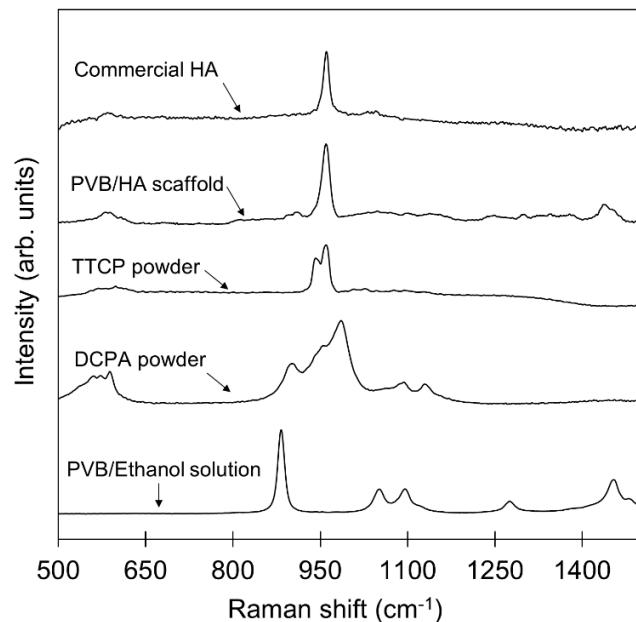
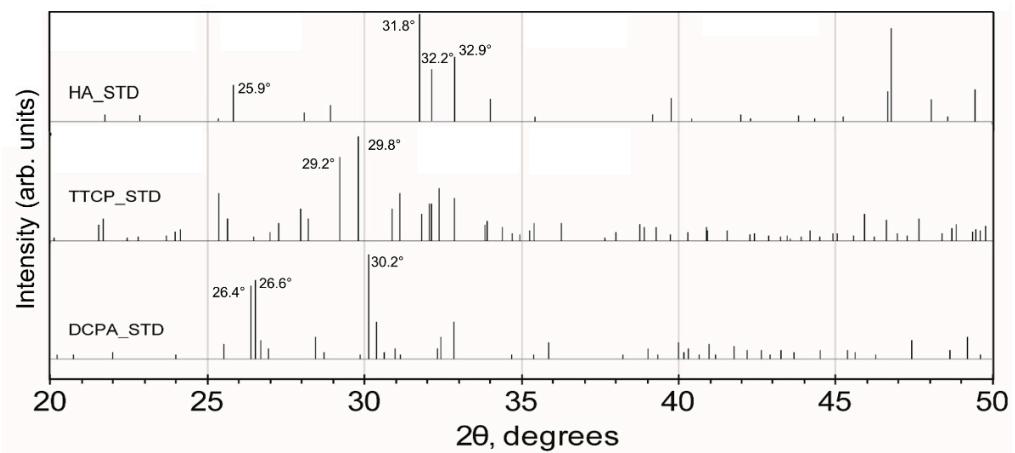


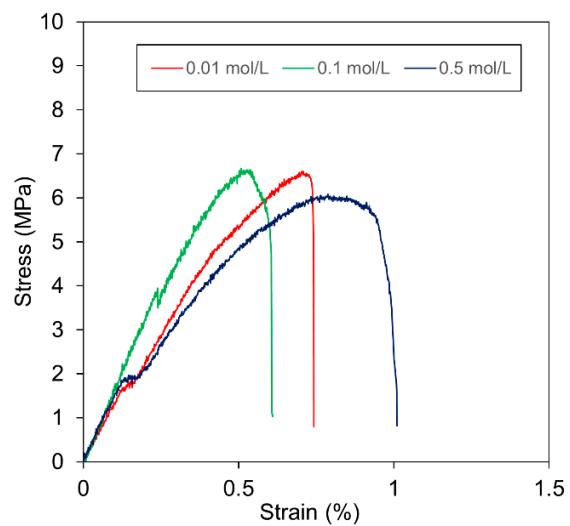
## Supplementary Materials



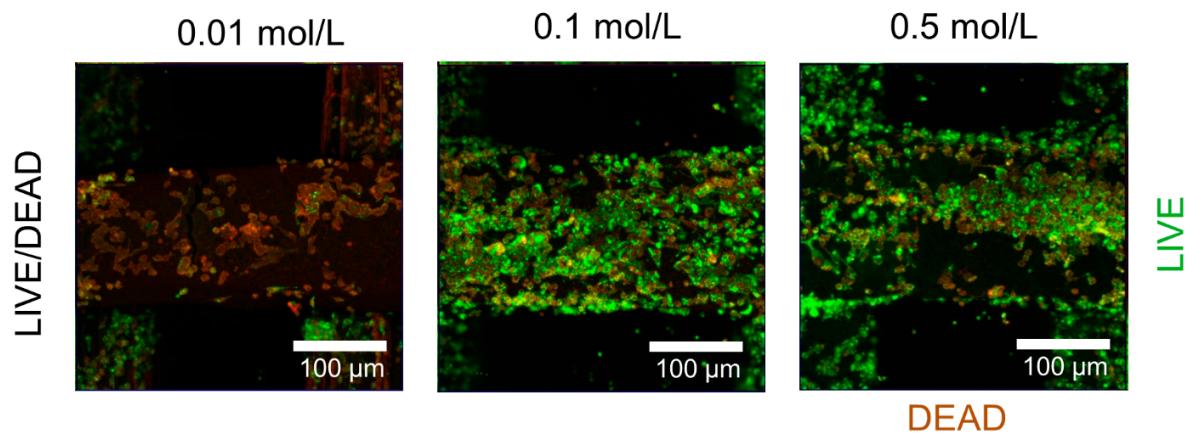
**Figure S1. Raman spectra for slurry and 3D printed components.** The focused Raman spectra on the fingerprint region of the spectrum between (500 to 1500) cm<sup>-1</sup> for commercial hydroxyapatite (HA); Poly vinyl butyral (PVB)/HA 3D printed scaffold; Tetracalcium phosphate (TTCP) powder; Dicalcium phosphate anhydrous (DCPA) powder; and PVB/Ethanol (EtOH) solution.



**Figure S2. XRD for slurry and 3D printed components.** Standard peaks in XRD of HA (blue), TTCP (green), DCPA (red) materials.



**Figure S3. Mechanical analysis of 3D printed scaffolds.** Strain-Stress curves of 3D printed scaffolds embedded in 0.01, 0.1 and 0.5 mol/L Na<sub>2</sub>HPO<sub>4</sub> solutions.



**Figure S4. Biocompatibility of 3D printed scaffolds.** The RAW cells lied on 3D printed scaffolds embedded in (0.01, 0.1 and 0.5) mol/L  $\text{Na}_2\text{HPO}_4$  solutions. Live and dead cells were stained for green and red, respectively (Scale Bar = 100  $\mu\text{m}$ ).

**Table S1. 3D printing parameters and apparent viscosity**

|                                     | <b>Values</b> |
|-------------------------------------|---------------|
| Needle diameter ( $\mu\text{m}$ )   | 210           |
| Needle length (mm)                  | 6.35          |
| Printing speed (mm/s)               | 5             |
| Fill Density (%)                    | 30            |
| Input flow (%)                      | 350           |
| syringe temp ( $^{\circ}\text{C}$ ) | 25            |
| plate temp ( $^{\circ}\text{C}$ )   | 25            |
| Lay-down pattern ( $^{\circ}$ )     | 0/90          |
| shear rate (1/s)                    | 65.58         |
| viscosity (Pa s)                    | 7.42          |

**Table S2. List of primers for RT-qPCR.** NFATC1, Nuclear Factor of Activated T Cells 1; TRAP, Tartrate-Resistant Acid Phosphatase; CTSK, Cathepsin K; OC-STAMP, Osteoclast Stimulatory Transmembrane Protein; DC-STAMP, Dendrocyte Expressed Seven Transmembrane Protein; 18S rRNA, have used in this study.

| Gene name      |         | Primer Sequence (5' - 3') | Amplicon Size |
|----------------|---------|---------------------------|---------------|
| human NFATC1   | Forward | CACCGCATCACAGGGAAGAC      | 119           |
|                | Reverse | GCACAGTCAATGACGGCTC       |               |
| human TRAP     | Forward | GACTGTGCAGATCCTGGGTG      | 122           |
|                | Reverse | GGTCAGAGAATACTGCCTCAAAG   |               |
| human CTSK     | Forward | ACACCCACTGGGAGCTATG       | 226           |
|                | Reverse | GACAGGGTACTTGAGTCCA       |               |
| human OC-STAMP | Forward | CACCCTGGGTATGGAGCAG       | 166           |
|                | Reverse | CTGGTGAGTGGTATTGAGGAGA    |               |
| human DC-STAMP | Forward | CGCTGCCTCCTGGATTATCAC     | 219           |
|                | Reverse | AAGCTTTGCCCTAGGTTG        |               |
| human 18S rRNA | Forward | CGGCTACCACATCCAAGGAA      | 169           |
|                | Reverse | GCTGGAATTACCGCGGCT        |               |