

SUPPLEMENTAL MATERIAL

Table S1

Listed in this table are the sequences (5' to 3') for primers used for cloning amplification of cDNAs representing *A.alcalica* MyoD1 and MyoD2. Products were cloned into pGEM T-Easy to make antisense cRNA probes for in situ hybridisation analysis.

Primer sets for generating probes	
AaMyoD1 forward	5' CCCACCGCTGATGATTTCTAT 3'
AaMyoD1 reverse	5' GCTTCGTCTTCTGGTTGTCT 3'
AaMyoD2 forward	5' ATGGATCTGTCAGACTTTCCCTTCGTT 3'
AaMyoD2 reverse	5' ACAAATGTTGCTGGACTCGGCAGACAG 3'

Table S2

Listed in this table are the sequences (5' to 3') for primers used for cloning full length HA-tagged products that were directionally cloned into the EcoR1 and Xba1 sites in pCS2+ to generate synthetic mRNAs for expression in *Xenopus*.

Primer sets for overexpression plasmids	
AaMyoD1 forward	5' AGAGAGGAATTCATGGAGTTGTCGGATATCTCTTTC 3'
AaMyoD1-HA reverse	5'AGAGAGTCTAGAAGCATAATCTGGAACATCATATGGATATAGGACTTGGTAAATCAGGTTGGGTCCTGGCTGCC 3'
AaMyoD2 forward	5' AGAGAGGAATTCATGGATCTGTCAGACTTTCCCTTCGTT 3'
AaMyoD2-HA reverse	5'AGAGAGTCTAGAAGCATAATCTGGAACATCATATGGATAACAAATGTTGCTGGACTCGGCAGACAG 3'
DrMyoD1 forward	5' AGAGAGGAATTCATGGAGTTGTCGGATATCC 5'
DrMyoD1-HA reverse	5'AGAGAGTCTAGAAGCATAAATCTGGAACATCATATGGATAAAGCACTTGATAAATGGTTTCC TG 3'

Table S3

Listed in this table are the sequences (5' to 3') for primers used for qPCR to measure transcriptional activity of MyoD1 proteins in *Xenopus* explants.

Primer sets for qPCR	
<i>X.laevis</i> act3 forward	5' TCACAACAGCTGAAAGGGAGAT 3'
<i>X.laevis</i> act3 reverse	5' AAGTCCAGAGCCACATAGGC 3'
<i>X.laevis</i> myh4 forward	5' GTGCGTTGTTTGATTCCCAAT 3'
<i>X.laevis</i> myh4 reverse	5' GCTGGTGGATGAGGAGATGGT 3'
<i>X.laevis</i> dicer forward	5' GGCTTTTACACATGCCTCTTACC 3'
<i>X.laevis</i> dicer reverse	5' GTCCAAAATTGCATCTCCAAG 3'

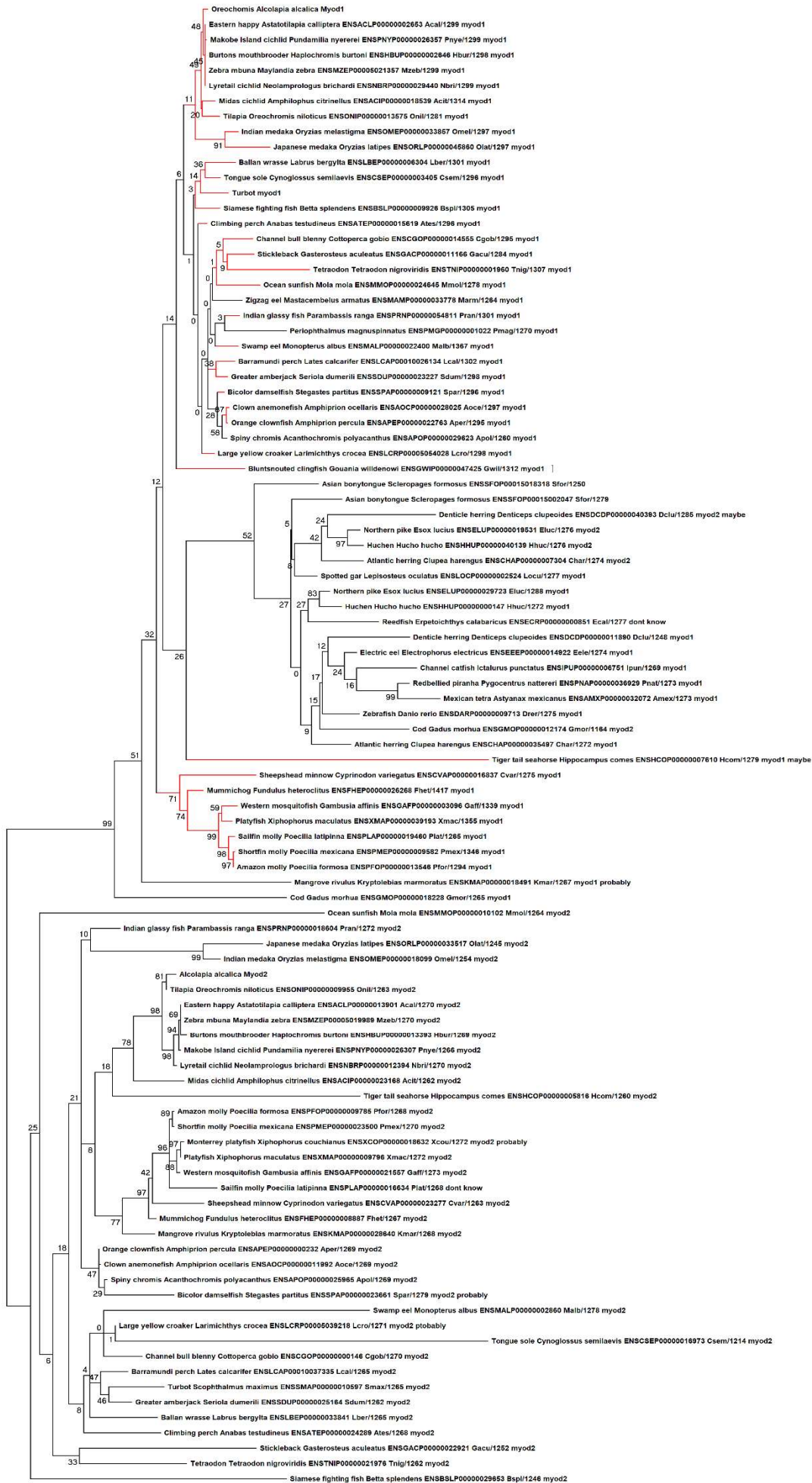
SUPPLEMENTAL FIGURES

O.alcalica	myoD1	MELS....DISFPIP.....TADDFYDDPCFNTSDMHFFEDLDPRLVHVG.LLKP
Danio	myoD1	MELS....DIPFPIP.....SADDFYDDPCFNTNDMHFFEDLDPRLVHVS.LLKP
Mouse	myoD1	MELLSPPLRDLIDLTPGDGSLCSFETADDFYDDPCFDSFPLDLRFEDLDPRLVHVGALLKPE
O.alcalica	myoD1	DSSSSSSSSPSSSSSSPSSLLHLHHHAEEVDEHVRAPSGHHQAGRCLLWACKACKRKT
Danio	myoD1	E.....HHHIE...DEHVRAPSGHHQAGRCLLWACKACKRKT
Mouse	myoD1	E...HAHFST.....AVHFGPGAREDEHVRAPSGHHQAGRCLLWACKACKRKT
O.alcalica	myoD1	NADRRKAATLRERRRLSKVNDAFETLKRCTTANPNQRLPKVEILRNAISYIESLQALLRG
Danio	myoD1	NADRRKAATMRERRRLSKVNDAFETLKRCTSTNPNQRLPKVEILRNAISYIESLQALLRS
Mouse	myoD1	NADRRKAATMRERRRLSKVNEAFETLKRCTSSNPNQRLPKVEILRNAIRYIEGLQALLRD
O.alcalica	myoD1	G.....QEDGFYP.....VLEHYSGSDSASSPLSNCSGDMTDFNGPTCQTTRRRSYD
Danio	myoD1QEDNYYP.....VLEHYSGSDSASSPSRNSCSGDMDFMGPTCQTTRRRNSYD
Mouse	myoD1	QDAAPPGAAAFYAPGPLPPGRGSEHYSGSDSASSPSRNSCSGMDYSGPPSPGPRRQNGYD
O.alcalica	myoD1	SSSYFSETPNGGLKLSERSSVSSSLDCLSSIVERISTDN..SSLPPADGPG.SPSTTTTTV
Danio	myoD1	SS.YFNDTPNADARNNKNSVSSSLDCLSSIVERISTETPACPVLSVPEGHEESPSPHEG
Mouse	myoD1	TA.YYSEAVR.ESRPGKSAAVSSSLDCLSSIVERISTDSFAAPALLLADAPPESEPGPPEG
O.alcalica	myoD1	P..VGEAGTATATAQVSSPTG..SQDPNLITYQVLI
Danio	myoD1	S..VLSDTGTTAPSTSCPQ...QQAQETIYQVL.
Mouse	myoD1	ASLSDTEQGTQTPSPDAAPQCPAGSNPNAIYQVL.

Supplemental Figure S1. Alignment of MyoD1 proteins from *Oreochromis alcalica*, *Danio rerio*, and *Mus musculus* showing highly conserved amino acid sequences in regions known to be essential for transcriptional activity (underlined): the transcriptional activation domain (TAD), Histidine/Cysteine (H/C) rich domain, and the bHLH domain. Residues implicated in protein stability are also conserved across the MyoD1 proteins and highlighted: putative phosphorylation targets of proline directed kinases (SP); Lysine (K) substrates for ubiquitin ligase; Tyrosines (Y) that can be targets of the kinase MEK1. Two additional sites for proline directed phosphorylation are found in the poly serine domain in *O.alcalica* MyoD1.

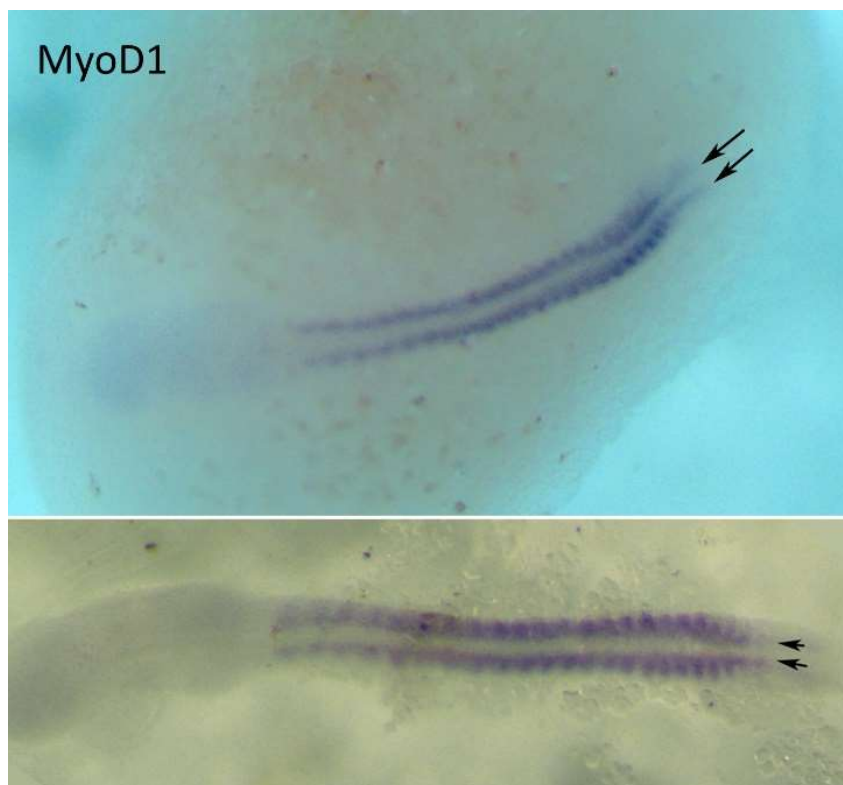
Amazon_molly_Poecilia_formosa
Ballan wrasse Labrus bergylta
Barramundi_perch Lates calcarifer
Bicolor_damselfish Stegastes partitus
Bluntnouted clingfish Gouania willdeno
Burtons_mouthbrooder Haplochromis burto
Channel_bull_blenny Cottoperca gobio
Climbing_perch Anabas testudineus
Clown_anemonefish Amphiprion ocellaris
Eastern_happy_Astatotilapia_Galliptera
Greater_amberjack_Seriola dumerili
Indian_glassy_fish Parambassis ranga
Indian_medaka Oryzias melastigma
Japanese_medaka Oryzias latipes
Large_yellow_croaker Larimichthys croce
Lyretail_cichlid Neolamprologus brichar
Makobe_Island_cichlid Pundamilia nyerere
Midas_cichlid Amphiprophus citrinellus
Mummichog Fundulus heteroclitus
Ocean_sunfish Mola mola
Orange_clownfish Amphiprion percula
Oreochromis Alcolapia alcalica
Platyfish Xiphophorus maculatus
Sheepshead_minnow Cyprinodon variegatus
Shortfin_molly Poecilia mexicana
Siamese_fighting_fish Betta splendens
Swamp_eel Monopterus albus
Tetraodon Tetraodon nigroviridis
Tilapia Oreochromis niloticus
Tongue_sole Cynoglossus semilaevis
Western_mosquitofish Gambusia affinis
Zebra mbuna Maylandia zebra

Supplemental Figure S2. Alignment of polyserine insert regions from multiple teleost MyoD1 proteins. The grey region is conserved in all vertebrate MyoD1 sequences as part of the N-terminal activation domain. The Histidine highlighted in green indicates the beginning of the H/C rich region, where conservation with all vertebrate MyoD1 proteins is re-established. Potential sites for proline directed kinase activity are indicated in yellow.

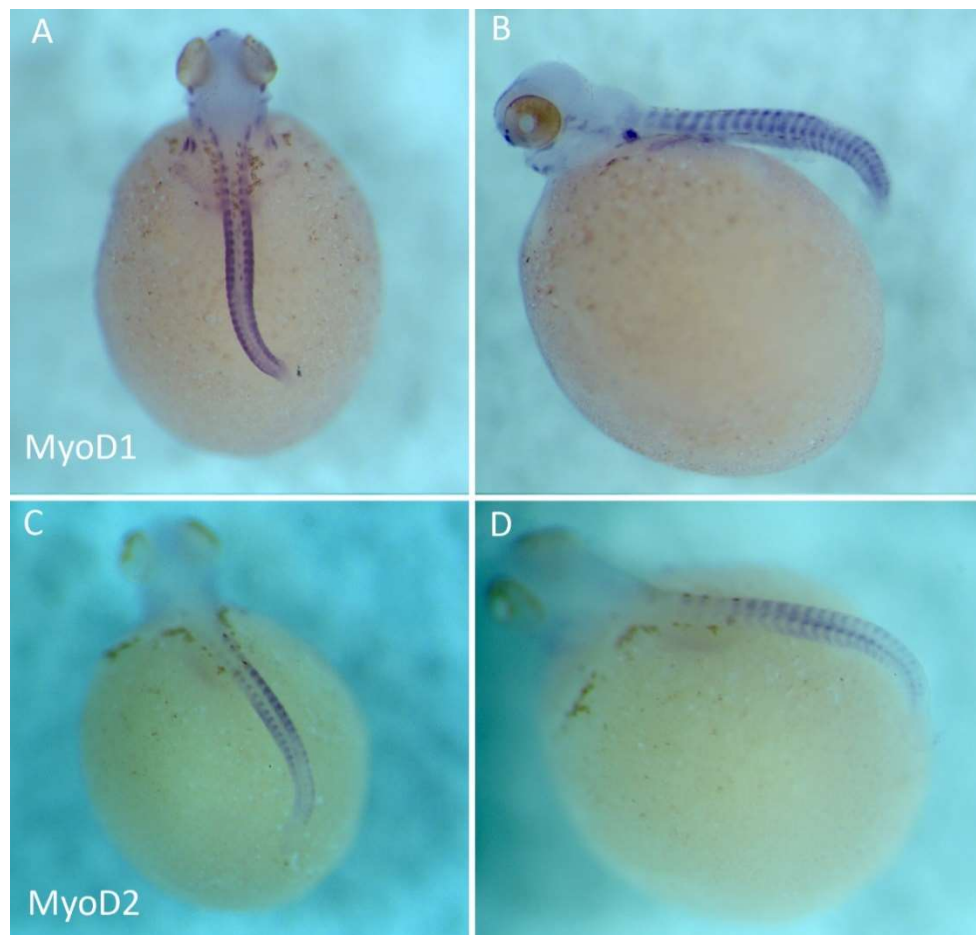


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Supplemental Figure S3 (previous page). Phylogenetic tree derived from the analysis of the amino acid sequences of 97 teleost MyoD proteins. MyoD1 and MyoD2 proteins were analysed using MEGAX using the JTT w/freq method and gamma distribution. 59 MyoD1 [36 with a poly-serine insertion] and 38 MyoD2 from 54 teleost species clade separately; where MyoD1 containing and not containing a poly-serine insertion clade as MyoD1. Analysis conducted on clades is supported by a bootstrap value of 99, scale bar represents a genetic distance of 0.5 amino acid substitutions per site. Those MyoD1 with a poly serine insertion are indicated by red branches red.



Supplemental Figure S4. The expression of *MyoD1* in pre-hatch stage *Oreochromis (Alcolapia) alcalica* embryos. The top panel shows a whole mount in situ hybridisation as a dorsal view, with anterior to the left. The bottom panel is a similar stage embryo as a flat-mount, where the yolk has been removed so the embryo could be flattened to improve visualisation of the posterior expression. The arrows in both panels indicate *MyoD1* expression in the putative adaxial region.



Supplemental Figure S5. Expression of MyoD1 (A,B) and MyoD2 (C,D) in stage *Oreochromis (Alcolapia) alcalica* embryos at approximately 3 days post fertilisation. A,C are dorsal views; B,D are lateral views with anterior to the left.