Pantothenate and CoA biosynthesis (2)
00270 Cysteine and methionine metabolism (1)	Metabolism of terpenoids and polyketides 00900 Terpenoid backbone biosynthesis (1)
00280 Valine, leucine and isoleucine degradation (1)	00981 Insect hormone biosynthesis (1)
00310 Lysine degradation (1)	00903 Limonene and pinene degradation (2)
00380 Tryptophan metabolism (1)	00281 Geraniol degradation (2)
Glycan biosynthesis and metabolism 00541 O-Antigen nucleotide sugar biosynthesis (1)	Xenobiotics biodegradation and metabolism 00362 Benzene degradation (4)
Metabolism of cofactors and vitamins 00780 Biotin metabolism (1)	00627 Aminobenzoate degradation (1)
Biosynthesis of other secondary metabolites 00521 Streptomycin biosynthesis (1)	00625 Chlороalkane and chlороalkene degradation (1)
00333 Prodigiosin biosynthesis (1)	00642 Ethylbenzene degradation (1)
<b>Environmental Information Processing</b>	00791 Atrazine degradation (1)
Membrane transport 02010 ABC transporters (3)	00930 Caprolactam degradation (2)
Signal transduction 02020 Two-component system (4)	00980 Metabolism of xenobiotics by cytochrome P450 (1)
04070 Phosphatidylinositol signaling system (1)	00982 Drug metabolism - cytochrome P450 (1)
<b>Cellular Processes</b>	00983 Drug metabolism - other enzymes (1)
Cell growth and death 04112 Cell cycle - Culobacter (2)	<b>Genetic Information Processing</b>
Cellular community - prokaryotes 02025 Biofilm formation - Pseudomonas aeruginosa (1)	Translation 00970 Aminoacyl-tRNA biosynthesis (1)
Cell motility 02030 Bacterial chemotaxis (2)	<b>Environmental Information Processing</b>
02040 Flagellar assembly (1)	Membrane transport 02010 ABC transporters (18)
<b>Organismal Systems</b>	Signal transduction 02020 Two-component system (3)
Endocrine system 04920 Adipocytokine signaling pathway (1)	<b>Cellular Processes</b>
03230 PPAR signaling pathway (2)	Transport and catabolism 04146 Peroxisome (2)
Digestive system 04975 Fat digestion and absorption (1)	Cell growth and death 04110 Cell cycle (1)
04724 Glutamatergic synapse (1)	04217 Necrosis (1)
04727 GABAergic synapse (1)	Cellular community - prokaryotes 02024 Quorum sensing (14)
Aging 04212 Longevity regulating pathway - worm (1)	<b>Organismal Systems</b>
Environmental adaptation 04714 Thermogenesis (1)	Endocrine system 04920 Adipocytokine signaling pathway (1)
<b>Human Diseases</b>	03230 PPAR signaling pathway (2)
Cancer: overview 05200 Pathways in cancer (1)	Digestive system 04975 Fat digestion and absorption (1)
05204 Chemical carcinogenesis - DNA adducts (1)	Nervous system 04724 Glutamatergic synapse (1)
05207 Chemical carcinogenesis - receptor activation (1)	04727 GABAergic synapse (1)
05208 Chemical carcinogenesis - reactive oxygen species (1)	Aging 04212 Longevity regulating pathway - worm (1)
Cancer: specific types 05225 Hepatocellular carcinoma (1)	Environmental adaptation 04714 Thermogenesis (1)
Cardiovascular disease 05418 Fluid shear stress and atherosclerosis (1)	
Endocrine disease 04936 Alcoholic liver disease (2)	
Drug resistance: antimicrobial 01501 beta-Lactam resistance (2)	
Drug resistance: antineoplastic 01524 Platinum drug resistance (1)	

**Figure S14: Scores plot of the multiblock PLS-DA analysis (DIABLO) of *A. armata* gametophyte (G), with developed cystocarps (GC), and tetrasporophyte (T) phases.**

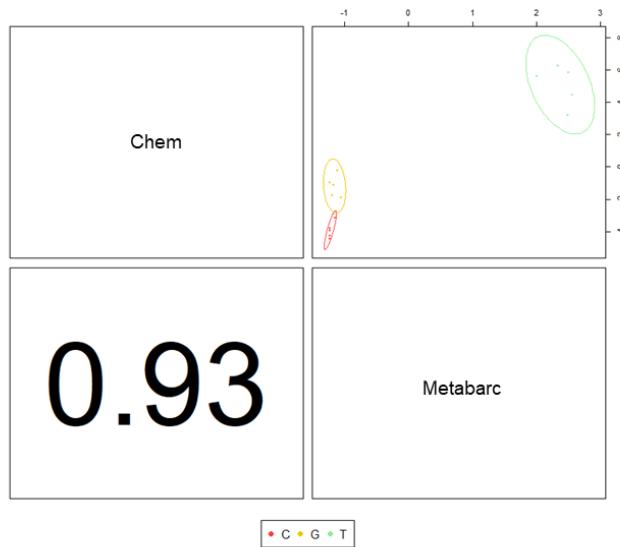


**Legend**

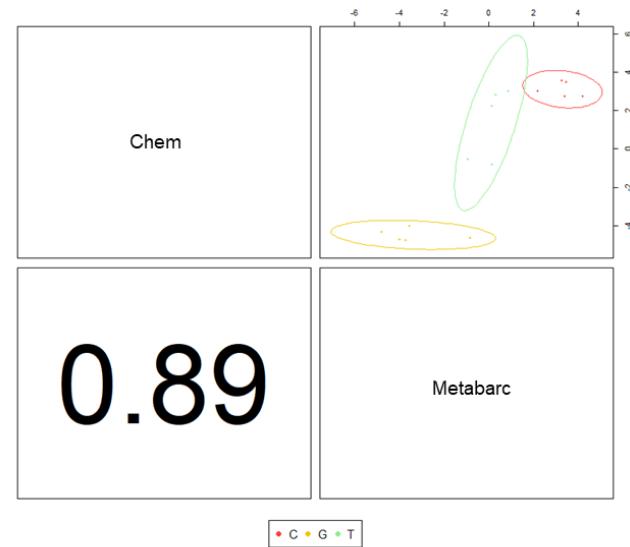
- |  |                      |
|--|----------------------|
| ● Gametophyte (G)                            | ○ Chemistry dataset  |
| ● Gametophyte with developed cystocarps (GC) | △ Microbiota dataset |
| ● Tetrasporophyte (T)                        | * Centroid           |

**Figure S15: Correlations between the first dimension (A) and the second dimension (B) of each dataset (Chem= Chemistry; Metabarc= Metabarcoding) for the two PLS models. Codes “C” represents samples of the gametophyte with developed cystocarps, “G” the gametophyte samples and “T” the tetrasporophyte samples.**

A. Dimension 1



B. Dimension 2



**Figure S16: Heatmap of significant ASV and metabolites of the three phases of *A. armata* (“GC” represents samples of the gametophyte with developed cystocarps, “G” the gametophyte samples and “T” the tetrasporophyte samples)**

