

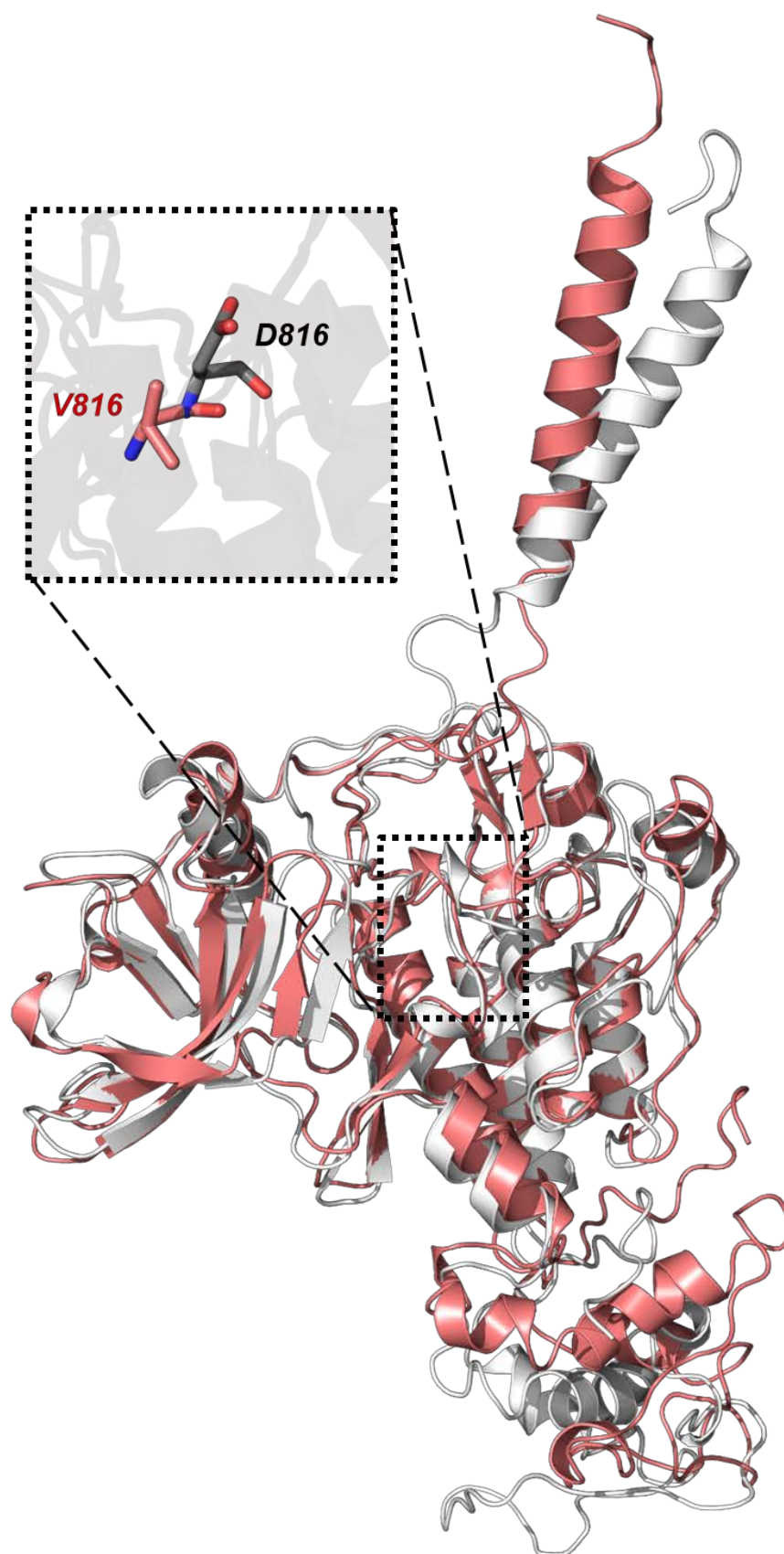
## **Supplementary Material**

### **Receptor Tyrosine Kinase KIT: Mutation-Induced Conformational Shift Promotes Alternative Allosteric Pockets**

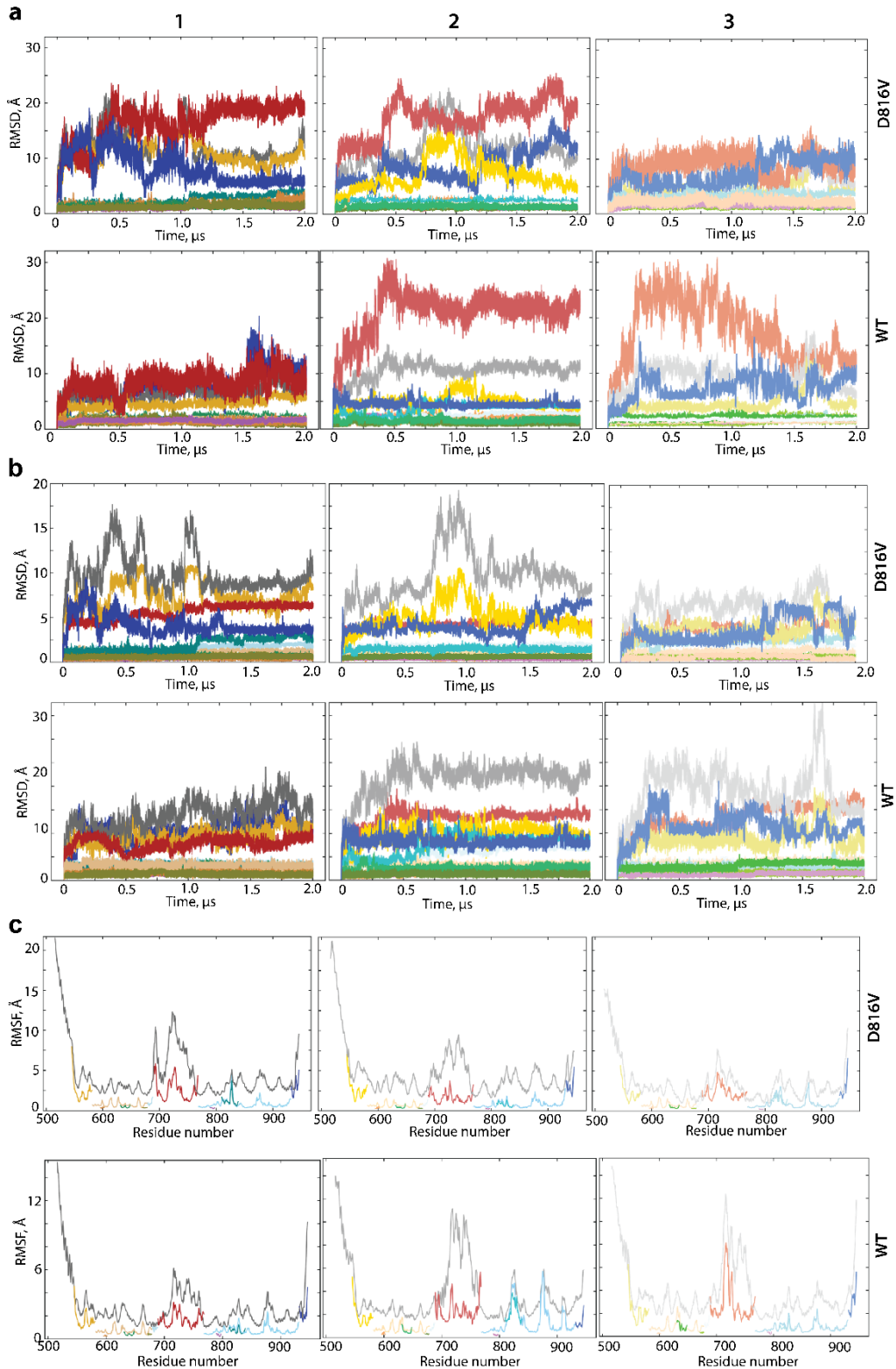
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**Figures S1-S6**

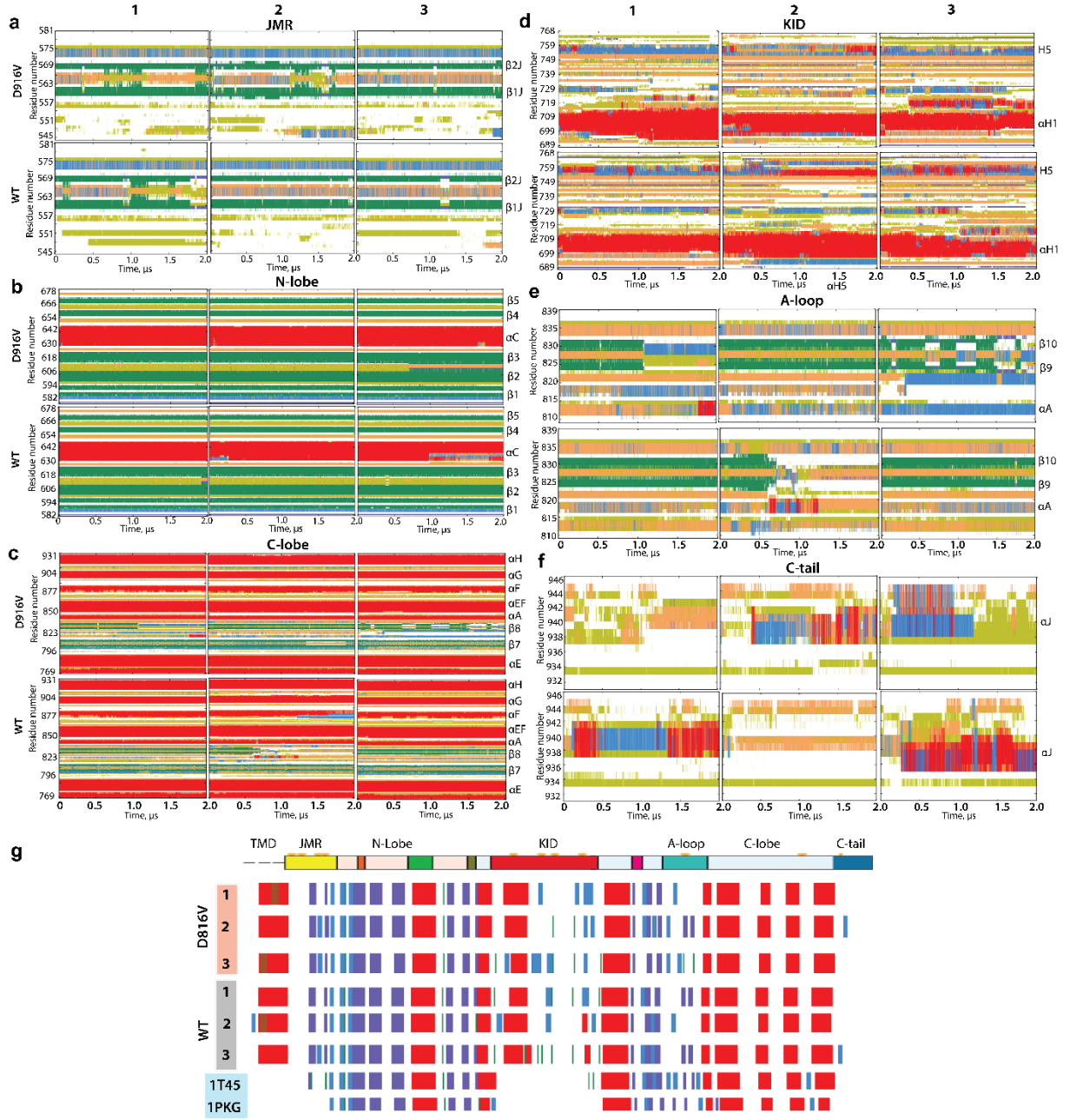
**Tables S1-S2**



**Figure S1.** Homology modelling of KIT<sup>D816V</sup> from the KIT<sup>WT</sup> full-length cytoplasmic domain model completed by the transmembrane helix (sequence I516-R946). Superimposition of KIT<sup>WT</sup> (in grey) taken at t=2  $\mu$ s of MD simulation, and KIT<sup>D816V</sup> (in pink) taken at t = 0  $\mu$ s. RMSDs calculated on C-alpha after fitting on tyrosine kinase domain is of 6.5 Å.

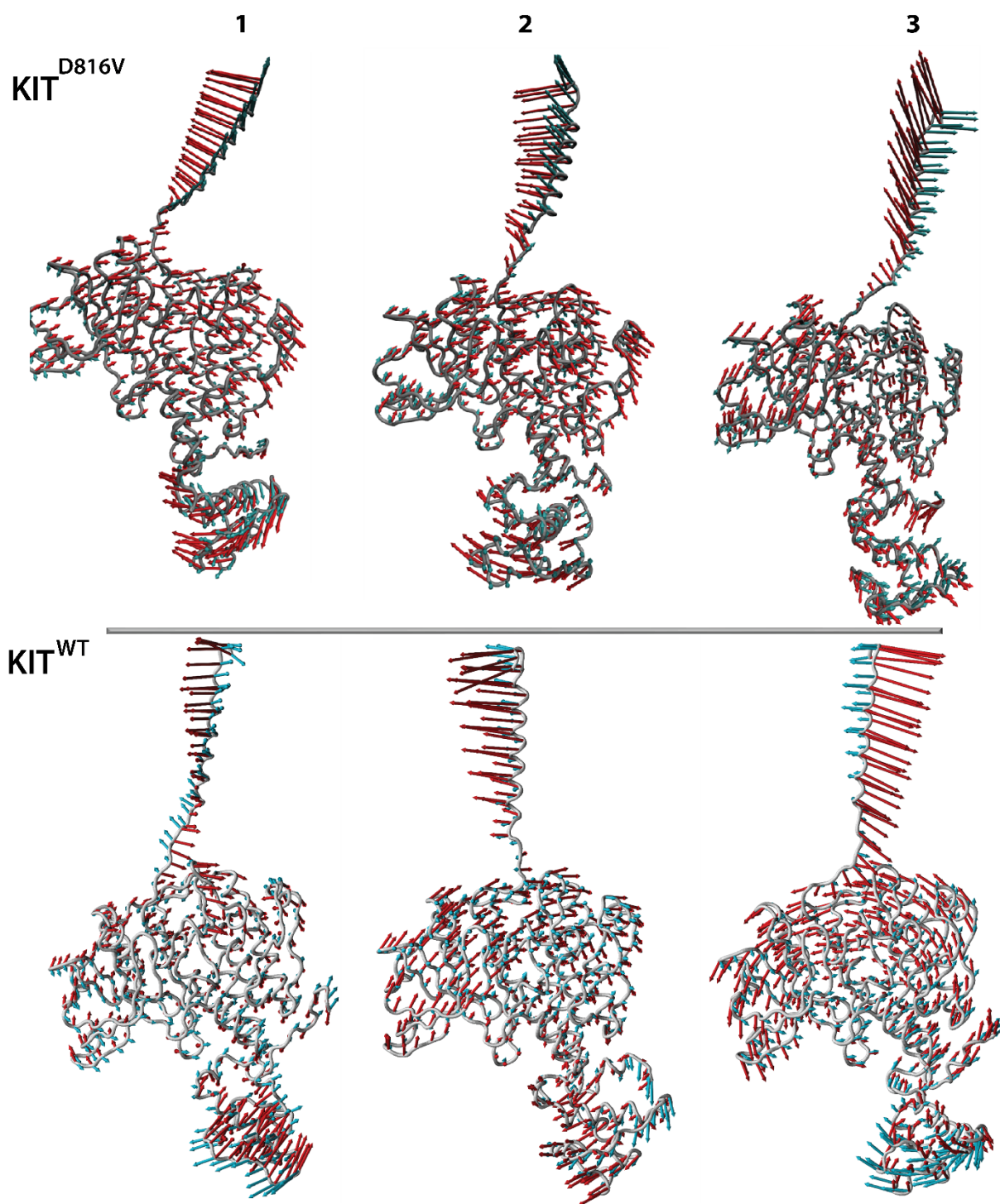


**Figure S2.** Molecular Dynamics (MD) simulations of the full-length cytoplasmic domain of KIT<sup>D816V</sup> and KIT<sup>WT</sup>. (a) RMSDs computed on the C $\alpha$  atoms after fitting on initial conformation (at  $t = 0$  ns) (1-3 replicas, each of 2  $\mu$ s) of kinase domain and each domain/region of KIT<sup>D816V</sup> (a) and KIT<sup>WT</sup> (b). (c) RMSFs computed on the C $\alpha$  atoms for MD conformations after the least-square fitting on the kinase domain initial conformation of KIT<sup>D816V</sup> (top panel) and KIT<sup>WT</sup>. (a-c) KIT is in grey, N-lobe in beige, C-lobe in blue, JMR in yellow, P-loop in orange,  $\alpha$ C-helix in green, hinge in olive, KID in red, C-loop in magenta, A-loop in teal, C-tail in dark blue for the 1-3 trajectories (replicas) of MD simulations. MD replicas 1-3 are distinguished by colour tonality, dark, lighter, and light.

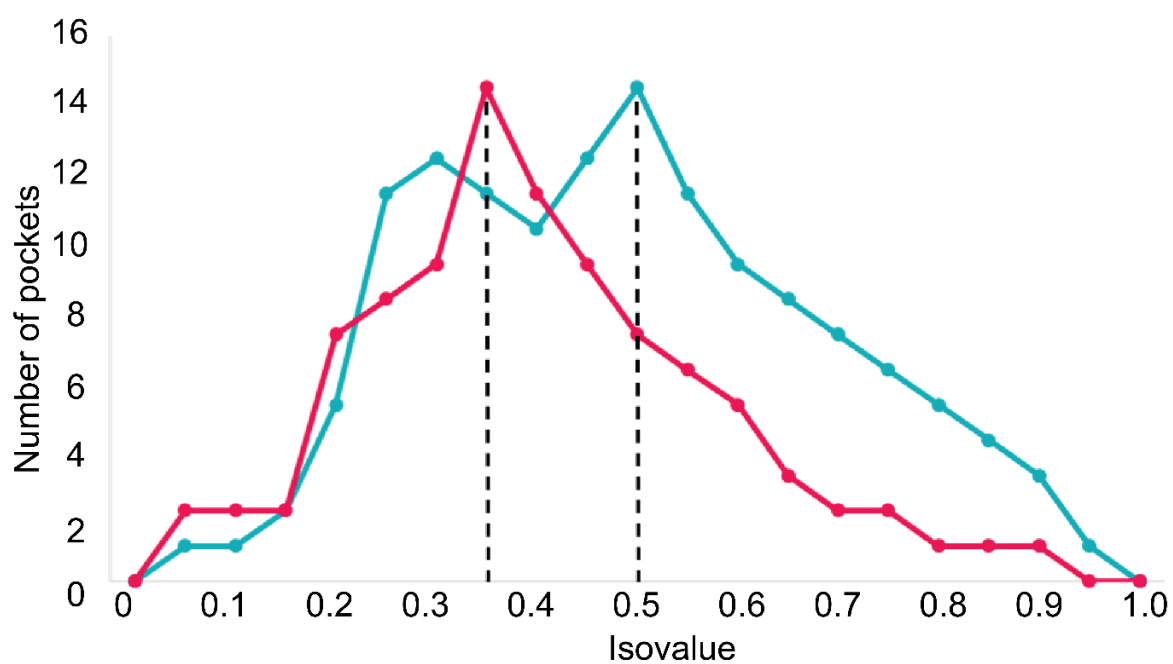


**Figure S3.** Folding of KIT<sup>D816V</sup> (top panels) and KIT<sup>WT</sup> (bottom panels). (a-f) The time-related evolution of the secondary structures of the entire, full-length KIT and per domain/region, as assigned by DSSP:  $\alpha$ -helices in red,  $3_{10}$ -helices in blue, parallel  $\beta$  strands in green, antiparallel  $\beta$  strands in dark blue, turns in orange, and bends in dark yellow. The three cMD replicas (1–3) were analysed individually. (g) The secondary structures— $\alpha$ H- (red),  $3_{10}$ -helices (light blue), and  $\beta$ -strands (dark blue)—assigned for a mean conformation of every MD trajectory (1–3) of KIT<sup>D816V</sup>, KIT<sup>WT</sup> and the crystallographic structures 1T45 (inactive KIT) and 1PKG (active KIT). (D) The secondary structures— $\alpha$ H- (red) and  $\beta$ -strands (dark blue)—assigned on the mean conformation of the concatenated trajectories.

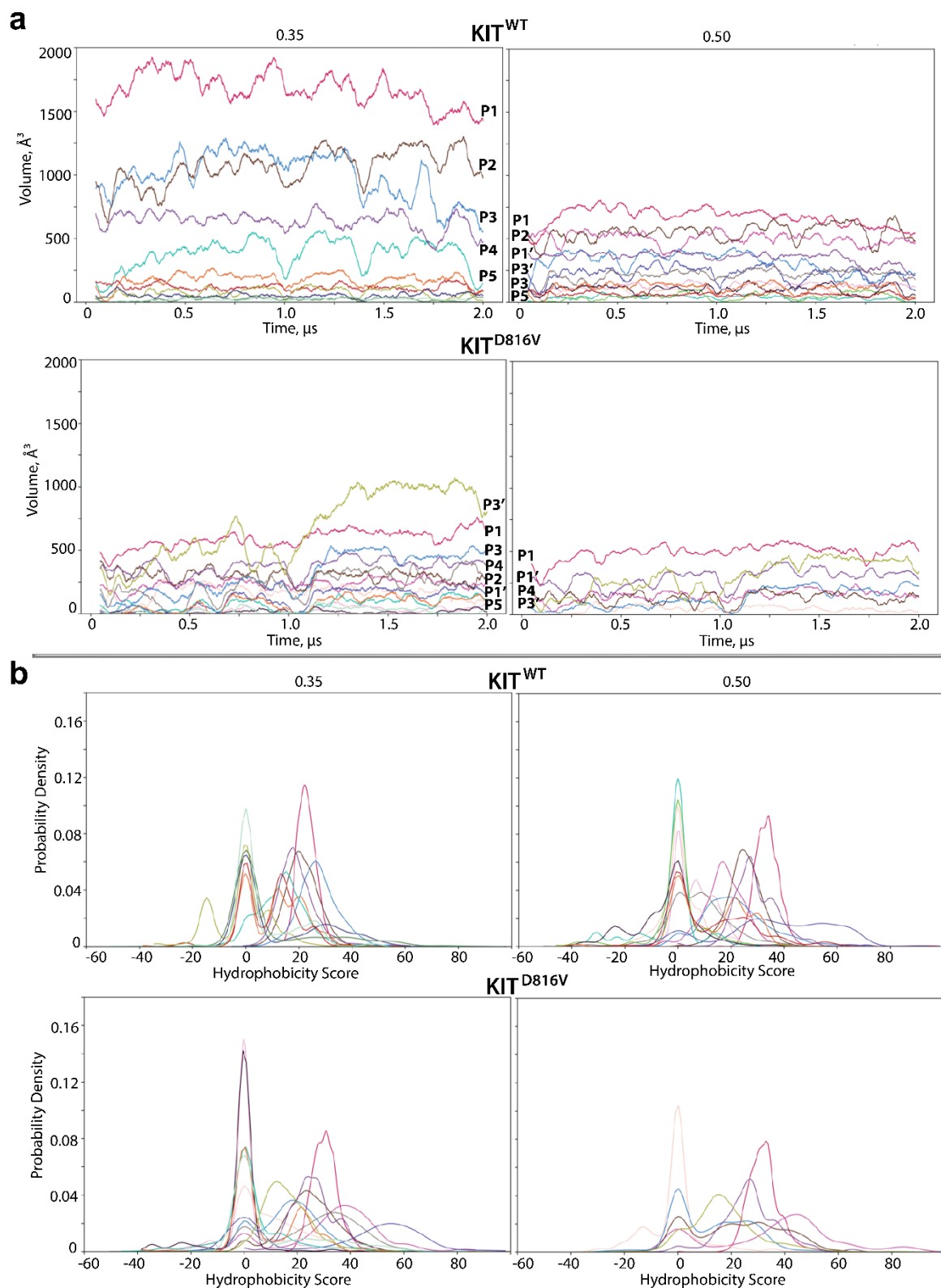




**Figure S4.** PCA of the MD conformations of KIT. Atomic components in PCA modes 1–2 are drawn as red (1<sup>st</sup> mode) and cyan (2<sup>nd</sup> mode) arrows, projected on the cartoon of  $KIT^{D816V}$  (top) and  $KIT^{WT}$  (bottom). PCA was performed on each individual trajectory of  $KIT^{D816V}$  and  $KIT^{WT}$ . A cut-off distance of 4 Å was used.



**Figure S5.** Search of the optimal criteria for pockets hunting. This step is carried out using the isovalue from 0 to 1.0 with a step of 0.5 for both proteins. KIT<sup>D816V</sup> and KIT<sup>WT</sup> are in red and cyan respectively.



**Figure S6.** The RTK KIT POCKETOME characterisation. **(a)** The pockets evolution over the simulation time in each KIT protein. **(b)** Probability of the pockets hydrophobicity defined as in (Monera *et al.*, 1995). The pockets are distinguished by colour as defined in Figure 8.

**Table S1.** Folding of the intrinsically disordered regions in KIT<sup>WT</sup> and KIT<sup>D816V</sup>

	KIT <sup>WT</sup> mean folding (%)			KIT <sup>D816V</sup> mean folding (%)		
	Helix	Strand	Total	Helix	Strand	Total
<b>JMR</b>	13 ± 4	14 ± 0	27 ± 4	10 ± 4	16 ± 2	29 ± 5
<b>KID</b>	30 ± 8	3 ± 2	33 ± 5	32 ± 5	2 ± 2	33 ± 7
<b>A-loop</b>	<b>3 ± 3</b>	<b>13 ± 9</b>	<b>17 ± 5</b>	<b>13 ± 5</b>	<b>9 ± 8</b>	<b>22 ± 9</b>
<b>C-tail</b>	13 ± 9	0	13 ± 9	15 ± 11	0	15 ± 11

**Table S2.** Pockets characterisation in KIT<sup>WT</sup> and KIT<sup>D816V</sup>. The numbering of the pocket (P1-P6) is presented as in Figure 8, characterised by the volume size ( $V_{\max}$ , Å<sup>3</sup>), measured in the frame (Frame) and formed by the number of residues (N res). Pockets were characterised by the Fpocket algorithm using isovalue of 0.35.

KIT <sup>WT</sup>				KIT <sup>D816V</sup>			
Pocket	$V_{\max}$ , Å <sup>3</sup>	Frame	N res	Pocket	$V_{\max}$ , Å <sup>3</sup>	Frame	N res
<b>P1</b>	2674.9	521	72	<b>P1/ P1'/ P1''</b>	993.7/574.2/218.0	1907	24/10/11
<b>P2</b>	1757.9	1649	38	<b>P2/P2'</b>	704.2/652.57	1850	44/9
<b>P3</b>	1818.9	668	43	<b>P3/P3'/P3''</b>	216.0/830.7/379.7	1199	4/25/8
<b>P4</b>	873.9	1499	20	<b>P4</b>	688.5	1760	23
<b>P5</b>	1076.9	1511	26	<b>P5</b>	382.8	1785	5
<b>P6</b>	421.4	724	11	<b>P6</b>	295.7	744	5