

Table S1: Translation fidelity in the control and Δ PSL strains

uL5 variant	<i>lacZ</i> variant	LacZ activity, Miller units	Standard deviation
wild type	WT	13943	± 455
wild type	+1 frameshift	16.2	± 1.8
wild type	-1 frameshift	3.0	± 0.2
wild type	Premature UAA	0.5	± 0.0
wild type	Premature UAG	6.9	± 0.5
wild type	Premature UGA	8.9	± 0.3
Δ PSL	WT	18703	± 2303
Δ PSL	+1 frameshift	19.4	± 0.6
Δ PSL	-1 frameshift	8.4	± 0.5
Δ PSL	Premature UAA	2.1	± 0.1
Δ PSL	Premature UAG	63.4	± 10.2
Δ PSL	Premature UGA	46.8	± 1.9

Table S2: Oligonucleotides used in the work

Name	Usage	Sequence	Source
AK07	The <i>lac</i> fusion check/sequencing	5'- GTGAACGCTCTCCTGAGTAGG	[30]
AK08b	The <i>lac</i> fusion check/sequencing	5'- CATTGCGCCATTCAGGCTGCGCAAC	[30]
AK45	Constructing <i>lac</i> fusions	5'-GACGCCCCGCATAAACTGCCAGGAATTAAGAGACGTTCTGAAATGAGCTGTTGACAATTAATCATCC	This work
AK46	Constructing <i>lac</i> fusions	5'-GACACTTTACCTCATAGCTGGCATAATCCACACATTATACGAGCCGGATGATTAATTGTCAACAGC	This work
AK47	Constructing <i>lac</i> fusions	5'- AGTCACGACGTTGTAAAACGACGGCCAGTGAATCCGTAATAGCCATTTGTGCAGCGAGTTC	This work
AK226	Constructing <i>lac</i> fusions	5'-GCTTAGCGTAATCATGGTGAGTTGAGCAGTAGATTTACG	This work
L5delta73-80F	73-80 deletion	5'-CACCAAAGCACGCAAAACAGGGCTATCCGATC	This work
L5delta73-80R	73-80 deletion	5'-GATCGGATAGCCCTGTTTGCCTGCTTTGGTG	This work
rplE34cat-F1	<i>rplE</i> knockout, forward	5'-CTACAATTCTGTCATGCAAGTCCCTCGGGTCGAGAAGATCATGGAGAAAAAATCACTGGAT	This work
rplEcat-R1	<i>rplE</i> knockout, reverse	5'-CTTTTCATTGATTGCTTAGCCATTTAGTAACCCTACCTTACGCCCGCCCTGCCACTC	This work
rplE-check-F	knockout verification, sequencing	5'-TGTAGGCTTTAGATTGAAGACGG	[2]
rplE-check-R	knockout verification, sequencing	5'-CGCTTTCAGTTCAGCGCGTTTCGC	[2]
rplE_rec_F	Restoring chromosomal <i>rplE</i>	5'-ATGGCGAAACTGCATGAT	This work
rplE_rec_R	Restoring chromosomal <i>rplE</i>	5'-CGGTGGTATATCCAGTGATTTTTTCTCCATGATCTTCTCCACTTACTTGCGGAACGGGAAG	This work

Table S3: *Escherichia coli* strains and plasmids used in the work

Name	Genotype/description	Reference/source
Strains		
OK391a	MS02 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -1 (wt)	this work
OK391b	MS02 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -2 (+1 frameshift)	this work
OK391c	MS02 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -3 (-1 frameshift)	this work
OK391d	MS02 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -4 (premature UAA)	this work
OK391e	MS02 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -5 (premature UAG)	this work
OK391f	MS02 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -6 (premature UGA)	this work
OK392a	MG1655 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -1 (wt)	this work
OK392b	MG1655 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -2 (+1 frameshift)	this work
OK392c	MG1655 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -3 (-1 frameshift)	this work
OK392d	MG1655 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -4 (premature UAA)	this work
OK392e	MG1655 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -5 (premature UAG)	this work
OK392f	MG1655 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZ</i> -6 (premature UGA)	this work
OK404a	OK392a <i>rplE1</i> :: <i>cat</i> (wild type uL5)	this work
OK404b	OK392b <i>rplE1</i> :: <i>cat</i> (wild type uL5)	this work
OK404c	OK392c <i>rplE1</i> :: <i>cat</i> (wild type uL5)	this work
OK404d	OK392d <i>rplE1</i> :: <i>cat</i> (wild type uL5)	this work
OK404e	OK392e <i>rplE1</i> :: <i>cat</i> (wild type uL5)	this work
OK404f	OK392f <i>rplE1</i> :: <i>cat</i> (wild type uL5)	this work
OK406a	OK392a <i>rplEΔ73-80</i> :: <i>cat</i> (uL5ΔPSL)	this work
OK406b	OK392b <i>rplEΔ73-80</i> :: <i>cat</i> (uL5ΔPSL)	this work
OK406c	OK392c <i>rplEΔ73-80</i> :: <i>cat</i> (uL5ΔPSL)	this work
OK406d	OK392d <i>rplEΔ73-80</i> :: <i>cat</i> (uL5ΔPSL)	this work
OK406e	OK392e <i>rplEΔ73-80</i> :: <i>cat</i> (uL5ΔPSL)	this work
OK406f	OK392f <i>rplEΔ73-80</i> :: <i>cat</i> (uL5ΔPSL)	this work
MG1655	<i>rph-1</i>	wild type <i>E. coli</i> [28]
MS02	<i>gal490 ldcI857 Δ(cro-bioA) lacI'</i> :: <i>bla-cat-sacB-lacZYA</i>	[30]
MS45	MS02 <i>lacI'</i> :: <i>bla-Ptrc-g32-lacZYA</i>	this work
MS45a	MS02 <i>lacI'</i> :: <i>bla-Ptrc</i> *- <i>g32-lacZYA</i> (G insertion into the -35 promoter sequence)	this work
MS128/pNK12	NM300 <i>ΔrplE34</i> :: <i>cat</i> (transformed with the complementing plasmid pNK12)	this work
MS129	MG1655 <i>rplE1</i> :: <i>cat</i> (wild type uL5)	this work

MS129a	MG1655 <i>rplE</i> Δ73-80:: <i>cat</i> (uL5ΔPSL)	this work
NM300	MG1655 <i>lacX74 mini-λ-Tet</i>	N. Majdalani (NIH)
Plasmids		
pACYC184	cloning vector, Tet ^R , Cam ^R	NEB, [26]
pNK12	arabinose-induced expression vector with cloned wild type <i>rplE</i> ORF, Amp ^R	[3]
pNK14	same as pNK12 with mutated <i>rplE</i> ORF encoding uL5ΔPSL	this work
pSG25	<i>lacZ</i> (wild type)	[7]
pSGlac7	<i>lacZ</i> (+1 frameshift)	[7]
pSG12DP	<i>lacZ</i> (-1 frameshift)	[7]
pSG853	<i>lacZ</i> (premature UAA stop codon)	[7]
pSG163	<i>lacZ</i> (premature UAG stop codon)	[7]
pSG3/4	<i>lacZ</i> (premature UGA stop codon)	[7]
