

Supporting Information for

Comparative Solution Equilibrium Studies on Anticancer

Estradiol-Based Conjugates and Their Copper Complexes

Éva A. Enyedy *, Anett Giricz, Tatsiana V. Petrasheuskaya, János P. Mészáros,
Nóra V. May, Gabriella Spengler, Ferenc Kovács, Barnabás Molnár and Éva Frank

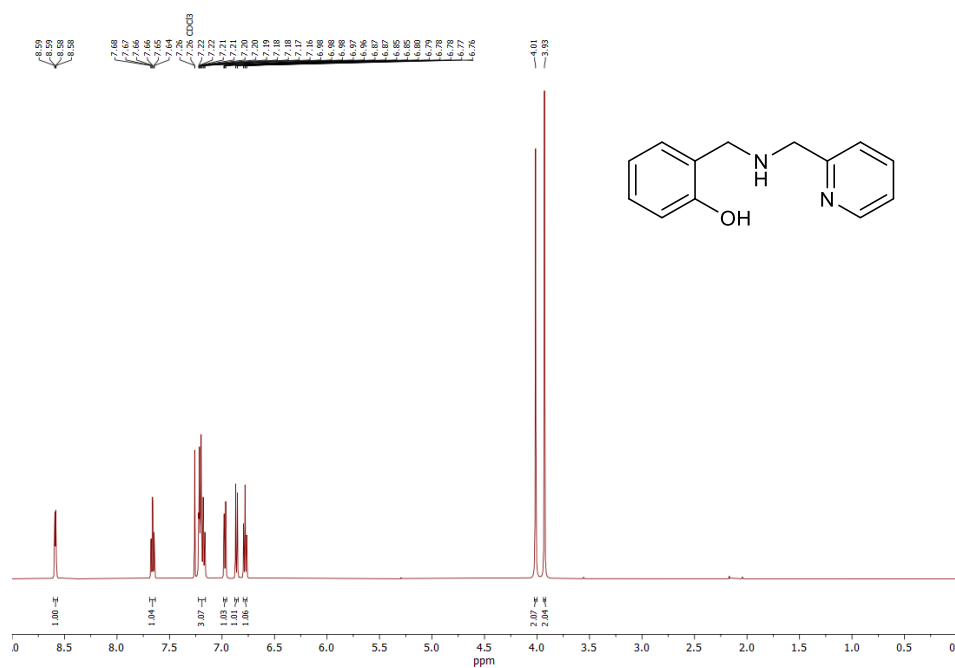


Figure S1. ¹H NMR spectrum of PMAP in CDCl₃.

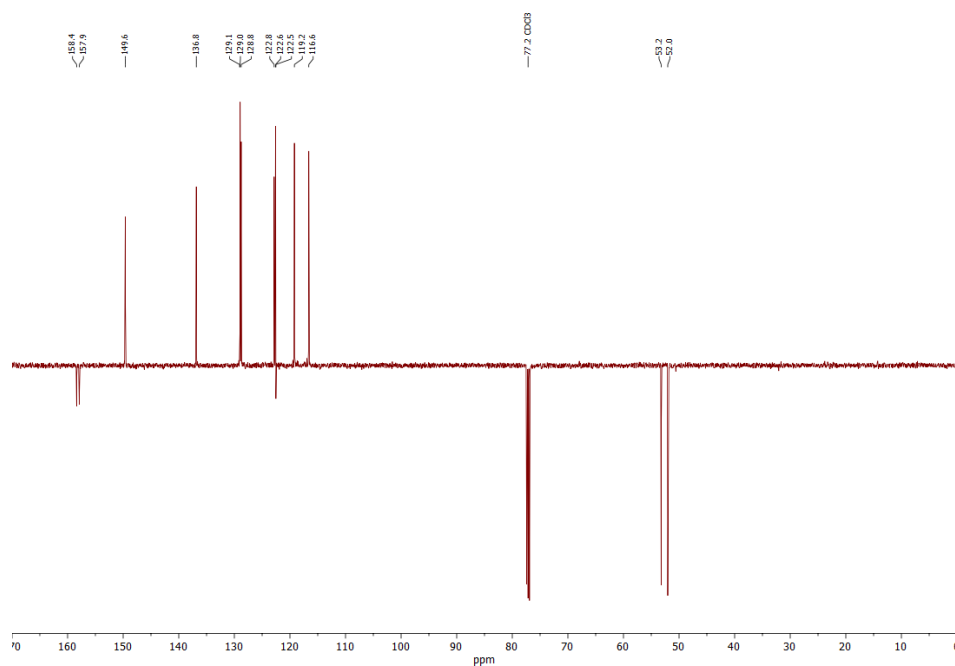


Figure S2. ¹³C NMR spectrum of PMAP in CDCl₃.

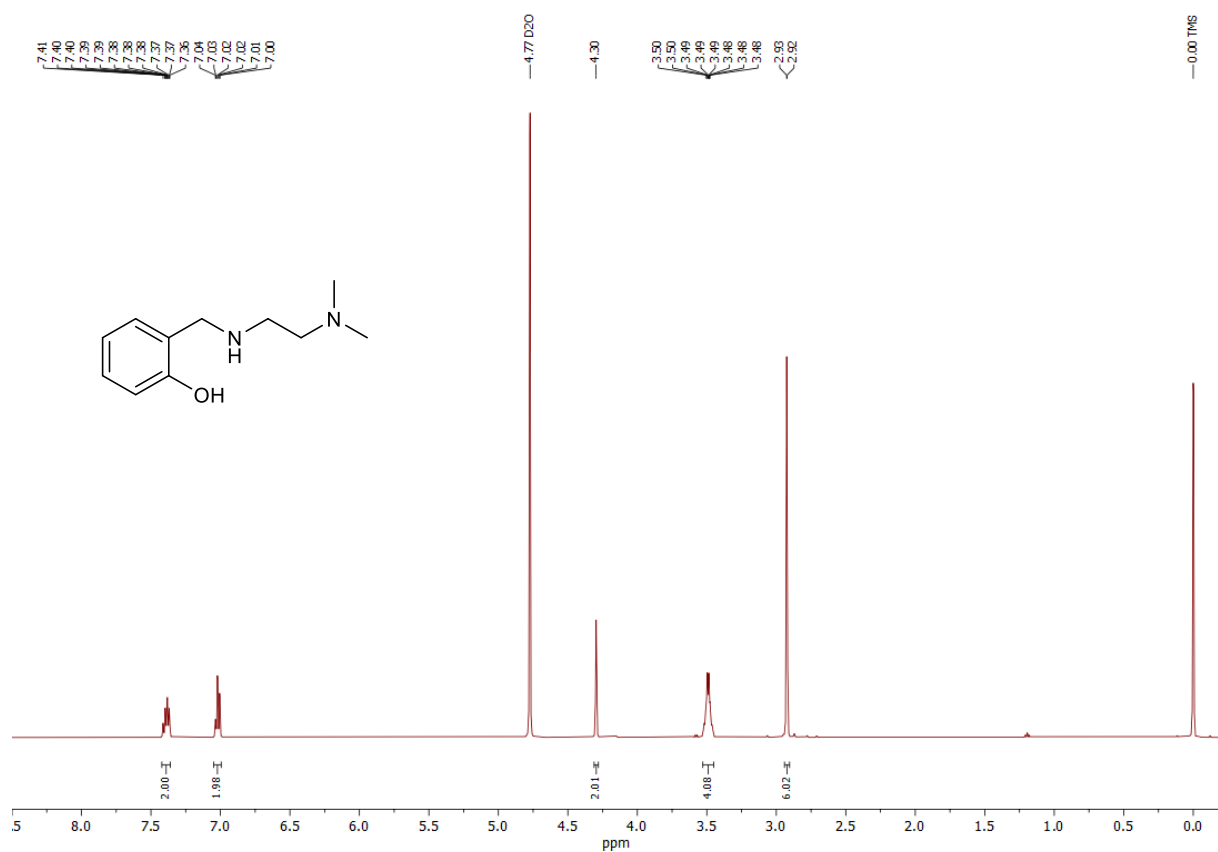


Figure S3. ¹H NMR spectrum of DMAP in D₂O.

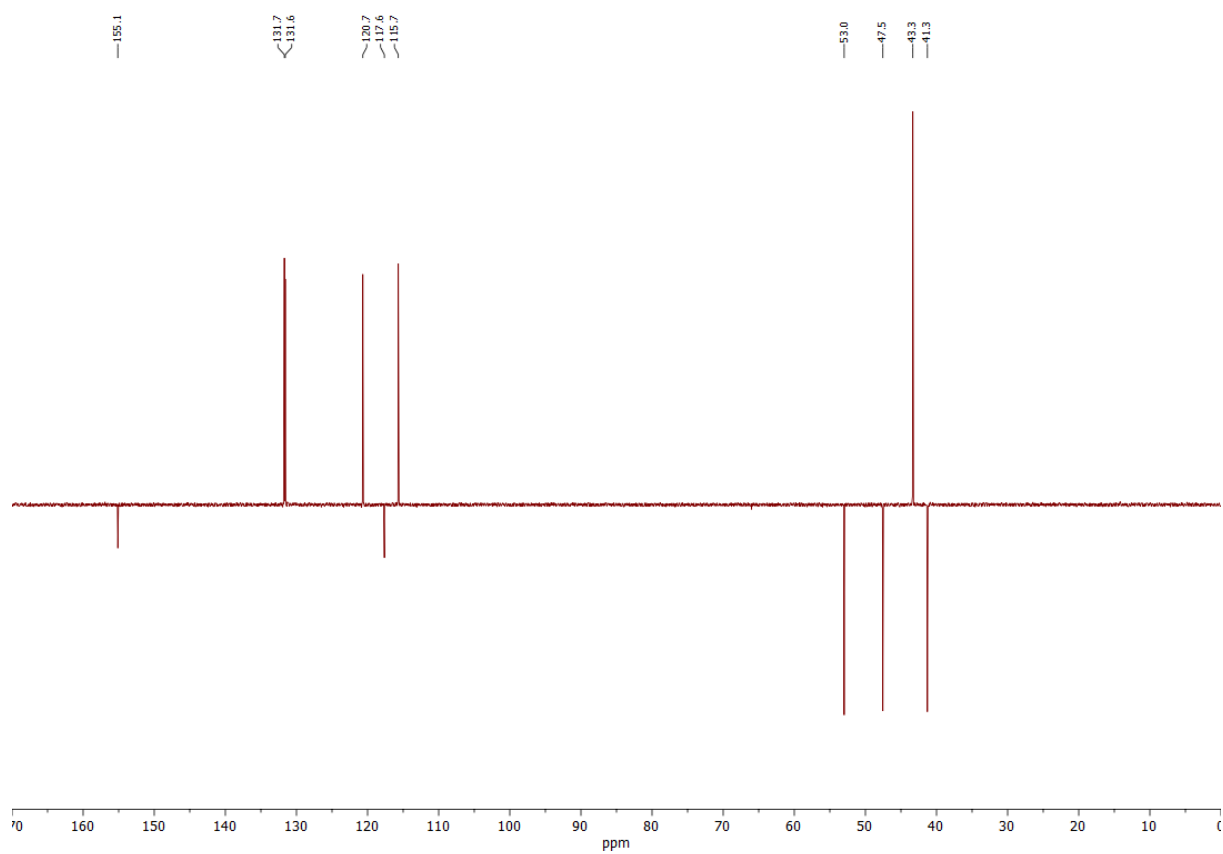


Figure S4. ¹³C NMR spectrum of DMAP in D₂O.

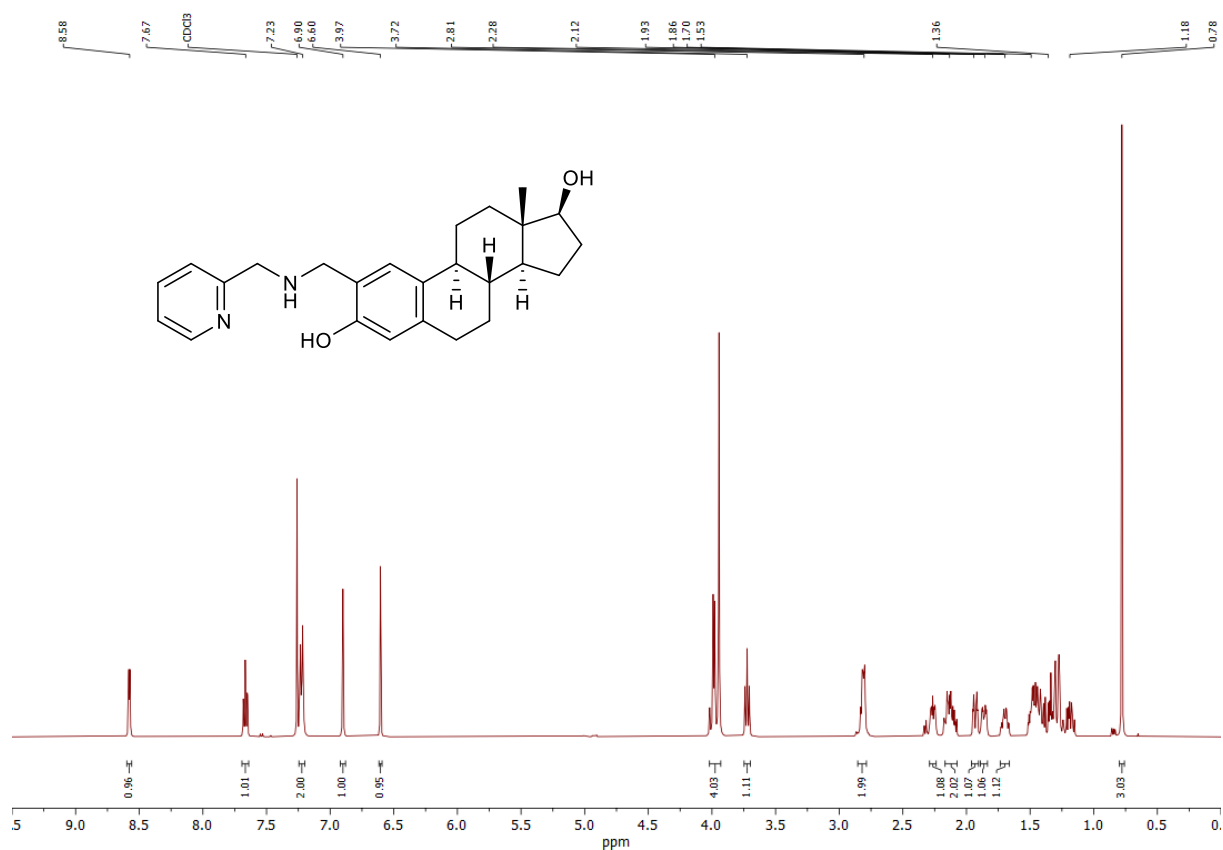


Figure S5. ¹H NMR spectrum of PMA-E2 in CDCl₃.

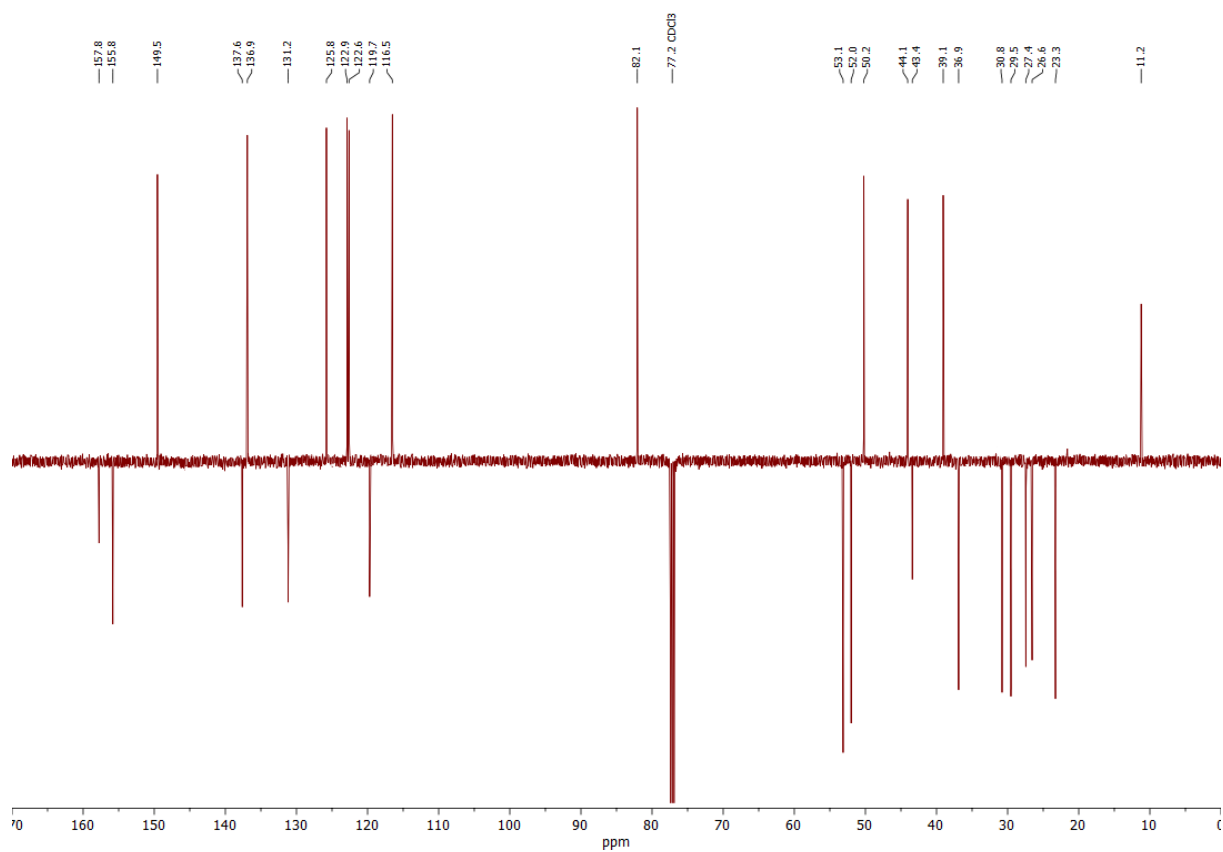


Figure S6. ¹³C NMR spectrum of PMA-E2 in CDCl₃.

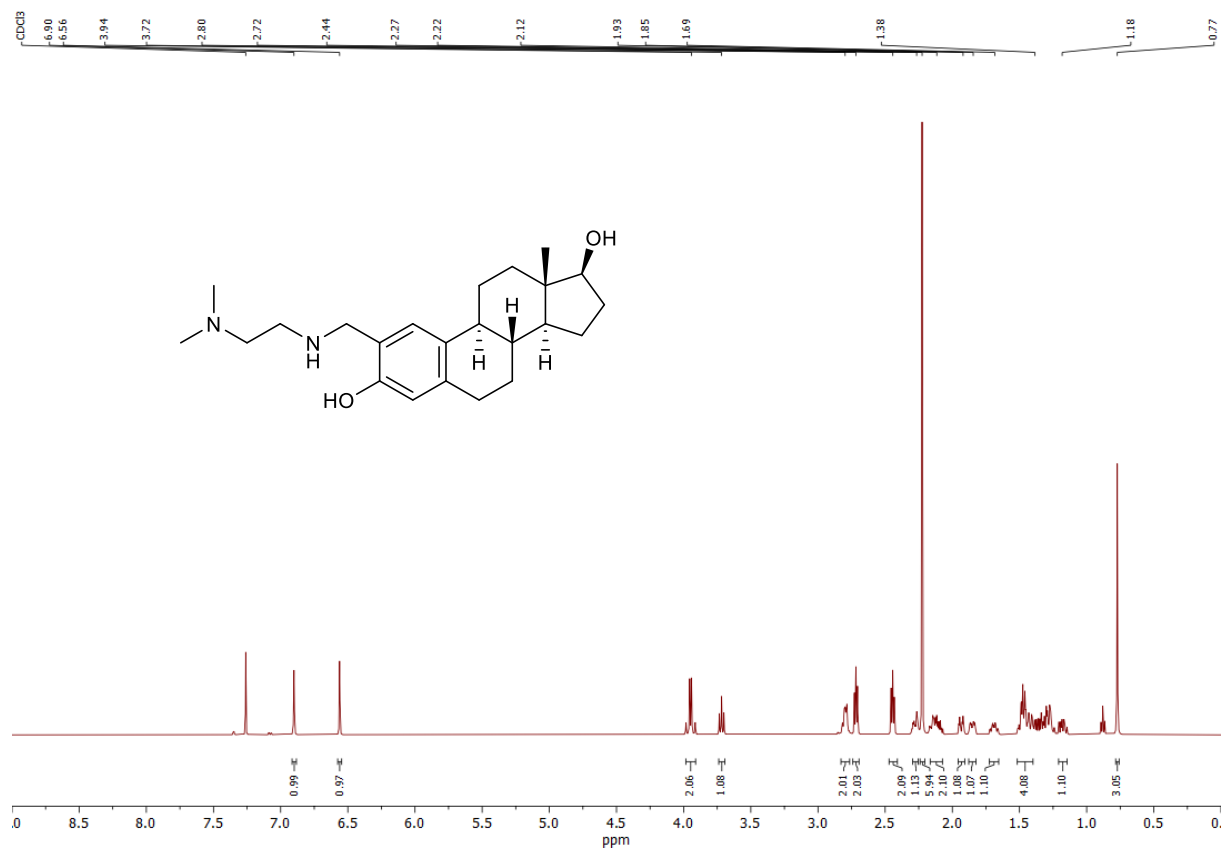


Figure S7. ¹H NMR spectrum of DMA-E2 in CDCl₃.

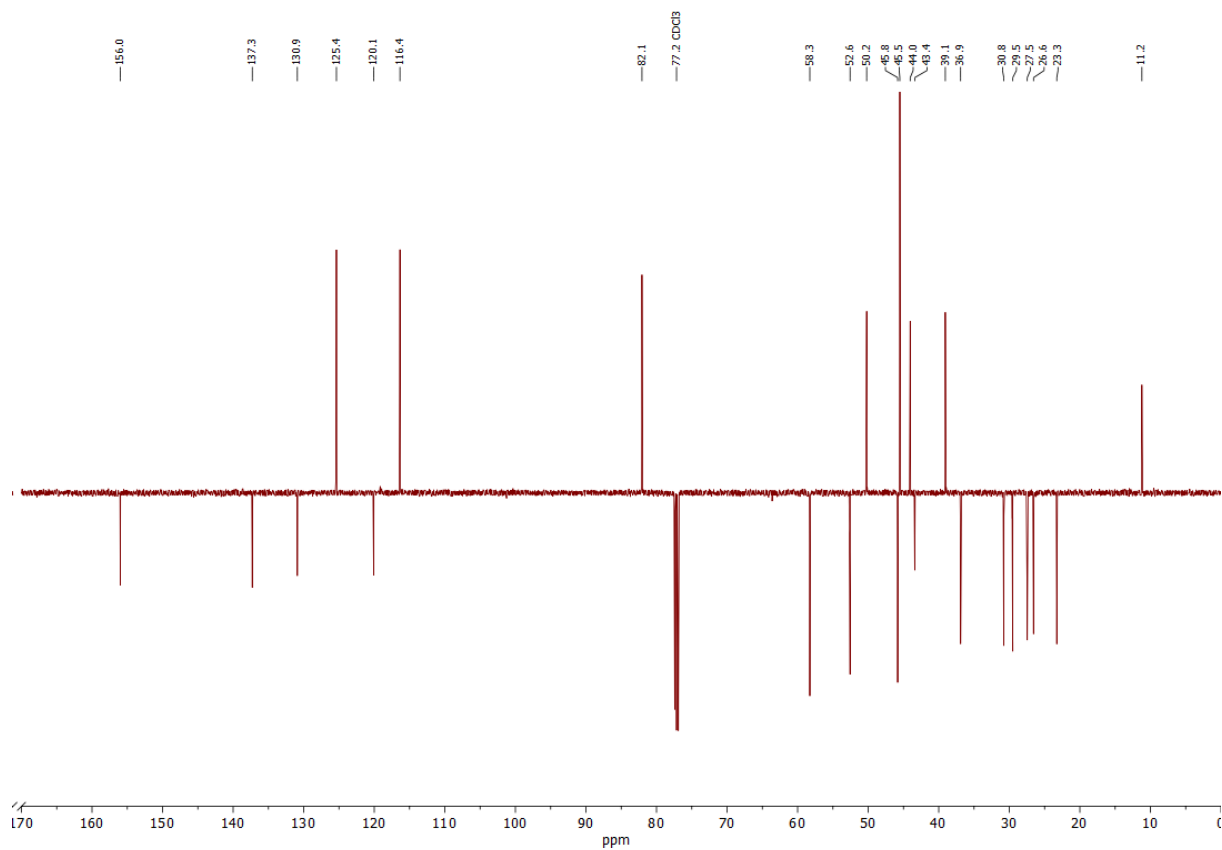


Figure S8. ¹³C NMR spectrum of DMA-E2 in CDCl₃.

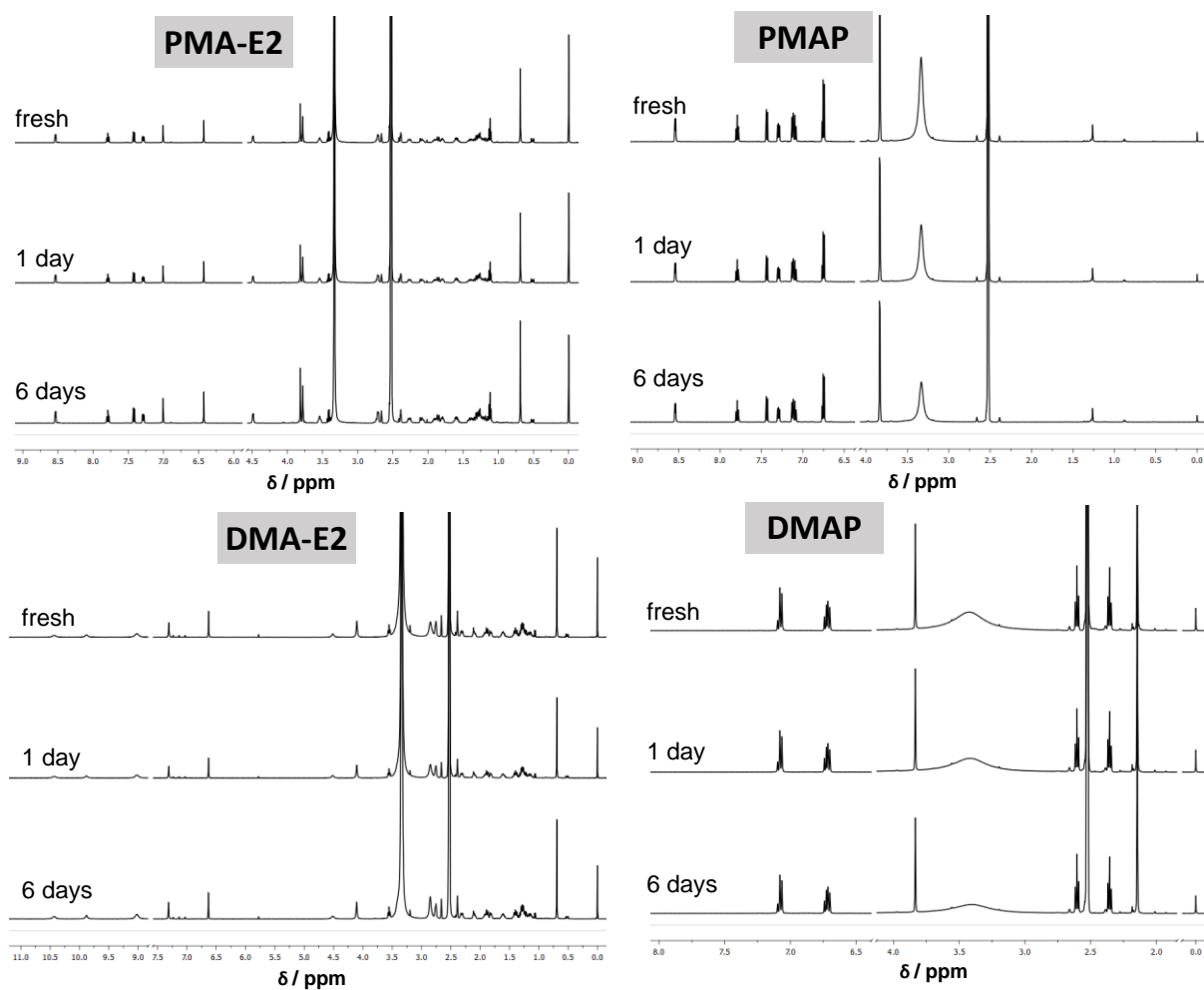


Figure S9. ^1H NMR spectra of the studied ligands in d_6 -DMSO in their fresh solution and after 1 and 6 days waiting time.

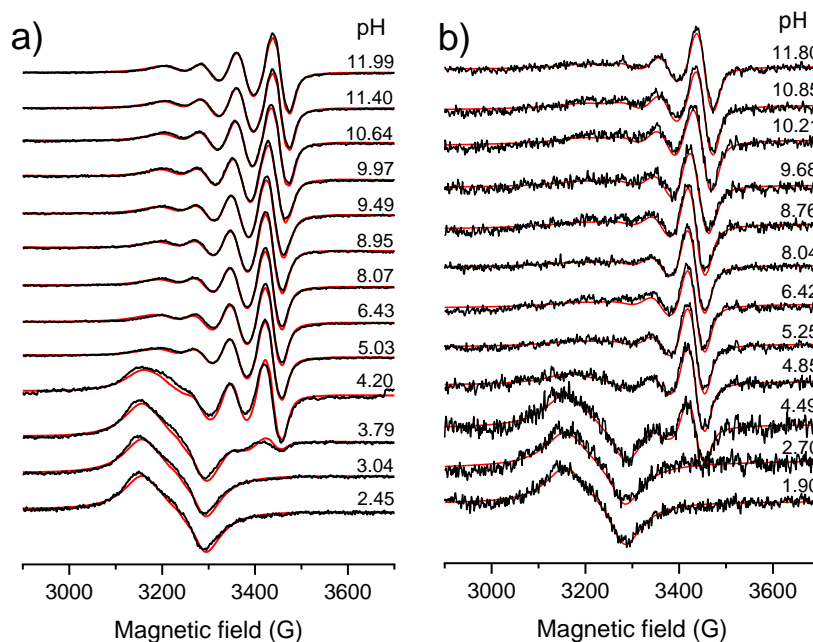


Figure S10. (a) pH-dependent experimental (black) and simulated (red) isotropic EPR spectra recorded for the Cu(II) – DMAP system at concentration $c_{\text{ligand}} = 3.0$ mM and $c_{\text{Cu(II)}} = 2.84$ mM, (b) and for Cu(II) – DMA-E2 system at concentration $c_{\text{ligand}} = 0.55$ mM and $c_{\text{Cu(II)}} = 0.43$ mM at 295 K. {30% (v/v) DMSO/H₂O; $I = 0.10$ M (KCl)}

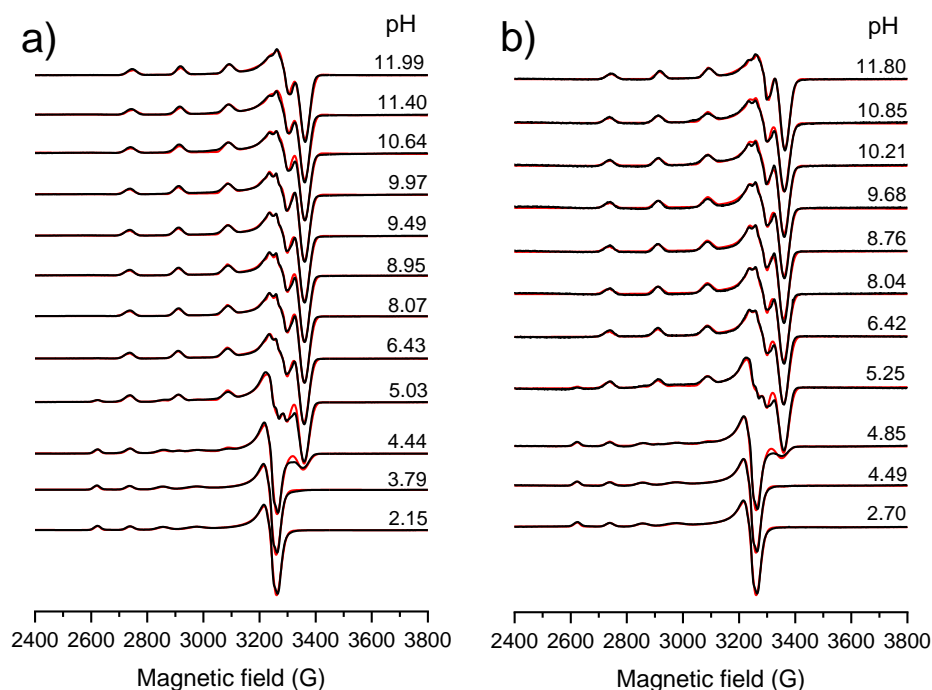


Figure S11. (a) pH-dependent experimental (black) and simulated (red) anisotropic EPR spectra recorded for Cu(II) – DMAP system at concentrations of $c_{\text{ligand}} = 3.0$ mM and $c_{\text{Cu(II)}} = 2.84$ mM, (b) and for Cu(II) – DMA-E2 system at concentrations of $c_{\text{ligand}} = 0.55$ mM and $c_{\text{Cu(II)}} = 0.43$ mM at 77 K. {30% (v/v) DMSO/H₂O; $I = 0.10$ M (KCl)}

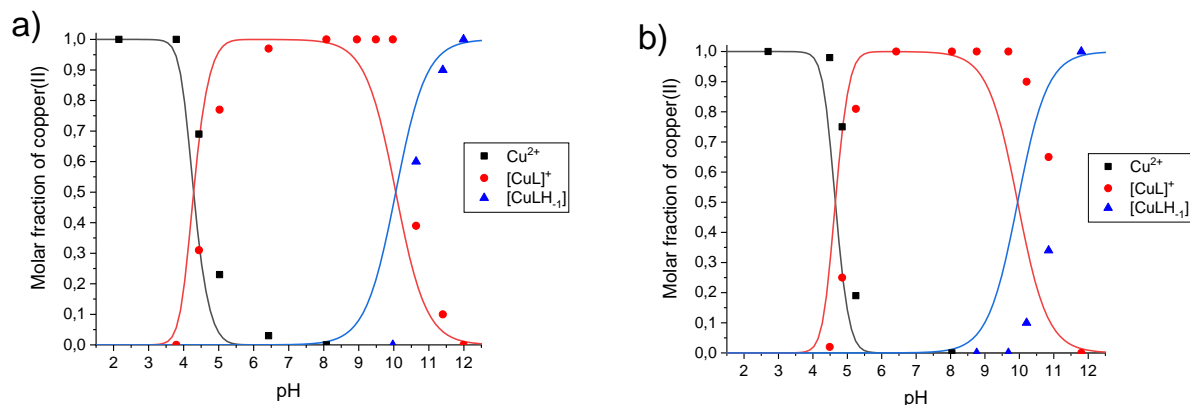


Figure S12. Concentration distribution curves at the concentrations of the EPR measurements using the formation constants obtained by pH-potentiometric (DMAP) or UV-vis spectrophotometric (DMA-E2) titrations (lines) and the component ratios (scatter) obtained by the simulation of frozen solution EPR spectra for (a) Cu(II) – DMAP and (b) Cu(II) – DMA-E2 systems.

Table S1. Electrochemical data collected for Cu(II) – DMAP, Cu(II) – DMA-E2 and Cu(II) – PMAP systems at pH 7.4 using 10 mV/s scan rate. {30% (v/v) DMF/H₂O, $c_{\text{ligand}} = 0.60$ or 0.55 mM, $c_{\text{Cu(II)}} = 0.50$ mM; glassy carbon working electrode, Pt auxiliary electrode, Ag/AgCl/KCl (3 M) reference electrode, 0.1 M KNO₃ supporting electrolyte}

| | DMAP | DMA-E2 | PMAP |
|------------------------|------|--------|------|
| E_c / mV | −452 | −486 | −450 |
| E_a / mV | −215 | −315 | −280 |
| ΔE / mV | 237 | 171 | 178 |
| $E_{1/2}$ / mV | −333 | −400 | −365 |
| $E_{1/2}$ / mV vs. NHE | −123 | −190 | −155 |
| $ i_d/i_a $ | 1.22 | 1.13 | 1.00 |

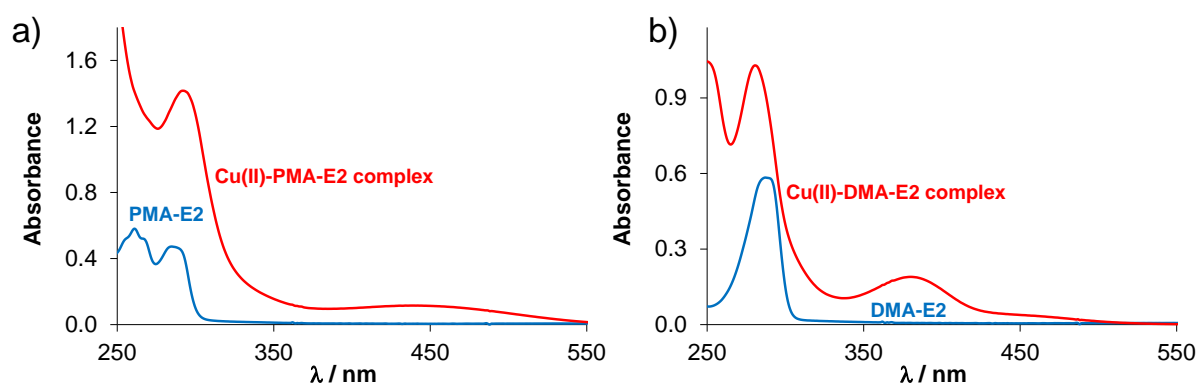


Figure S13. (a) UV-vis absorption spectra of PMA-E2 and its Cu(II) complex and (b) DMA-E2 and its Cu(II) complex in methanol. $\{c_{\text{PMA}} = 170 \mu\text{M}$; $c_{\text{Cu(II)-PMA}} = 230 \mu\text{M}$; $c_{\text{DMA}} = 160 \mu\text{M}$; $c_{\text{Cu(II)-DMA}} = 110 \mu\text{M}$; $\ell = 1 \text{ cm}\}$

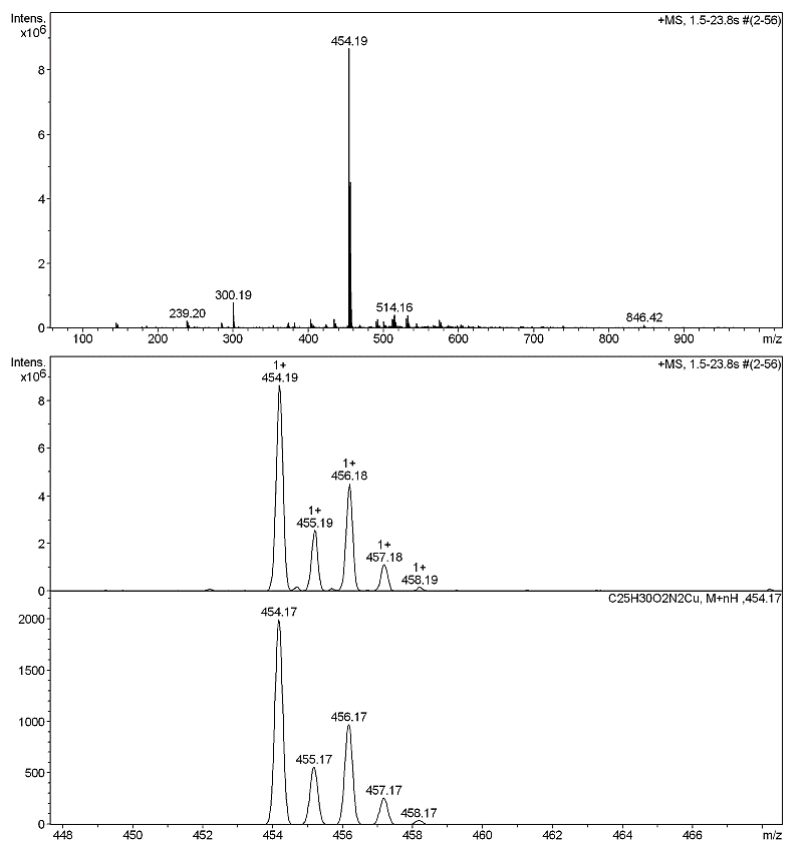


Figure S14. Low-resolution ESI-MS spectra of Cu(II) complex of PMA-E2 [CuLCl]. Sample was prepared in methanol (m/z [CuL]⁺ found 454.19, calcd. 454.17 for C₂₅H₃₁N₂O₂Cu).

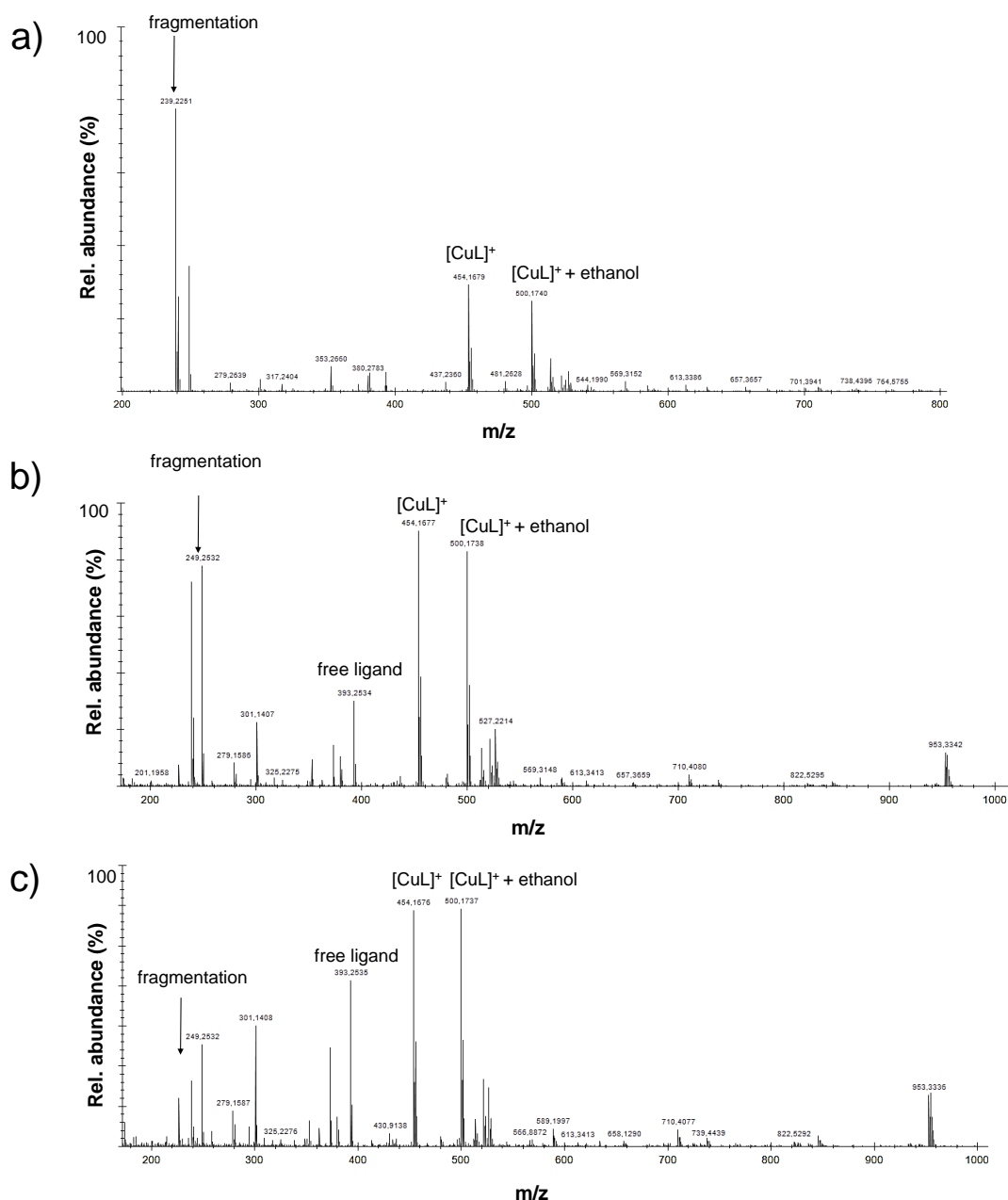


Figure S15. High-resolution ESI-MS spectra of Cu(II) complex of PMA-E2 [CuLCl]. Sample was prepared in methanol. **(a)** Scan time: 0.63 min. (m/z [CuL]⁺ found 454.1679, calcd. 454.1682 for C₂₅H₃₁N₂O₂Cu; [CuL]⁺ + ethanol found 500.1740, calcd. 500.2100 for C₂₅H₃₁N₂O₂Cu + C₂H₅OH). **(b)** Scan time: 0.68 min. (m/z [CuL]⁺ found 454.1677, calcd. 454.1682 for C₂₅H₃₁N₂O₂Cu; [CuL]⁺ + ethanol found 500.1738, calcd. 500.2100 for C₂₅H₃₁N₂O₂Cu + C₂H₅OH). **(c)** Scan time: 0.73 min. (m/z [CuL]⁺ found 454.1676, calcd. 454.1682 for C₂₅H₃₁N₂O₂Cu; [CuL]⁺ + ethanol found 500.1737, calcd. 500.2100 for C₂₅H₃₁N₂O₂Cu + C₂H₅OH).

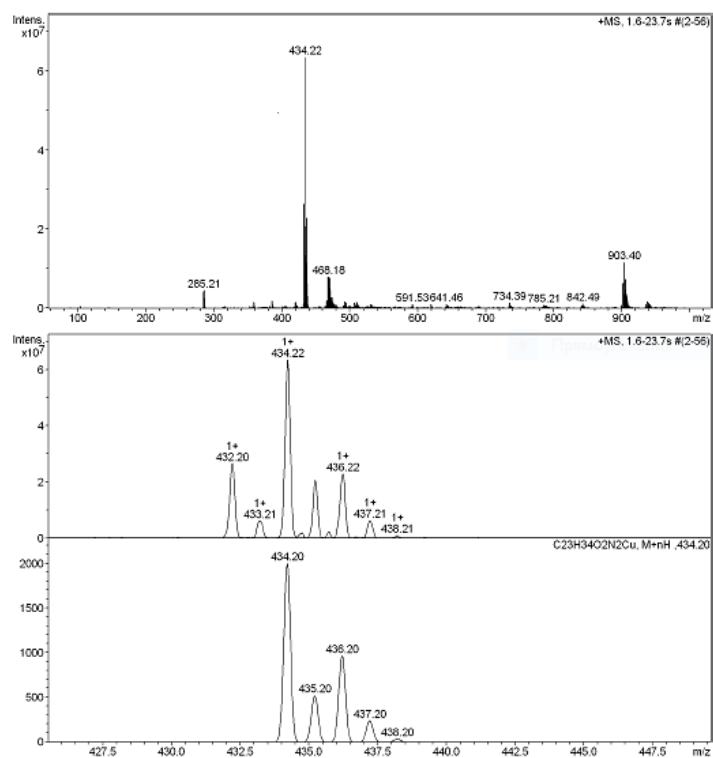


Figure S16. Low-resolution ESI-MS spectra of Cu(II) complex of DMA-E2 [CuLCl]. Sample was prepared in methanol (m/z [CuL]⁺ found 434.22, calcd. 434.20 for $C_{23}H_{35}N_2O_2Cu$).

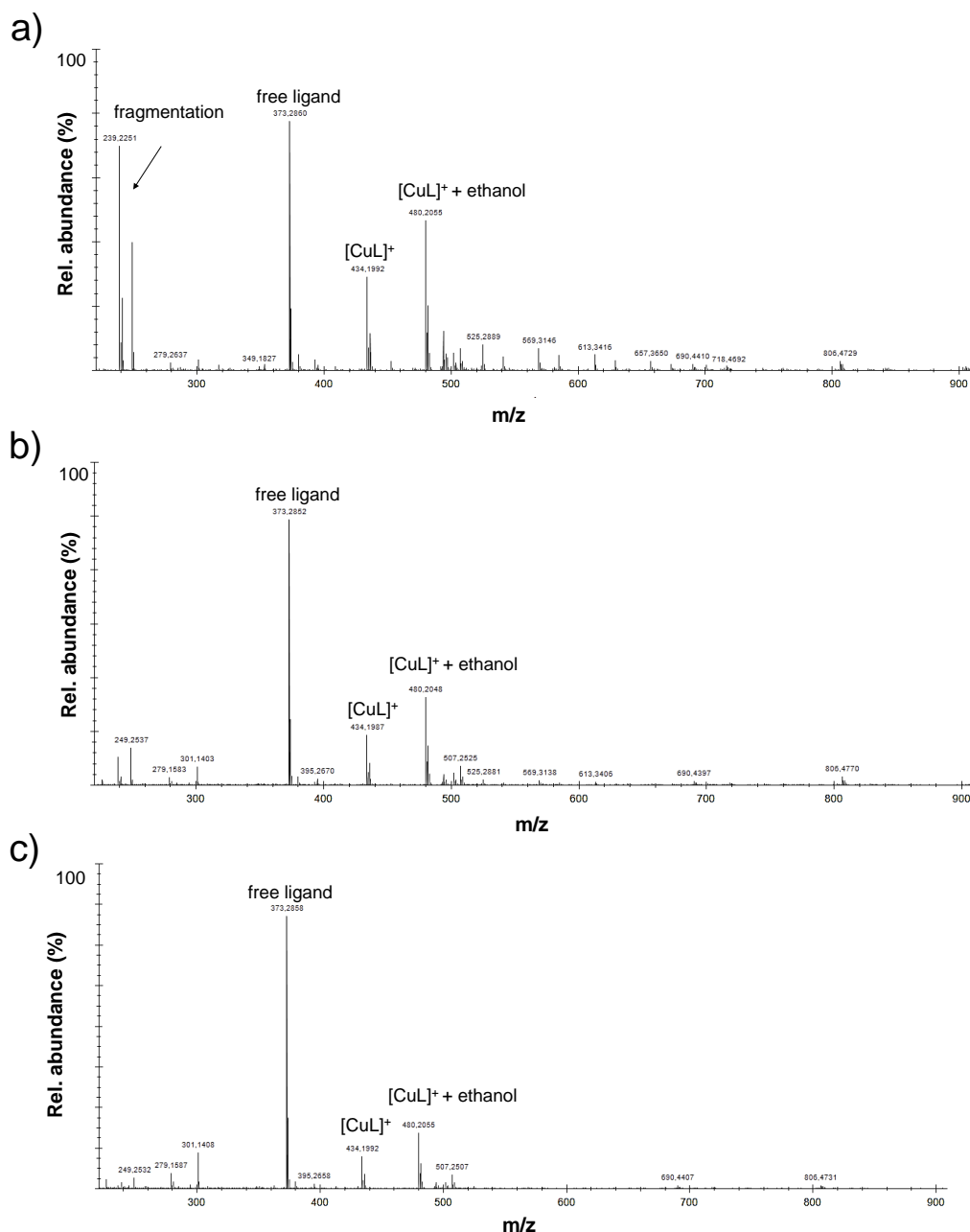


Figure S17. High-resolution ESI-MS spectra of Cu(II) complex of DMA-E2 [CuLCl]. Sample was prepared in methanol. **(a)** Scan time: 0.28 min. (m/z [CuL]⁺ found 434.1992, calcd. 434.1995 for C₂₃H₃₅N₂O₂Cu; [CuL]⁺ + ethanol found 480.2055, calcd. 480.2413 for C₂₃H₃₅N₂O₂Cu + C₂H₅OH). **(b)** Scan time: 0.33 min. (m/z [CuL]⁺ found 434.1987, calcd. 434.1995 for C₂₃H₃₅N₂O₂Cu; [CuL]⁺ + ethanol found 480.2048, calcd. 480.2413 for C₂₃H₃₅N₂O₂Cu + C₂H₅OH). **(c)** Scan time: 0.38 min. (m/z [CuL]⁺ found 434.1987, calcd. 434.1992 for C₂₃H₃₅N₂O₂Cu; [CuL]⁺ + ethanol found 480.2055, calcd. 480.2413 for C₂₃H₃₅N₂O₂Cu + C₂H₅OH).

NOTE: It should be noted that the DMA-E2 complex appeared exclusively in the low-resolution ESI-MS spectrum, whereas the signal of the free ligand was found in the high-resolution spectra (increased signal with increased scan time), which may be due to dissociation of the Cu(II) complex under the condition of the measurement.

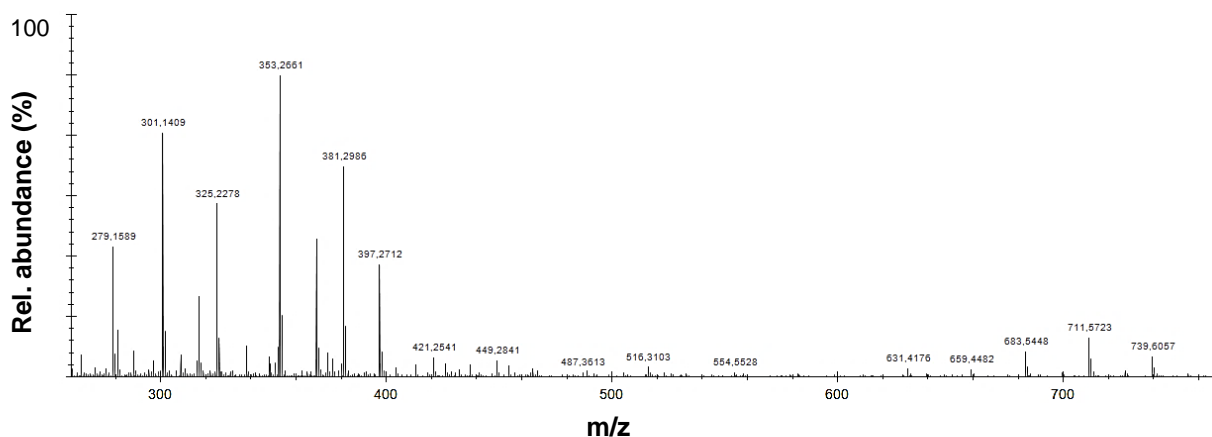
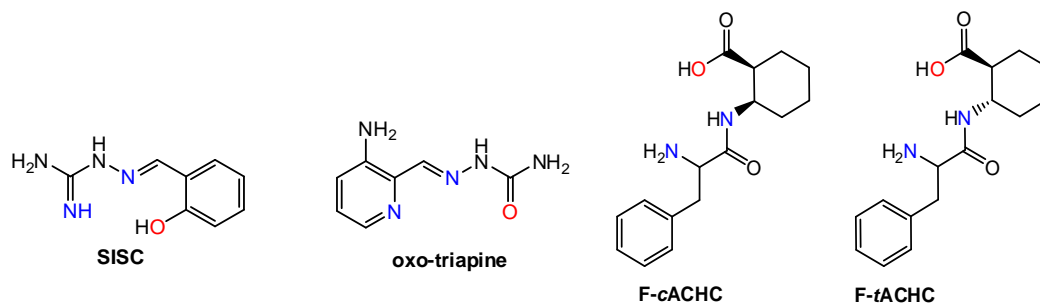


Figure S18. Background spectrum for the high resolution ESI-MS measurements.

Table S2. EPR parameters^a of the complexes determined at 77 K (anisotropic) from the EPR spectra recorded for the isolated Cu(II) complex of DMA-E2 and PMA-E2, together with the parameters of [CuL]⁺ and [CuLH-1] complexes of DMAP and reference data for Cu(II) complexes of salicylaldehyde (2-(2-hydroxybenzylidene)hydrazinecarboximidamide (SISC) [1], (E)-2-((3-aminopyridin-2-yl)methylene)hydrazinecarboxamide (oxo-triapine) [2], and the dipeptides F-cACHC [3] and F-tACHC [3] bearing (N,N,O) coordination mode.



| | complex | coordination mode | g_x | g_y | g_z | A_x | A_y | A_z | g_0^{calcd} |
|---------------------|--------------------|---|-------|-------|-------|-------|-------|-------|---------------|
| DMA-E2 | isolated | (N,N,O ⁻) | 2.055 | 2.055 | 2.244 | 27 | 27 | 177 | 2.118 |
| PMA-E2 | isolated | (N,N,O ⁻) | 2.051 | 2.051 | 2.256 | 20 | 20 | 178 | 2.119 |
| DMAP | [CuL] ⁺ | (N,N,O ⁻) | 2.050 | 2.050 | 2.248 | 24 | 24 | 181 | 2.116 |
| | [CuLH-1] | (N,N ⁻ ,O ⁻) | 2.047 | 2.047 | 2.242 | 23 | 23 | 176 | 2.112 |
| SISC | [CuL] ⁺ | (N,N,O ⁻) | 2.034 | 2.056 | 2.248 | 19 | 22 | 174 | 2.113 |
| | [CuLH-1] | (N,N,O ⁻)(OH ⁻) | 2.045 | 2.048 | 2.245 | 14 | 21 | 176 | 2.113 |
| oxo-triapine | [CuL] ⁺ | (N,N,O ⁻) | 2.054 | 2.054 | 2.255 | 16 | 16 | 156 | 2.121 |
| F-cACHC | [CuLH-1] | (N,N,O ⁻) | 2.038 | 2.053 | 2.237 | 23 | 19 | 191 | 2.109 |
| F-tACHC | [CuLH-1] | (N,N,O ⁻) | 2.040 | 2.057 | 2.252 | 18 | 21 | 182 | 2.116 |

^a The coupling values are given in 10^{-4} cm^{-1} , the relaxation parameters are in G. The experimental error was ± 0.001 for g and $\pm 1 \times 10^{-4} \text{ cm}^{-1}$ for A parameters. ^b Calculated by the equation $g_0 = (g_x + g_y + g_z)/3$.

References

1. Dömötör, O.; May, N.V.; Gál, G.T.; Spengler, G.; Dobrova, A.; Arion, V.B.; Enyedy, É.A. Solution Equilibrium Studies on Salicylidene Aminoguanidine Schiff Base Metal Complexes: Impact of the Hybridization with L-Proline on Stability, Redox Activity and Cytotoxicity. *Molecules* **2022**, *27*, 2044.
2. Enyedy, É.A.; May, N.V.; Pape, V.F.S.; Heffeter, P.; Szakács, G.; Keppler, B.K.; Kowol, C.R. Complex formation and cytotoxicity of Triapine derivatives: A comparative solution study on the effect of the chalcogen atom and NH-methylation. *Dalton Trans.* **2020**, *49*, 16887–16902.
3. Nagy, N.V.; Doorslaer, S.V.; Szabó-Plánka, T.; Rompaey, S.V.; Hamza, A.; Fülöp, F.; Tóth, G.K.; Rockenbauer, A. Copper (II)-binding ability of stereoisomeric *cis*- and *trans*-2-aminocyclohexanecarboxylic acid–l-phenylalanine dipeptides. A combined cw/pulsed EPR and DFT study. *Inorg. Chem.* **2012**, *51*, 1386–1399.