

# Macrophage Cell Membrane Coating on Piperine-Loaded MIL-100(Fe) Nanoparticles for Breast Cancer Treatment

Christian Rafael Quijia<sup>1</sup>, Geovana Navegante<sup>2</sup>, Rafael Miguel Sábio<sup>1</sup>, Valeria Valente<sup>2</sup>, Alberto Ocaña<sup>3</sup>, Carlos Alonso-Moreno<sup>4</sup>, Regina Célia Galvão Frem<sup>5</sup>, and Marlus Chorilli<sup>1\*</sup>

<sup>1</sup> Department of Drugs and Medicines, School of Pharmaceutical Sciences of São Paulo State University (UNESP), Rodovia Araraquara Jau, Km 01 – s/n – Campos Ville, 14800-903 Araraquara, Sao Paulo, Brazil. E-mail: christianqui@hotmail.com; rafael.m.sabio@unesp.br; marlus.chorilli@unesp.br

<sup>2</sup> Laboratory of Molecular and Cell Biology, School of Pharmaceutical Sciences, Department of Clinical Analysis, São Paulo State University (Unesp), Araraquara, Brazil. E-mail: geonavegante@gmail.com; valenteval@gmail.com

<sup>3</sup> Experimental Therapeutics Unit, Hospital Clínico San Carlos, IdISSC, Fundación Jiménez Díaz, START, 28040 Madrid, Spain. E-mail: alberto.ocana@salud.madrid.org

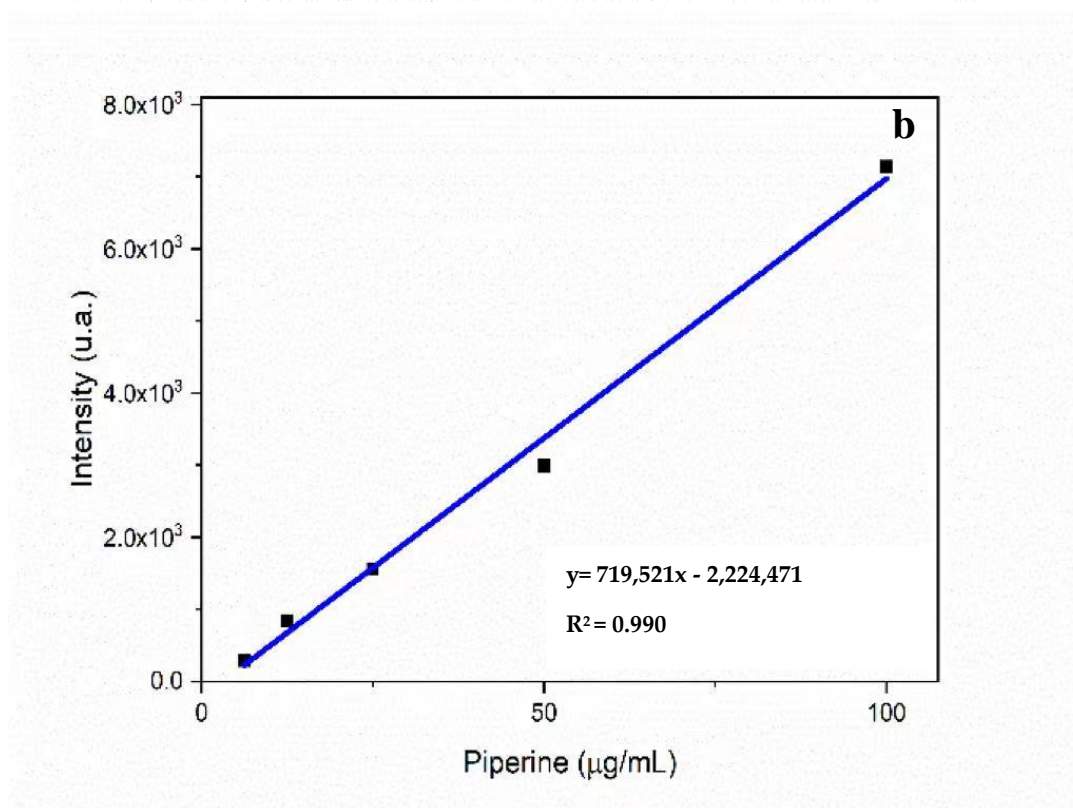
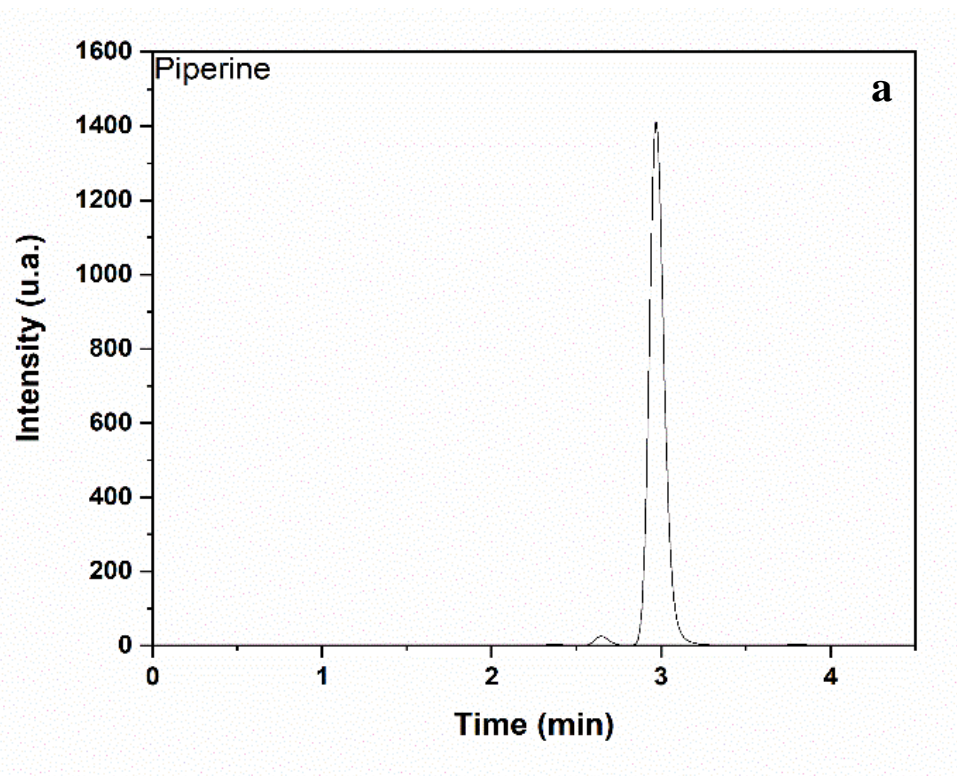
<sup>4</sup> Unidad NanoDrug, Facultad de Farmacia, Universidad de Castilla-La Mancha, 02008 Albacete, Spain. E-mail: Carlos.AMoreno@uclm.es

<sup>5</sup> Institute of Chemistry, São Paulo State University (UNESP), Prof. Francisco Degni 55, 14800-060 Araraquara, Sao Paulo, Brazil. E-mail: rcgrem@gmail.com

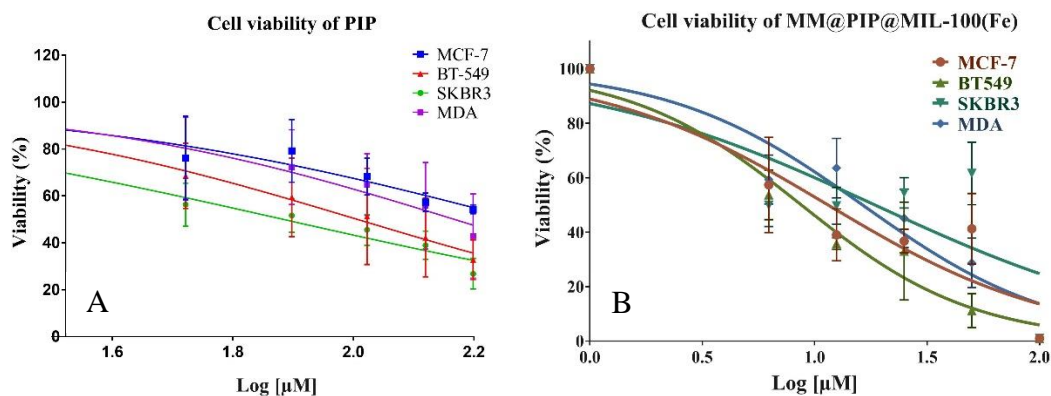
\*Corresponding author at Department of Drugs and Medicines, School of Pharmaceutical Sciences of São Paulo State University (UNESP), Rodovia Araraquara Jau, Km 01 – s/n – Campos Ville, 14800-903 Araraquara, Sao Paulo, Brazil. E-mail address: marlus.chorilli@unesp.br

## HPLC assay for PIP

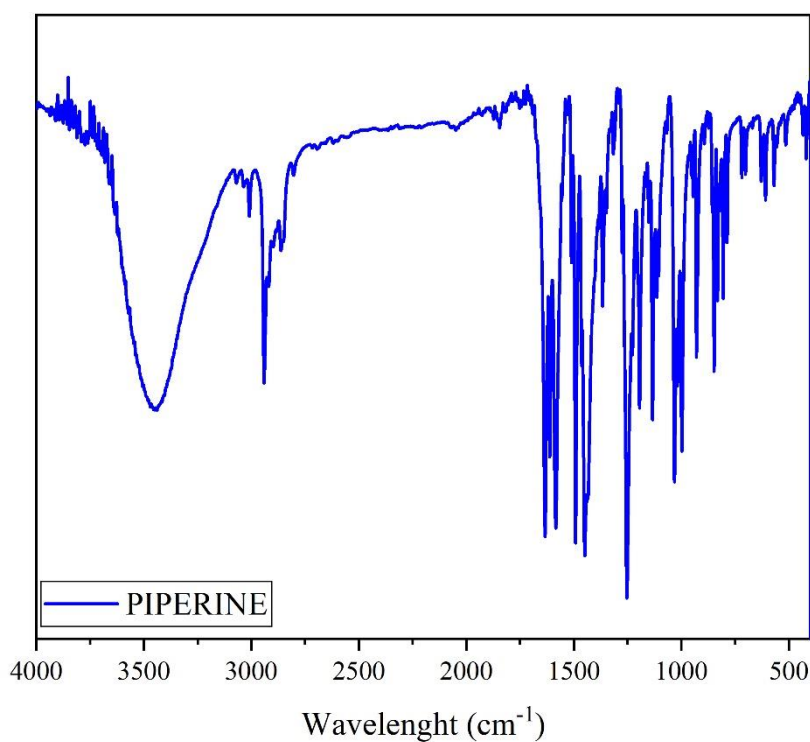
The calibration curve was linear by analyzing a series of PIP concentrations in ethanol from 6.25 to 100  $\mu\text{g mL}^{-1}$  with a correlation coefficient of  $R^2=0.990$  (**Figure S1**). The limit of detection (LOD) and limit of quantification (LOQ) were found to be 0.25 and 0.77  $\mu\text{g mL}^{-1}$ , respectively. Standard samples were prepared and injected in triplicates on three successive days.



**Figure S1.** (a) Chromatogram of PIP (concentration = 6.25 to 100  $\mu\text{g mL}^{-1}$ ) in mobile phase (methanol/water (75:25)) and (b) calibration curve of PIP quantified by HPLC.



**Figure S2.** Cytotoxic activity of piperine (A) and MM@PIP@MIL-100 (Fe)- (B). The results represented cell viability and refer to the averages of three independent experiments (mean  $\pm$  standard deviation). Cells were treated with 6.25 to 100  $\mu\text{M}$  and incubated for 48 h. Viability was analyzed by nonlinear regression in GraphPad Prism version 7.0 (GraphPad Software, San Diego, CA, USA).



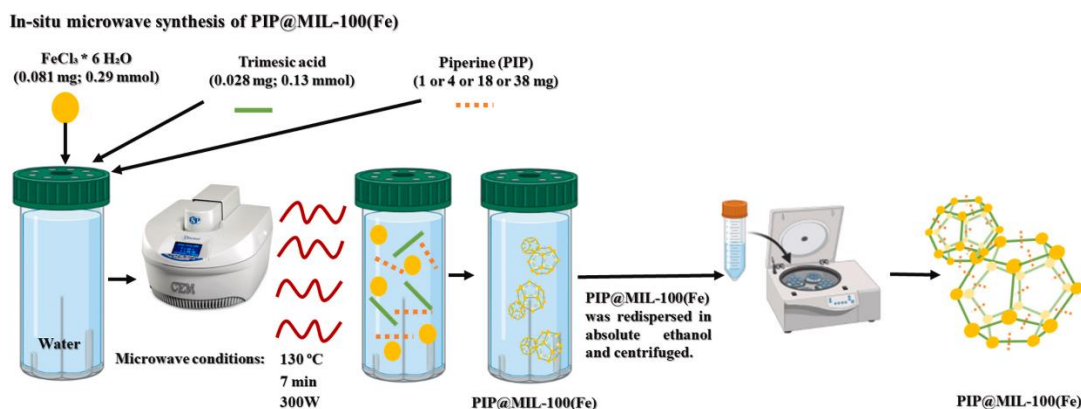
**Figure S3.** Fourier-transform infrared (FT-IR) spectrum analysis of PIP (blue line).

### Synthesis of MIL-100(Fe)

The synthesis of MIL-100(Fe) was performed using a previously established method. [1] In this method, iron (III) chloride hexahydrate (0.081 g; 0.29 mmol) and BTC (0.028 g; 0.13 mmol) were dissolved in distilled water (1 mL). The resulting mixture was heated to 130 °C for 30 seconds and maintained at this temperature for 7 minutes (300 W) using a Mars-5 microwave reactor (CEM, US) with a maximum power output of 300 W and a frequency of 60 Hz. The MIL-100(Fe) product was obtained by centrifugation at 5600g for 8 minutes. To activate the solid, it was redispersed in absolute ethanol (1 mL) and subjected to centrifugation at 5600 g for 8 minutes. This process was repeated for a total of 4 cycles. The resulting orange solid was recovered with a yield of 98% (40 mg). Finally, the nanostructure was stored in pure ethanol at 4 °C for preservation.

### PIP@MIL-100(Fe) in situ

We conducted the microwave synthesis of PIP@MIL-100(Fe) on-site following a previously described method, but with a variation. In this case, we introduced different concentrations of PIP (1, 4, 18, and 38 mg) into the initial synthesis solution. After the synthesis, the resulting PIP@MIL-100(Fe) was separated by centrifugation at 5600g for 8 minutes, and the sample with the most favorable chemical properties was chosen for further investigation. To continue the research, the recovered PIP@MIL-100(Fe) was then dispersed again in absolute ethanol (1 mL), subjected to centrifugation at 5600g for 8 minutes, and this process was repeated four times. As a result, an orange solid (40 mg, 96% yield) was obtained. To maintain the integrity of the nanostructure, it was stored in absolute ethanol at a temperature of 4 °C until it was ready to be used (see Fig. S4).



**Figure S4.** We performed the in-situ microwave synthesis of PIP@MIL-100(Fe) by dissolving BTC (0.028 mg; 0.13 mmol), iron (III) chloride hexahydrate (0.081 mg; 0.29 mmol), and various concentrations of PIP (1; 4; 18; and 38 mg) in 1 mL of distilled water. The reaction mixture was then subjected to microwave irradiation at 300 W, heating it to 130 °C for 30 seconds, and maintaining this temperature for 7 minutes. Next, the obtained PIP@MIL-100(Fe) was dispersed again in 1 mL of absolute ethanol, followed by centrifugation at 5600 g for 8 minutes. This centrifugation and redispersion process was repeated four times. As a result, an orange solid (40 mg, 96% yield) was recovered.

### REFERENCE BIBLIOGRAPHIC

- [1] C. R. Quijia *et al.*, "In situ synthesis of piperine-loaded MIL-100 (Fe) in microwave for breast cancer treatment," vol. 75, p. 103718, 2022.