

Design, Synthesis, and Antifungal/Anti-oomycete Activities of Novel 1,2,4-Triazole Derivatives Containing Carboxamide Fragments

Jiali Wang [†], Haoran Shi [†] and Aidang Lu ^{*}

School of Chemical Engineering and Technology, Hebei University of Technology, Tianjin 300401, China; galizhu999@163.com (J.W.); shr15130128422@163.com (H.S.)

^{*} Correspondence: luaidang@hebut.edu.cn; Tel. +86-22-60202812

Contents

Section S1: Detailed bioassay procedures for the in vitro antifungal/anti-oomycete activities	3
Section S2: Calculation procedures for molecular docking research	3
Section S3: Copies of NMR spectra (Figures S1–S74)	4
Figures S1–S2. ¹H NMR and ¹³C NMR spectra of 2	4
Figures S3–S4. ¹H NMR and ¹³C NMR spectra of 3	5
Figures S5–S6. ¹H NMR and ¹³C NMR spectra of 4	6
Figures S7–S8. ¹H NMR and ¹³C NMR spectra of 5a	7
Figures S9–S10. ¹H NMR and ¹³C NMR spectra of 5b	8
Figures S11–S12. ¹H NMR and ¹³C NMR spectra of 5c	9
Figures S13–S14. ¹H NMR and ¹³C NMR spectra of 5d	10
Figures S15–S16. ¹H NMR and ¹³C NMR spectra of 5e	11
Figures S17–S18. ¹H NMR and ¹³C NMR spectra of 5f	12
Figures S19–S20. ¹H NMR and ¹³C NMR spectra of 5g	13
Figures S21–S22. ¹H NMR and ¹³C NMR spectra of 5h	14
Figures S23–S24. ¹H NMR and ¹³C NMR spectra of 5i	15
Figures S25–S26. ¹H NMR and ¹³C NMR spectra of 5j	16
Figures S27–S28. ¹H NMR and ¹³C NMR spectra of 5k	17
Figures S29–S30. ¹H NMR and ¹³C NMR spectra of 5l	18

Figures S31–S32. ^1H NMR and ^{13}C NMR spectra of 5m	19
Figures S33–S34. ^1H NMR and ^{13}C NMR spectra of 5n	20
Figures S35–S36. ^1H NMR and ^{13}C NMR spectra of 6a	21
Figures S37–S38. ^1H NMR and ^{13}C NMR spectra of 6b	22
Figures S39–S40. ^1H NMR and ^{13}C NMR spectra of 6c	23
Figures S41–S42. ^1H NMR and ^{13}C NMR spectra of 6d	24
Figures S43–S44. ^1H NMR and ^{13}C NMR spectra of 6e	25
Figures S45–S46. ^1H NMR and ^{13}C NMR spectra of 6f	26
Figures S47–S48. ^1H NMR and ^{13}C NMR spectra of 6g	27
Figures S49–S50. ^1H NMR and ^{13}C NMR spectra of 6h	28
Figures S51–S52. ^1H NMR and ^{13}C NMR spectra of 6i	29
Figures S53–S54. ^1H NMR and ^{13}C NMR spectra of 6j	30
Figures S55–S56. ^1H NMR and ^{13}C NMR spectra of 6k	31
Figures S57–S58. ^1H NMR and ^{13}C NMR spectra of 6l	32
Figures S59–S60. ^1H NMR and ^{13}C NMR spectra of 6m	33
Figures S61–S62. ^1H NMR and ^{13}C NMR spectra of 7a	34
Figures S63–S64. ^1H NMR and ^{13}C NMR spectra of 7b	35
Figures S65–S66. ^1H NMR and ^{13}C NMR spectra of 7c	36
Figures S67–S68. ^1H NMR and ^{13}C NMR spectra of 7d	37
Figures S69–S70. ^1H NMR and ^{13}C NMR spectra of 74	38
Figures S71–S72. ^1H NMR and ^{13}C NMR spectra of 7f	39
Figures S73–S74. ^1H NMR and ^{13}C NMR spectra of 7g	40

Section S1. Detailed bioassay procedures for the in vitro antifungal/anti-oomycete activities

The fungicidal activities of compounds were evaluated in mycelial growth tests conducted in artificial media against 7 plant pathogens at a rate of 50 µg/mL. Each test compound was dissolved in a suitable amount of acetone and diluted with water containing 0.1% TW-80 to a concentration of 500 µg/mL. To each petri dish was added 1 mL of the test solution and 9 mL of culture medium to make a 50 µg/mL concentration of the test compound, while in another petri dish was added 1 mL distilled water containing 0.1% TW-80 and 9 mL of culture medium as a blank control. A 4 mm diameter of hyphal growth was cut using a hole puncher on a growing fungal culture and the hyphae were moved to the petri dish containing the test compound. Each assay was performed three times. The dishes were stored in controlled environment cabinets (24 ± 1 °C) for 4 days, after which the diameter of mycelial growth was measured and the percentage inhibition was calculated using the following equation: Percentage inhibition (%) = (averaged diameter of mycelia in blank controls—averaged diameter of mycelia in medicated tablets)/(averaged diameter of mycelia in blank controls—4 mm) × 100 [43].

Section S2. Calculation procedures for molecular docking research

The calculation procedures for molecular docking research consist of four steps [44].

Receptor Preparation. The 3D crystal structure of C-14α demethylase (PDB code: 3L4D) was downloaded from the protein data bank (PDB) and this was used as the receptor for molecular docking. Water molecules were removed from the target protein and hydrogen atoms were added using AutoDock Tools prior to molecular docking.

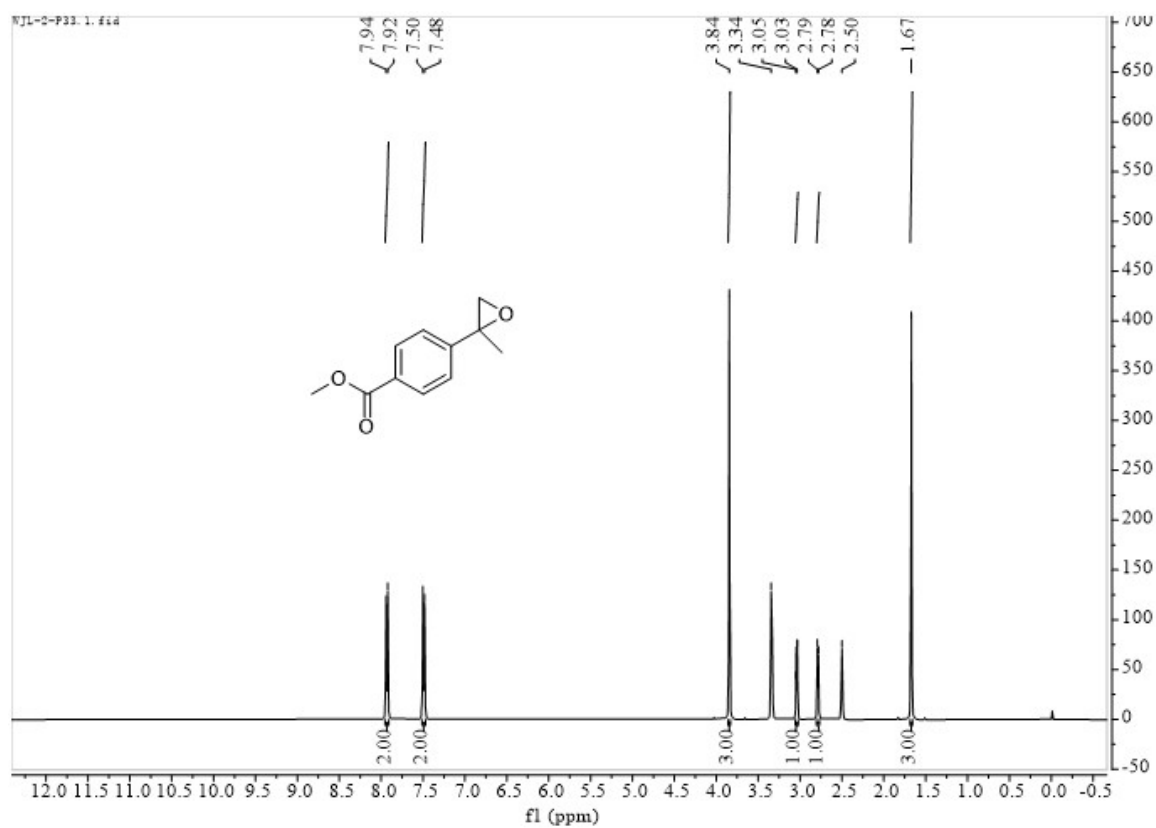
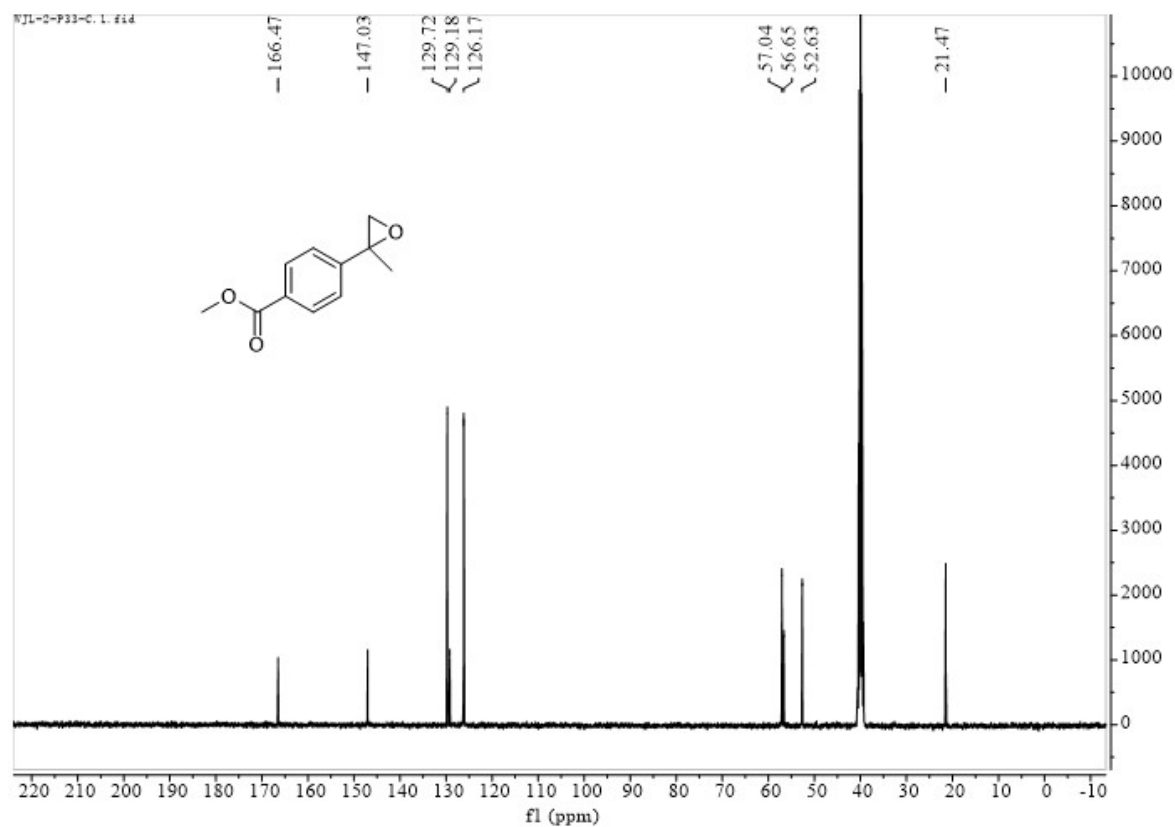
Ligand preparation. Target compounds are drawn using ChemOffice 2015 as ligands followed by management of its conformer and the minimisation process.

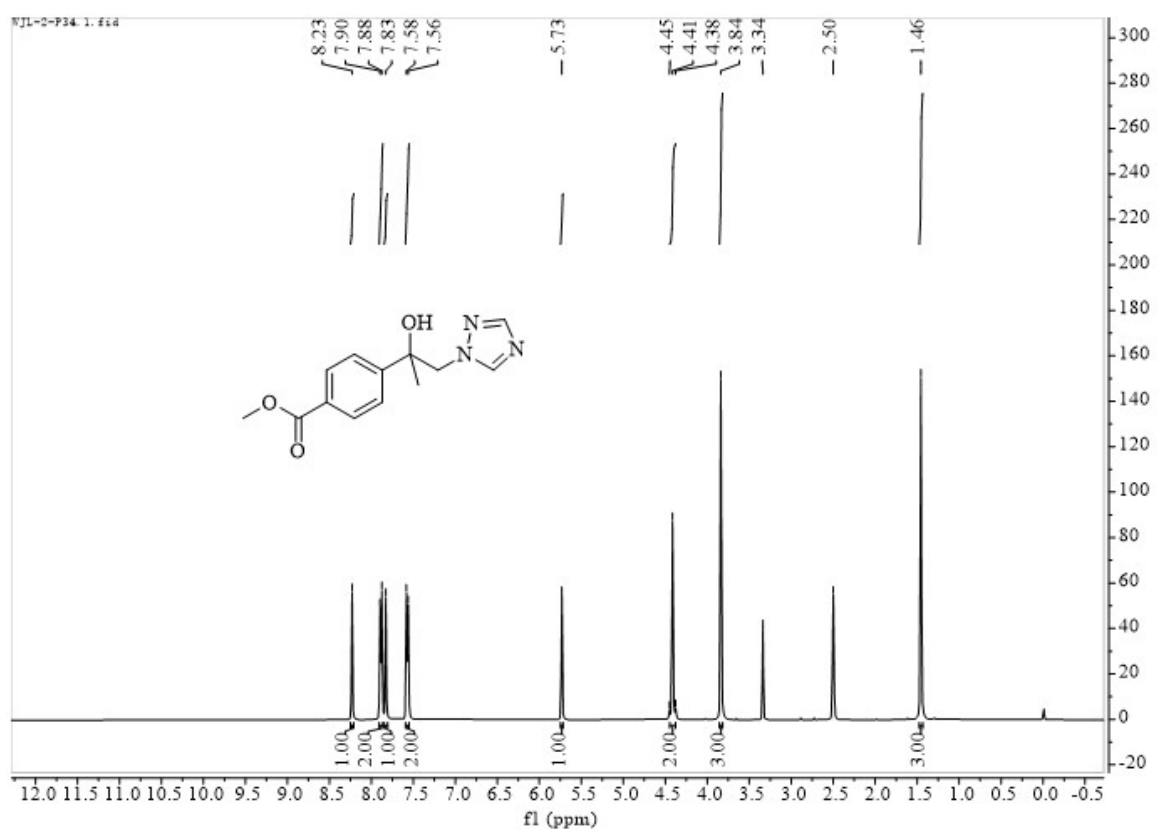
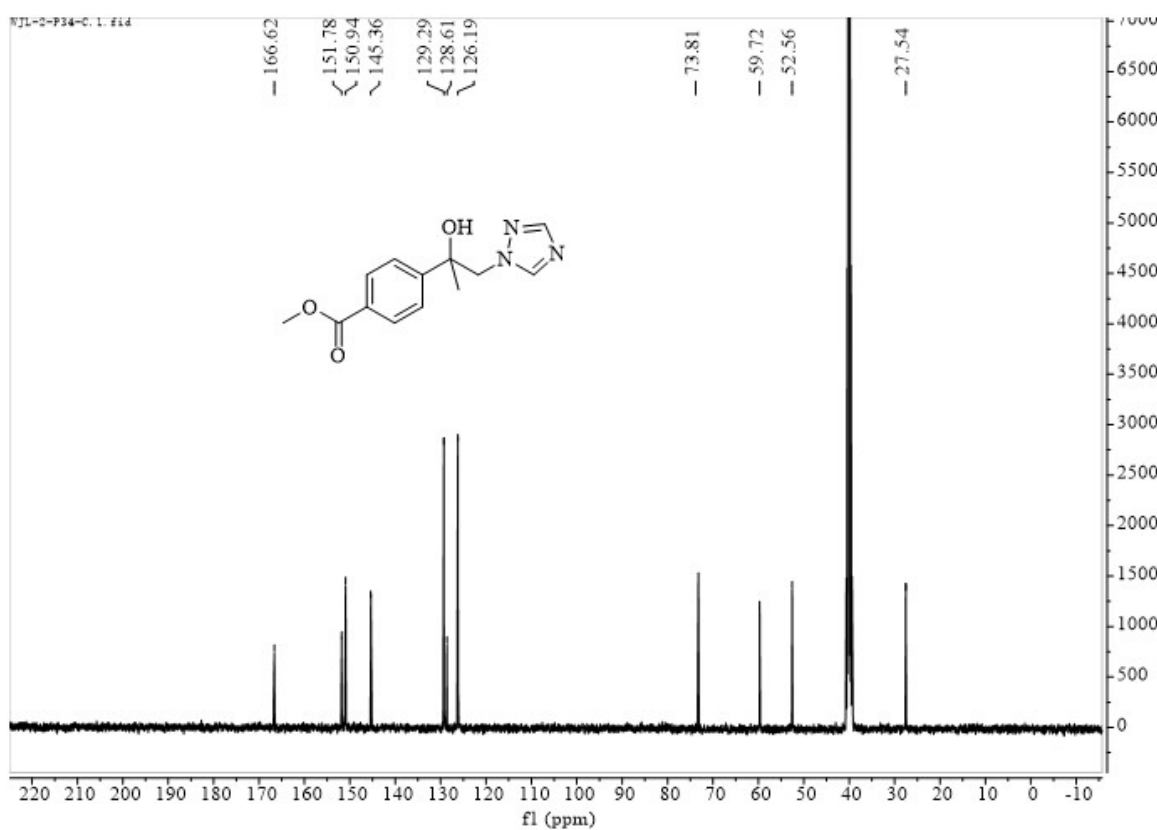
Molecular Docking Using AutoDock Vina. The input files for AutoDock Vina were prepared using AutoDock Tools. The protein was placed in a grid box (grid parameters: center x = 38.01, center y = -33.901, center z = -28.152, size x = 102.63, size y = 76.506, size z = 117.58), using AutoDock Vina at 1.00 Å to define the binding site. The docking procedure was performed using the instructed command prompts.

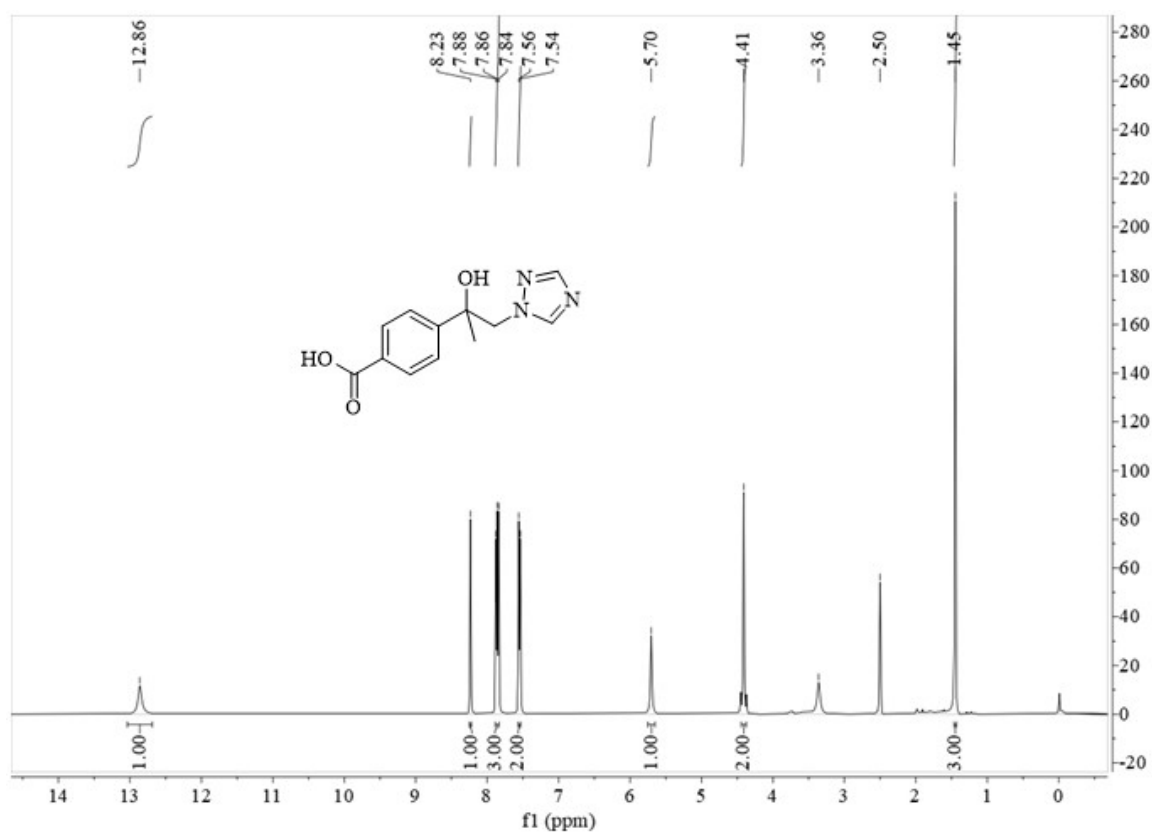
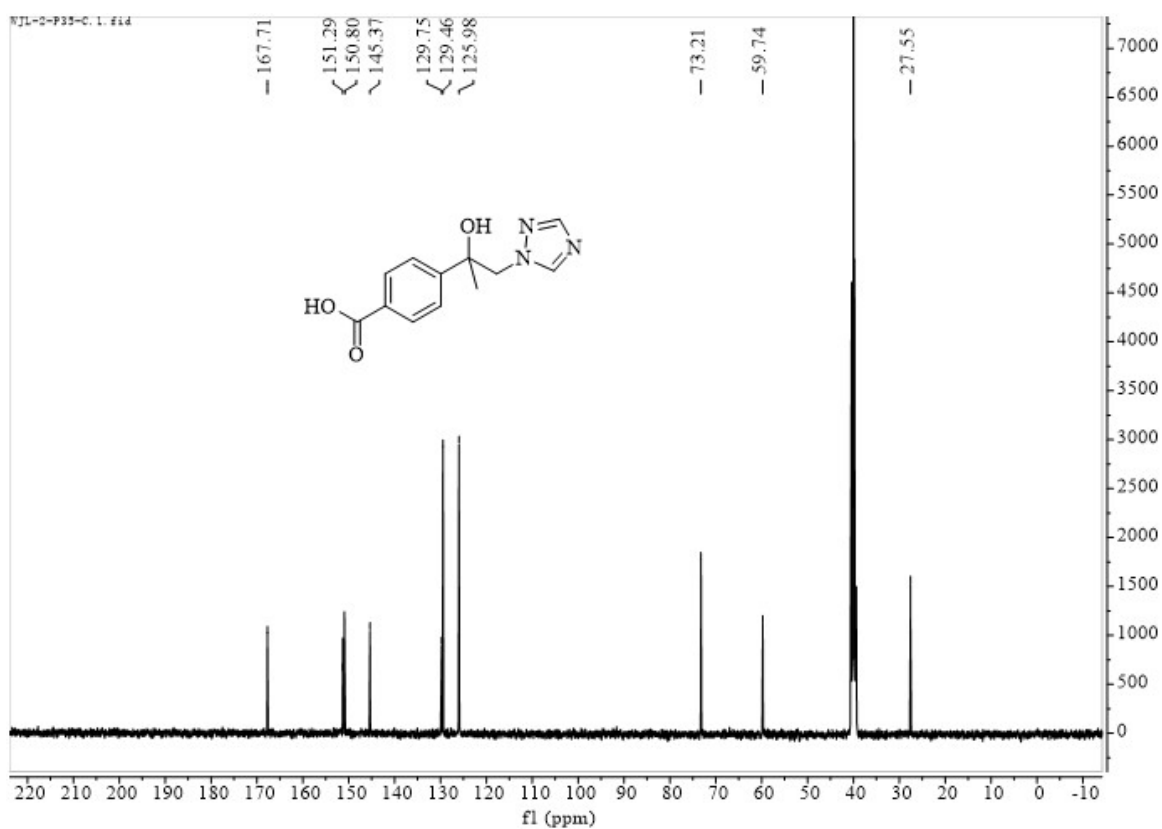
Analyzing and Output Visualisation using PyMOL. The docking poses were ranked according to their docking scores. The scoring function in Auto Dock was used to predict the binding affinity of one ligand to the receptor molecule. The conformation with the lowest binding affinity was selected for further analysis after the docking process. The docking results included the locations of hydrogen bonds and closely interacting residues were performed by PyMOL software.

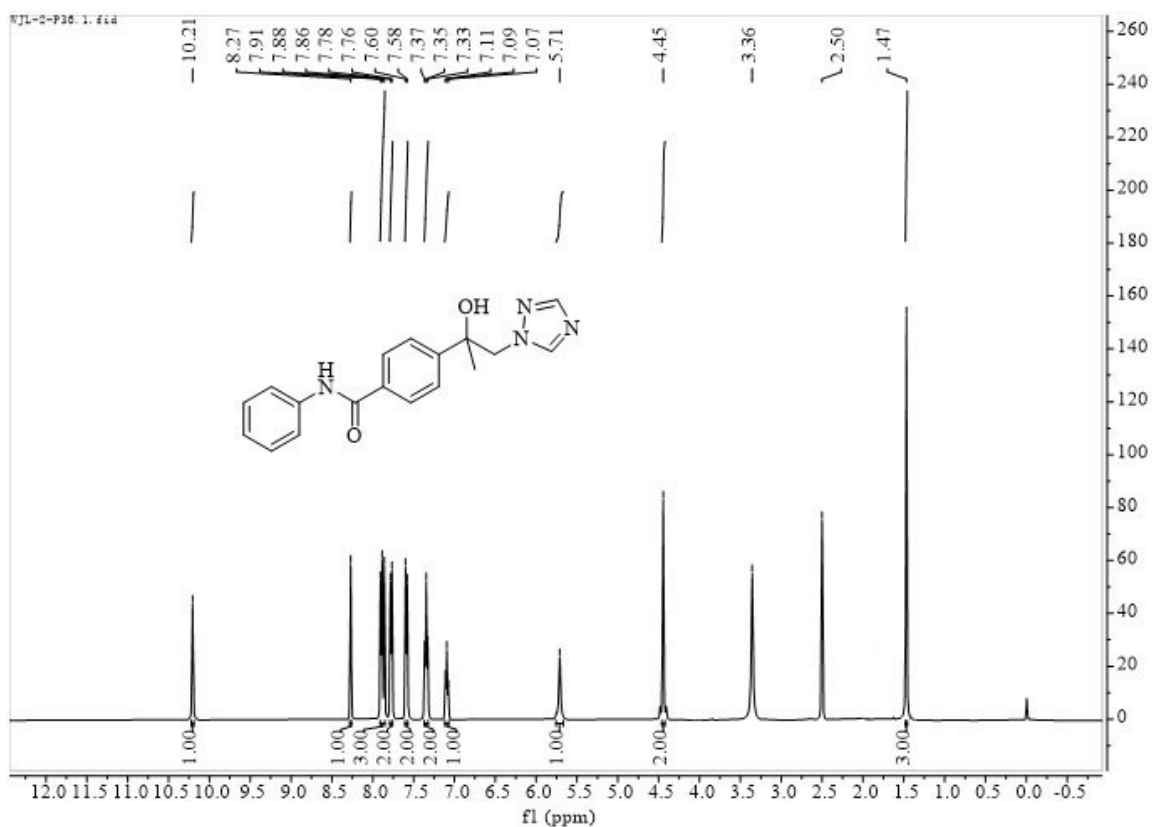
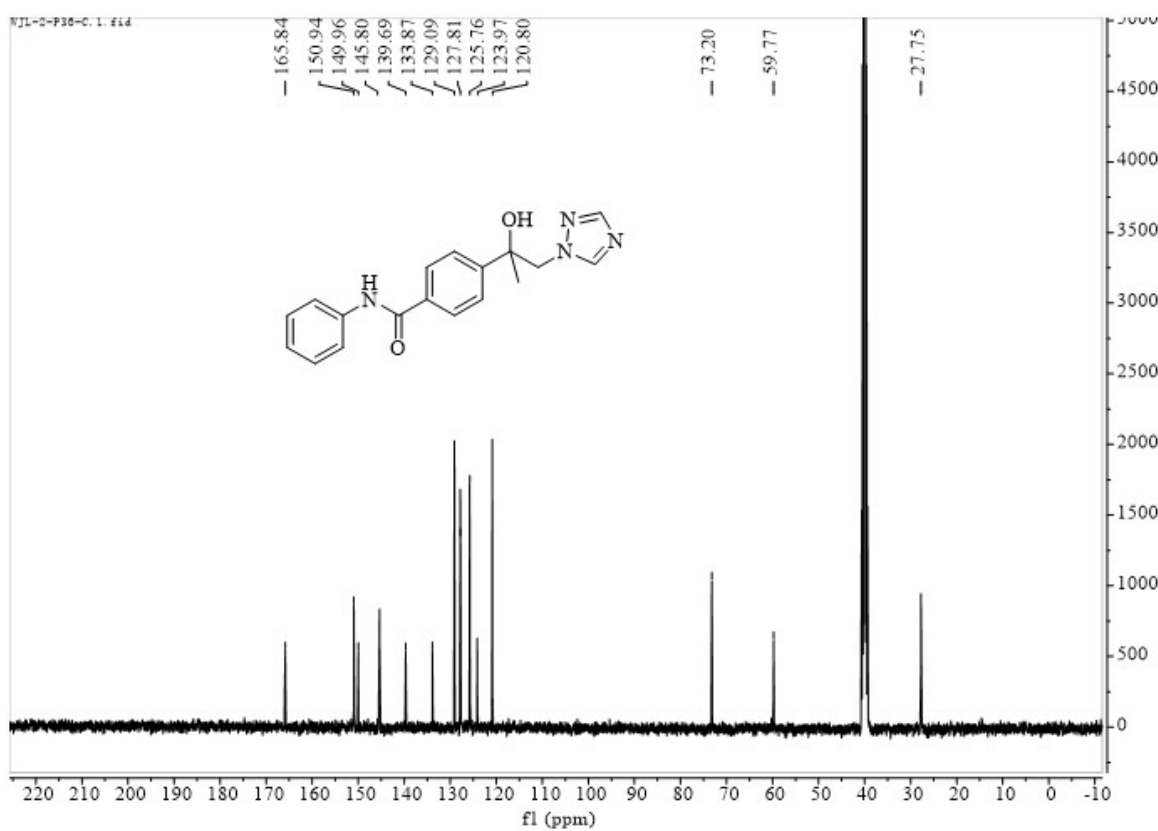
References

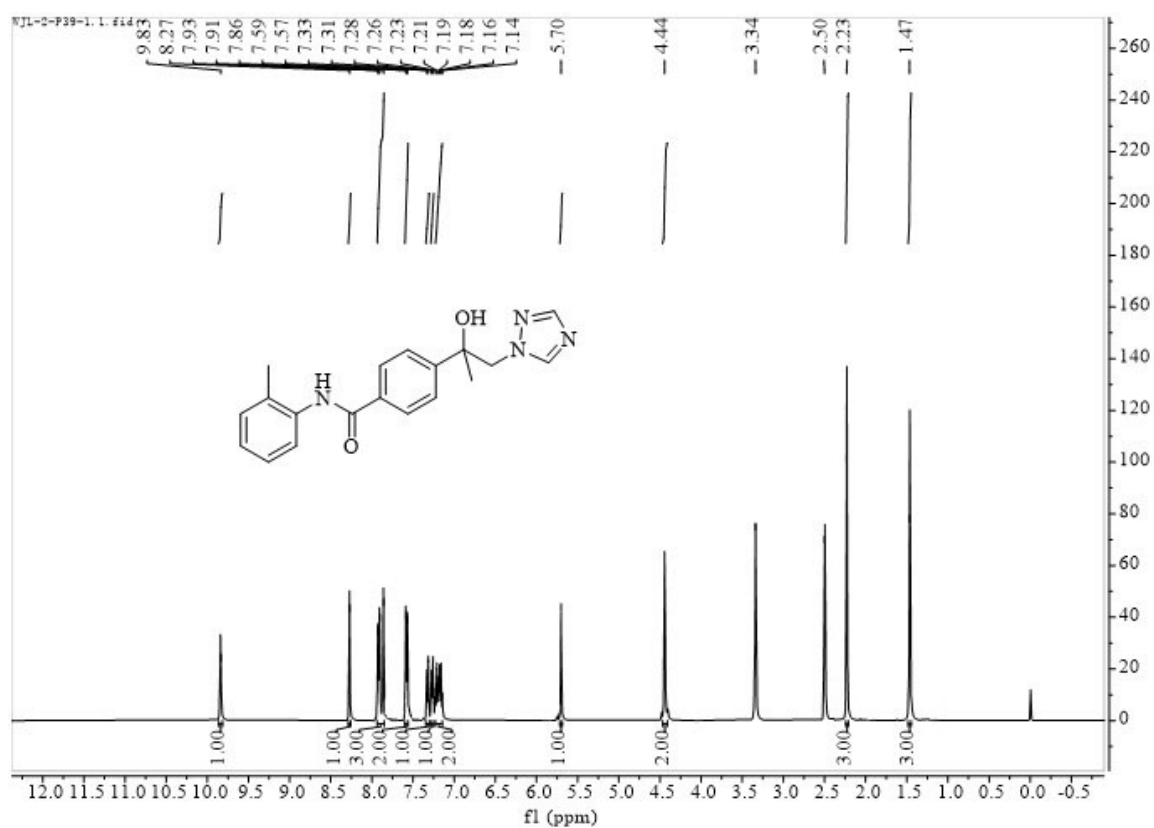
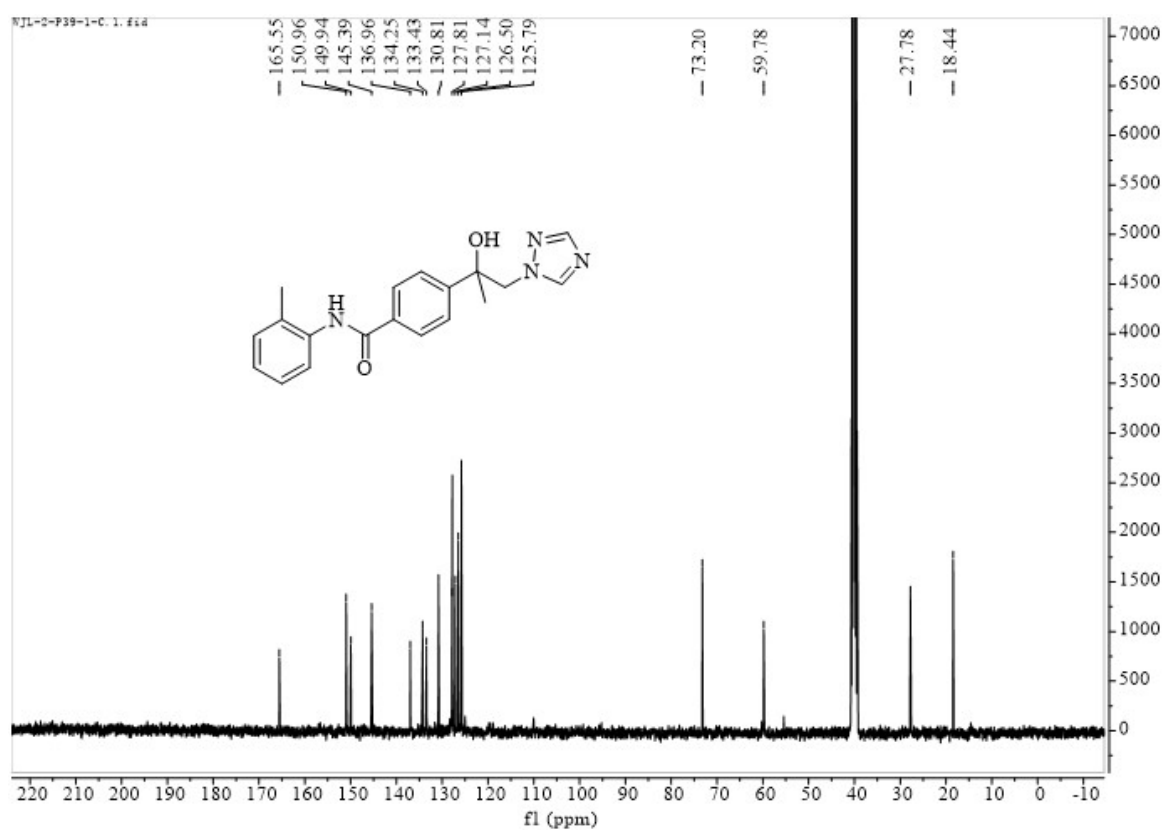
43. Zhao, H.P.; Liu, Y.X.; Cui, Z.P.; Beattie, D.; Gu, Y.C.; Wang, Q.M. Design, synthesis, and biological activities of arylmethylamine substituted chlorotriazine and methylthiotriazine compounds. *J. Agric. Food Chem.* **2011**, *59*, 11711–11717.
44. Seyedi, S.S.; Shukri, M.; Hassandarvish, P.; Oo, A.; Muthu, S.E.; Abubakar, S.; Zandi, K. Computational approach towards exploring potential anti-chikungunya activity of selected flavonoids. *Sci. Rep.* **2016**, *6*, 24027.

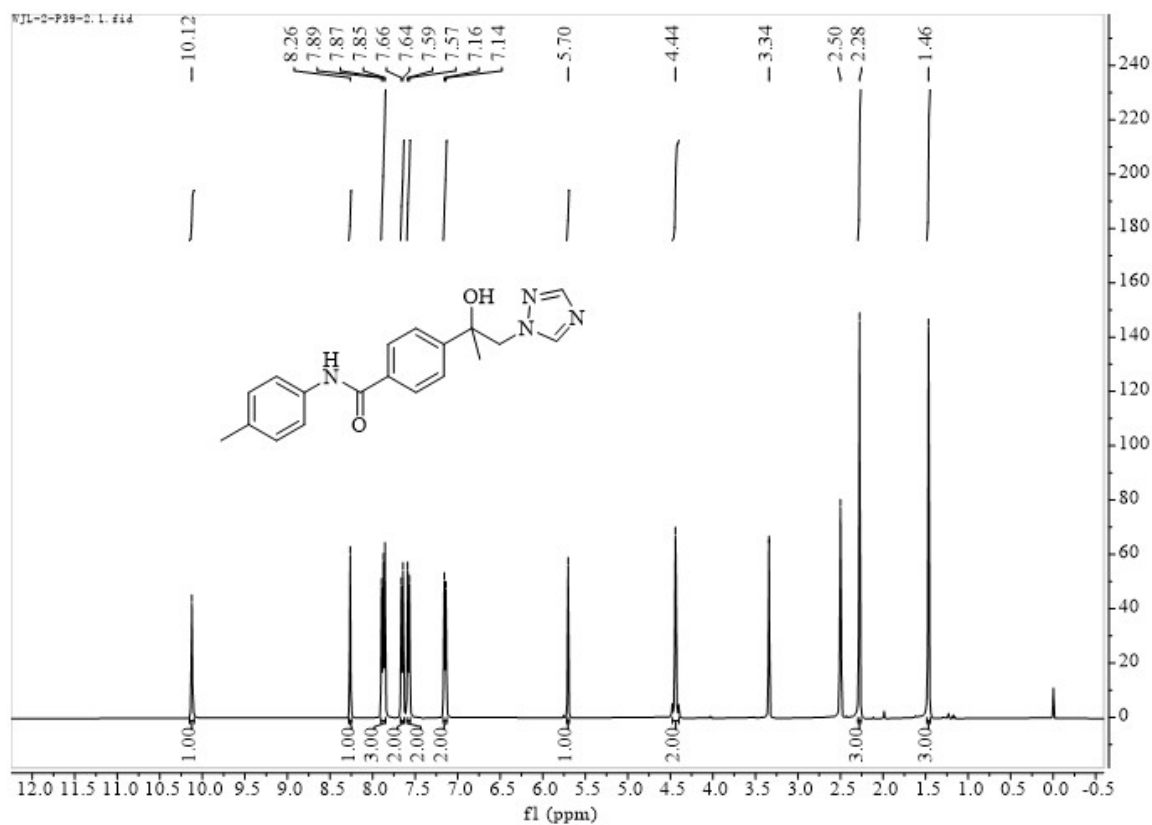
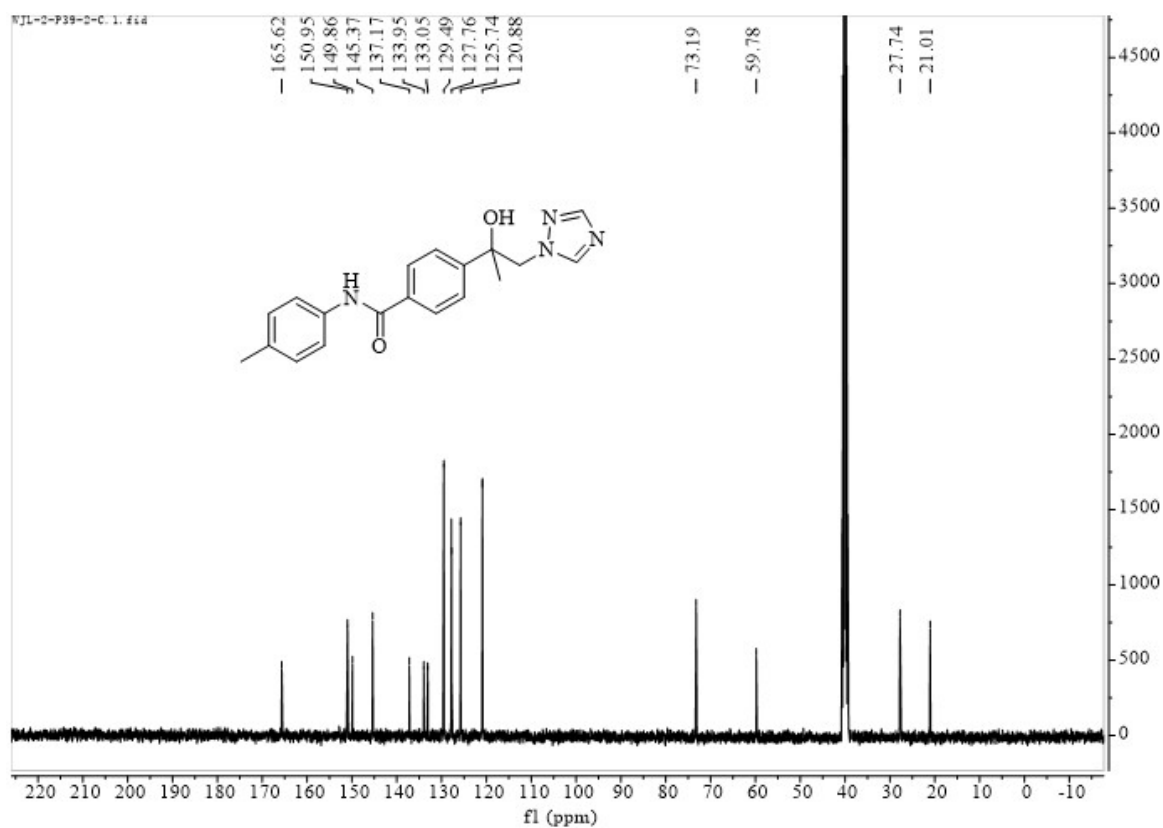
Section S3. Figures S1–S74. ^1H , and ^{13}C NMR spectra of 2–7Figure S1. ^1H NMR spectrum of 2Figure S2. ^{13}C NMR spectrum of 2

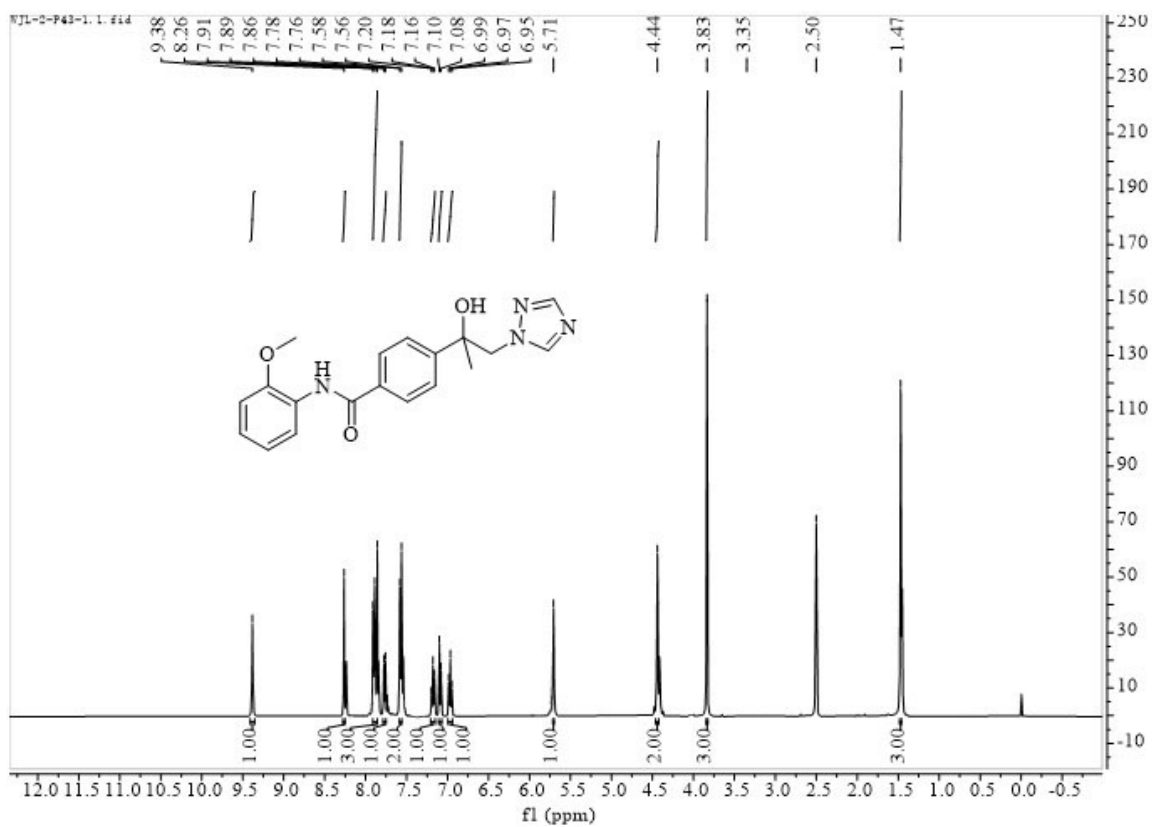
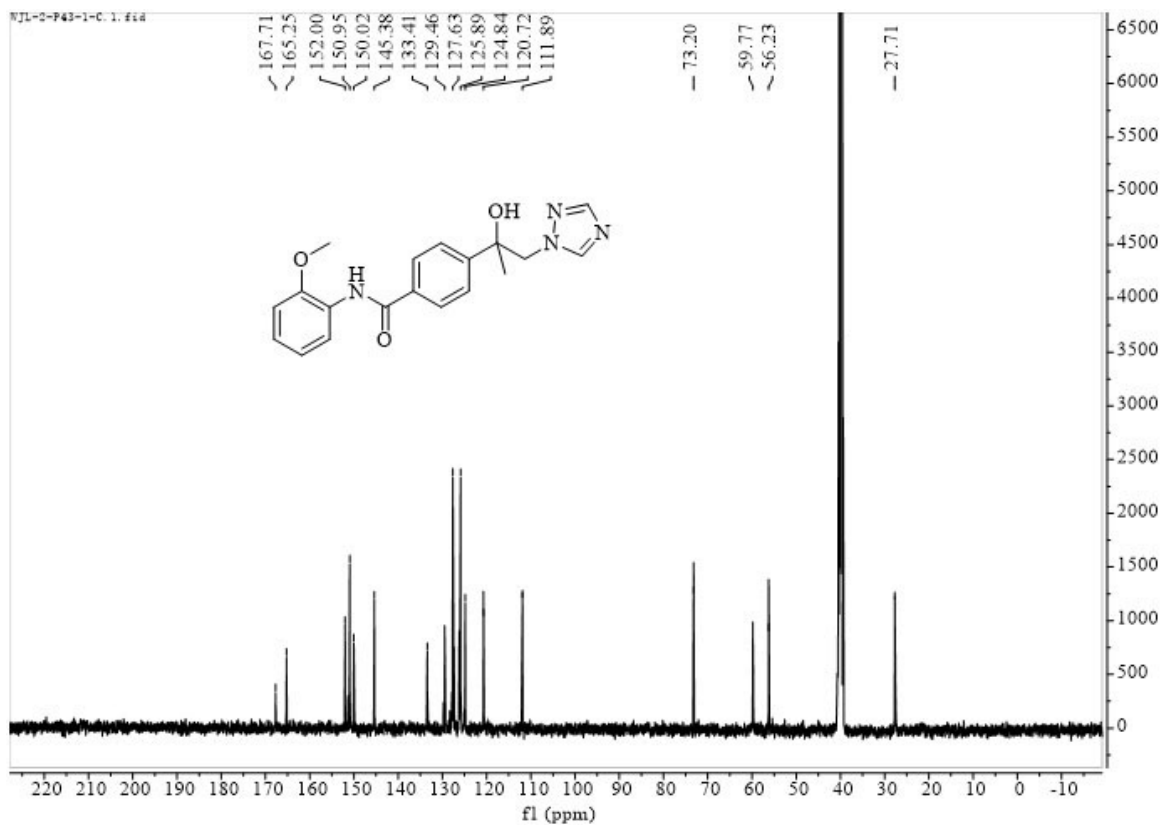
Figure S3. ^1H NMR spectrum of 3Figure S4. ^{13}C NMR spectrum of 3

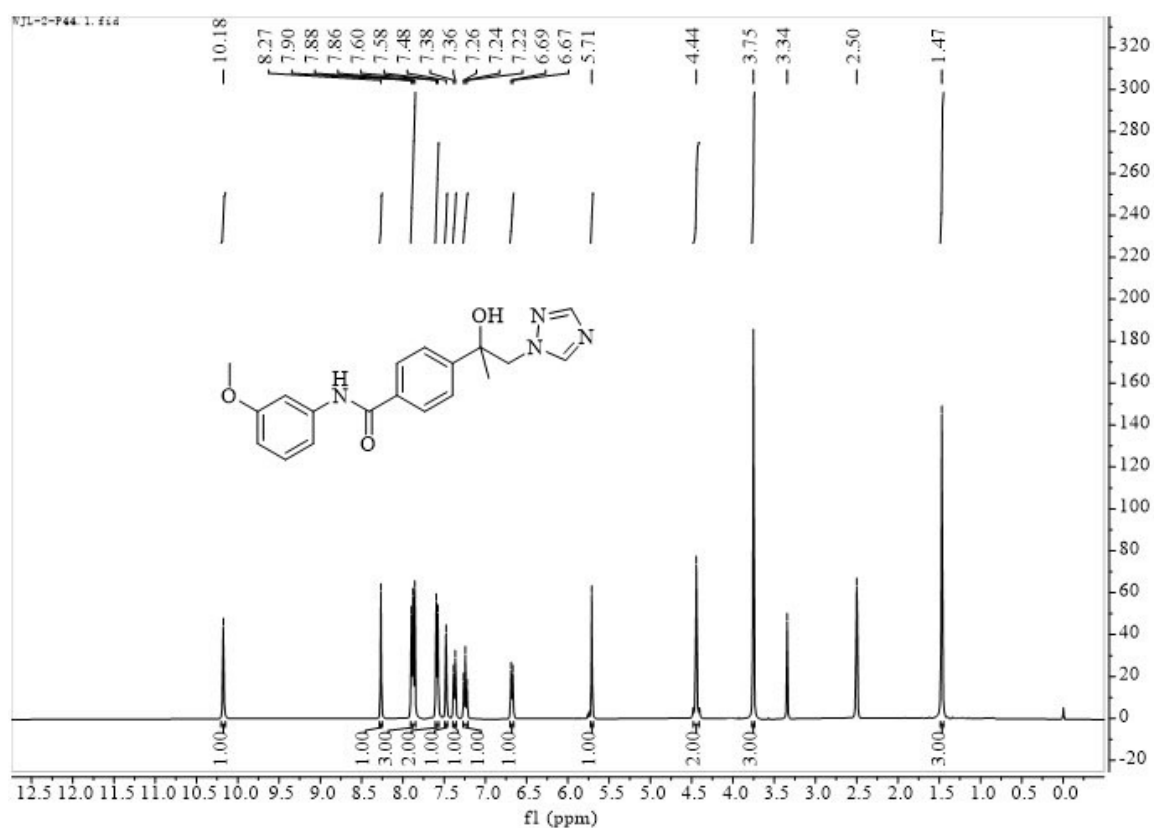
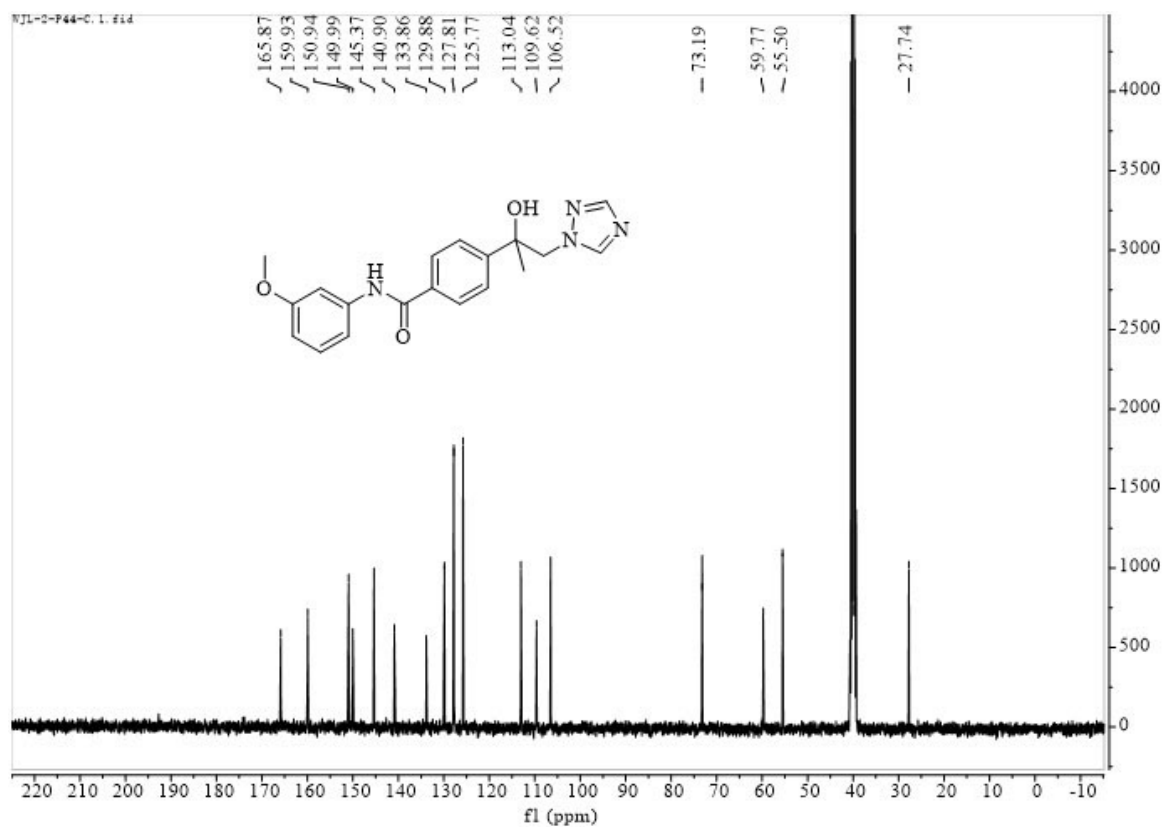
Figure S5. ¹H NMR spectrum of 4Figure S6. ¹³C NMR spectrum of 4

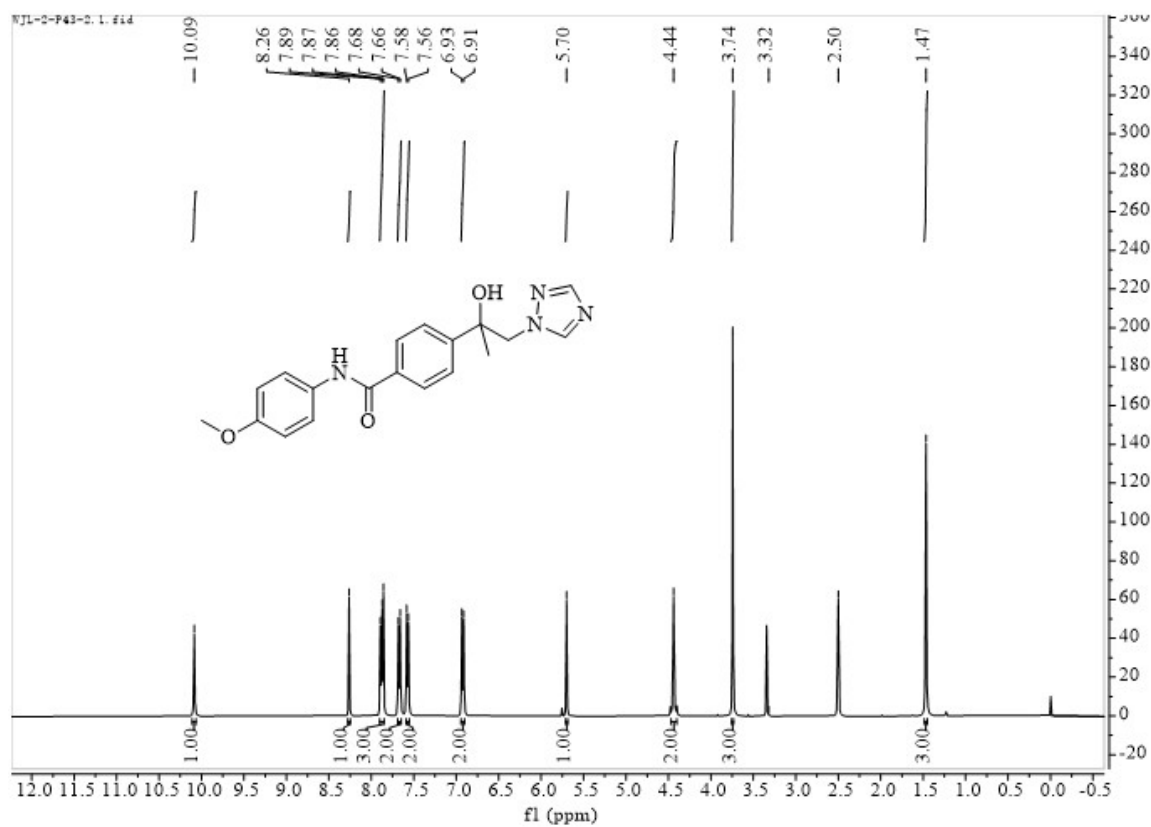
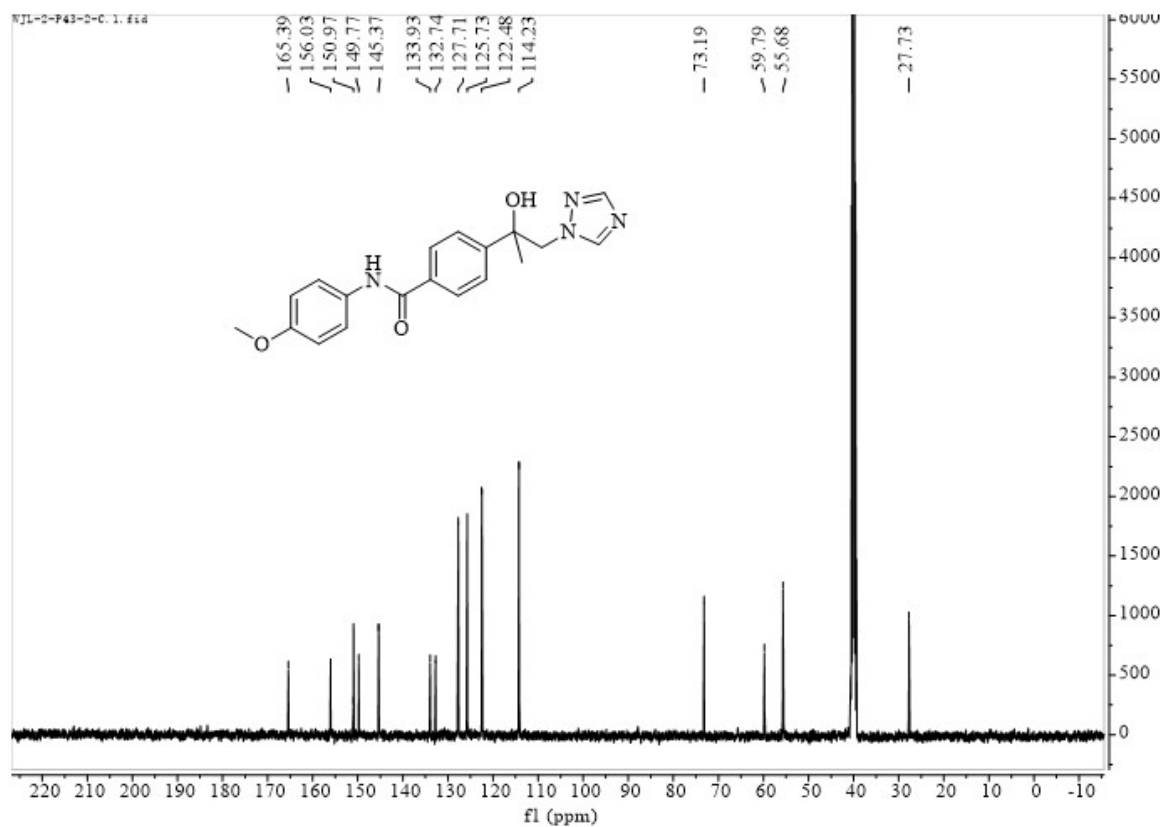
Figure S7. ¹H NMR spectrum of 5aFigure S8. ¹³C NMR spectrum of 5a

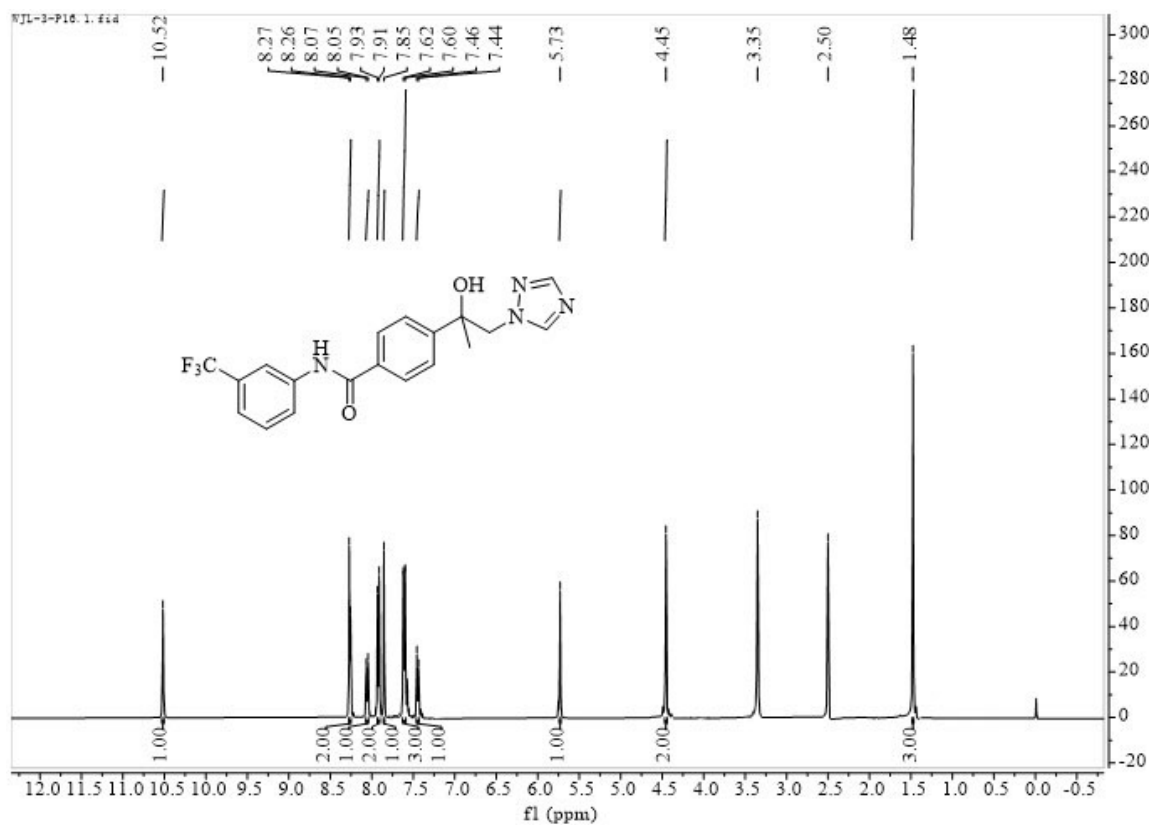
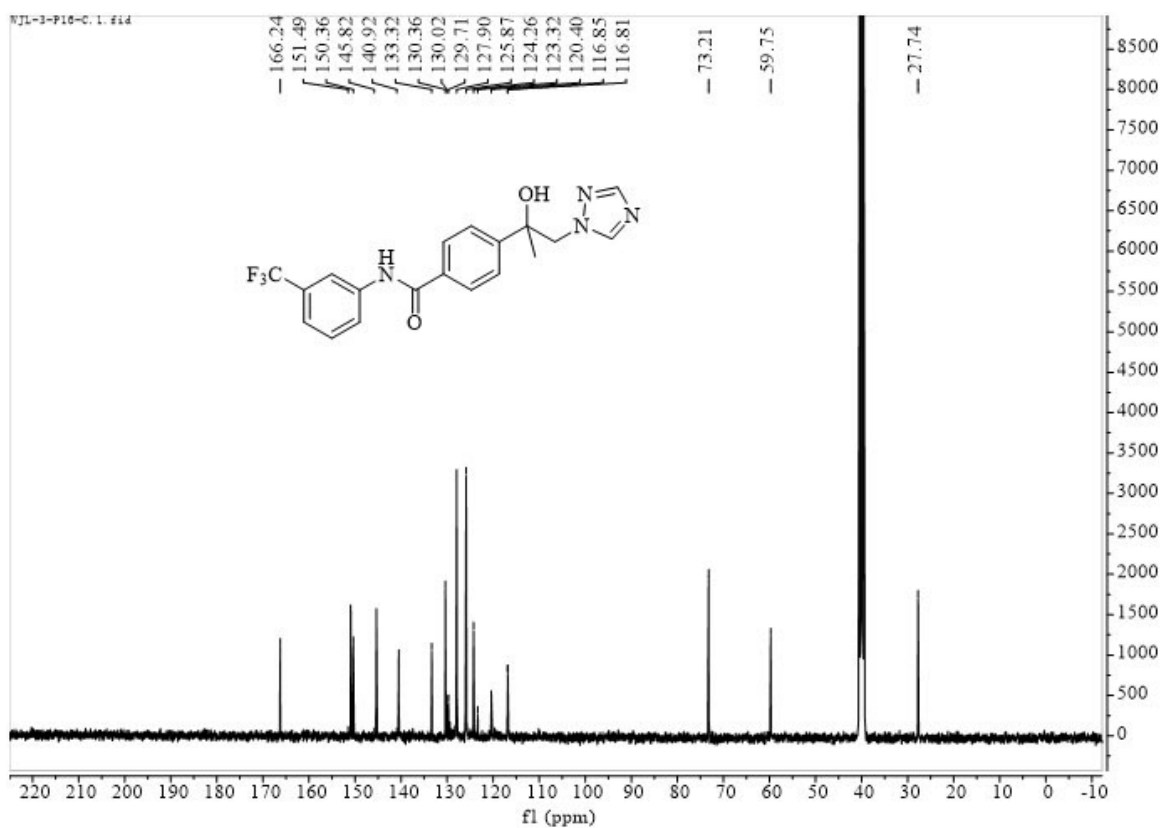
Figure S9. ¹H NMR spectrum of **5b**Figure S10. ¹³C NMR spectrum of **5b**

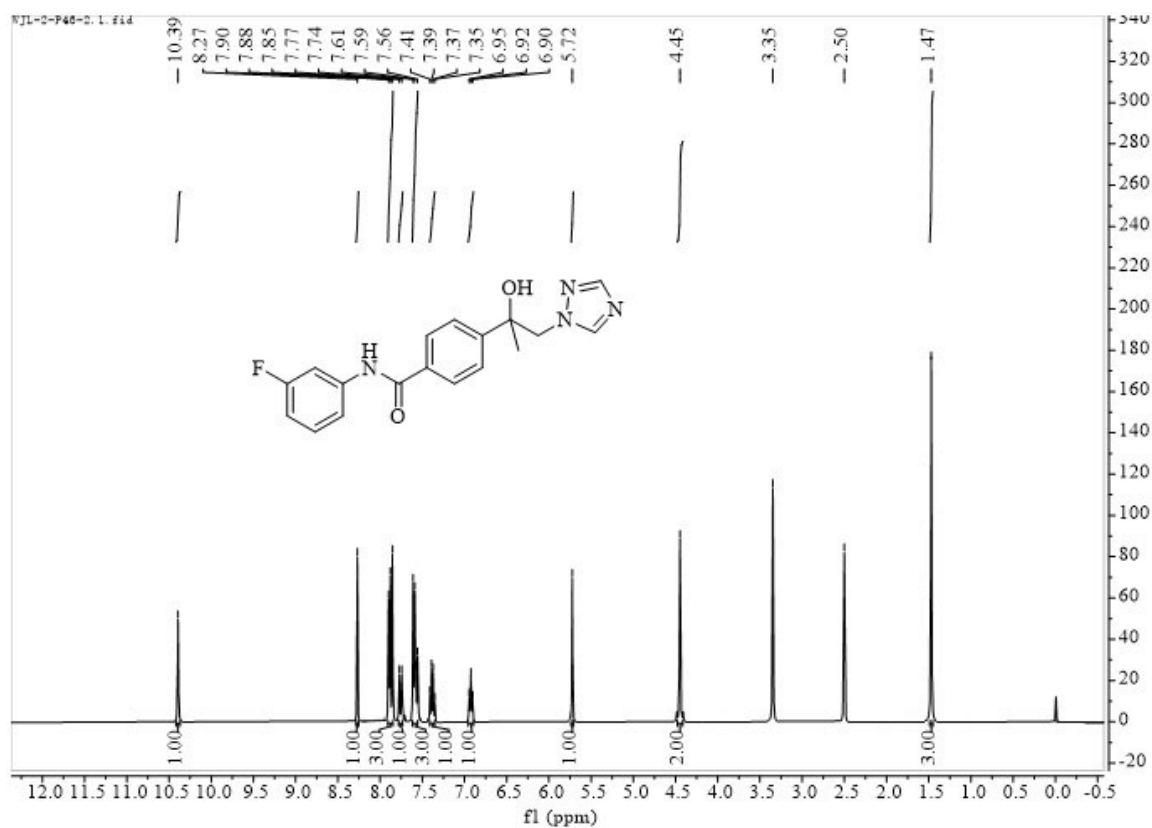
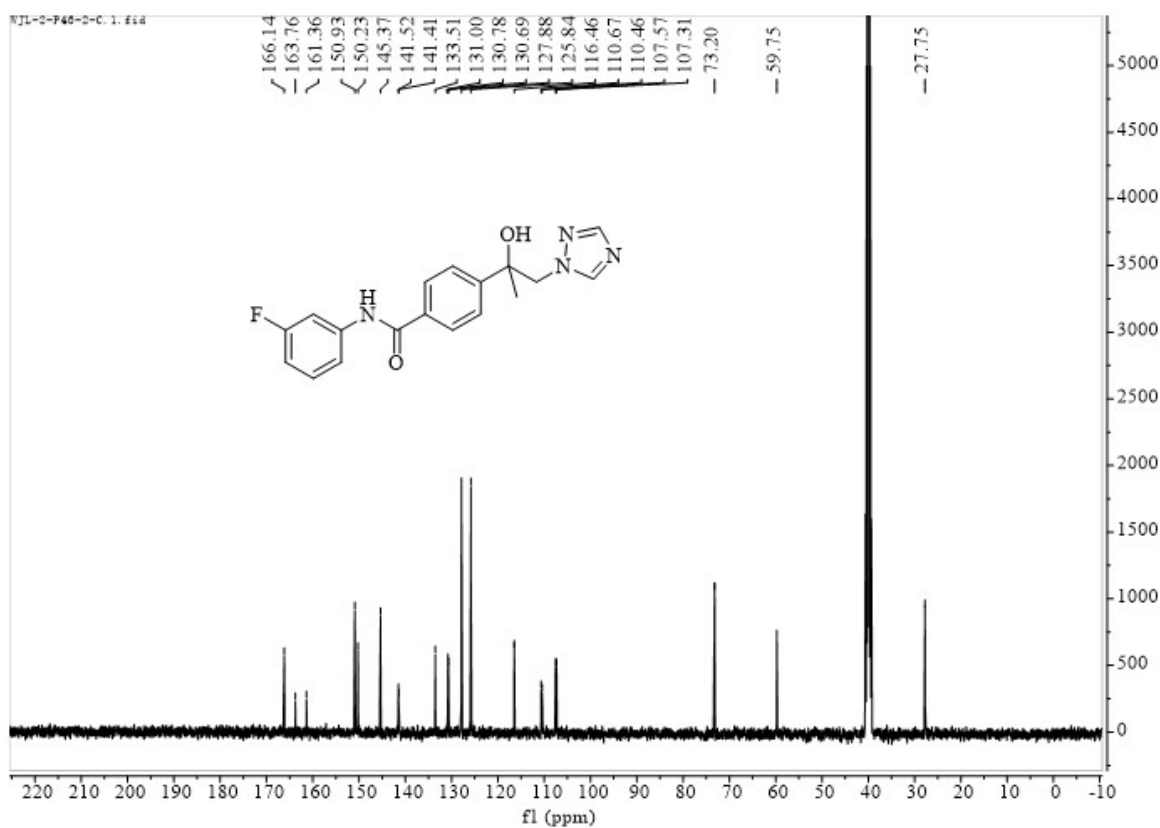
Figure S11. ^1H NMR spectrum of 5cFigure S12. ^{13}C NMR spectrum of 5c

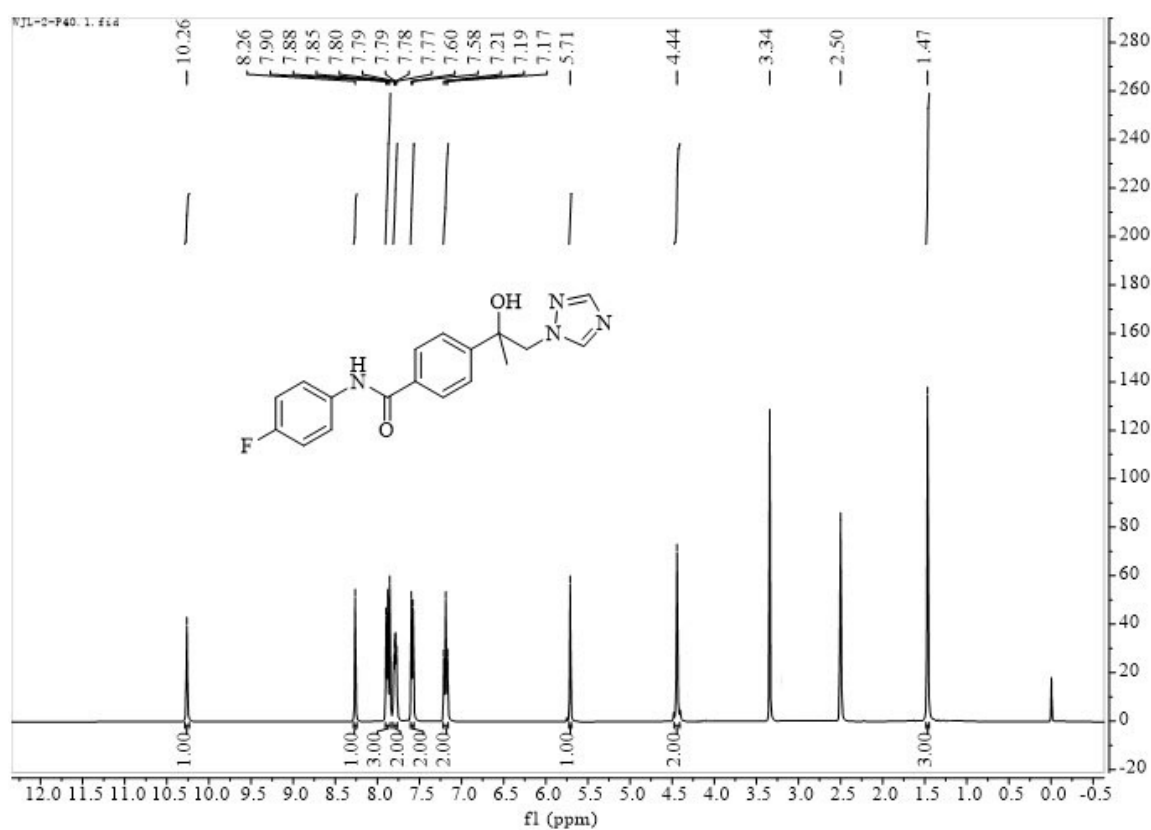
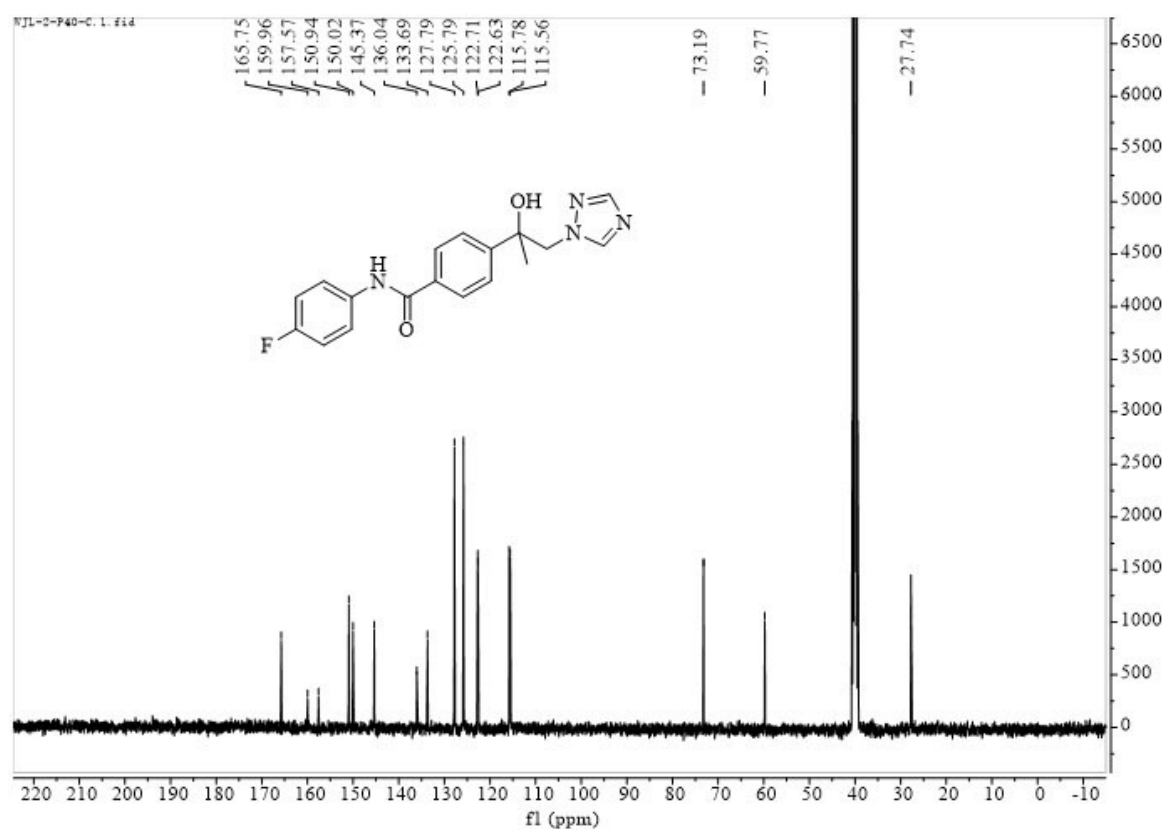
Figure S13. ¹H NMR spectrum of 5dFigure S14. ¹³C NMR spectrum of 5d

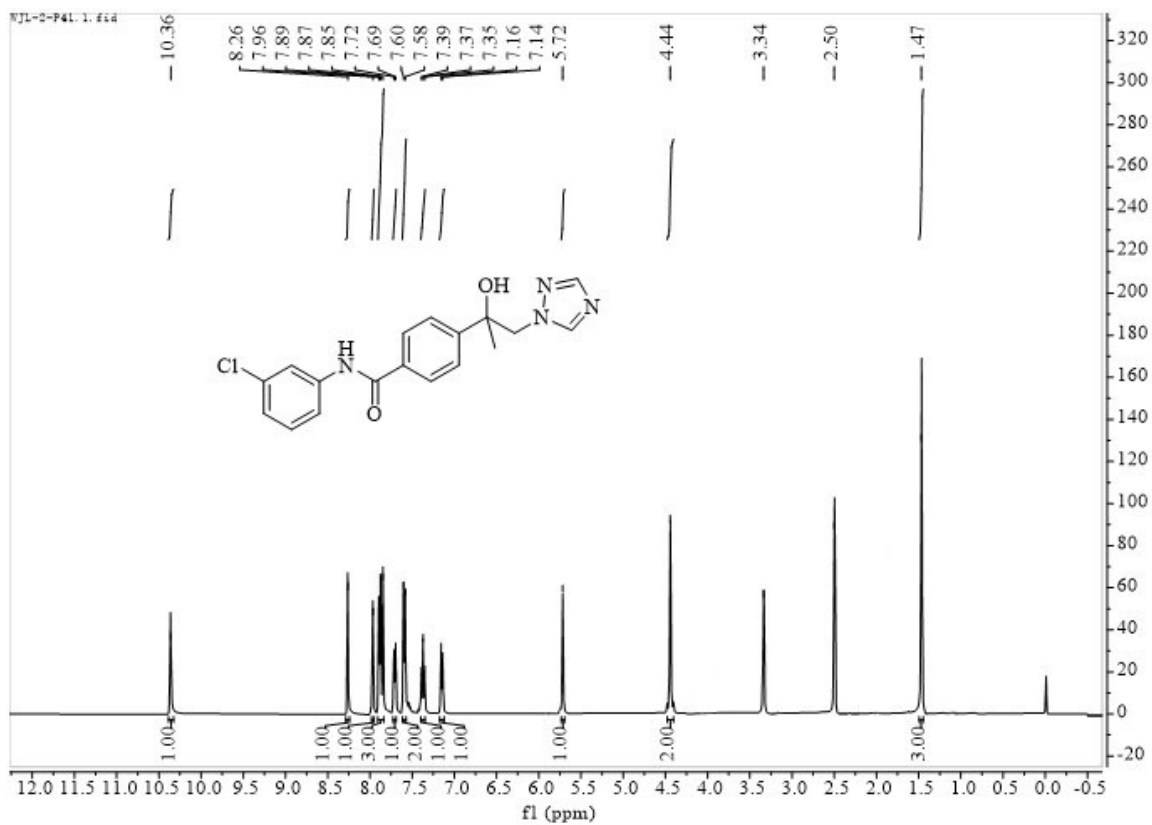
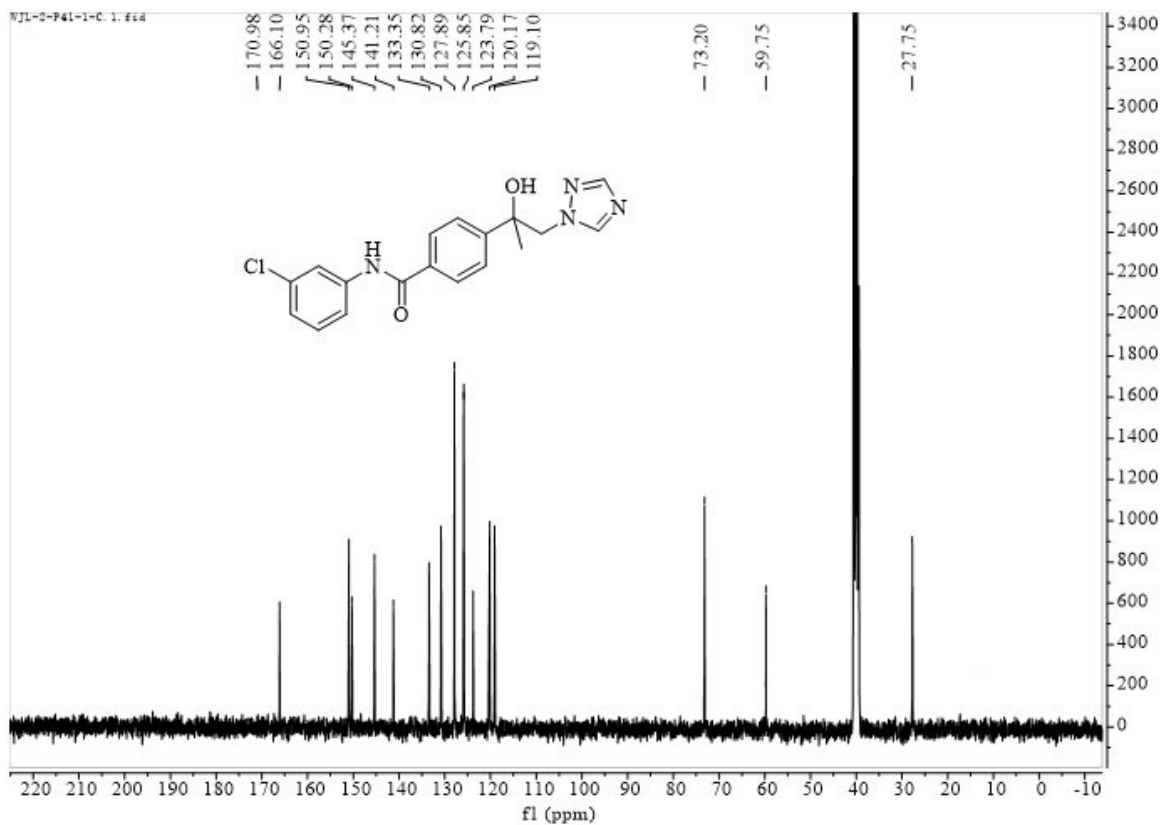
Figure S15. ^1H NMR spectrum of 5eFigure S16. ^{13}C NMR spectrum of 5e

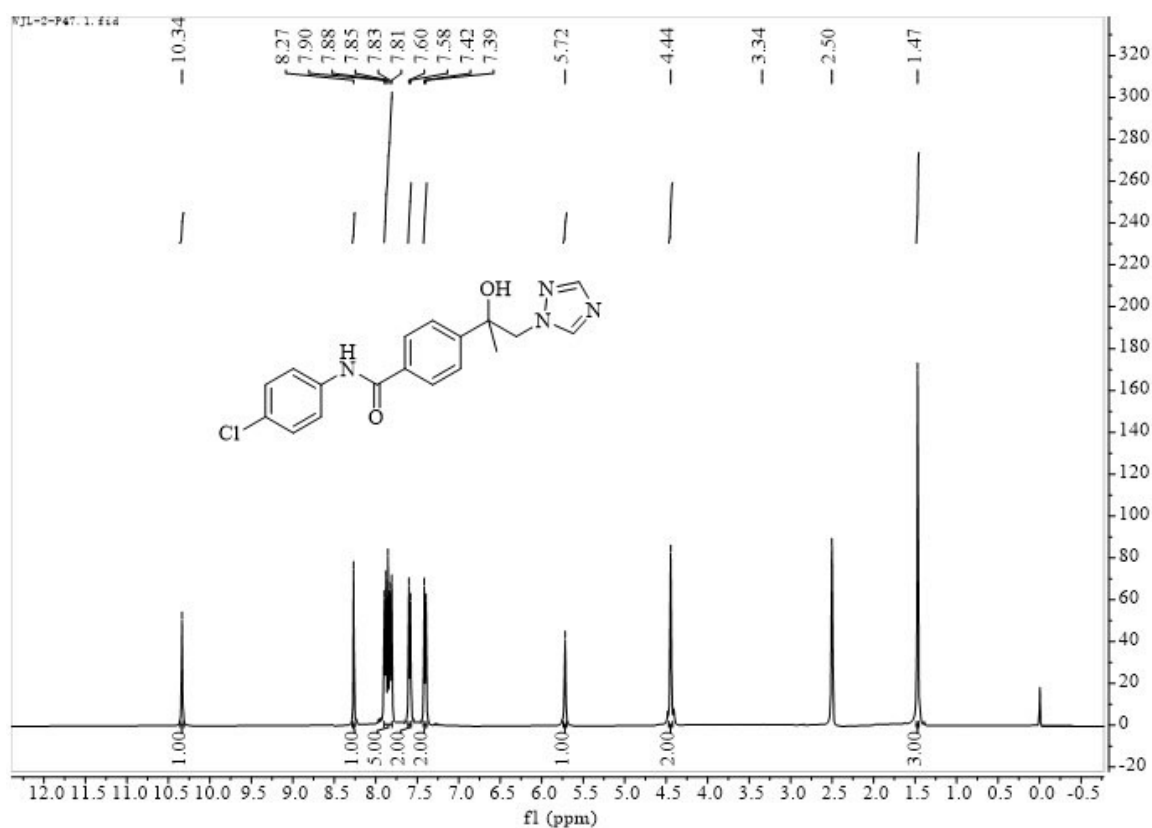
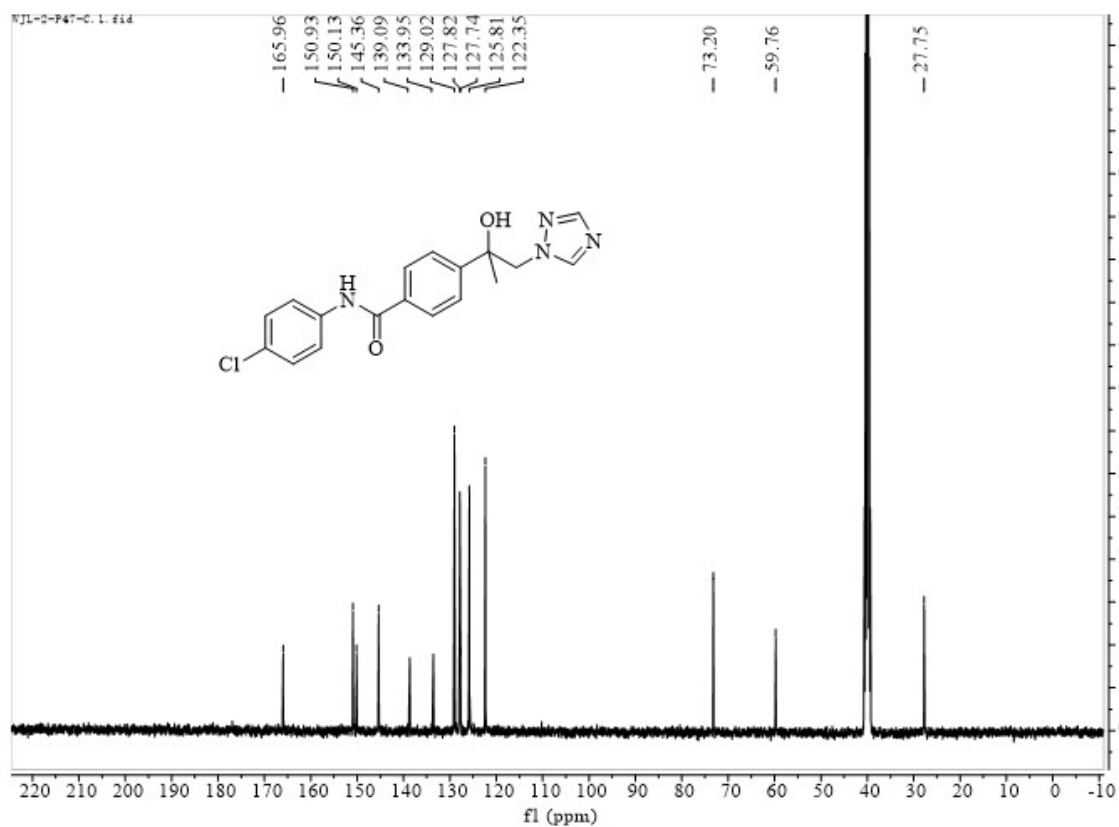
Figure S17. ^1H NMR spectrum of 5fFigure S18. ^{13}C NMR spectrum of 5f

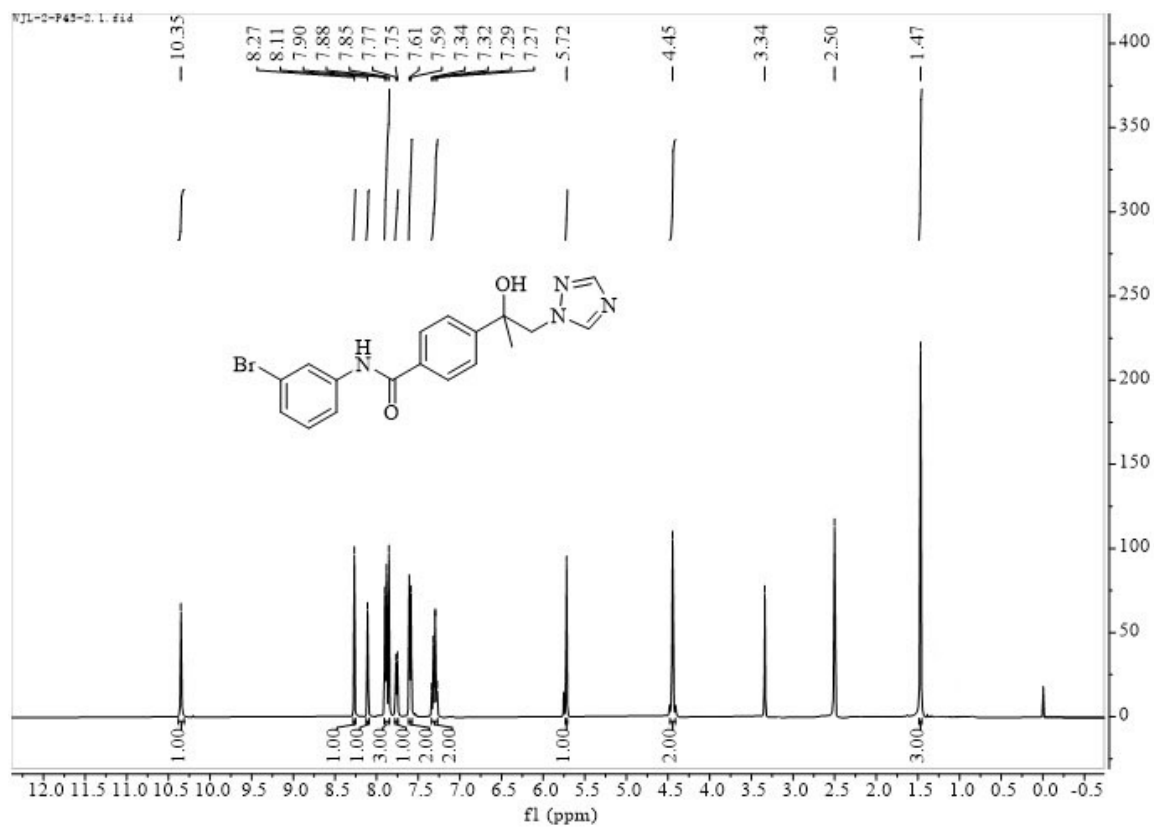
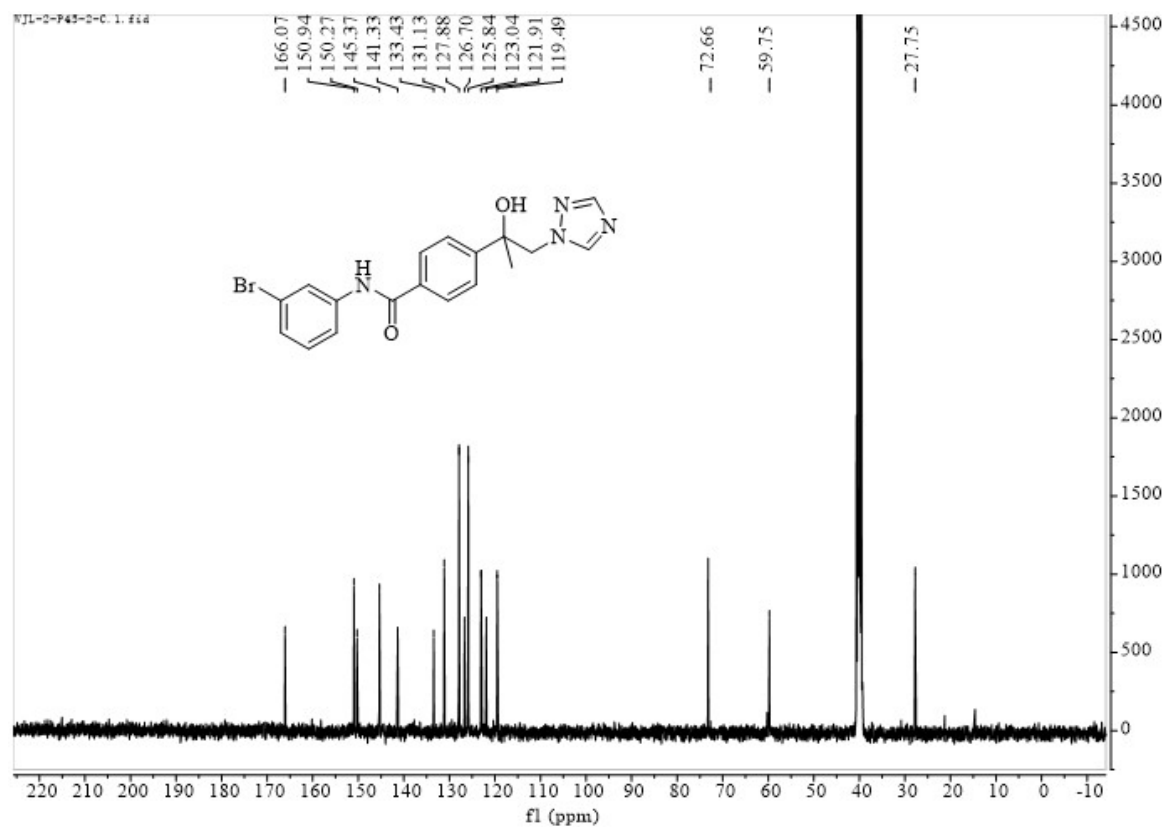
Figure S19. ¹H NMR spectrum of **5g**Figure S20. ¹³C NMR spectrum of **5g**

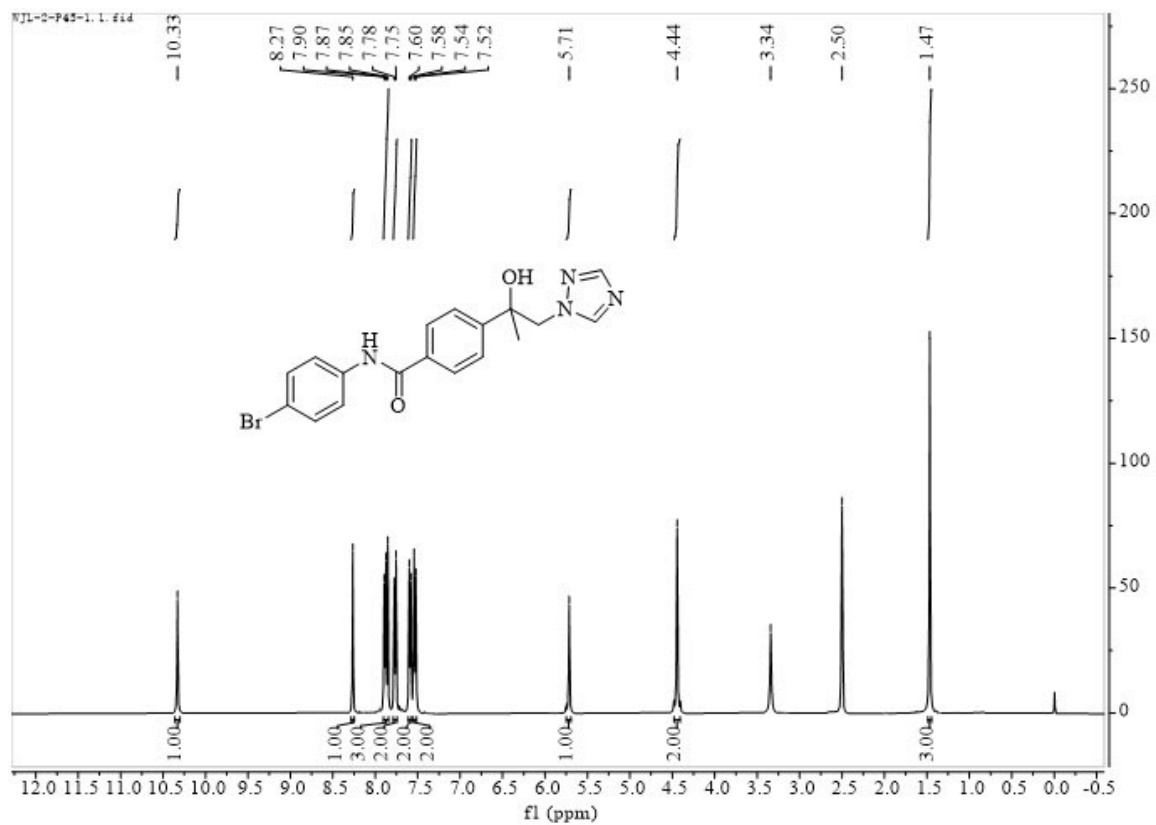
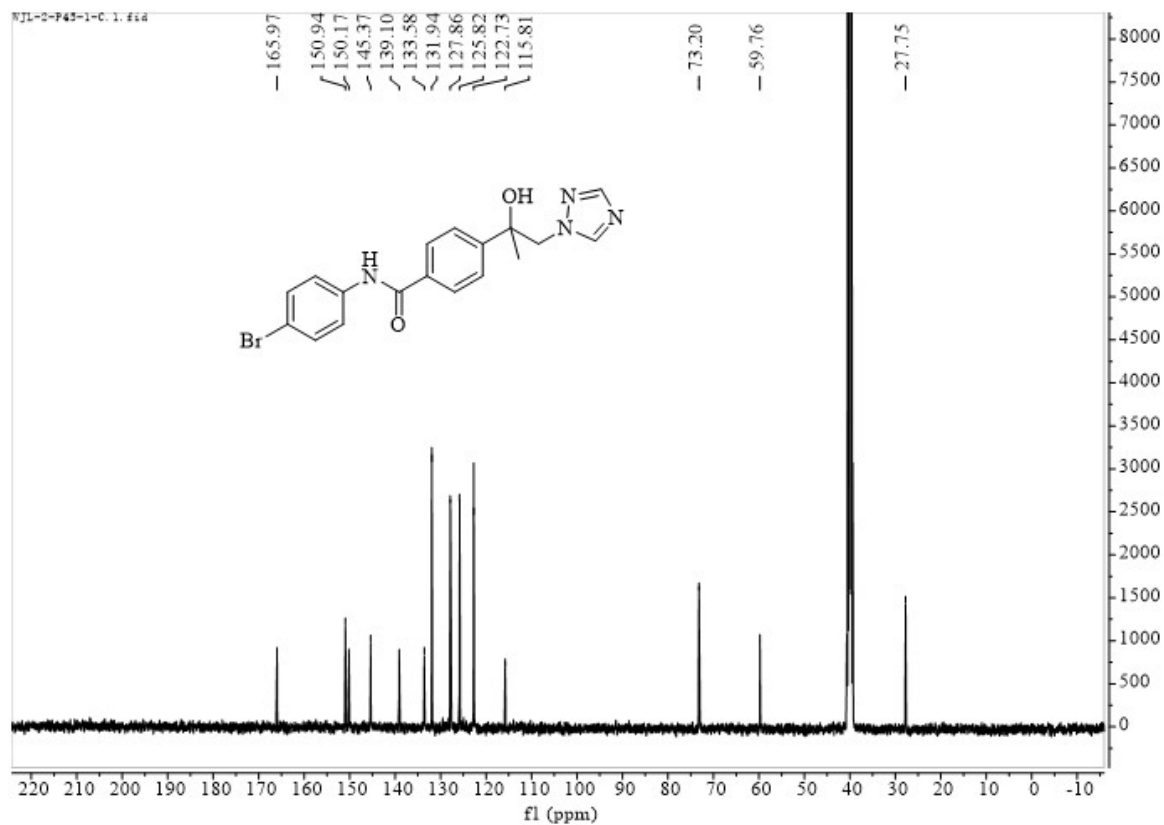
Figure S21. ^1H NMR spectrum of 5hFigure S22. ^{13}C NMR spectrum of 5h

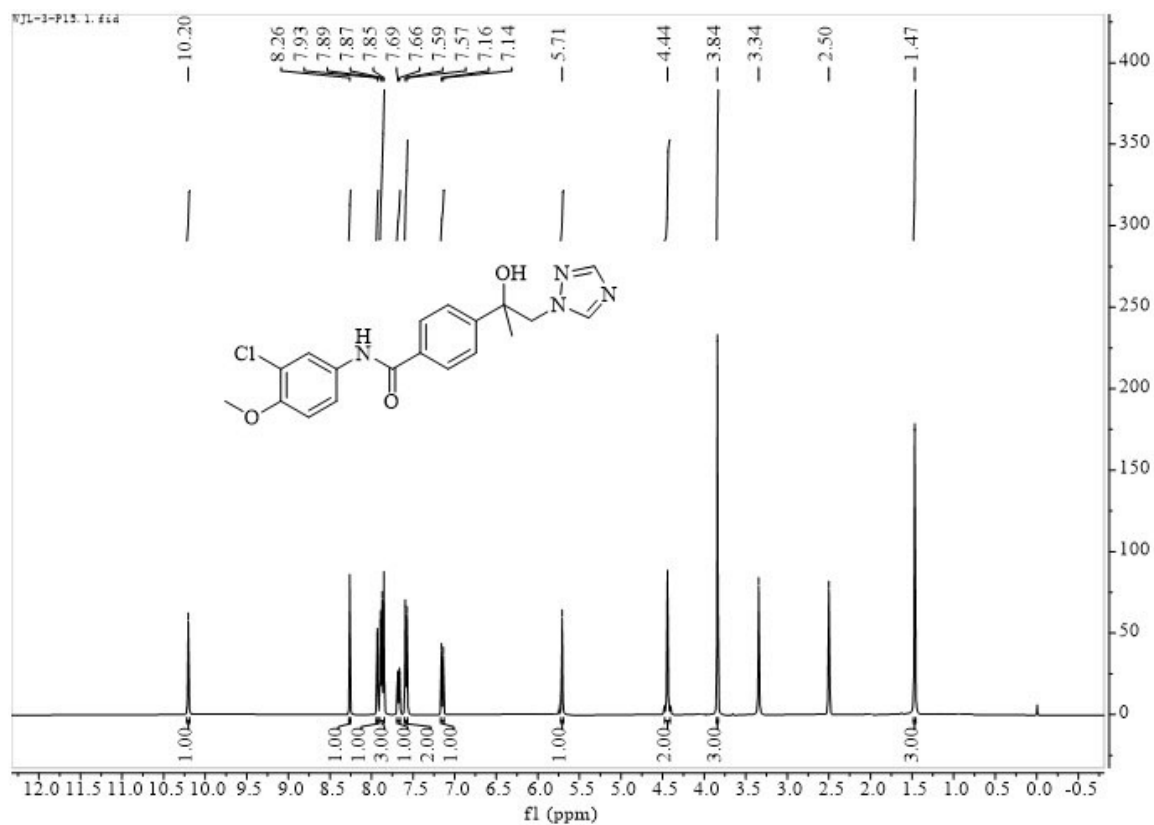
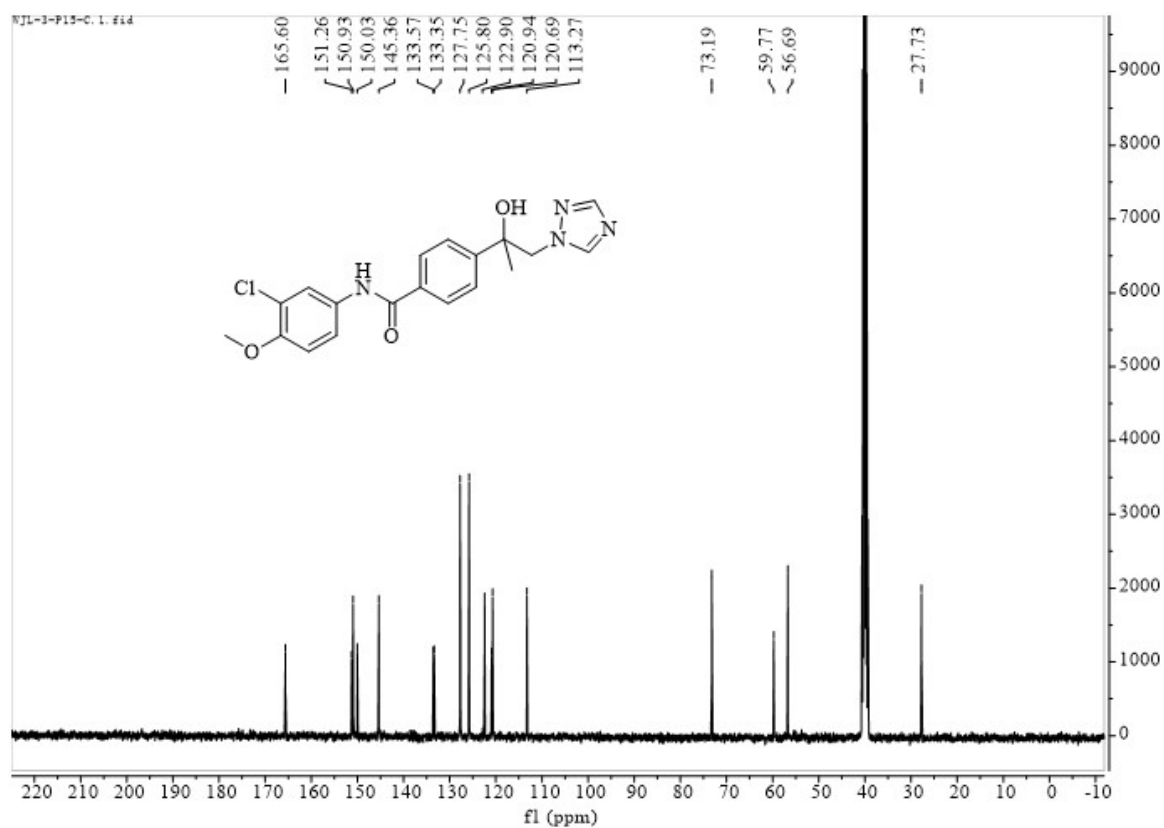
Figure S23. ¹H NMR spectrum of **5i**Figure S24. ¹³C NMR spectrum of **5i**

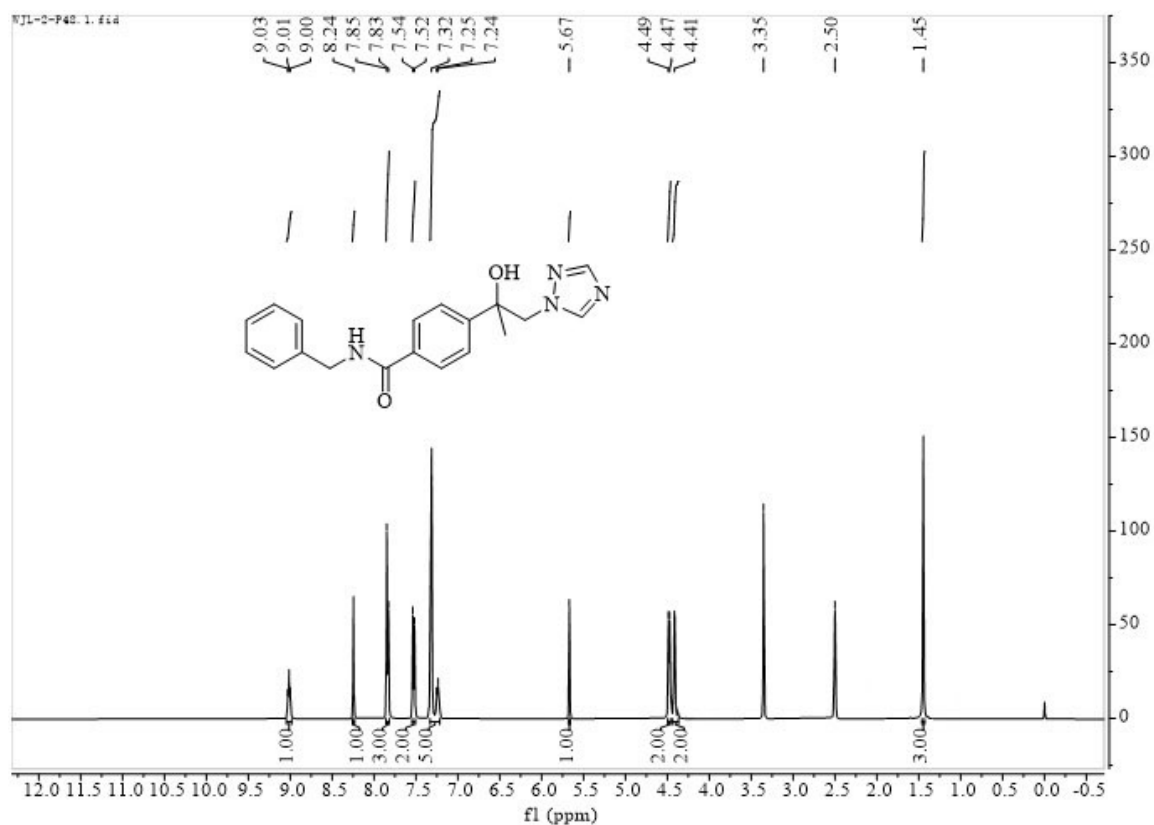
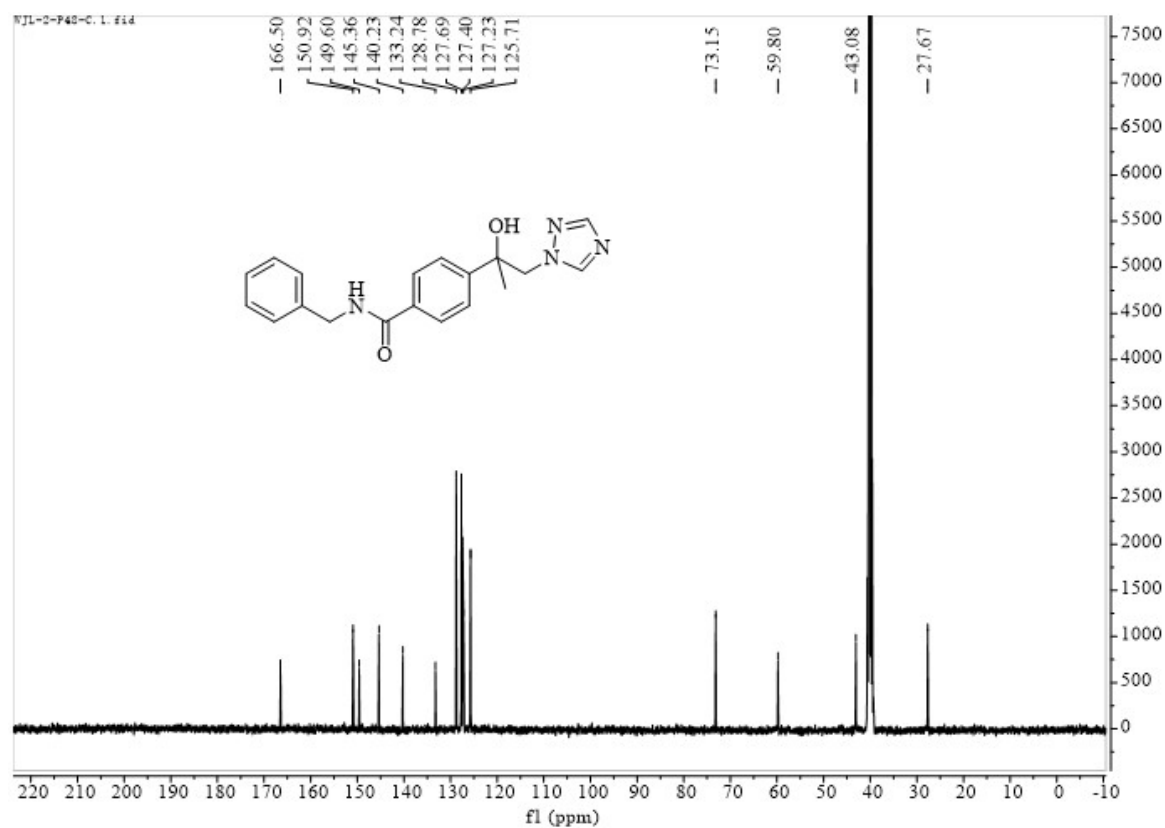
Figure S25. ^1H NMR spectrum of 5jFigure S26. ^{13}C NMR spectrum of 5j

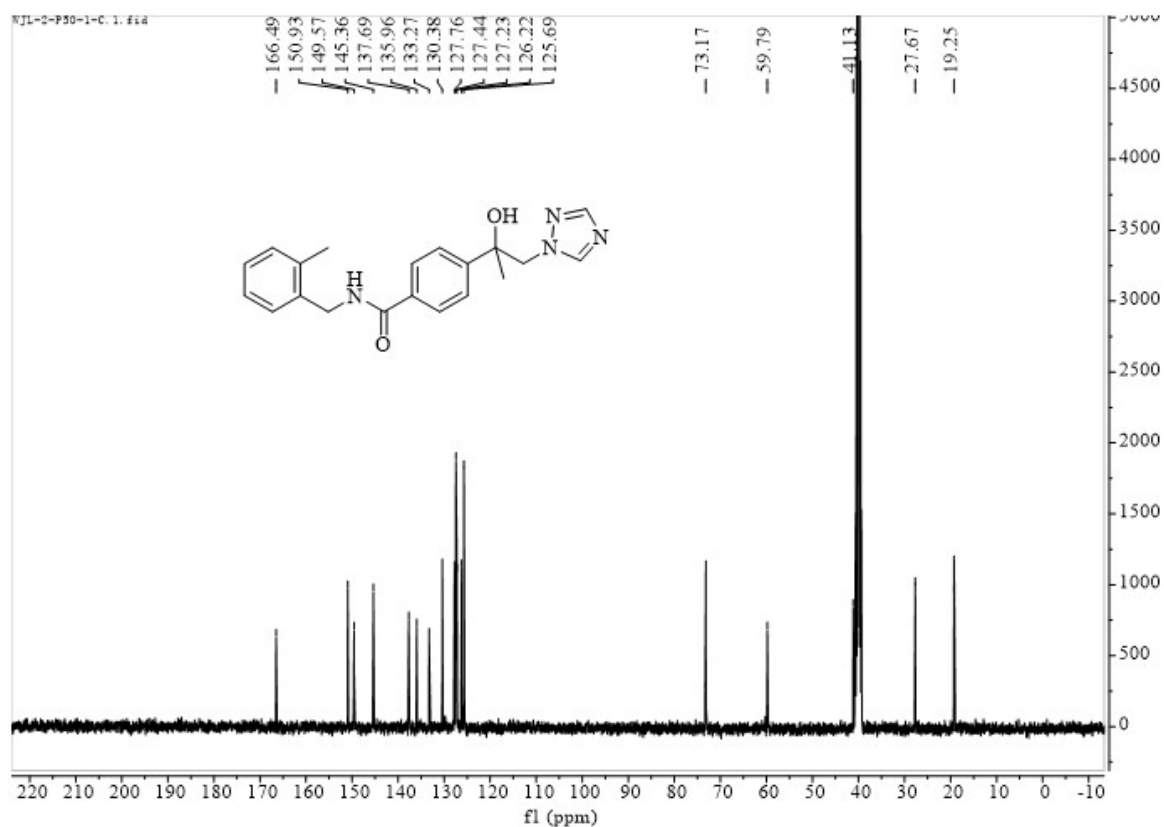
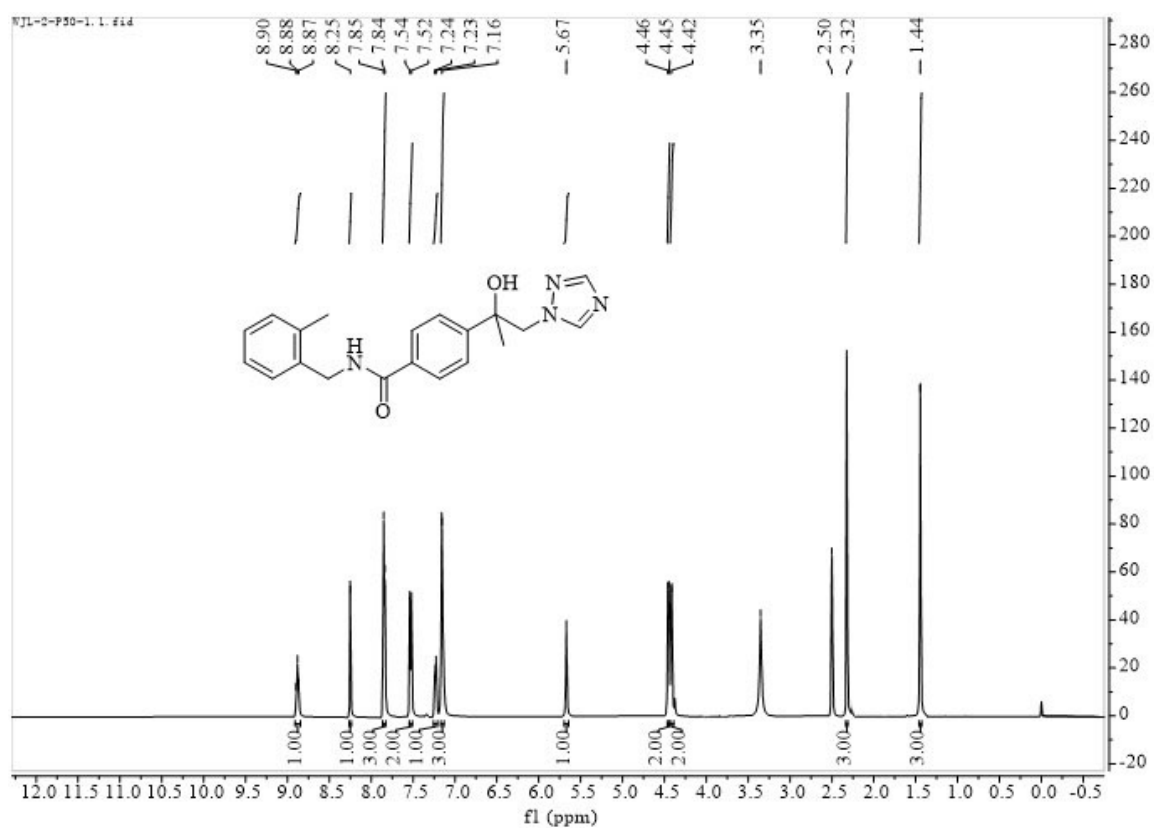
Figure S27. ¹H NMR spectrum of 5kFigure S28. ¹³C NMR spectrum of 5k

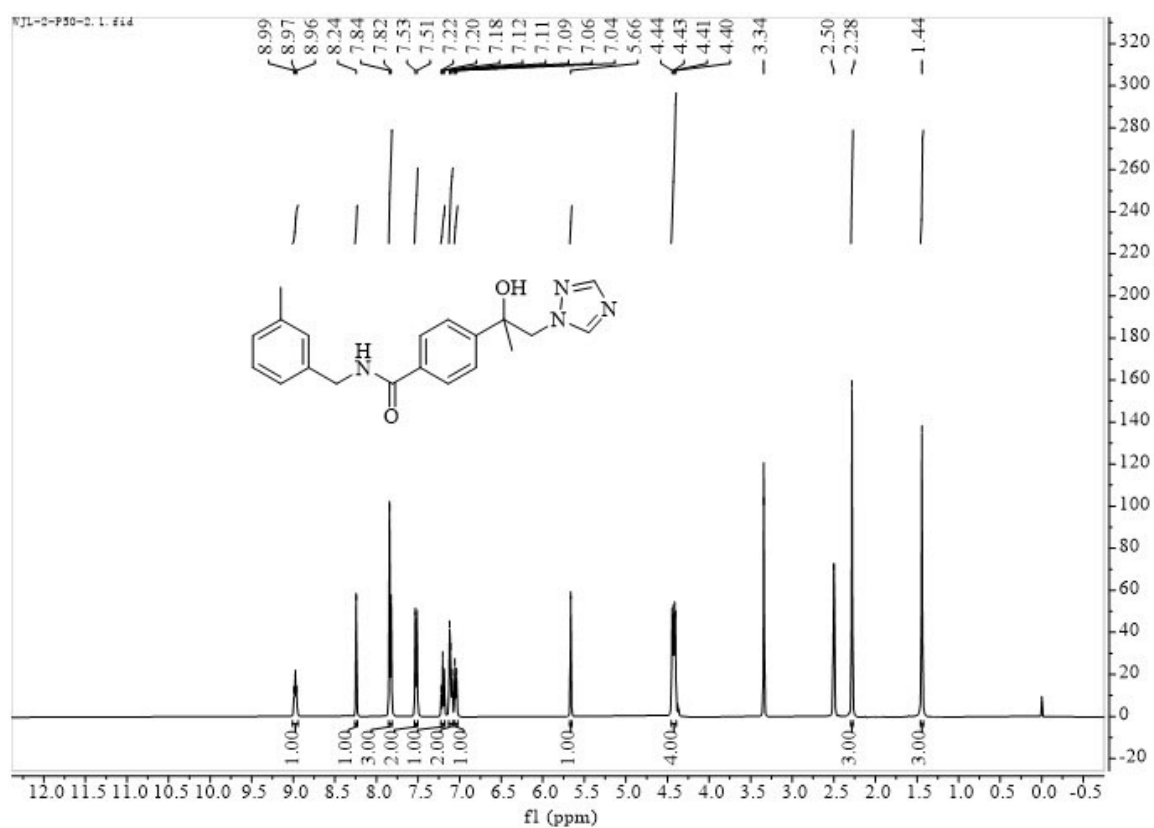
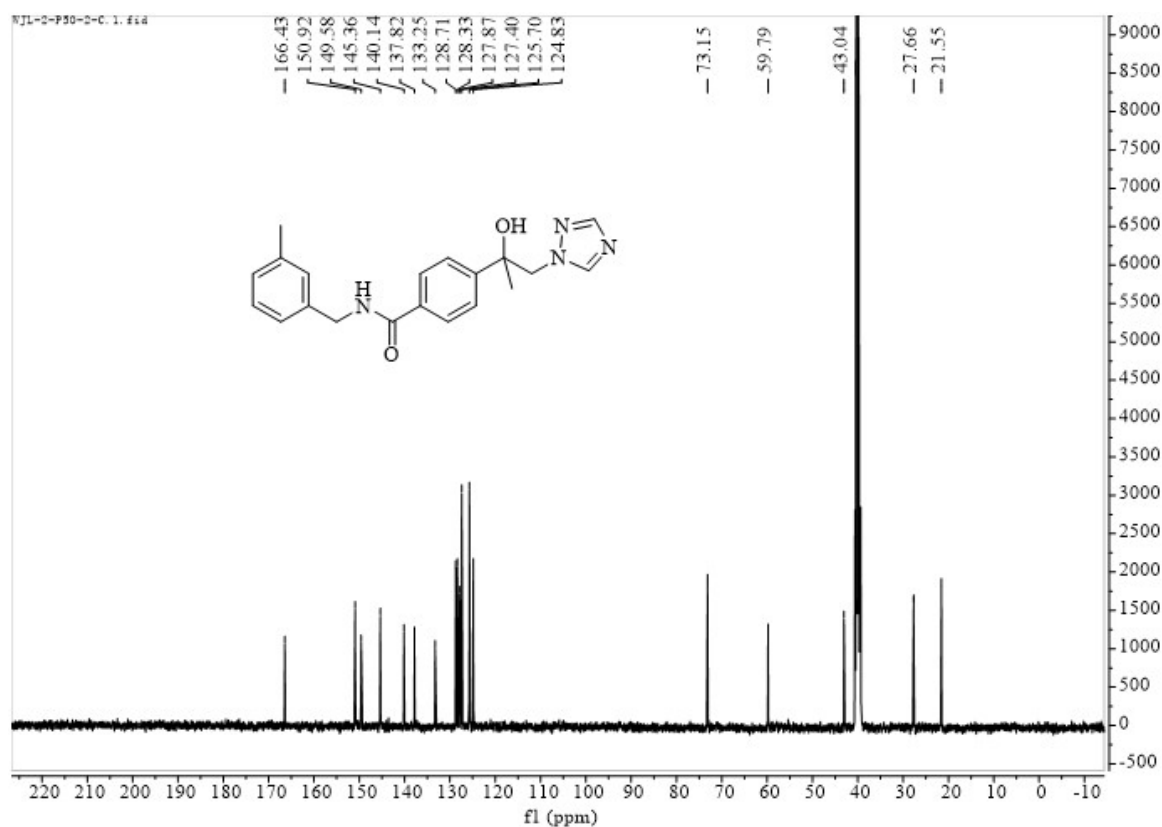
Figure S29. ^1H NMR spectrum of 51Figure S30. ^{13}C NMR spectrum of 51

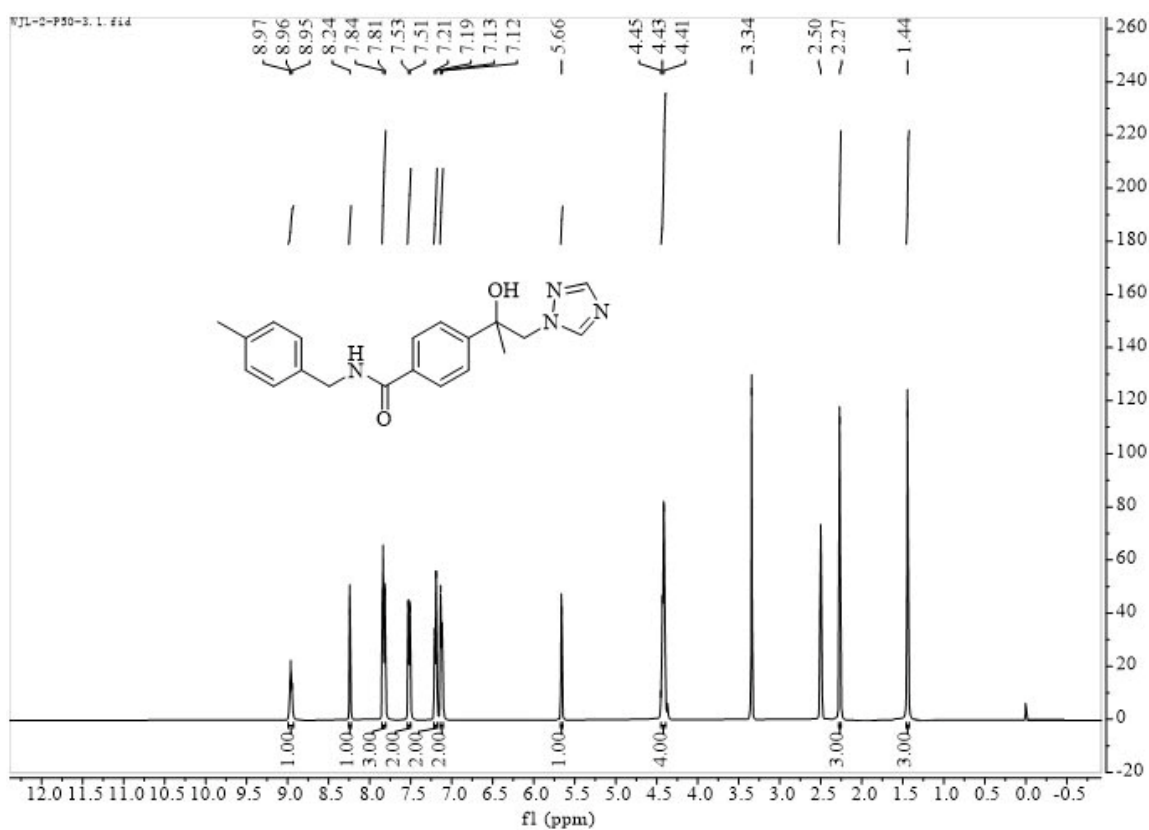
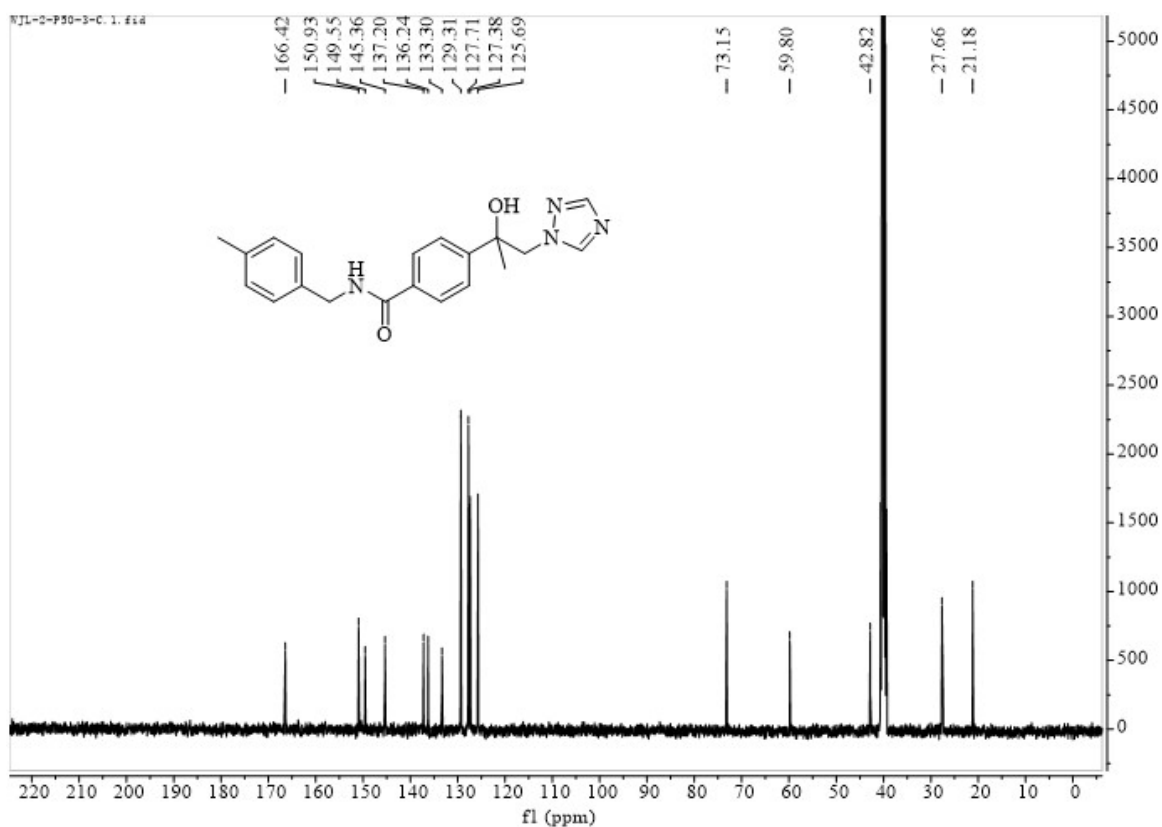
Figure S31. ¹H NMR spectrum of 5mFigure S32. ¹³C NMR spectrum of 5m

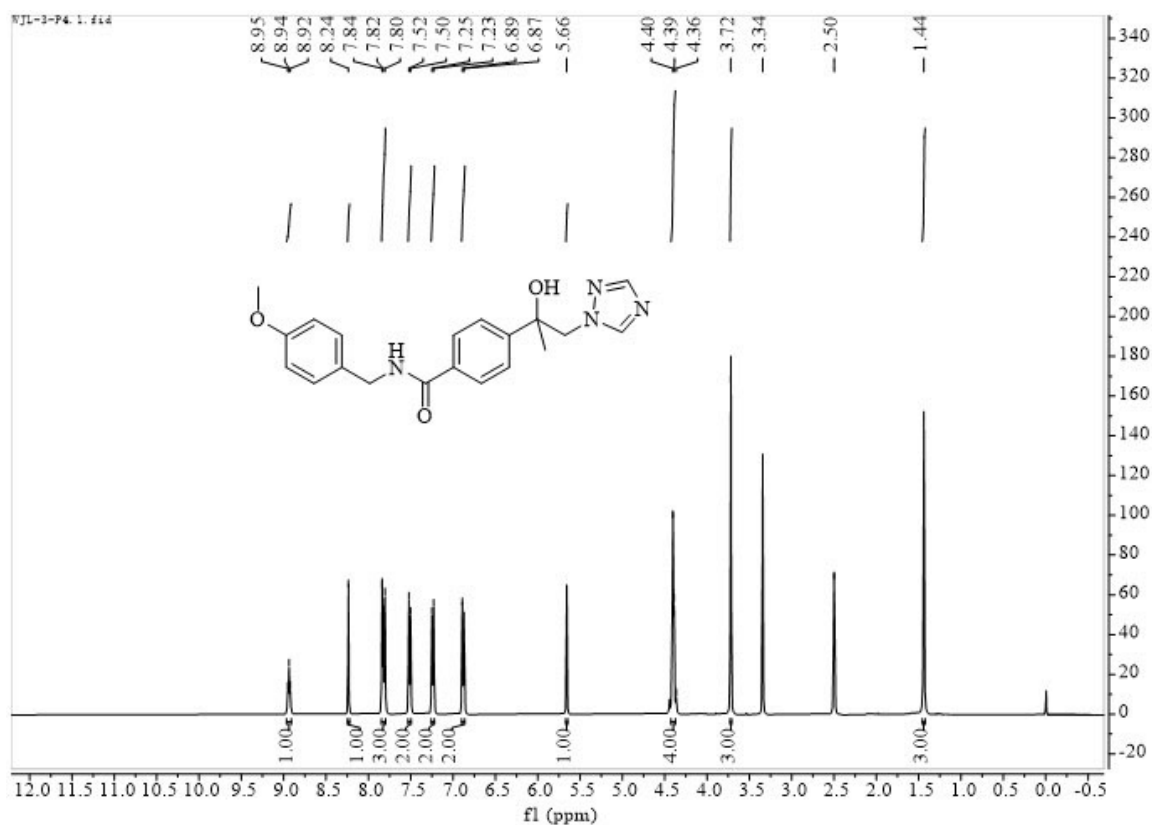
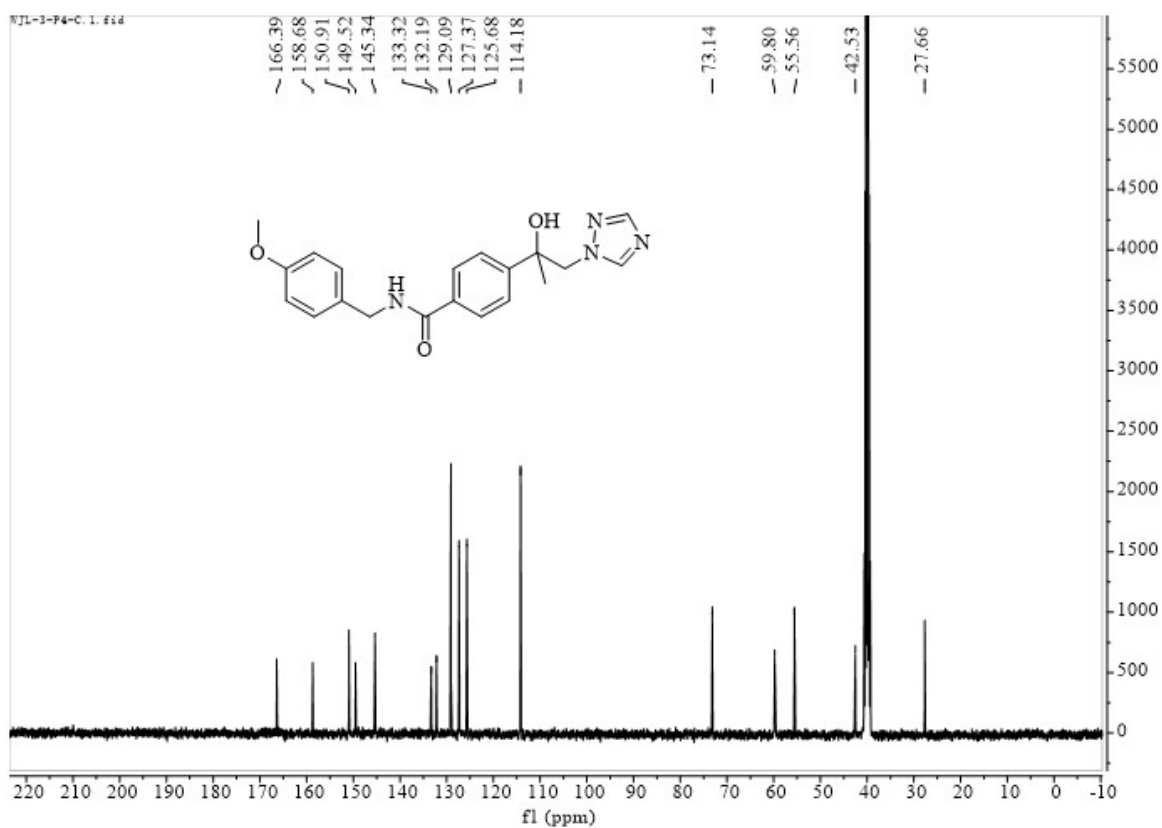
Figure S33. ¹H NMR spectrum of **5n**Figure S34. ¹³C NMR spectrum of **5n**

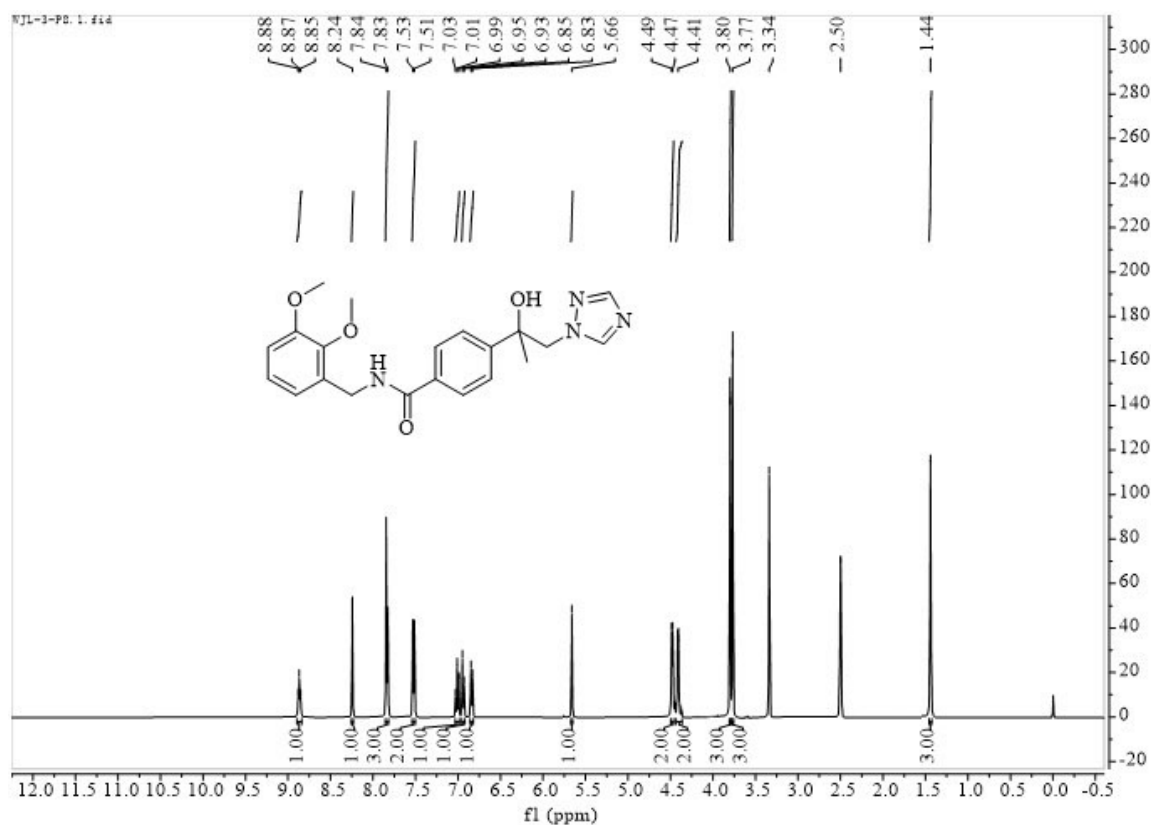
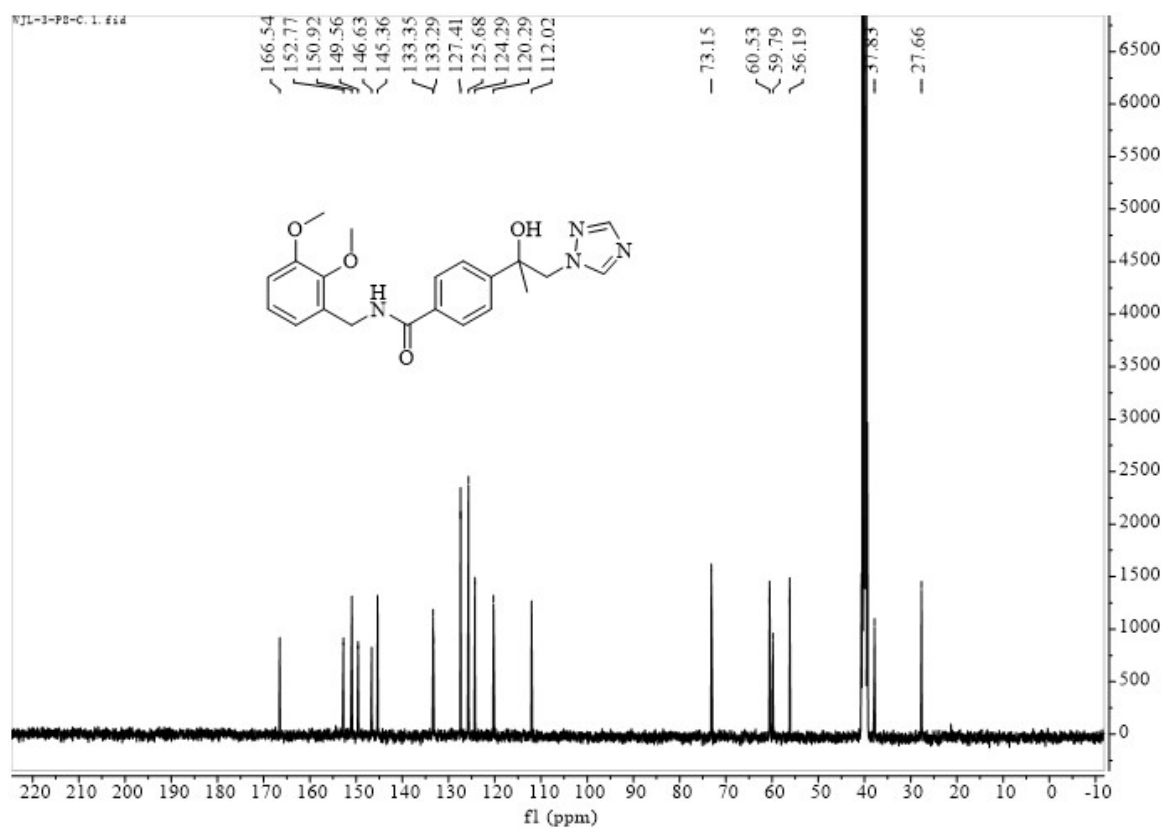
Figure S35. ^1H NMR spectrum of 6aFigure S36. ^{13}C NMR spectrum of 6a

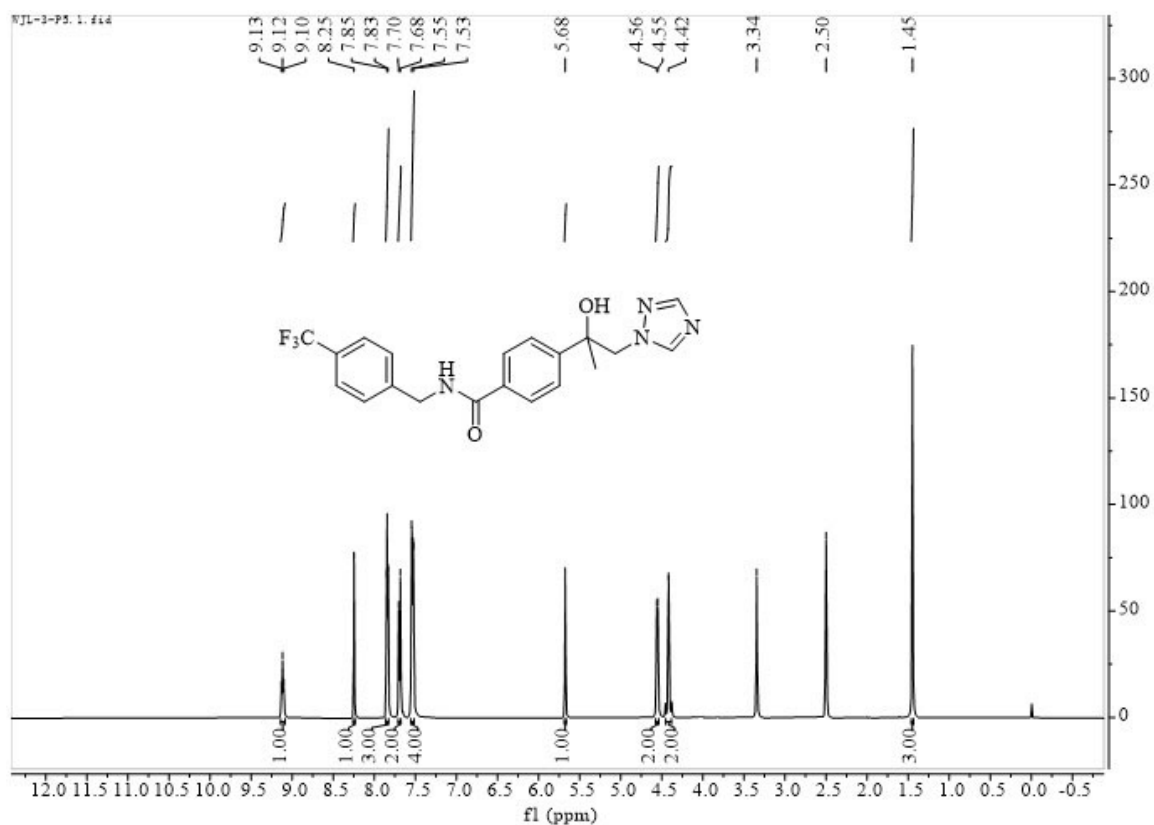
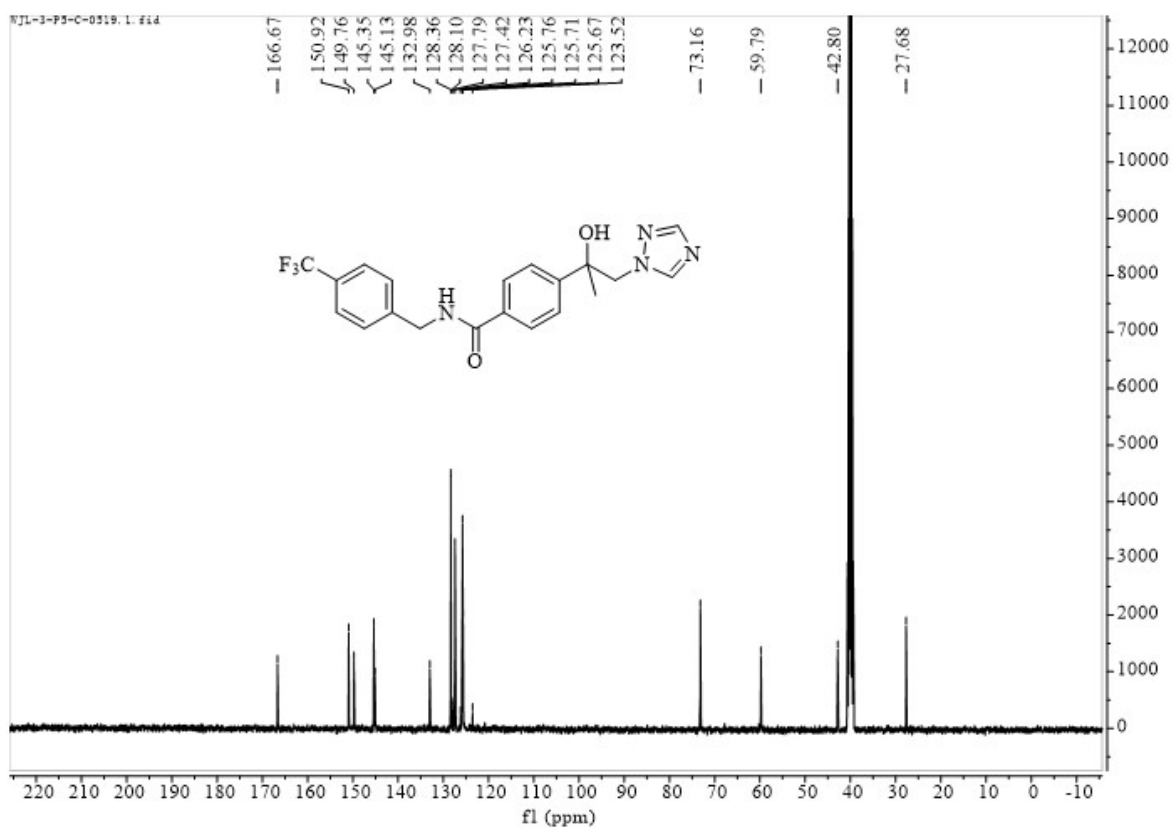


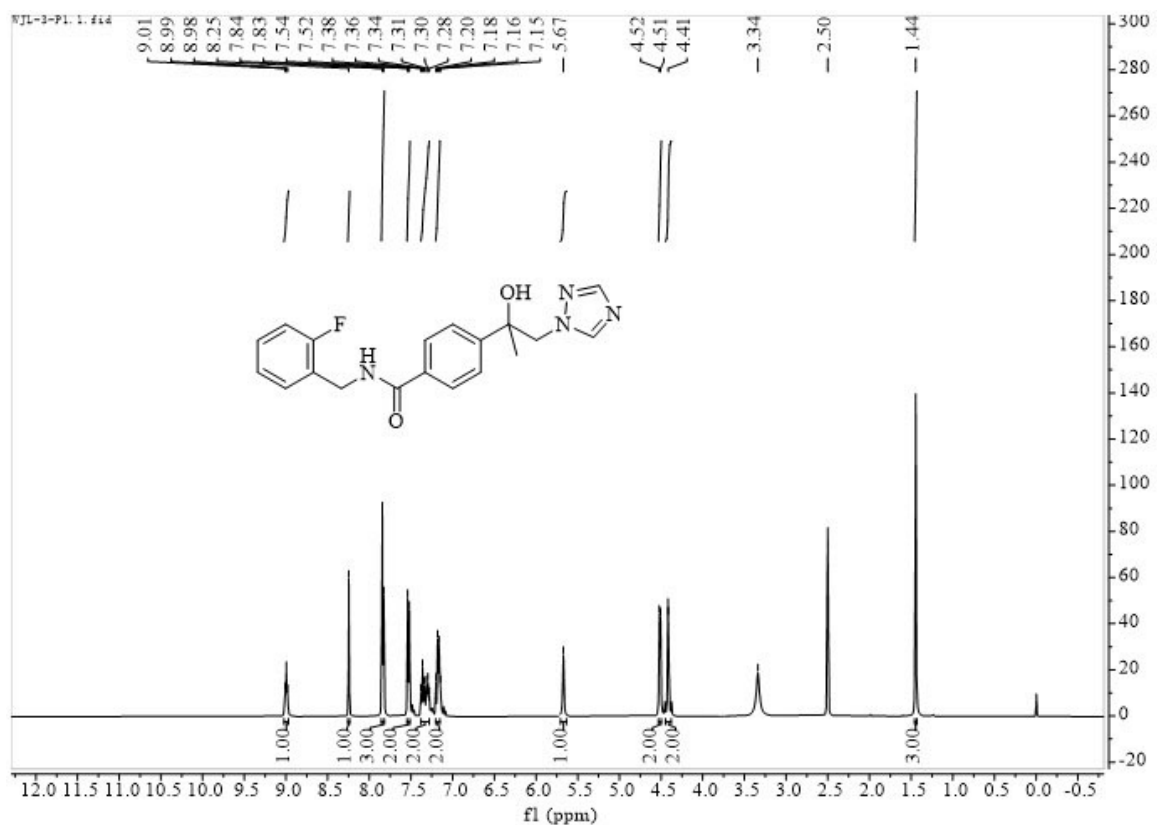
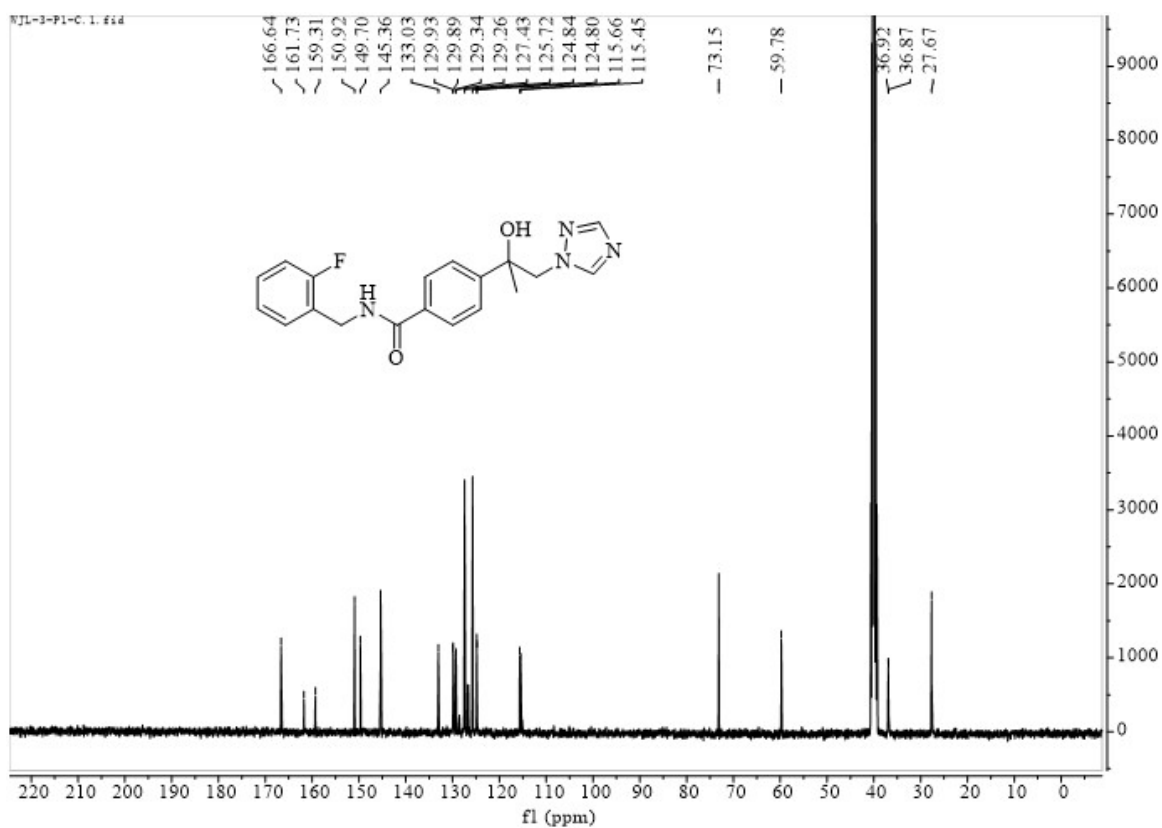
Figure S39. ^1H NMR spectrum of 6cFigure S40. ^{13}C NMR spectrum of 6c

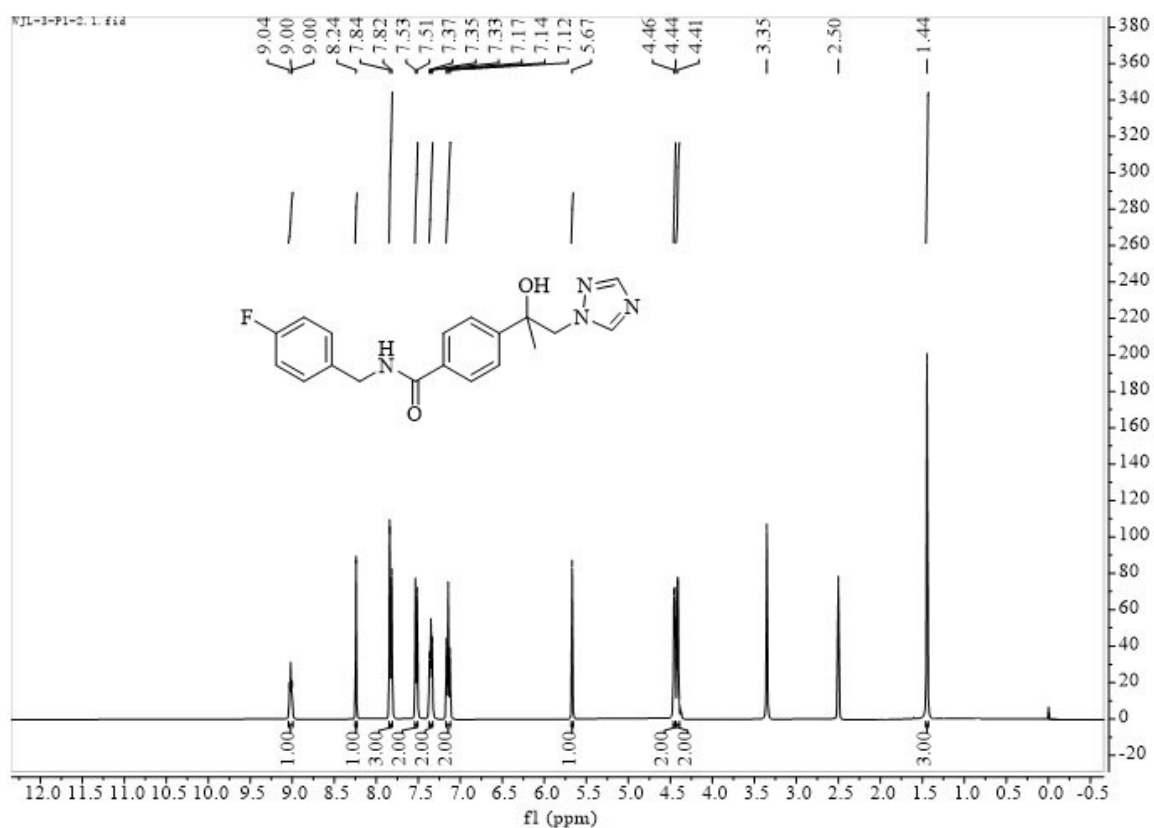
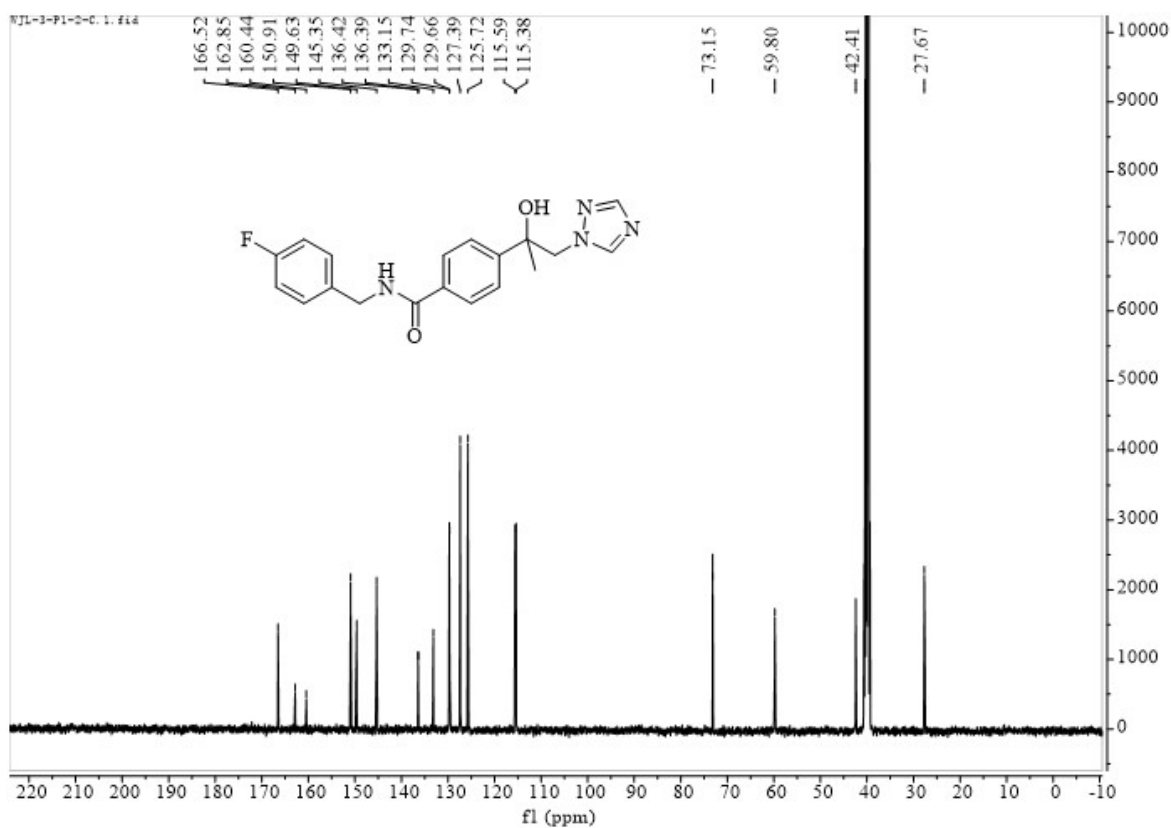
Figure S41. ^1H NMR spectrum of 6dFigure S42. ^{13}C NMR spectrum of 4d

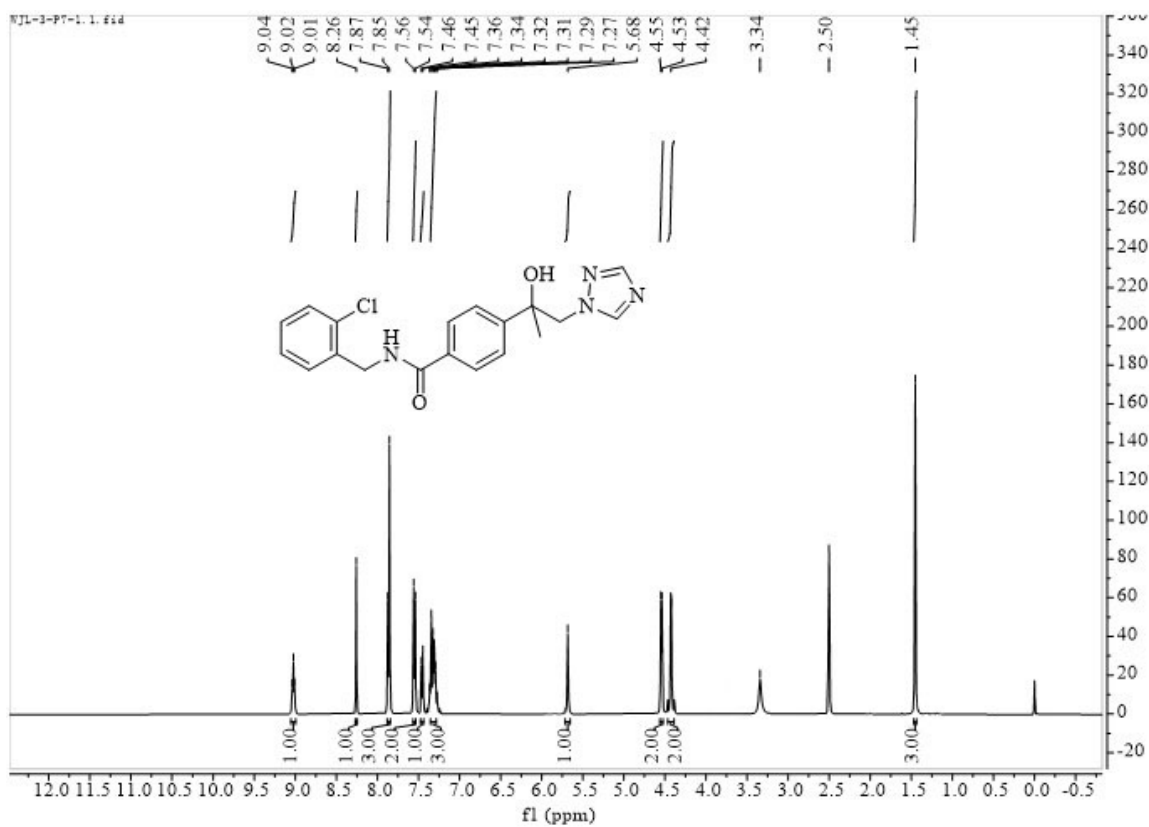
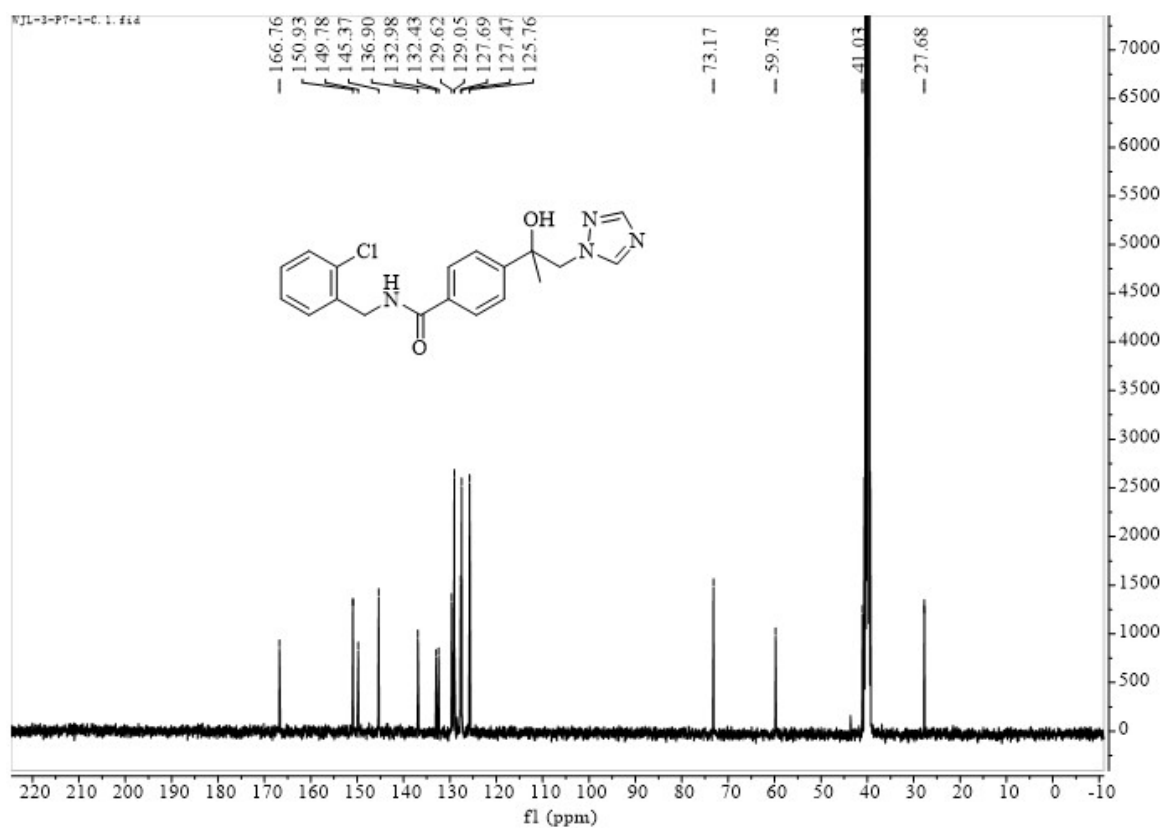
Figure S43. ^1H NMR spectrum of 6eFigure S44. ^{13}C NMR spectrum of 6e

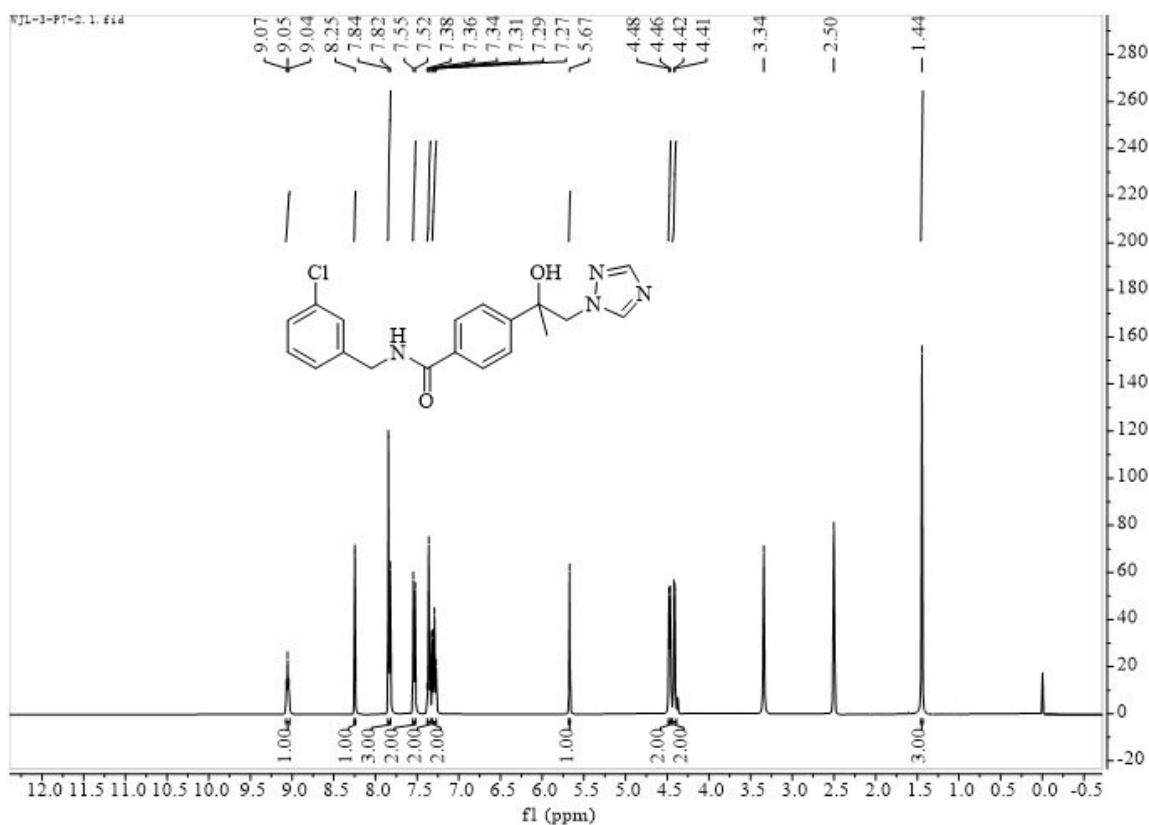
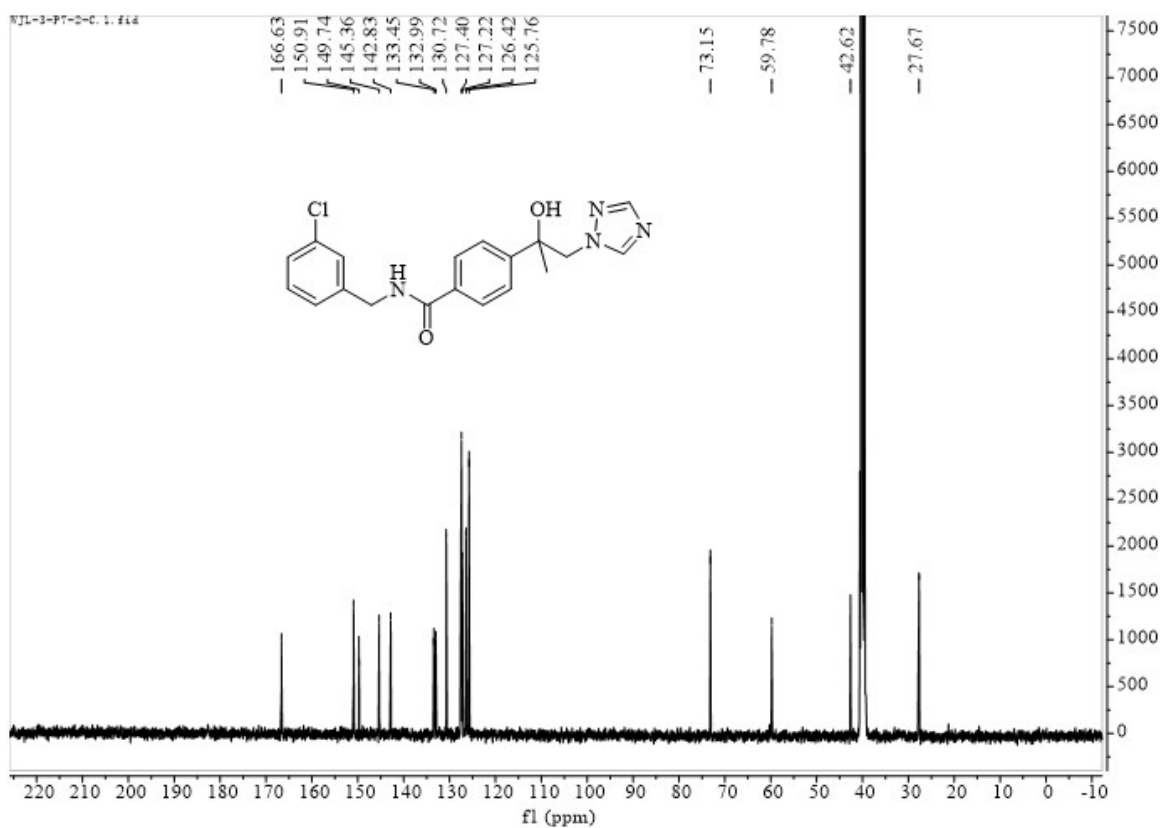
Figure S45. ^1H NMR spectrum of 6fFigure S46. ^{13}C NMR spectrum of 6f

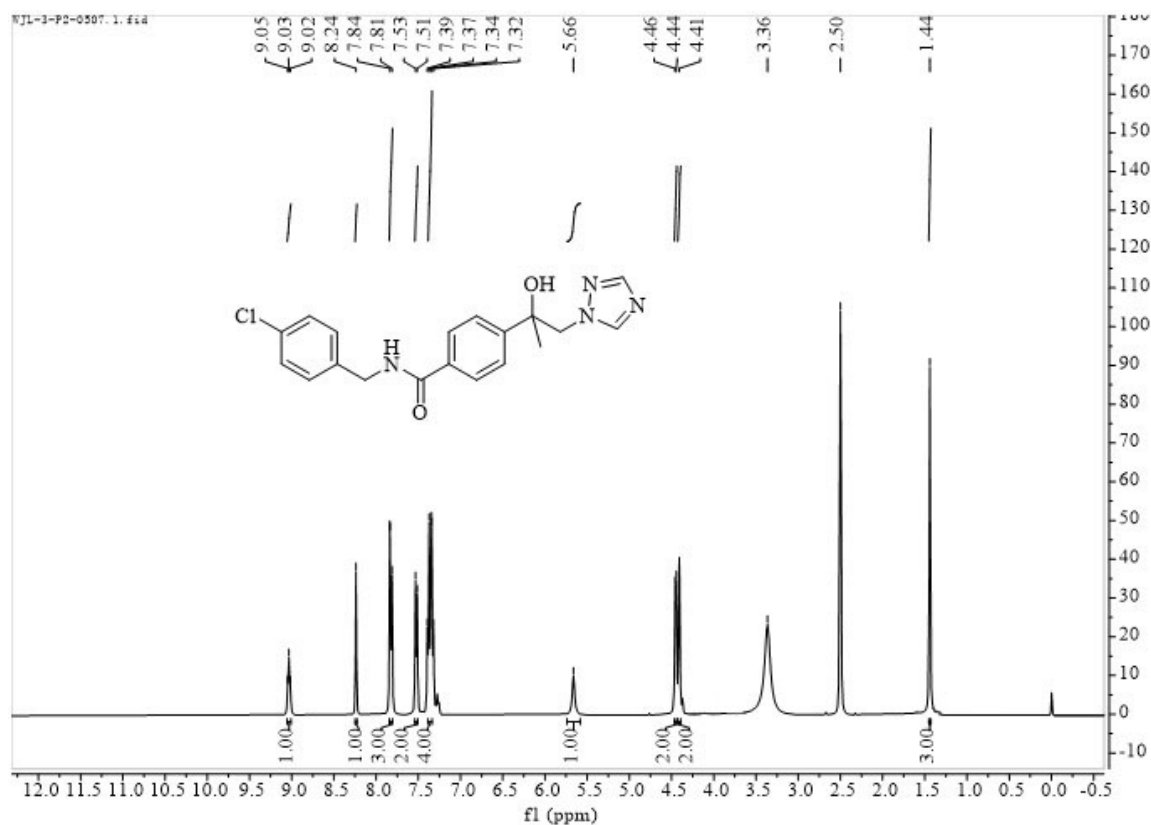
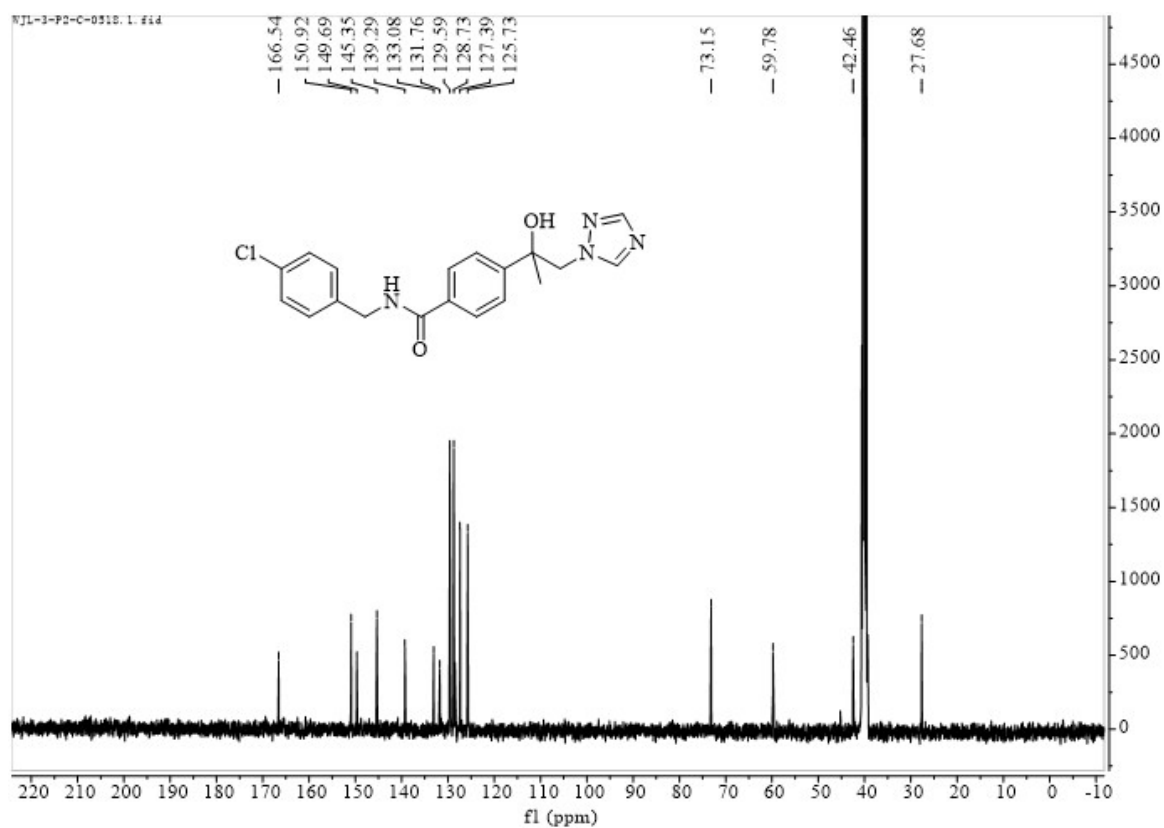
Figure S47. ^1H NMR spectrum of 6gFigure S48. ^{13}C NMR spectrum of 6g

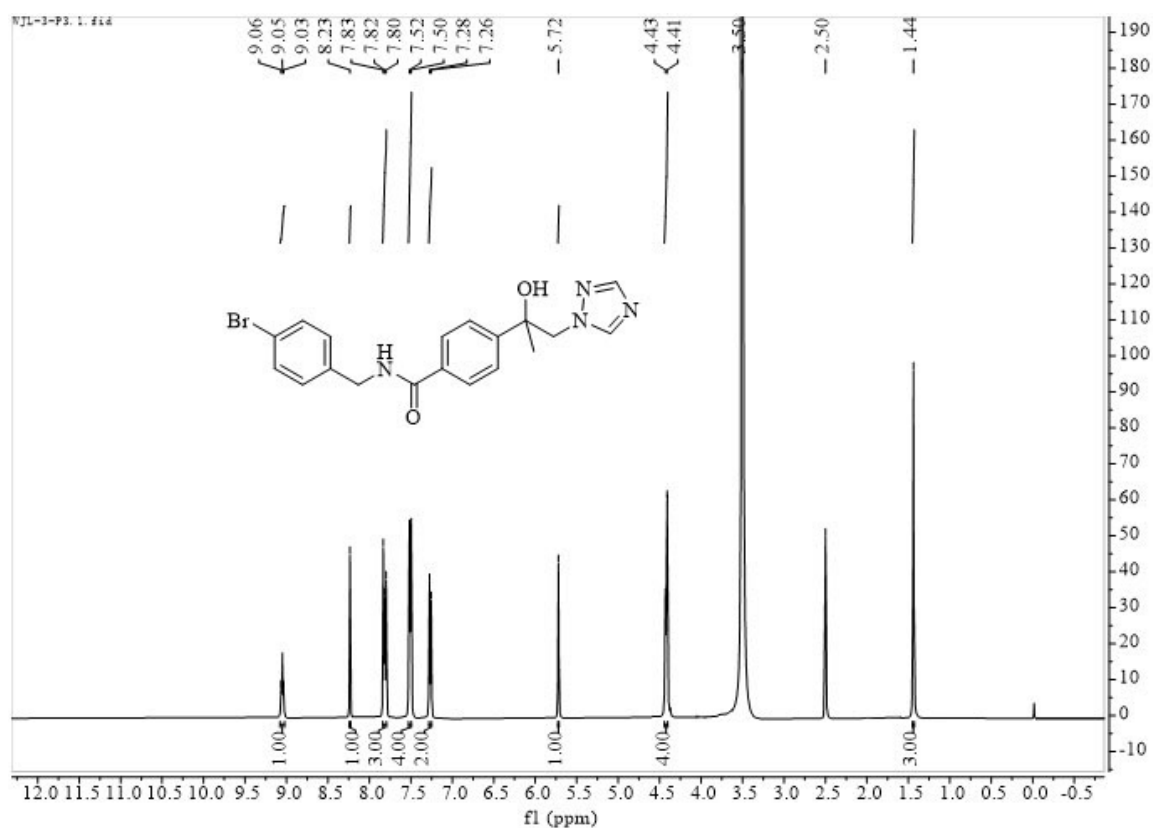
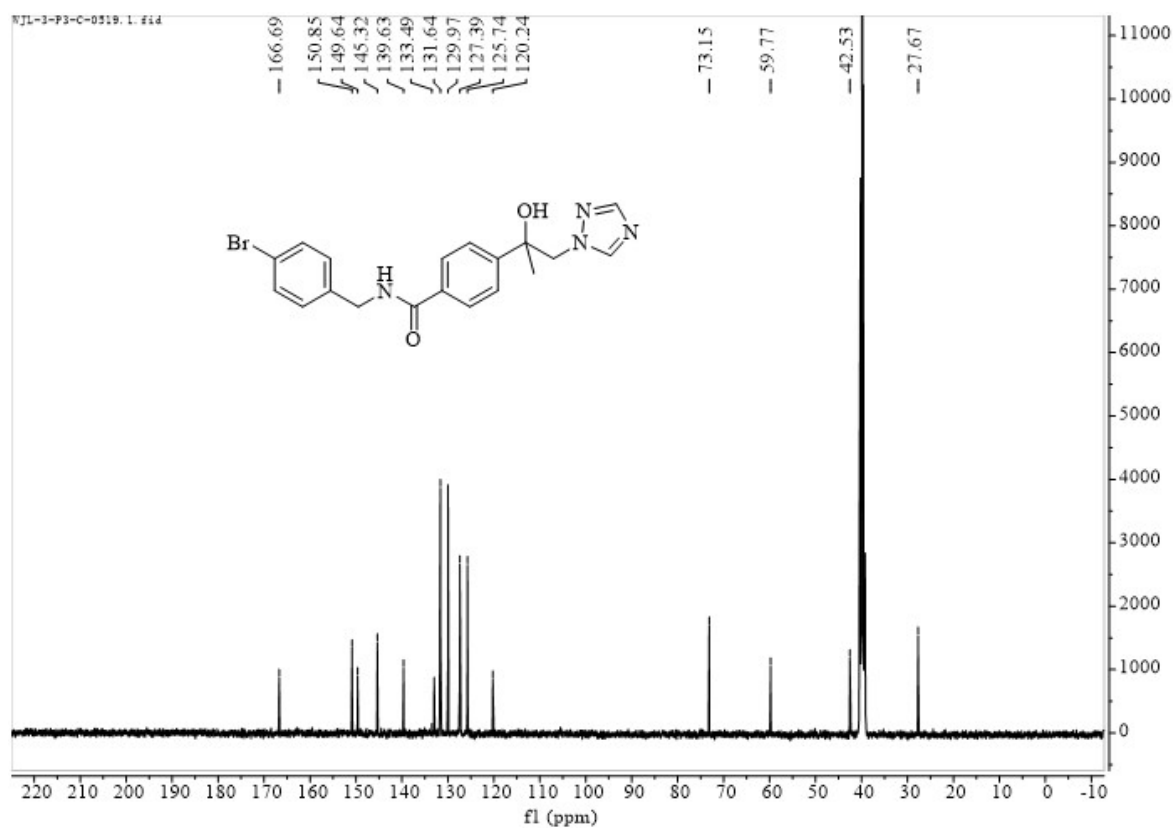
Figure S49. ^1H NMR spectrum of 6hFigure S50. ^{13}C NMR spectrum of 6h

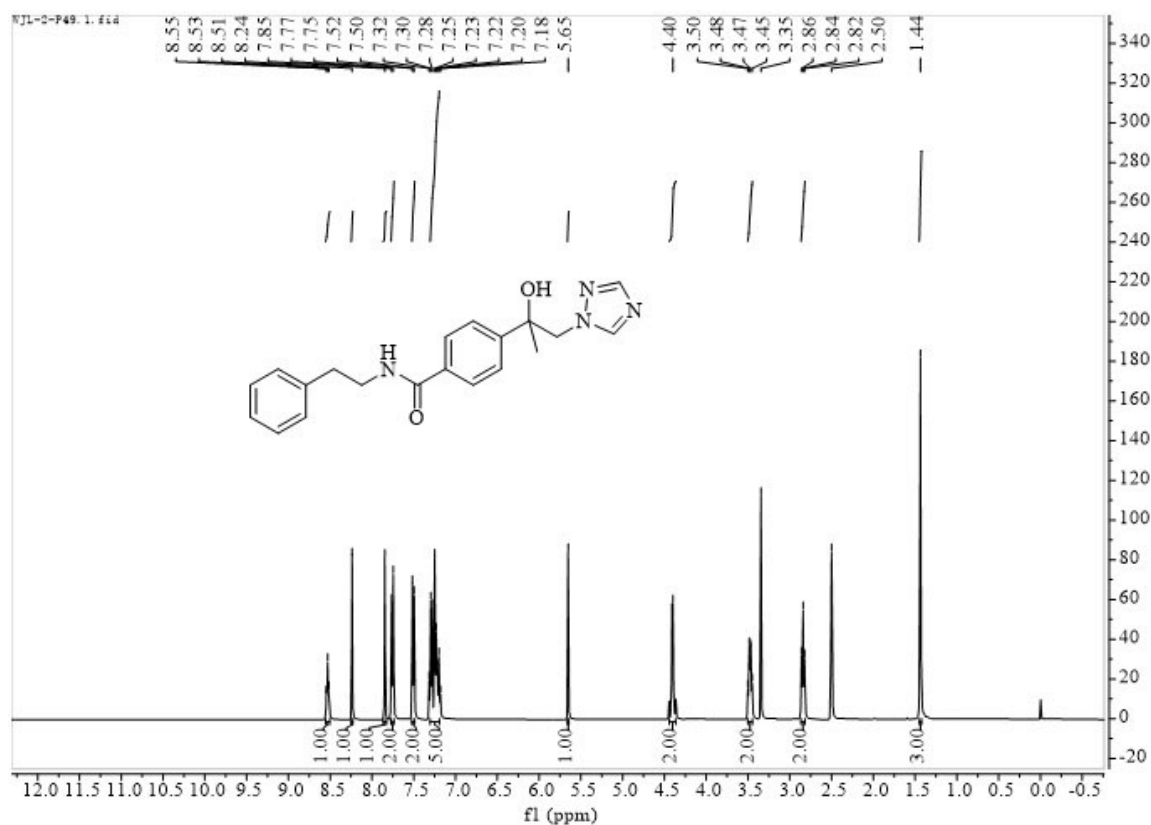
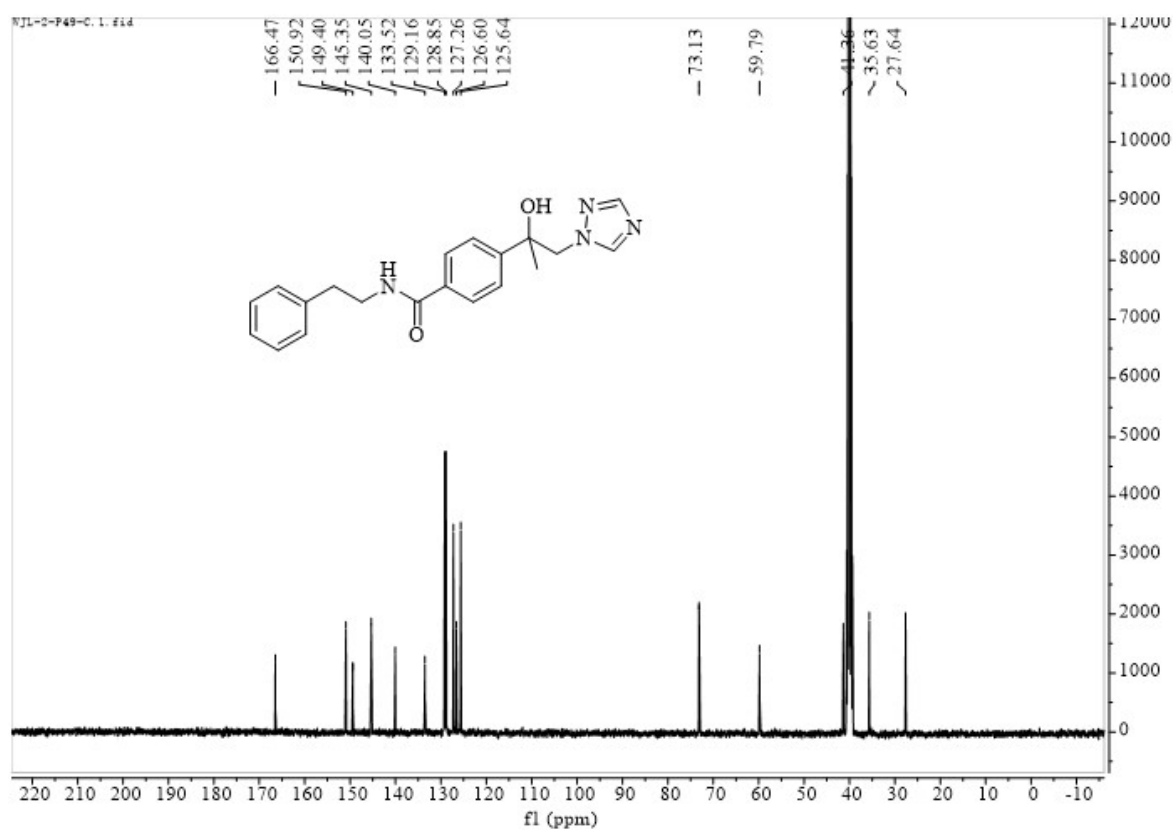
Figure S51. ¹H NMR spectrum of 6iFigure S52. ¹³C NMR spectrum of 6i

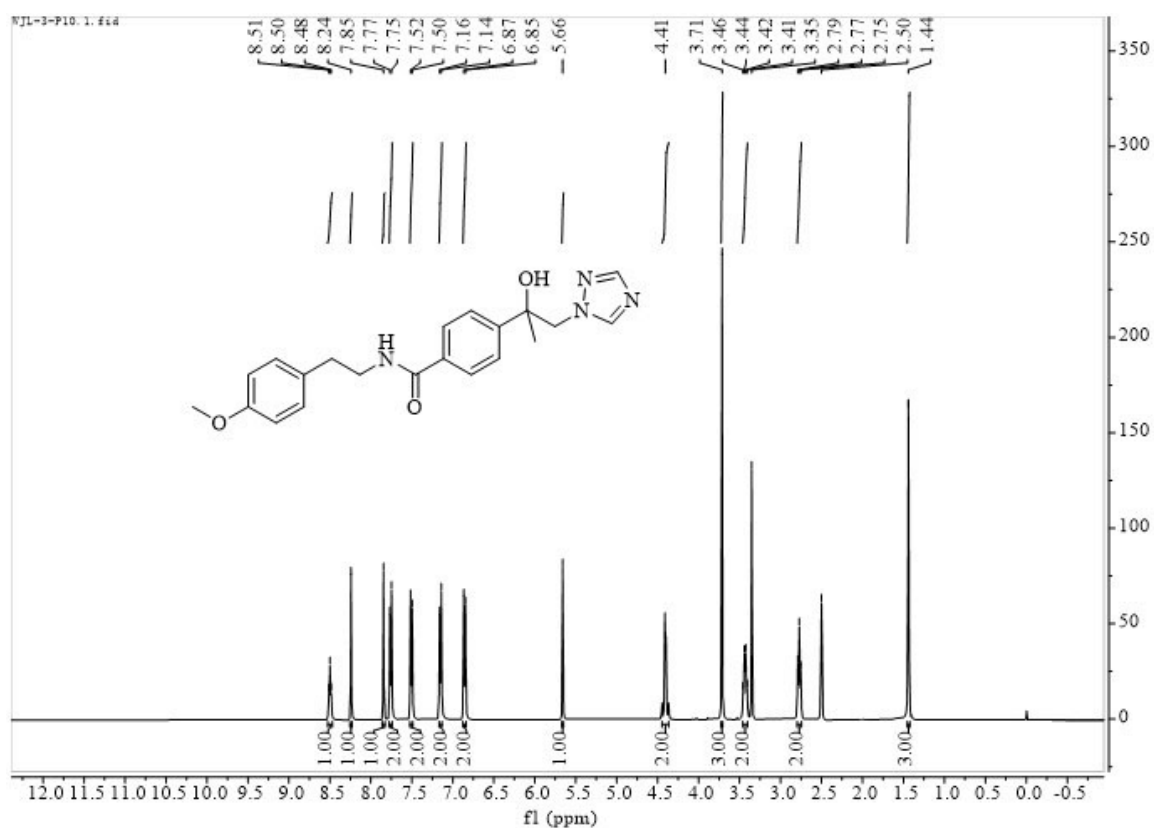
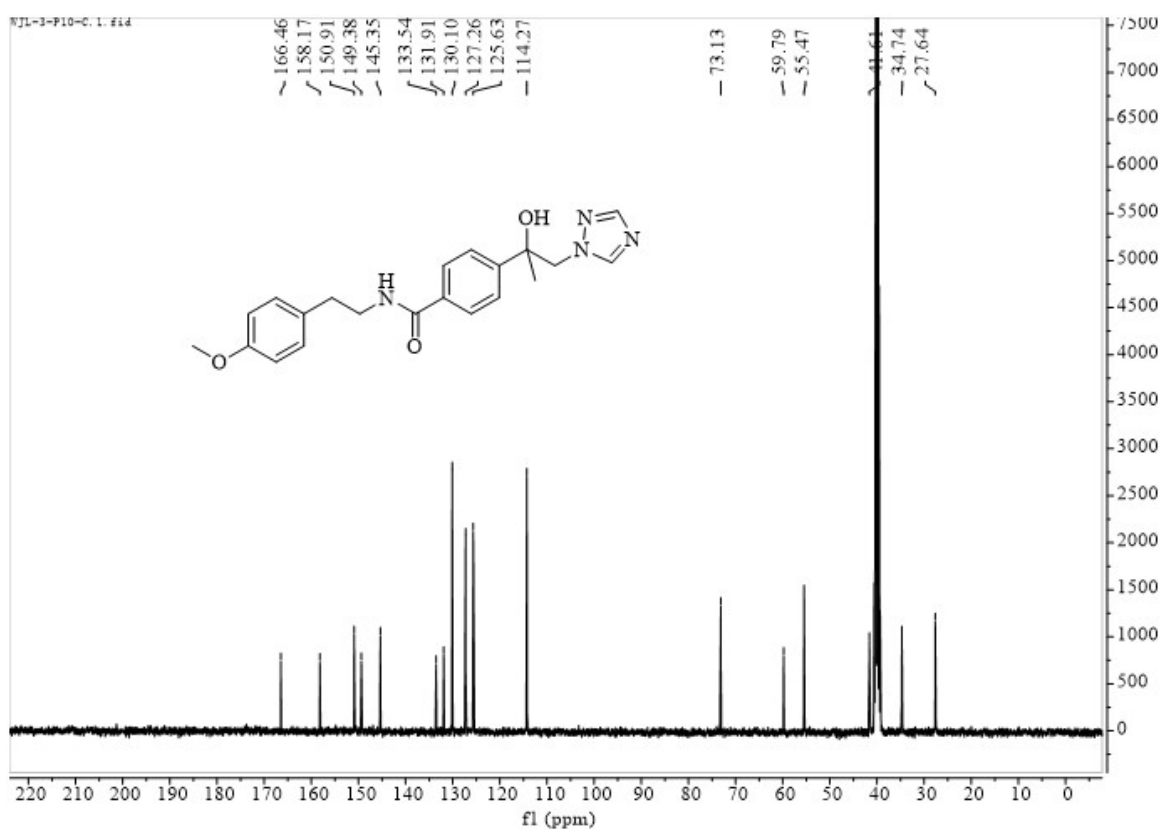
Figure S53. ^1H NMR spectrum of 6jFigure S54. ^{13}C NMR spectrum of 6j

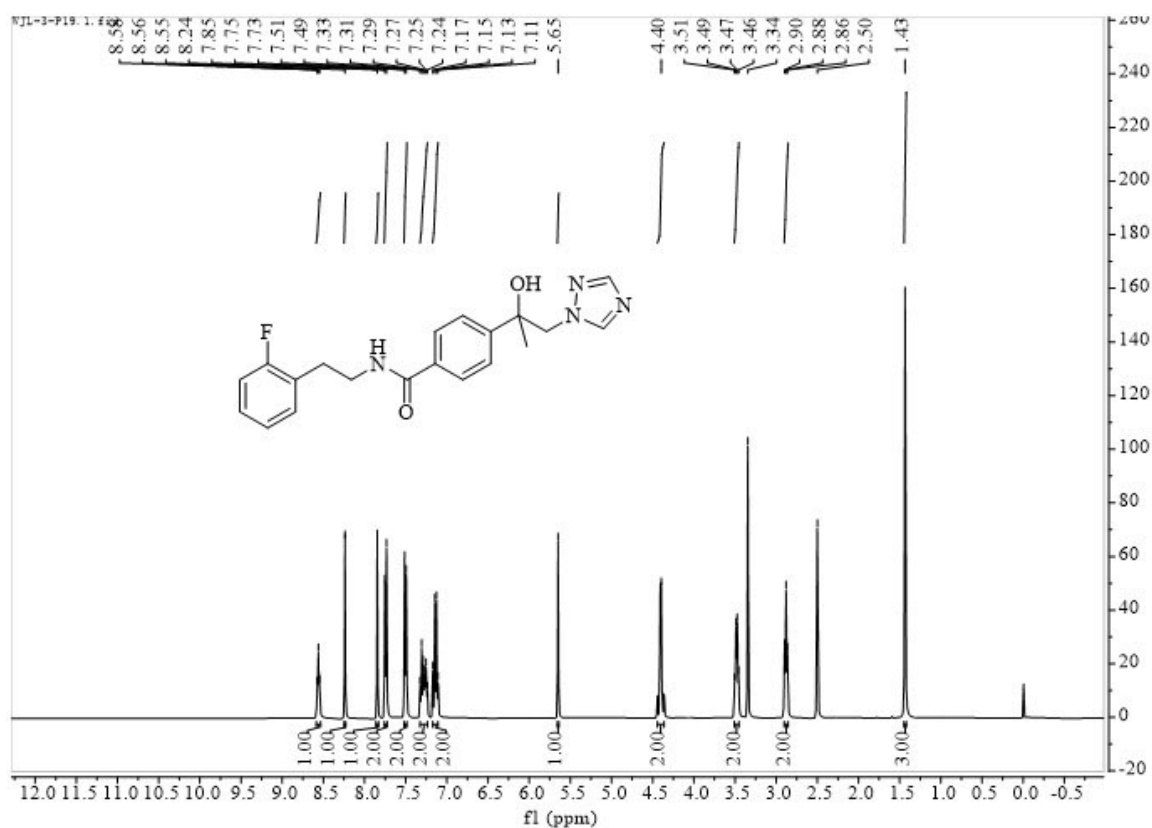
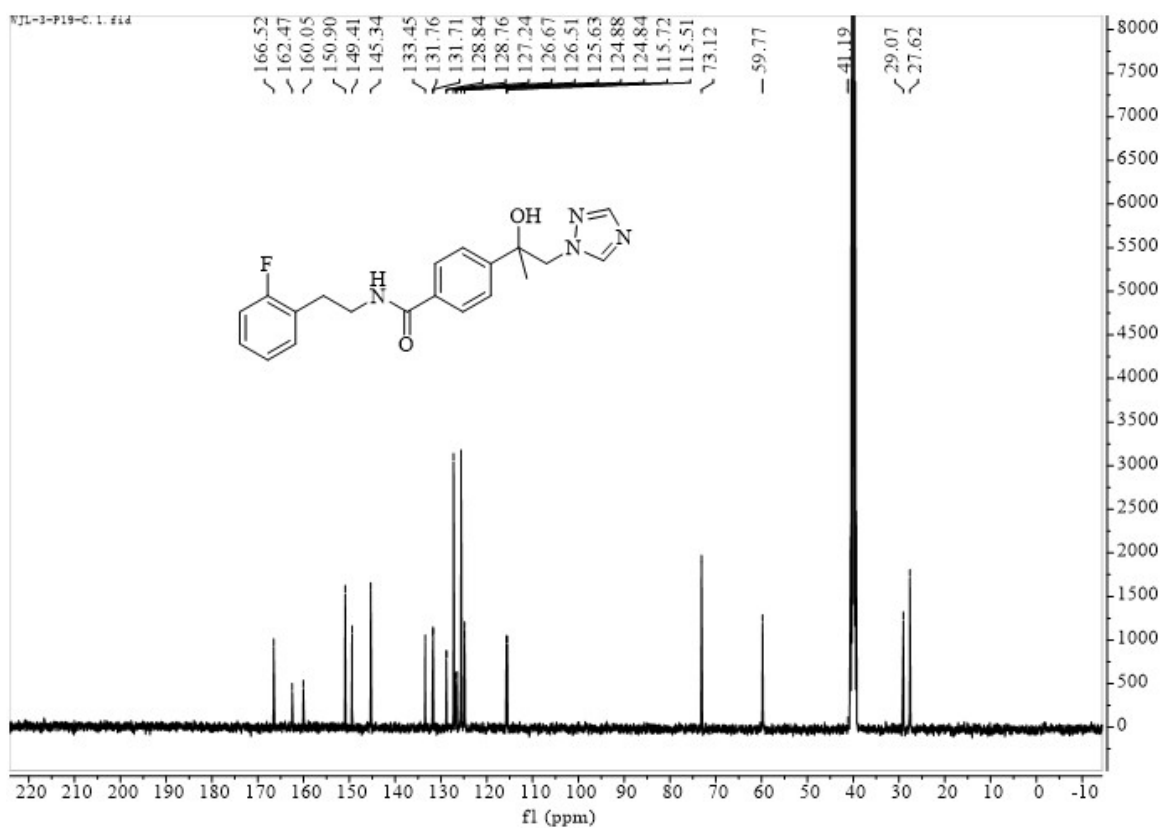
Figure S55. ^1H NMR spectrum of 6kFigure S56. ^{13}C NMR spectrum of 6k

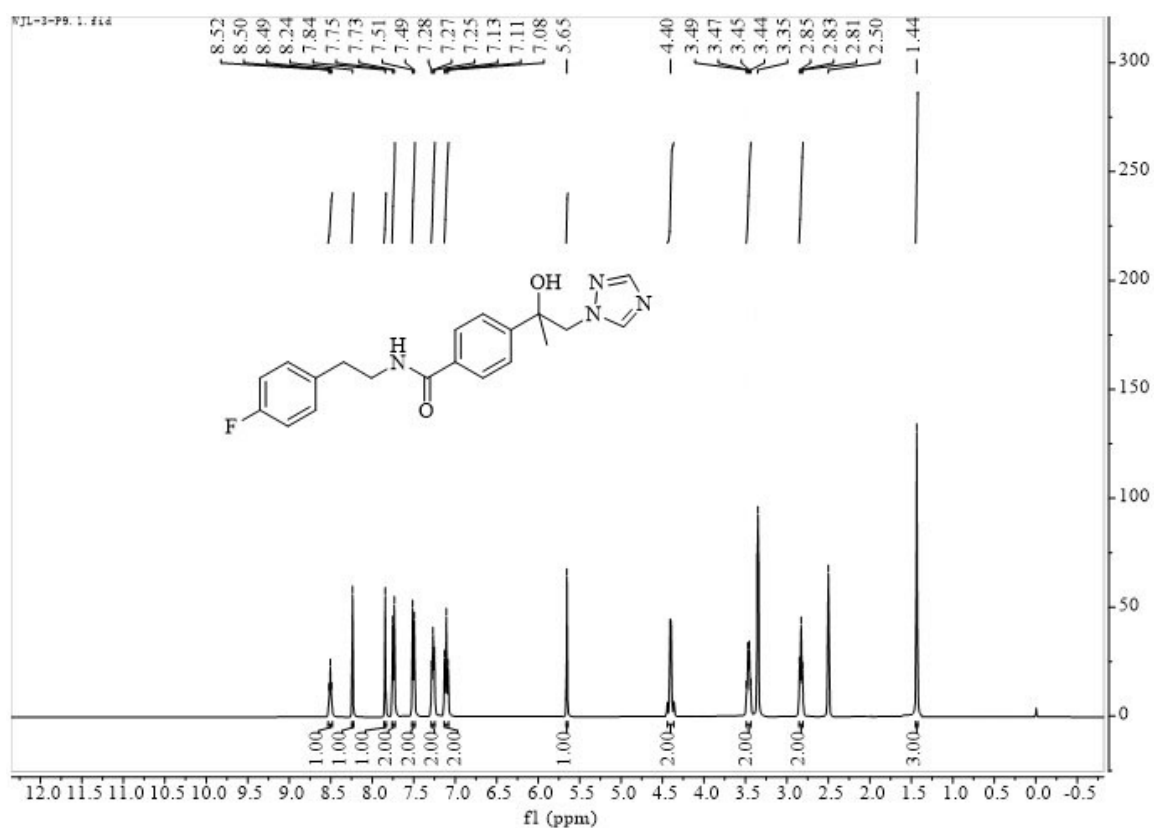
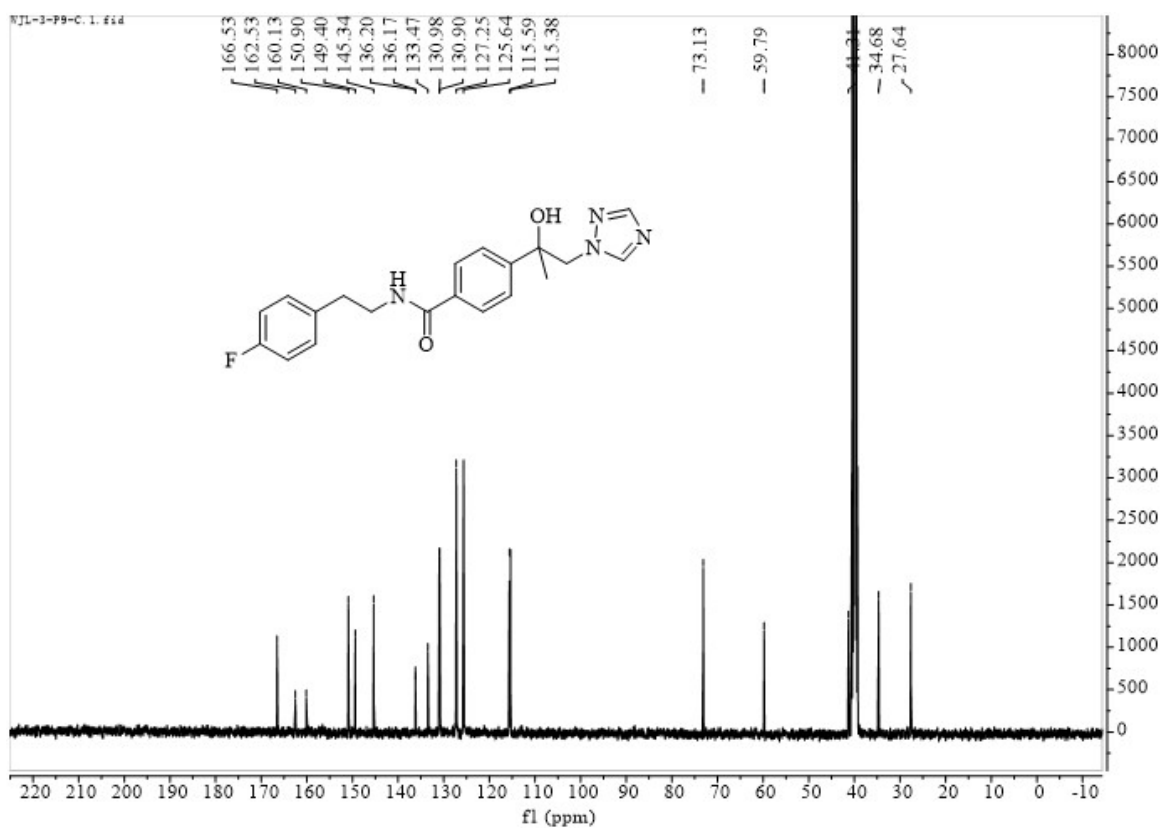
Figure S57. ^1H NMR spectrum of 61Figure S58. ^{13}C NMR spectrum of 61

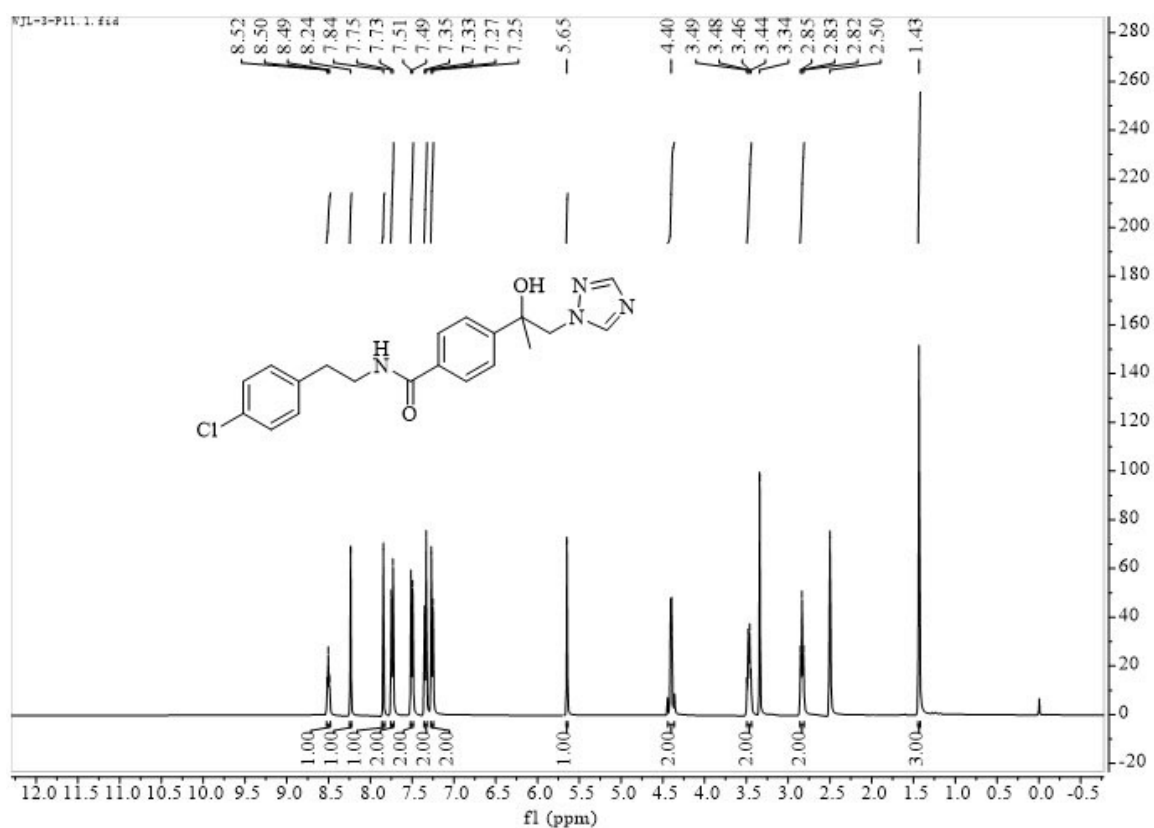
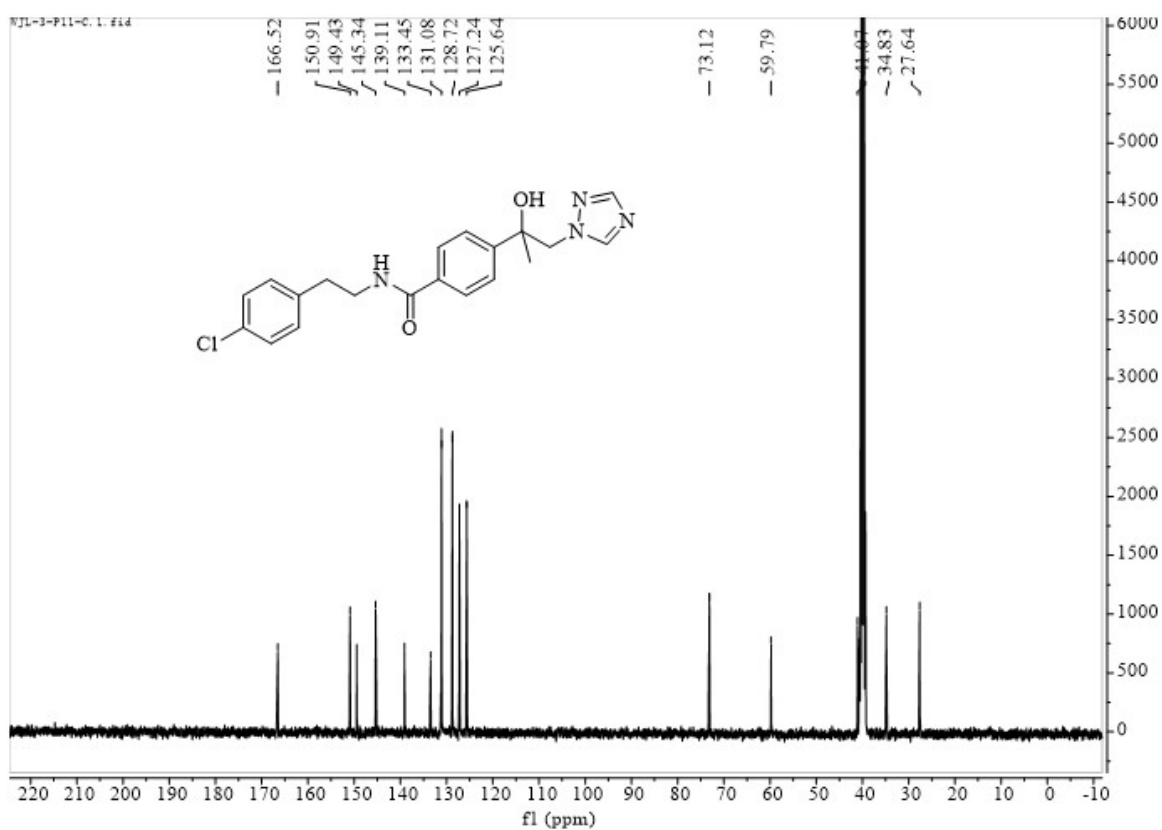
Figure S59. ^1H NMR spectrum of 6mFigure S60. ^{13}C NMR spectrum of 6m

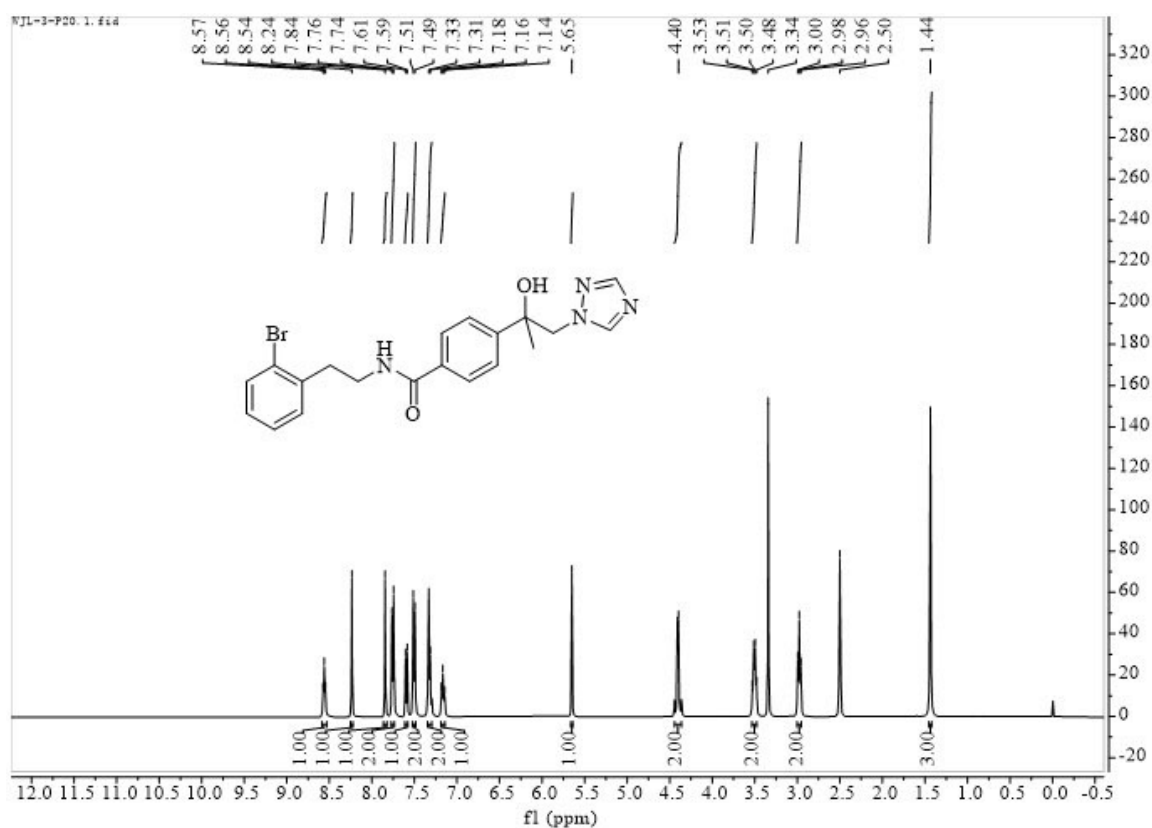
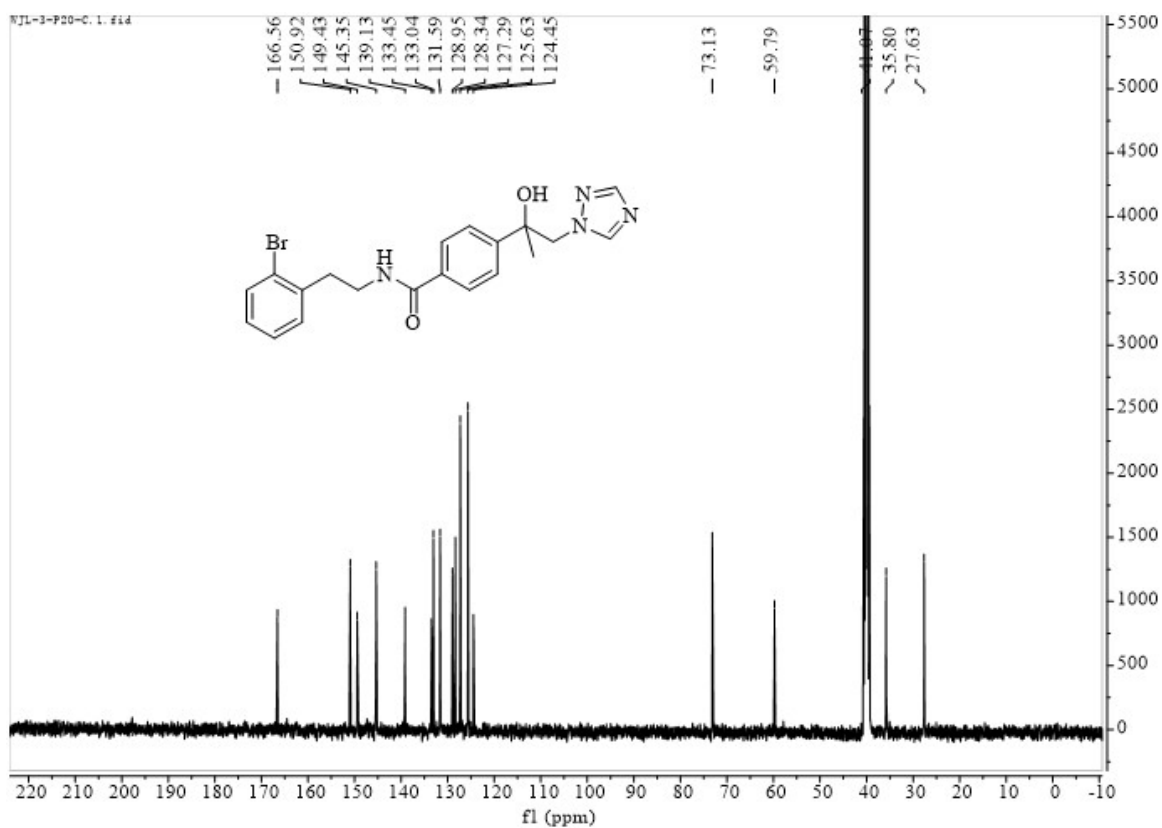
Figure S61. ¹H NMR spectrum of 7aFigure S62. ¹³C NMR spectrum of 7a

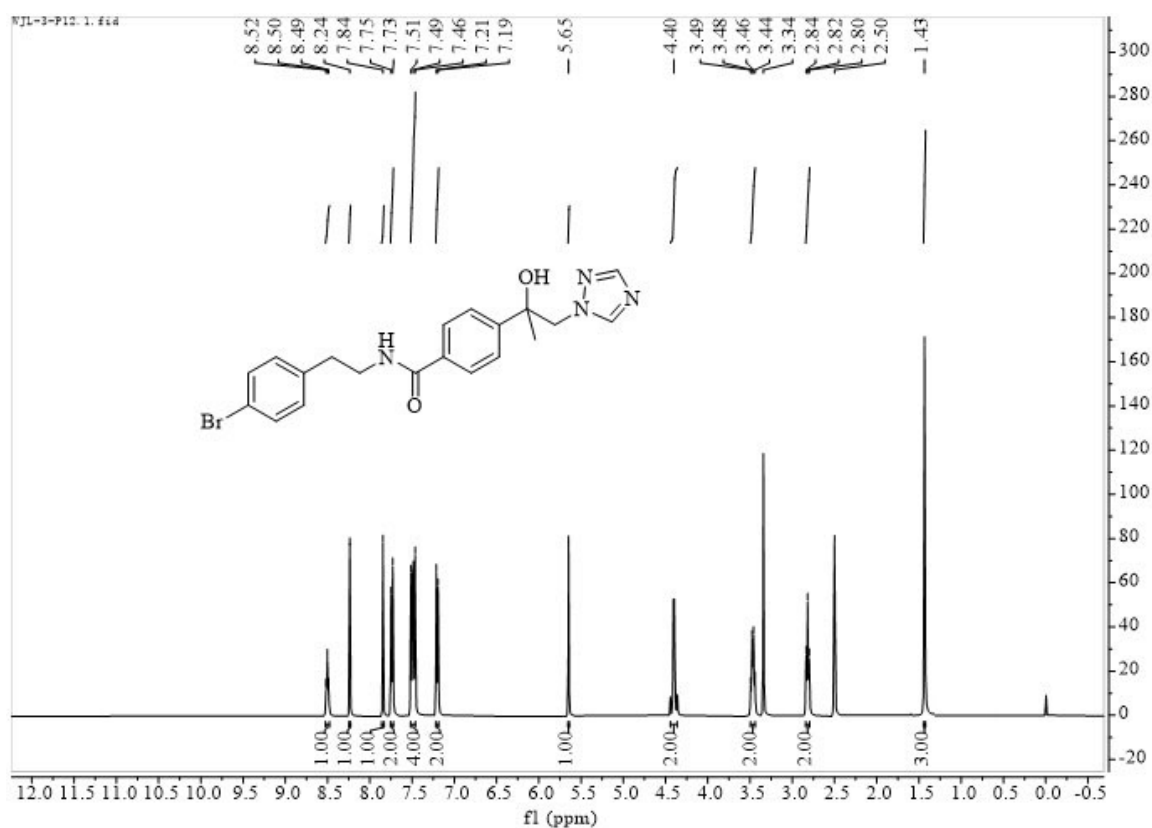
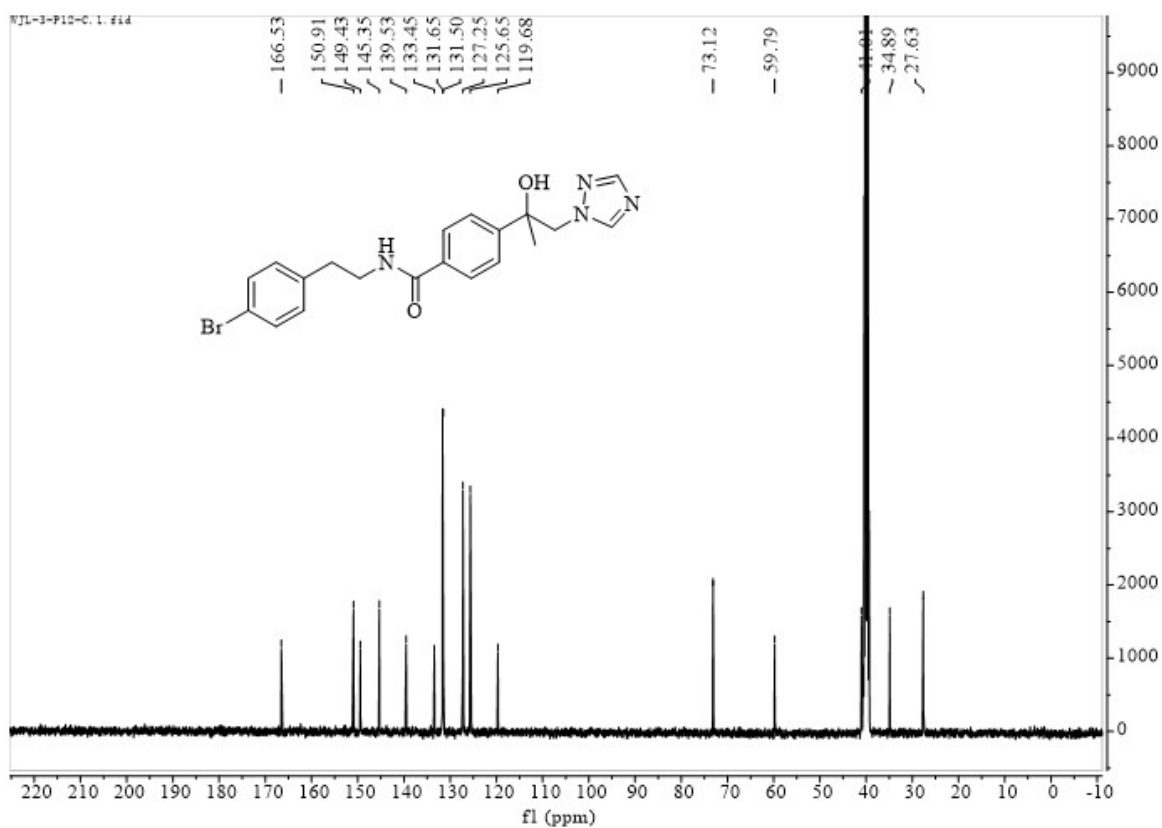
Figure S63. ¹H NMR spectrum of **7b**Figure S64. ¹³C NMR spectrum of **7b**

Figure S65. ¹H NMR spectrum of 7cFigure S66. ¹³C NMR spectrum of 7c

Figure S67. ^1H NMR spectrum of 7dFigure S68. ^{13}C NMR spectrum of 7d

Figure S69. ¹H NMR spectrum of **7e**Figure S70. ¹³C NMR spectrum of **7e**

Figure S71. ^1H NMR spectrum of 7fFigure S72. ^{13}C NMR spectrum of 7f

Figure S73. ^1H NMR spectrum of 7gFigure S74. ^{13}C NMR spectrum of 7g