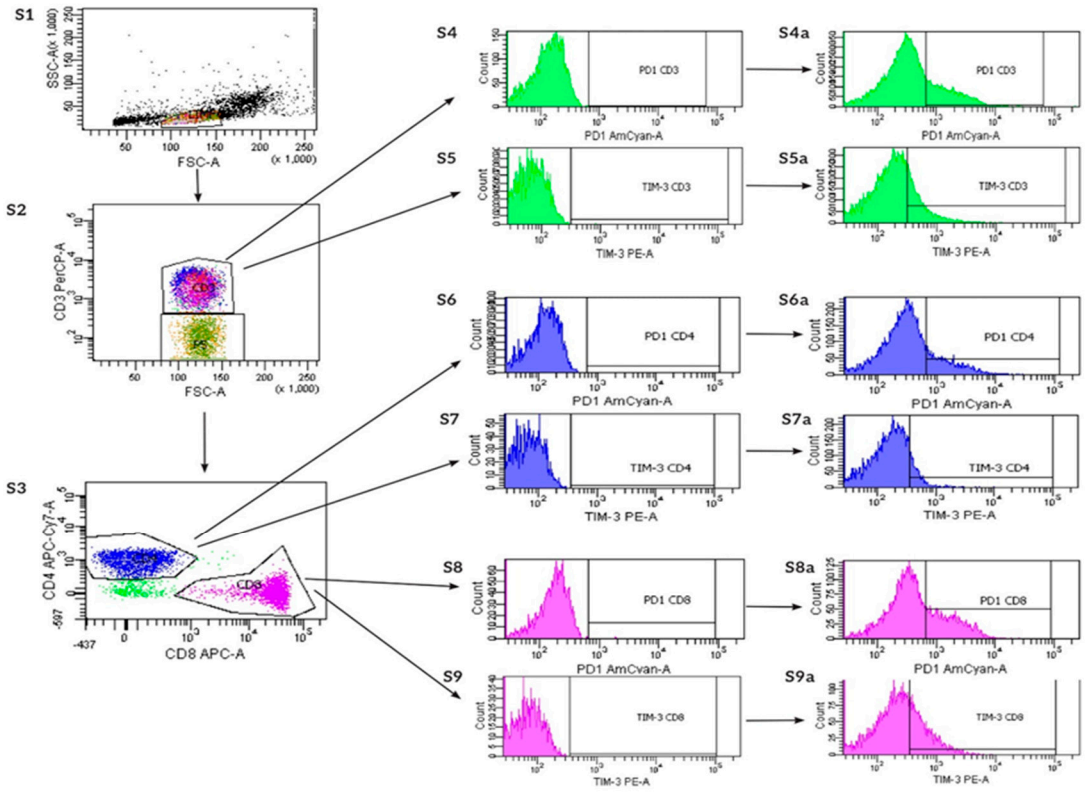

Figure S1. Graph 1-Gating strategy: lymphocytes were identified on FSC/SSC dot plot S1) as cells within 50-100 x 10^3 on FSC and below 50 x 10^3 on SSC. S2) CD3 positive cells (T lymphocytes were identified as events with PreCP fluorescence above 10^3 on logarithmic scale) S3) CD4 positive cells (T helper lymphocytes were identified as events with APC-Cy7 fluorescence above 10^2 on logarithmic scale) and CD8 positive cells (T cytotoxic lymphocytes were identified as events with APC fluorescence above 10^3 on logarithmic scale) S4-S9)Cut-off values for PD1 (AmCyan channel) and TIM3 (PE channel) considering each studied population (total T cell, T helper and T cytotoxic cells) were established basing on fluorescence minus one (FMO) experiments, performed each time test was run. S4a-S9a) PD1 and Tim-3 positive populations were identified with fluorescence higher than fluorescence minus one, therefore positive.

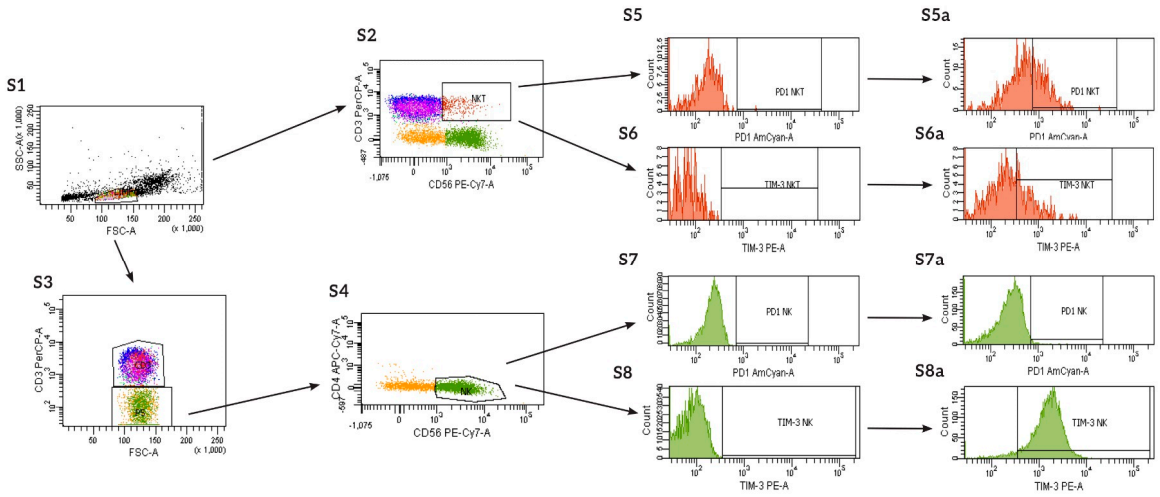

Figure S2. Graph 2-Gating strategy: lymphocytes were identified on FSC/SSC dot plot S1) as cells within 50-100 x 10^3 on FSC and below 50 x 10^3 on SSC, S2) NKT cells were identified as CD56 positive(fluorescence above 10^3 Pe-Cy-7 channel, logarithmic scale) and CD 3 positive (fluorescence above 10^3 PerCP channel, logarithmic scale), S3) CD3 negative cells

(non- T lymphocytes were identified as events with PreCP fluorescence below 10^3 on logarithmic scale), S4) NK cells were identified as CD56 positive(fluorescence above 10^3 Pe-Cy-7 channel, logarithmic scale) and negative for CD3(S3), S5-S8)Cut-off values for PD1 (AmCyan channel) and TIM3 (PE channel) considering each studied population (NKT and NK cells) were established basing on fluorescence minus one (FMO) experiments, performed each time test was run. S5a-S8a) PD1 and Tim-3 positive populations were identified with fluorescence higher than fluorescence minus one, therefore positive.