



Supplementary Information

Rapid Antibacterial Activity Assessment of Chimeric Lysins

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Table S1. Information of SH3 domains used in this study.

Abbreviated name	Sequence identity to K _{SH3} (%)	Genbank Protein Full name	Genbank accession No.	Amino acid Region	Amino acid Length
L25(K _{SH3})	100	LysK [<i>Staphylococcus phage</i> P4W]	AFN38929	409-495	87
ClyC _{SH3}	94.2	Lysin [<i>Staphylococcus phage</i> PALS_1]	QDJ97474.1	123-209	87
vB _{SH3}	93.1	N-acetylmuramoyl-L-alanine amidase [<i>Staphylococcus phage</i> vB_Sau_Clo6]	ARM69344.1	409-495	87
tras14 _{SH3}	40.2	endolysin [<i>Staphylococcus phage</i> trsa14]	AWH13066.1	395-481	87
ClyO _{SH3}	11.4	PlySs2 family phage lysin [<i>Streptococcus suis</i>]	WP_024417665.1	162-245	84
Lsp _{SH3}	9.1	Lysostaphin [<i>Staphylococcus simulans</i>]	WP_013012297.1	369-452	84
2638A _{SH3}	6.8	endolysin [<i>Staphylococcus phage</i> 2638A]	YP_239818.1	401-486	86
2958 _{SH3}	4.5	endolysin [<i>Staphylococcus phage</i> phi2958PVL]	YP_002268027.1	405-484	80

Table S2. Information of lysostaphin and chimeric lysins used in this study.

Protein name	Genbank accession No.	Amino acid Region	Amino acid Length	Reference
Lysostaphin	WP_013012297.1	207-452	246	[34]
Lsp		207-368	162	
ClyC	AGO49867.1	1-171	171	[24]
	InsertVD	172-173	2	
	QDJ97474.1	102-209	108	
ClyO	WP_258391128.1	1-157	157	[33]
	InsertG	158	1	
	WP_024417665.1 (R195C)	146-245	100	

Table S3. Analyses of genetic traits related to expression levels of chimeric lysins.

Chimeric lysin	RI	Frequency of amino acid (%)		Translation stalling codon pair ^c	Free energy ^d (kcal/mol)
		Low expression-related amino acid ^a /codon ^b	high expression-related amino acid		
ALS2-dA-L21	1.00	52.3 (11/21)/ 14.2 (3/21)	52.3% (11/21)	PP, PG	-14.4/-213.2
ALS2-dA-L24	0.35	50.0 (12/24)/12.5 (3/24)	50.0% (12/24)	PP, PG	-16.3/-213.6
ALS2-dA-L25	0.95	52.0 (13/25)/16.0 (4/25)	52.0% (13/25)	PP, PG	-16.5/-219.2
ALS2-dA-L26	0.93	50.0 (13/26)/15.3 (4/26)	50.0% (13/26)	PP, PG	-15.1/-214.8
ALS2-dA-L27	0.52	48.1 (13/27)/14.8 (4/27)	48.1% (13/27)	PP, PG	-15.2/-213.9
ALS2-dA-L28	0.76	46.4 (13/28)/14.2 (4/28)	46.4% (13/28)	PP, PG	-16.3/-216.0
ALS2-dA-L31	0.46	45.1 (14/31)/12.9 (4/31)	45.1% (14/31)	PP, PG	-23.4/-222.7
ALS2-dA-L34	0.23	47.0 (16/34)/11.7 (4/34)	47.0% (16/34)	PP, PG	-25.6/-221.7
ALS2-dA-L35	0.57	48.5 (17/35)/14.2 (5/35)	48.5% (17/35)	PP, PG	-28.5/-227.0
ALS2-dA-L38	0.33	44.7 (17/38)/13.1 (5/38)	44.7% (17/38)	PP, PG	-29.6/-227.8
ALS2-dA-L25	0.32	54.0 (47/87) 19.5 (17/87)	11.5% (10/87)	PP, PG	-85.7/-219.2
ALS2-dA-ClyC _{SH3}	0.40	54.0 (47/87) 16.0 (14/87)	11.5% (10/87)	-	-66.0/-203.4
ALS2-dA-vB _{SH3}	0.36	55.1 (48/87) 18.3 (16/87)	11.5% (10/87)	-	-64.3/-197.8
ALS2-dA-tras14 _{SH3}	0.57	48.2 (42/87) 16.0 (14/87)	18.4% (16/87)	-	-76.9/-210.1
ALS2-dA-ClyO _{SH3}	0.64	44.0 (37/84) 8.3 (7/84)	15.4% (13/84)	-	-81.7/-208.3
ALS2-dA-Lsp _{SH3}	0.91	46.4 (39/84) 15.4 (13/84)	19.0% (16/84)	-	-59.6/-187.5
ALS2-dA-2638A _{SH3}	1.00	40.6 (35/86) 9.3 (8/86)	30.2% (26/86)	-	-54.4/-187.3
ALS2-dA-2958 _{SH3}	0.09	50.0 (40/80) 12.5 (10/80)	10.0% (8/80)	-	-56.1/-184.9
ALS2-dA-L25	0.29	50.6% (115/227)/16.7% (38/227)	13.2% (30/227)	PP, PG SH3	-1.3/-219.2
Lysostaphin	1.00	46.9% (116/247)/15.4% (38/247)	19.0% (47/247)	-	-4.6/-214.4
Lsp-ClyC _{SH3}	0.10	49.6% (124/250)/15.6% (39/250)	16.4% (41/250)	-	-4.6/-224.8
ClyC	0.17	47.6% (134/281)/11.7% (33/281)	19.9% (56/281)	-	-3.6/-241.8
ClyO	2.08	46.9% (121/258)/13.2% (34/258)	15.9% (41/258)	-	-10.0/-245.5

^a The frequency of L, S, R, A, P, G, C, W, and I in linker, SH3 domain, insert gene and initial~16codon.^b The frequency of AGG (R), CGU (R), CGA (R), AGU (S), GGU (G), AGA (R), UGU (C), CGG (R), CCC (P), AUA (I) in linker, SH3 domain and complete coding region.^c Ribosome stalling codon pairs: RR (CGG-AGG, AGG-CGG, CGA-CGG, CGG-AGG), PP (CCA-CCC, CCG-CCA, CCT-CCG, CCG-CCG, CCT-CCT, CCA-CCG), KR (AAG-AGG), LR (CTG-AGG), PG (CCG-GGT, CCG-GGG), VG (GTG-GGA), and QD (CAG-GAT).^d Predicted free energy of mRNA secondary structure of chimeric lysin by RNA fold. Partial regions: linker, SH3 domain, and first 16-dodons.

Table S4. PCR primers used for amplification of 10 linker candidates

Primer name	Sequence (5' -3')	Usage
P T7-F	TCGAAATTAATACGACTCACTATAG	IVTC/TL
T term-R	GAGGGGATCCAGATCTTGGGAT	amplification for all
P_gCHAP-L-F	CTTTAAGAAGGAGATATACATATGG- CATCATCTTTTAATCACC	Linker & SH3 domain
P_gCHAP-L-R	GTGATTAAAA- GATGATGCCATATGTATATCTCCTTCTTAAAG	SOE-PCR for all
K _{SH3} _T-L-F	GTTGCATGGGGTGTTTTCAAAAAGGGTCATCATCAC- CATCAC	Linker & SOE-PCR
K _{SH3} _T-L-R	GTGATGGTGTATGATGACCCTTTT- GAAAACACCCCATGCAAC	for all
gCHAP_L21-L-F	GCTATGCATTCATCCATTCTGCATCTACTCCGGCAAC	SOE-PCR
gCHAP_L21-L-R	GTTGCCGGAGTAGATGCAGAATGGATGAATGCATAGC	for linker21
gCHAP_L24-L-F	GCTATGCATTCATCCATAAAACTTCTTCTGCATCTAC	SOE-PCR
gCHAP_L24-L-R	GTAGATGCAGAAGAAGTTTTATGGATGAATGCATAGC	for linker24
gCHAP_L25-L-F	GCTATGCATTCATCCATGGTAAACTTCTTCTGCATC	SOE-PCR
gCHAP_L25-L-R	GATGCAGAAGAAGTTTTACCATGGATGAATGCATAGC	for linker25
gCHAP_L26-L-F	GCTATGCATTCATCCATGACGGTAAACTTCTTCTGC	SOE-PCR
gCHAP_L26-L-R	GCAGAAGAAGTTTTACCGTCATGGATGAATGCATAGC	for linker26
gCHAP_L27-L-F	GCTATGCATTCATCCATAAAGACGGTAAACTTCTTC	SOE-PCR
gCHAP_L27-L-R	GAAGAAGTTTTACCGTCTTTATGGATGAATGCATAGC	for linker27
gCHAP_L28-L-F	GCTATGCATTCATCCATGTTAAAGACGGTAAACTTC	SOE-PCR
gCHAP_L28-L-R	GAAGTTTTACCGTCTTTAACATGGATGAATGCATAGC	for linker28
gCHAP_L31-L-F	GCTATGCATTCATCCATTCTACTGTCGTAAAGAC	SOE-PCR
gCHAP_L31-L-R	GTCTTTAACGACAGTAGAATGGATGAATGCATAGC	for linker31
gCHAP_L34-L-F	GCTATGCATTCATCCATACTTCTTCTTCTACTGTCG	SOE-PCR
gCHAP_L34-L-R	CGACAGTAGAAGAAGAAGTATGGATGAATGCATAGC	for linker34
gCHAP_L35-L-F	GCTATGCATTCATCCATGGTACTTCTTCTTCTACTG	SOE-PCR
gCHAP_L35-L-R	CAGTAGAAGAAGAAGTACCATGGATGAATGCATAGC	for linker35
gCHAP_L38-L-F	GCTATGCATTCATCCATATGGACAAGGGTACTTCTTC	SOE-PCR
gCHAP_L38-L-R	GAAGAAGTACCCTTGTCATATGGATGAATGCATAGC	for linker38

*Underlined letters indicate the sequence of gCHAP fragments.

Table S5. PCR primers used for amplification 8 SH3 domain linker

Primer name	Sequence (5' -3')	Usage
ALS2-dA-ClyC _{SH3} -L-F	<u>GAAAAACCAGTACGGTACTT</u> GGTACAAACCGGAATCG	SOE-PCR for ALS2-dA-ClyC _{SH3}
ALS2-dA-ClyC _{SH3} -L-R	CGATTCCGGTTTGTACCA <u>AGTACCGTACTGGTTTTTC</u>	
ALS2-dA-vB _{SH3} -L-F	<u>GAAAAACCAGTACGGTACTT</u> GGTACAAACCGGAAAAACG	SOE-PCR for ALS2-dA-vB _{SH3}
ALS2-dA-vB _{SH3} -L-R	CGTTTTCCGGTTTGTACCA <u>AGTACCGTACTGGTTTTTC</u>	
ALS2-dA-tras14 _{SH3} -L-F	<u>GAAAAACCAGTACGGTACTT</u> ACTACATGGAAGAAAGC	SOE-PCR for ALS2-dA- tras14 _{SH3}
ALS2-dA-tras14 _{SH3} -L-R	GCTTTCTTCCATGTAGTA <u>AGTACCGTACTGGTTTTTC</u>	
ALS2-dA-ClyO _{SH3} -L-F	<u>GAAAAACCAGTACGGTACTC</u> GTTTCCTATCGCGAGAC	SOE-PCR for ALS2-dA-ClyO _{SH3}
ALS2-dA-ClyO _{SH3} -L-R	GTCTCGCGATAGGAACGAGTACCGTACTGGTTTTTC	
ALS2-dA-Lsp _{SH3} -L-F	<u>GAAAAACCAGTACGGTACTCT</u> GTATAAATCTGAATCTG	SOE-PCR for ALS2-dA-Lsp _{SH3}
ALS2-dA-Lsp _{SH3} -L-R	CAGATTCAGATTTATACAGAGTACCGTACTGGTTTTTC	
ALS2-dA-2638A _{SH3} -L-F	<u>GAAAAACCAGTACGGTACTT</u> GGTATAAAGCTGAACATG	SOE-PCR for ALS2-dA- 2638A _{SH3}
ALS2-dA-2638A _{SH3} -L-R	CATGTTTCAGCTTTATACCA <u>AGTACCGTACTGGTTTTTC</u>	
ALS2-dA-2958 _{SH3} -L-F	<u>GAAAAACCAGTACGGTACTG</u> GAAATTATACCGTCGC	SOE-PCR for ALS2-dA-2958 _{SH3}
ALS2-dA-2958 _{SH3} -L-R	GCGACGGTATAATTTC <u>AGTACCGTACTGGTTTTTC</u>	

*Underlined letters indicate the sequence of gCHAP fragments.

Table S6. PCR primers used for amplification of various chimeric lysins

Primer name	Sequence (5' -3')	Usage
P_Lsp-L-F	CTTTAAGAAGGAGATATACATatgGCGGCAACGCATGAAC	SOE-PCR for Lysostaphin
P_Lsp-L-R	GTTTCATGCGTTGCCGCcatATGTATATCTCCTTCTTAAAG	
Lsp_T-L-F	CTGTGGGGTACTATCAAAAAGGGTCATCATCACCATC	SOE-PCR for Lysostaphin & Lsp_ClyC _{SH3}
Lsp_T-L-R	GATGGTGATGATGACCCTTTTTGATAGTACCCACAG	
P_ClyC-L-F	CTTTAAGAAGGAGATATACATATGCAAGCAAACTAAC	SOE-PCR for ClyC
P_ClyC-L-R	GTTAGTTTTGCTTGCATATGTATATCTCCTTCTTAAAG	
ClyC_T-L-F	CCTGGGGAACATTCAAGAAGGGTCATCATCACCATC	
ClyC_T-L-R	GATGGTGATGATGACCCTTCTTGAATGTTCCCCAGG	
P_ClyO-L-F	CTTTAAGAAGGAGATATACATATGGCACTGCCTAAAACGG	SOE-PCR for ClyO
P_ClyO-L-R	CCGTTTTAGGCAGTGCCATATGTATATCTCCTTCTTAAAG	
ClyO_T-L-F	CTTGGGGTACATTTAAAAAGGGTCATCATCACCATC	
ClyO_T-L-R	GATGGTGATGATGACCCTTTTTAAATGTACCCCAAG	
Lsp_ClyC-L-F	GAAAACTAACAAATATGGTACTTGGTACAAACCGGAATCG	SOE-PCR for Lsp_ClyC _{SH3}
Lsp_ClyC-L-R	CGATTCCGGTTTGTACCAAGTACCATATTTGTTAGTTTTTC	

*small case letters indicate nucleotides added artificially.

Table S7. PCR primers used for amplification of pET101/D/TOPO vector

Primer name	Sequence (5' -3')	Usage
gCHAP_CACC-F	caccATGGCATCATCTTTTAATC	ALS2-dA-L25 amplification
K _{SH3} -R	TTTGAAAACACCCCATGCAAC	ALS2-dA-L25 amplification
ClyC _{SH3} -R	CTTGAATGTTCCCCAGGCAAC	ALS2-dA-ClyC _{SH3} amplification

*small case letters indicate nucleotides added artificially.

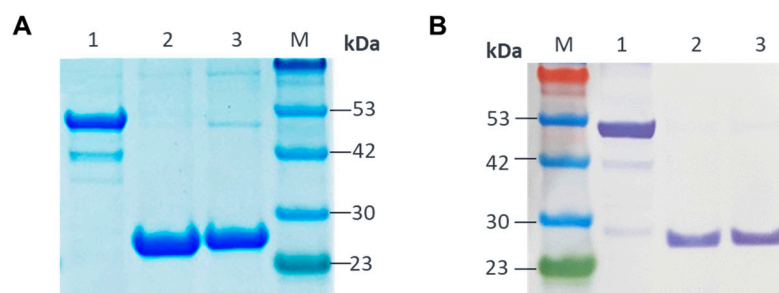


Figure S1. SDS-PAGE and Western blotting of *E. coli* expressed chimeric lysins. **(A)** SDS-PAGE analysis of purified protein using 4–12% polyacrylamide gel. Lane M: protein molecular weight marker; Lane 1: purified protein of ALS2; Lane 2: purified protein of ALS2-dA-L25; Lane 3: ALS2-dA-ClyC_{SH3}. For SDS-PAGE analysis, 5 μ L of each purified protein, **(B)** Western blot analysis of expressed protein by six histidine antibody. The molecular weight of the protein inserted into the pET/D/101 vector is approximately 52.4 kDa and 28.31 kDa. Lane M: protein molecular weight marker (PM2700 ExcelBand™ 3-color Broad Range Protein Marker); Lane 1: Protein expressed ALS2; Lane 2: Protein expressed ALS2-dA-L25; Lane 3: Protein expressed ALS2-dA-ClyC_{SH3}. For western blot analysis, 20 μ L of each induced protein.

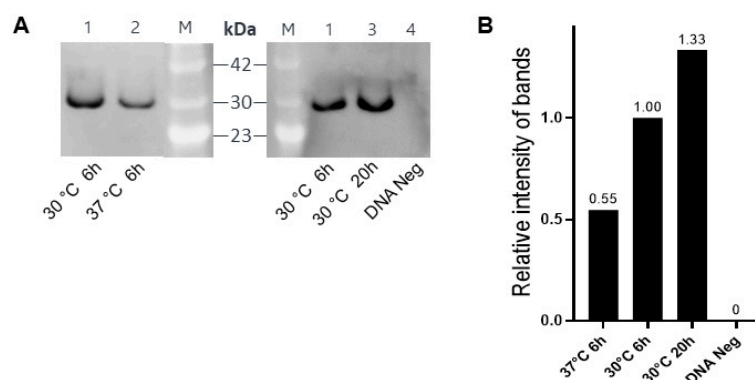


Figure S2. Comparison of expression levels of Lsp-ClyC_{SH3} in different temperature and incubation times. **(A)** Western blotting of Lsp-ClyC_{SH3} using anti-histidine antibody and ECL. Lane M: protein molecular weight marker; Lane 1: Expressed at 30°C for 6h; Lane 2: Expressed at 37°C for 6h; Lane 3: Expressed at 30°C for 20h; Lane 4: DNA-negative sample. **(B)** The relative intensity of the Lsp-ClyC_{SH3} protein bands. The ratio of protein expression compared to Lsp-ClyC_{SH3} expressed at 30°C for 6h was measured using Image J 1.53