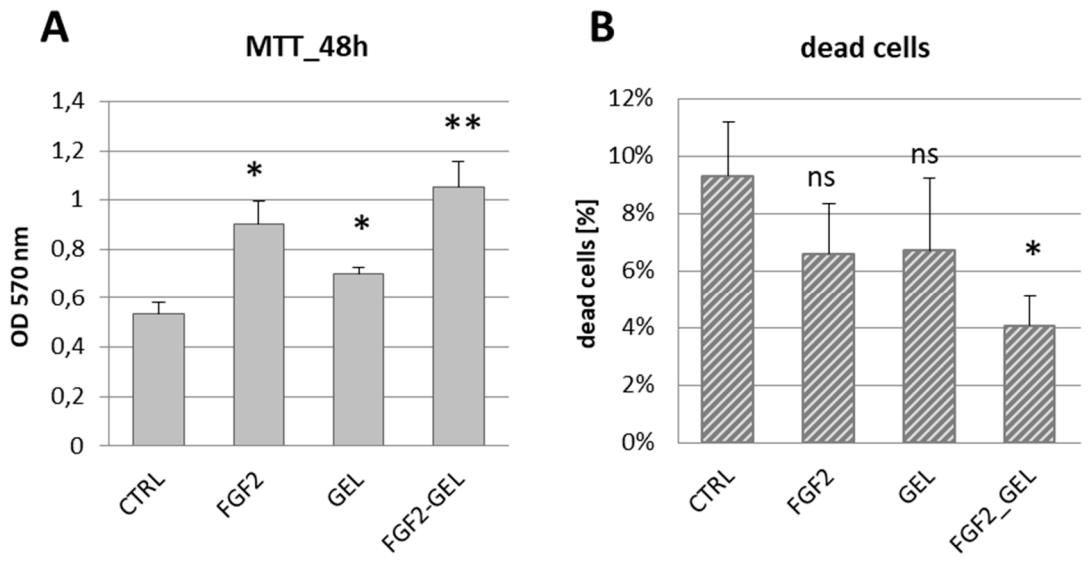


Supplementary Figure S1. The effect of different culture conditions on viability and proliferation of canine umbilical cord-derived cells. A) MTT assay, B) dead cell proportion assessment based on Trypan blue staining. FGF2 - cells cultured on uncoated surface in growth medium (GM) supplemented with 2.5ng/mL fibroblast growth factor 2, GEL – cells cultured in GM on a surface coated with porcine gelatine, FGF2_GEL – cells cultured on gelatine coated dishes in GM with FGF2; ns – statistically not significant, *, $p<0.05$; **,$p<0.01$ in comparison to control (using Wilcoxon test or t-Student for related data test depending on data distribution), n=6-7.

Supplementary Table S1. List of reagents used for identification of canine UC-derived cells.

	Clone	Fluorochrome	Cat no	Antibody specificity
CD44	IM-7	PE	#12-0441-82	used in canine cells [1,2]
CD90	YKIX337.217	APC	#17-5900-42	Anti dog
CD45	YKIX716.13	FITC	#11-5450-41	Anti dog
CD11b	M1/70	V450	#560456	used in canine cells [3]

Human MSCs were identified using a ready set of antibodies from kit: BD Stemflow™ hMSC Analysis Kit.

Supplementary Table S2. List of reagents used for analysis of lymphocytes proliferation in both human and canine samples.

Cell origin	antigen	Clone	Fluorochrome	Cat no
human	CD3	SK7	PerCP	#557851
	CD4	SK3	APC Cy7	#341115
	CD8	SK1	APC	#345775
	Viability	NA	V450	#L23105
	proliferation	NA	FITC (CFSE)	#C34570
canine	CD3	CA17.2A12	FITC	#MCA1774F
	CD4	YKIX302.9	PE-Cy7	#17-5040-42
	CD8	YCATE55.9	PE	#MCA1039PE
	Viability	NA	V450	#L23105
	proliferation	NA	APC	#C34564

NA- not applicable

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