

Herpes Simplex Virus Infection Alters the Immunological Properties of Adipose-Tissue-Derived Mesenchymal Stem-like Cells

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Supplementary Information

Materials and methods

Differentiation

To do the trilineage differentiation, after the trypsinization of the cells, they were counted using EVE automatic cell counter. NanoEntek (NanoEntek, Seoul, Korea) and 50.000 cells were seeded in 24-well plates. The isolated cells were applied to commercially available Gibco's StemPro® Adipogenesis, Osteogenesis, and Chondrogenesis Differentiation Kits according to the manufacturer's guidelines (Gibco). After 24 hours, the medium was replaced with a differentiation medium. After 21 days, cells were fixed with 4% methanol-free formaldehyde (Molar Chemicals, Hungary) for 20 min at RT. In the differentiated adipocytes, Nile red staining was used to detect the lipid-laden particles. Alizarin red staining was applied to quantify the mineral deposits during osteogenesis. And Toluidine blue staining was used to label the chondrogenic mass formed by AD-MSC.

RNA-seq analysis

enrichplot: Yu G (2022). *enrichplot: Visualization of Functional Enrichment Result*. R package version 1.16.1. <https://yulab-smu.top/biomedical-knowledge-mining-book/>.

GOSemSim:

- Guangchuang Yu. Gene Ontology Semantic Similarity Analysis Using GOSemSim. In: Kidder B. (eds) Stem Cell Transcriptional Networks. Methods in Molecular Biology. 2020. 2117:207-215. Humana, New York, NY. doi:10.1007/978-1-0716-0301-7_11

- Guangchuang Yu, Fei Li, Yide Qin, Xiaochen Bo, Yibo Wu, Shengqi Wang. GOSemSim: an R package for measuring semantic similarity among GO terms and gene products Bioinformatics 2010. 26(7):976-978. doi:10.1093/bioinformatics/btq064

rrvgo:

Sayols S (2020). *rrvgo: a Bioconductor package to reduce and visualize Gene Ontology terms*. <https://ssayols.github.io/rrvgo>.

EnhancedVolcano:

Blighe K. Rana S. Lewis M (2022). *EnhancedVolcano: Publication-ready volcano plots with enhanced coloring and labeling*. R package version 1.14.0

Donor	Age	Sex (M/F)
1	40	M
2	35	M
3	34	M
4	47	M
5	51	F
6	53	F
7	42	F
8	25	M
9	65	F
10	36	M
11	46	F
12	53	M
13	41	F
14	39	F

Table S1. Details of donors used in this study.

Surface marker	Mean (% of positive cells)	SEM (+/-)
CD6	2.58	0.88
CD29	98.74	0.25
CD31	0.05	0.03
CD34	0.00	0.00
CD40	3.14	2.35
CD44	90.39	2.15
CD47	95.79	0.64
CD49A	93.61	1.51
CD49D	29.04	13.51
CD50	8.31	7.27
CD51	87.12	2.49
CD54	17.49	6.22
CD61	38.45	15.59
CD73	98.80	0.19
CD90	92.45	0.89
CD105	90.59	3.09
CD106	0.47	0.43
CD112	89.39	1.90
CD144	49.02	12.66
CD146	1.82	1.11
CD166	96.22	0.44
CD271	14.87	10.84
CD304	0.85	0.75
CD325	26.81	12.96
HLA-DR	0.69	0.24

Table S2. Surface marker pattern of in vitro cultured ADMSCs.

Multicolor flow cytometry carried out surface markers' profiling of the in vitro cultured ADCMSCs. Cells showed high expression of well-known MSC markers such as CD29. CD44. CD73. CD90. and CD105 (Data shown are mean \pm SEM, N=14).

Top Tox Functions			
Strain	Name	p-value range	# Molecules
Assays: Clinical Chemistry and Hematology			
HSV-1 KOS	Increased Levels of Hematocrit	5.46×10^{-06} - 5.46×10^{-06}	33
	Increased Levels of Red Blood Cells	4.79×10^{-01} - 1.86×10^{-04}	33
	Decreased Levels of Albumin	6.24×10^{-01} - 1.50×10^{-01}	7
	Increased Levels of Albumin	5.58×10^{-01} - 1.50×10^{-01}	4
	Increased Levels of Alkaline Phosphatase	2.01×10^{-01} - 1.53×10^{-01}	17
HSV-1- 532	Increased Levels of Hematocrit	5.24×10^{-03} - 5.24×10^{-03}	13
	Increased Levels of Red Blood Cells	2.13×10^{-01} - 3.96×10^{-02}	12
	Decreased Levels of Albumin	3.02×10^{-01} - 5.82×10^{-02}	1
	Increased Levels of Albumin	5.13×10^{-01} - 5.82×10^{-02}	2
	Increased Levels of Alkaline Phosphatase	2.59×10^{-01} - 1.33×10^{-01}	8
HSV-2	Increased Levels of ALT	9.26×10^{-02} - 9.26×10^{-02}	1
	Increased Levels of Alkaline Phosphatase	2.26×10^{-01} - 2.26×10^{-01}	1
Cardiotoxicity			
HSV-1 KOS	Cardiac Enlargement	1.00×10^{00} - 1.08×10^{-03}	159
	Cardiac Infarction	1.00×10^{00} - 1.29×10^{-03}	79
	Congenital Heart Anomaly	1.00×10^{00} - 3.85×10^{-03}	83
	Cardiac Hypoplasia	4.79×10^{-01} - 5.66×10^{-03}	17
	Cardiac Necrosis/Cell Death	5.25×10^{-01} - 1.21×10^{-02}	65
HSV-1- 532	Cardiac Enlargement	1.00×10^{00} - 3.38×10^{-03}	55
	Cardiac Necrosis/Cell Death	1.00×10^{00} - 9.75×10^{-03}	24
	Cardiac Dilation	1.00×10^{00} - 1.45×10^{-02}	26
	Cardiac Inflammation	1.00×10^{00} - 1.56×10^{-02}	12
	Cardiac Proliferation	2.82×10^{-01} - 1.68×10^{-02}	17
HSV-2	Cardiac Dilation	4.74×10^{-01} - 3.23×10^{-03}	3
	Cardiac Enlargement	4.74×10^{-01} - 3.23×10^{-03}	4
	Cardiac Proliferation	7.54×10^{-02} - 1.67×10^{-02}	3
	Cardiac Dysfunction	2.56×10^{-02} - 2.56×10^{-02}	1
	Cardiac Stenosis	1.77×10^{-01} - 3.18×10^{-02}	1
Hepatotoxicity			
HSV-1 KOS	Liver Hyperplasia/Hyperproliferation	1.00×10^{00} - 1.69×10^{-37}	1617
	Hepatocellular carcinoma	5.10×10^{-01} - 2.11×10^{-14}	489
	Liver Necrosis/Cell Death	4.79×10^{-01} - 1.19×10^{-04}	78
	Liver Inflammation/Hepatitis	1.00×10^{00} - 1.22×10^{-04}	116
	Liver Fibrosis	1.00×10^{00} - 1.76×10^{-04}	107
HSV-1- 532	Liver Hyperplasia/Hyperproliferation	1.00×10^{00} - 1.55×10^{-16}	638
	Hepatocellular carcinoma	1.00×10^{00} - 4.77×10^{-05}	183
	Liver Inflammation/Hepatitis	1.00×10^{00} - 6.83×10^{-05}	44
	Liver Steatosis	1.00×10^{00} - 6.83×10^{-05}	50
	Liver Necrosis/Cell Death	3.47×10^{-01} - 1.28×10^{-04}	36
HSV-2	Liver Inflammation/Hepatitis	2.18×10^{-01} - 1.25×10^{-08}	11
	Liver Steatosis	2.18×10^{-01} - 1.25×10^{-08}	10
	Liver Necrosis/Cell Death	1.13×10^{-01} - 1.98×10^{-03}	5
	Liver Damage	1.08×10^{-01} - 2.24×10^{-03}	5
	Liver Failure	3.23×10^{-03} - 3.23×10^{-03}	1
Nephrotoxicity			
HSV-1 KOS	Renal Necrosis/Cell Death	1.00×10^{00} - 7.51×10^{-06}	152
	Renal Proliferation	1.00×10^{00} - 2.94×10^{-05}	80
	Glomerular Injury	1.00×10^{00} - 4.22×10^{-04}	125
	Renal Inflammation	1.00×10^{00} - 4.22×10^{-04}	87
	Renal Nephritis	1.00×10^{00} - 4.22×10^{-04}	87
HSV-1- 532	Glomerular Injury	1.00×10^{00} - 1.96×10^{-04}	58
	Renal Inflammation	4.51×10^{-01} - 1.96×10^{-04}	43

	Renal Nephritis	4.51×10^{-01} - 1.96×10^{-04}	43
	Kidney Failure	1.00×10^{00} - 5.98×10^{-04}	28
	Renal Fibrosis	1.65×10^{-01} - 5.98×10^{-04}	19
HSV-2	Glomerular Injury	5.58×10^{-01} - 7.18×10^{-06}	9
	Renal Inflammation	3.18×10^{-02} - 7.18×10^{-06}	9
	Renal Nephritis	3.18×10^{-02} - 7.18×10^{-06}	9
	Renal Damage	6.87×10^{-02} - 7.54×10^{-04}	3
	Renal Tubule Injury	6.87×10^{-02} - 7.54×10^{-04}	3

Table S3. Top TOX functions determined by IPA

Top Regulator Effect Networks				
Strain	ID	Regulators	Disease & Functions	Consistency Score
HSV1-KOS	1	SPDEF	Migration of cells	3.5
	2	Eldr	Cell proliferation of tumor cell lines	3.464
	3	LARP1	Cell death of tumor	3.328
	4	LARP1	Cell death of tumor cells	3.328
	5	LARP1	Necrosis of tumor	3.328
HSV1- 532	1	poly rI:rC-RNA	Replication of Herpesviridae	3.474
	2	IRF3	Replication of Herpesviridae	3.317
	3	IFNA2	Replication of Herpesviridae	3.175
	4	IRF7	Transactivation	3.162
	5	IRF7	Transactivation of RNA	3.162
HSV-2	1	SP110	Viral Infection	3.015
	2	IFNG	Cell death of epithelial cells	3.0
	3	IRF3	Cell death of epithelial cells	3.0
	4	Interferon alpha	Cell death of epithelial cells	3.0
	5	NONO	Cell death of epithelial cells	3.0
Top Networks				
HSV1-KOS	ID	Associated Network Functions		Score
	1	Cell Death and Survival. Cardiovascular Disease. Cellular Movement		35
	2	Cell Morphology. Cellular Function. and Maintenance. Metabolic Disease		35
	3	Cancer. Cell Death. and Survival. Organismal Injury. and Abnormalities		35
	4	Post-Translational Modification. Protein Folding. Developmental Disorder		35
	5	Cell-To-Cell Signaling and Interaction. Cellular Assembly and Organization. Tissue Development		33
HSV1- 532	1	Cellular Assembly and Organization. DNA Replication. Recombination. and Repair. Cellular Function and Maintenance		50
	2	Cellular Assembly and Organization. Cellular Function. and Maintenance. Molecular Transport		47
	3	RNA Damage and Repair. Cellular Function and Maintenance. Dermatological Diseases. and Conditions		47
	4	RNA Post- Transcriptional Modification. Cancer. DNA Replication. Recombination. and Repair		44
	5	RNA Post- Transcriptional Modification. Endocrine System Disorders. Hereditary Disorder		44
HSV2	1	Immunological Disease. Inflammatory Disease. Organismal Injury. and Abnormalities		45
	2	Antimicrobial Response. Inflammatory Response. Cell Signaling		36
	3	Antimicrobial Response. Inflammatory Response. Hereditary Disorder		34
	4	Immunological Disease. Inflammatory Disease. Organismal Injury. and Abnormalities		28
	5	Cell-To-Cell Signaling and Interaction. Cellular Growth and Proliferation. Connective Tissue Development. and Function		19
Top Tox Lists				
HSV1-KOS	Name		p-value	Overlap
	Mitochondrial Dysfunction		6.00x10 ⁻¹²	36.3 % 62/171
	Renal Necrosis/Cell Death		2.30x10 ⁻⁰⁸	23.1 % 152/659
	Liver Necrosis/Cell Death		1.39x10 ⁻⁰⁴	22.5 % 78/347

HSV1- 532	Recovery from Ischemic Acute Renal Failure (Rat)	3.33x10 ⁻⁰⁴	57.1 % 8/14
	Aryl Hydrocarbon Receptor Signaling	3.80x10 ⁻⁰⁴	25.5 % 41/161
	Renal Necrosis/Cell Death	1.34x10 ⁻⁰⁴	9.4 % 62/659
	Liver Necrosis/Cell Death	5.75x10 ⁻⁰⁴	10.4 % 36/347
	Mitochondrial Dysfunction	9.76x10 ⁻⁰⁴	12.3 % 21/171
	Recovery from Ischemic Acute Renal Failure (Rat)	7.12x10 ⁻⁰³	28.6 % 4/14
HSV2	Increases Cardiac Proliferation	1.10x10 ⁻⁰²	12.8 % 11/86
	Increases Renal Nephritis	6.23x10 ⁻⁰⁵	6.1 % (4/66)
	Increases Liver Hepatitis	3.36x10 ⁻⁰⁴	3.9 % (4/102)
	Increases Depolarization of Mitochondria and Mitochondrial Membrane	1.52x10 ⁻⁰³	11.1 % (2/18)
	Liver Necrosis/Cell Death	5.33x10 ⁻⁰³	1.4 % (5/347)
	Increases Glomerular Injury	1.75x10 ⁻⁰²	1.8 % (3/169)
Top Analysis-Ready Molecules			
Expr Log Ratio			
HSV1-KOS	Molecules	Expr. Value	Chart
	CH507_528H121	18.890	↑
	CXCL11	10.522	↑
	RP3_453C1214	9.921	↑
	PRAMEF1 (includes others)	9.282	↑
	CCDC194	9.279	↑
	SOX3	9.120	↑
	ZSCAN4	8.982	↑
	CXCL10	8.889	↑
	DIO3	8.811	↑
	RP11_235E173	8.755	↑
	TMEM205	-5.128	↓
	GPR68	-4.857	↓
	SCARA3	-4.829	↓
	GREM2	-4.816	↓
	ISLR	-4.570	↓
	PSAT1	-4.459	↓
	EMILIN1	-4.235	↓
	ZER1	-4.177	↓
	SIL1	-4.170	↓
	MORN2	-4.139	↓
HSV1-532	CH507_528H121	20.970	↑
	CXCL11	10.067	↑
	CXCL10	8.526	↑
	OASL	8.087	↑
	CCDC194	7.748	↑
	UNCX	7.579	↑
	RP3_453C1214	7.339	↑
	DIO3	7.231	↑
	USP18	6.974	↑
	RSAD2	6.936	↑
	BRD3OS	-2.800	↓
	GREM2	-2.784	↓
	TMEM37	-2.295	↓
	CDK5	-2.286	↓
	RPS6KA2	-2.171	↓
	ITGB1BP1	-2.154	↓
	PTPN9	-2.059	↓
	TMEM35A	-2.047	↓
	GSTA4	-2.044	↓












	PLPP3	-2.013	
HSV2	CXCL11	9.671	
	CH507_528H121	9.041	
	CXCL10	7.910	
	OASL	7.060	
	IDO1	6.643	
	MX1	6.268	
	CMPK2	6.250	
	CCDC194	6.106	
	IFIT2	5.878	
	RSAD2	5.534	

Table S4. Top networks and analysis-ready molecules determined by IPA. Green arrows indicate downregulation and red arrows indicate upregulation of genes.

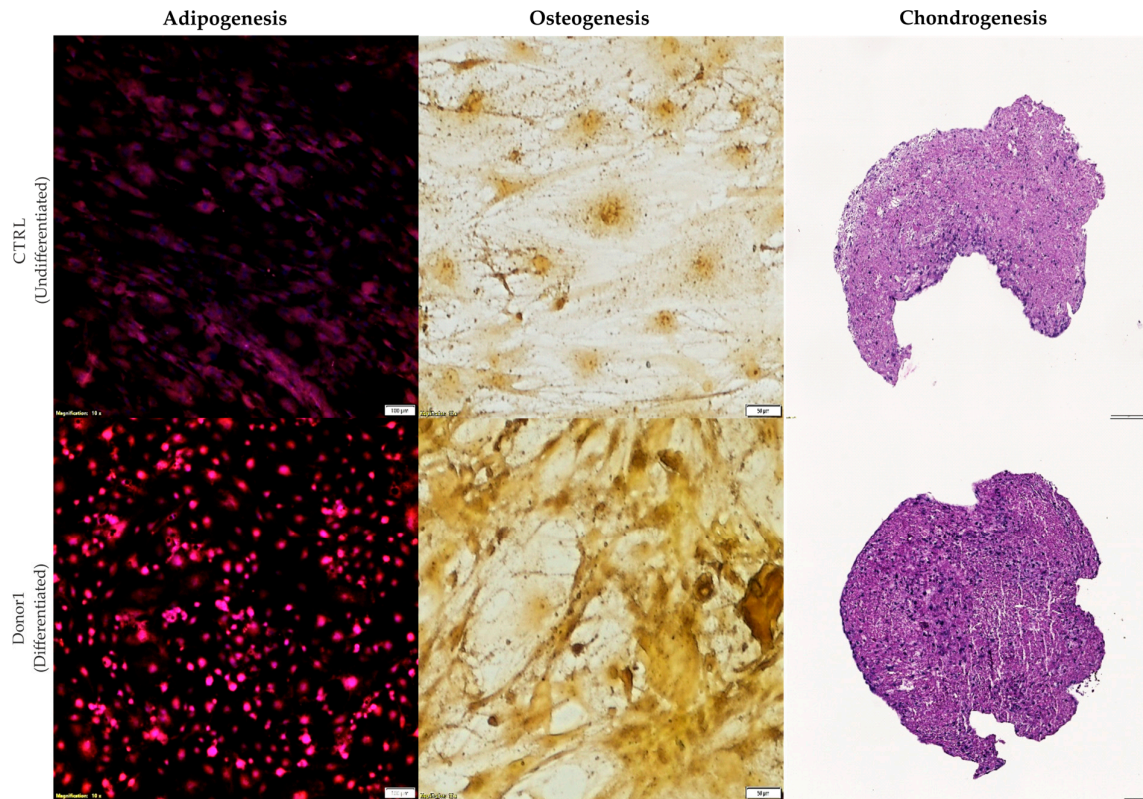
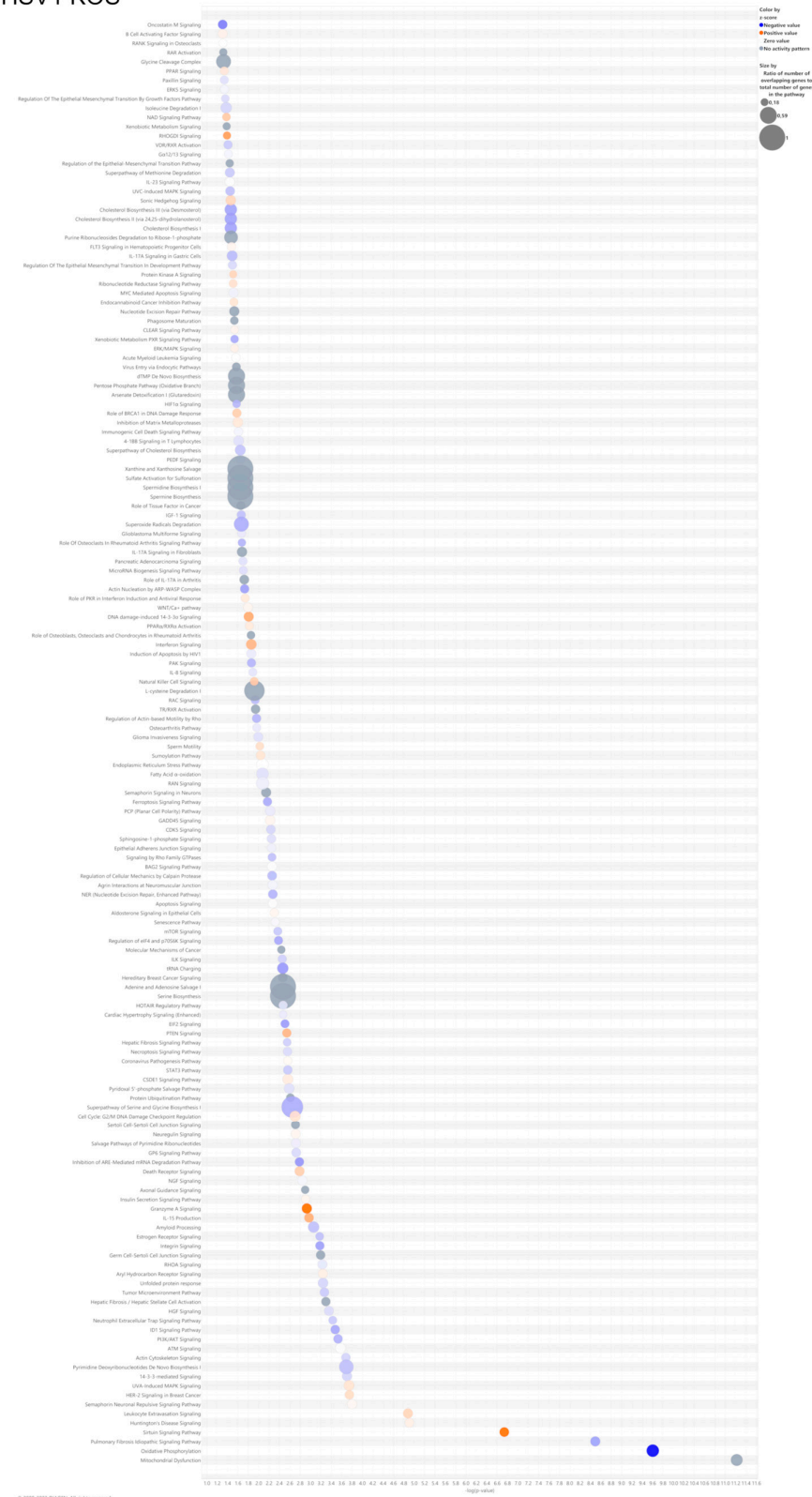


Figure S1. Validation of the in vitro differentiation potential of AD-MSC

MSCs isolated from adipose tissue were able to differentiate into adipose tissue in vitro, as revealed by Nile-red staining of lipid droplets in the cells. In the case of bone-directed differentiation, calcium deposits were detected by alizarin red staining. The extracellular matrix appearance characteristic of cartilage tissue was shown by metachromasia of toluidine blue staining. Results from a representative study are shown.(CT: control culture without differentiation cell culture media). Scale bar: For Adipogenesis 100um, for Osteogenesis 50 um and for chondrogenesis 200um.

HSV1 KOS



HSV1 532

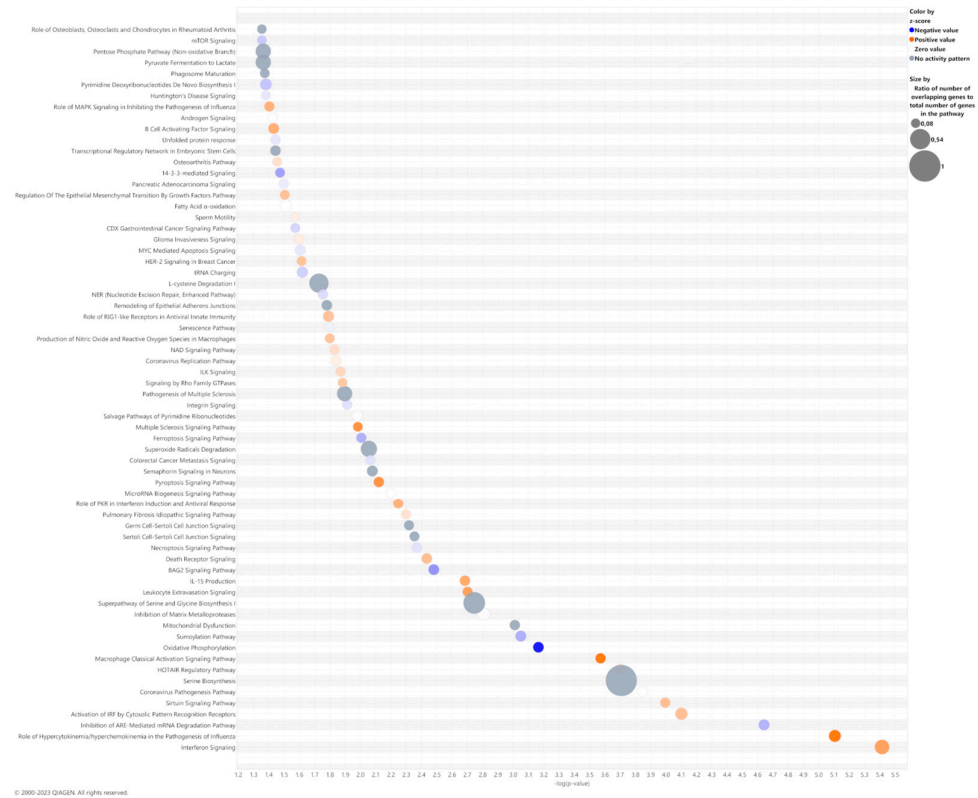


Figure S4. Canonical pathways in HSV-1 532 infected ADMSC
Bubble chart of canonical pathways initiated by HSV-1 532 infection in vitro. Pathways are determined by IPA.

HSV2

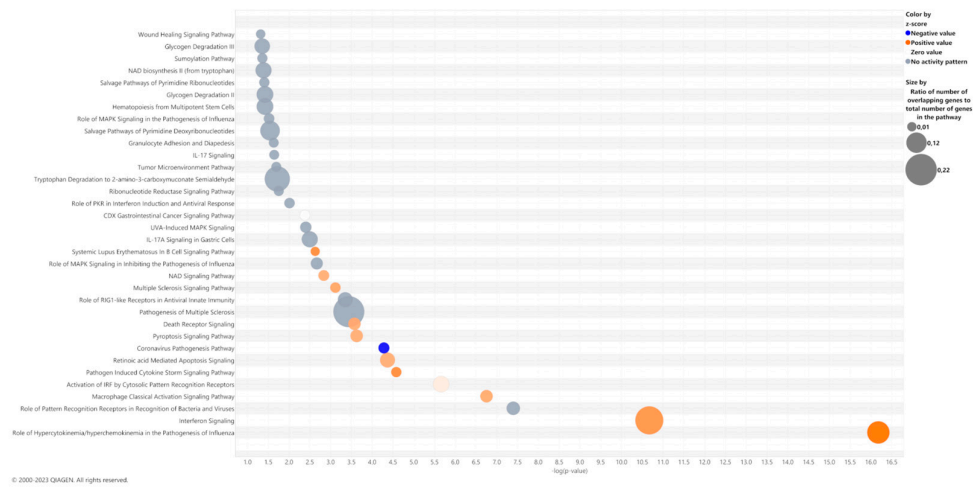


Figure S5. Canonical pathways in HSV-2 infected ADMSC
Bubble chart of canonical pathways initiated by HSV-2 infection in vitro. Pathways are determined by IPA.

HSV1 KOS

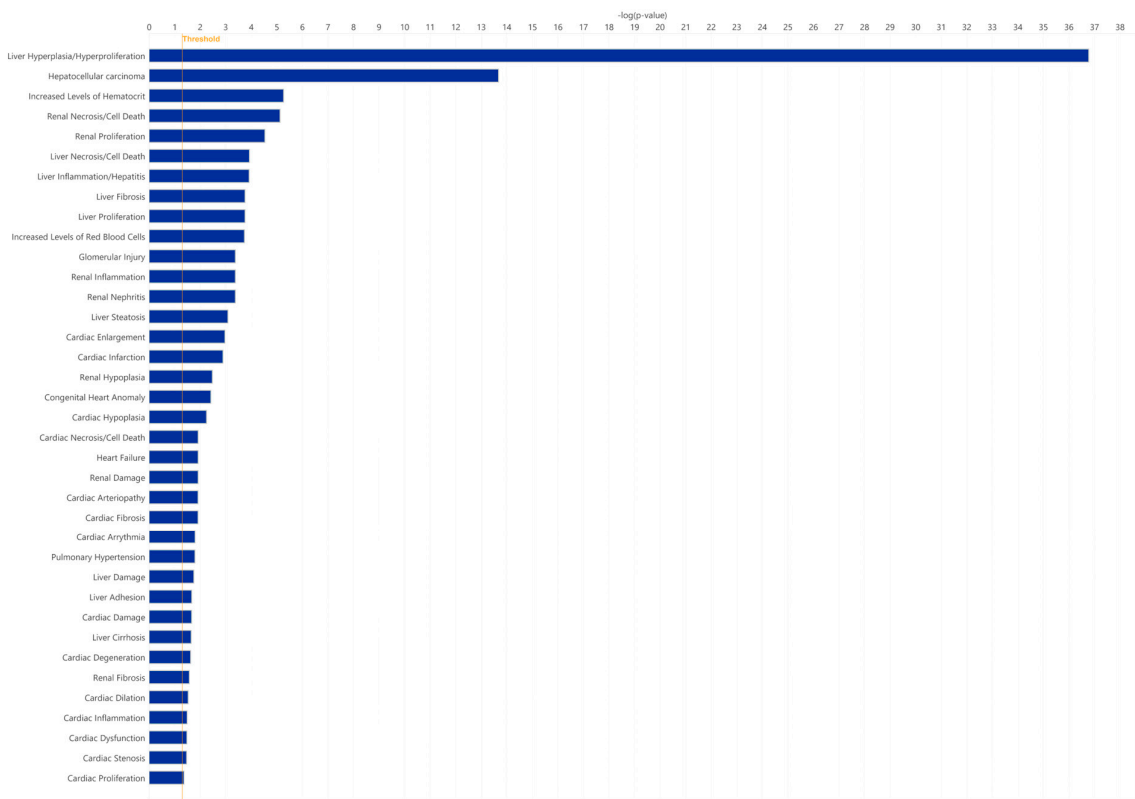
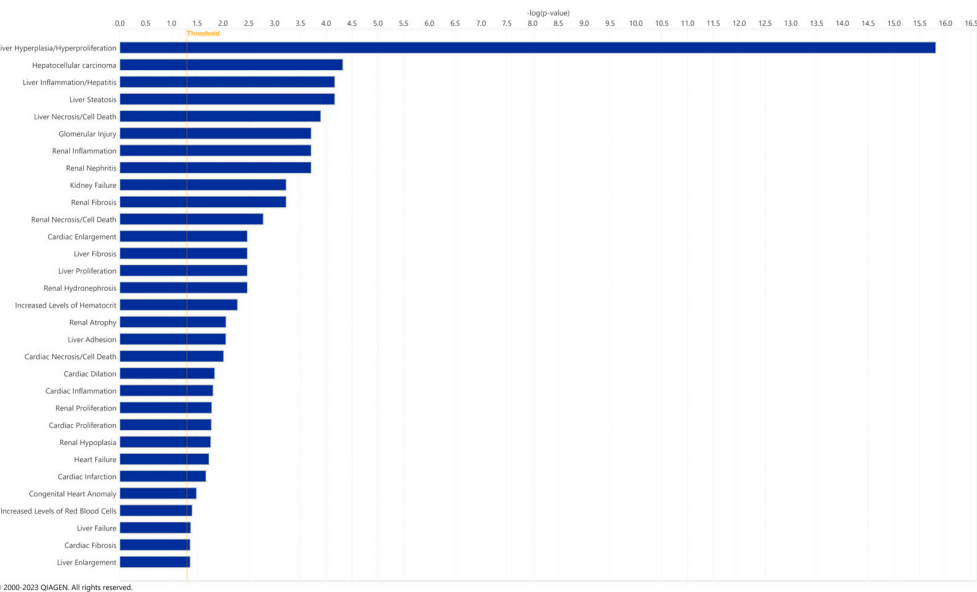


Figure S6. TOX pathways in HSV-1 KOS infected ADMSC
Bar chart of TOX pathways activated by in vitro HSV-1 KOS infection in AD-MSC. Pathways were determined by IPA.

HSV1 532



HSV2

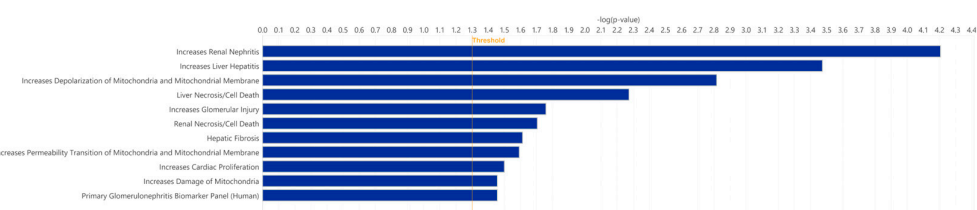
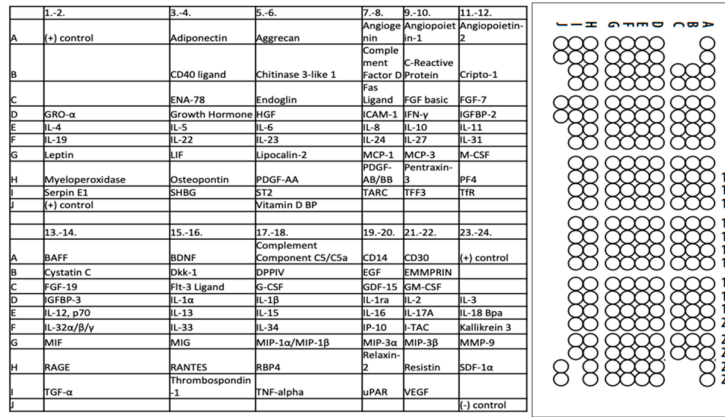
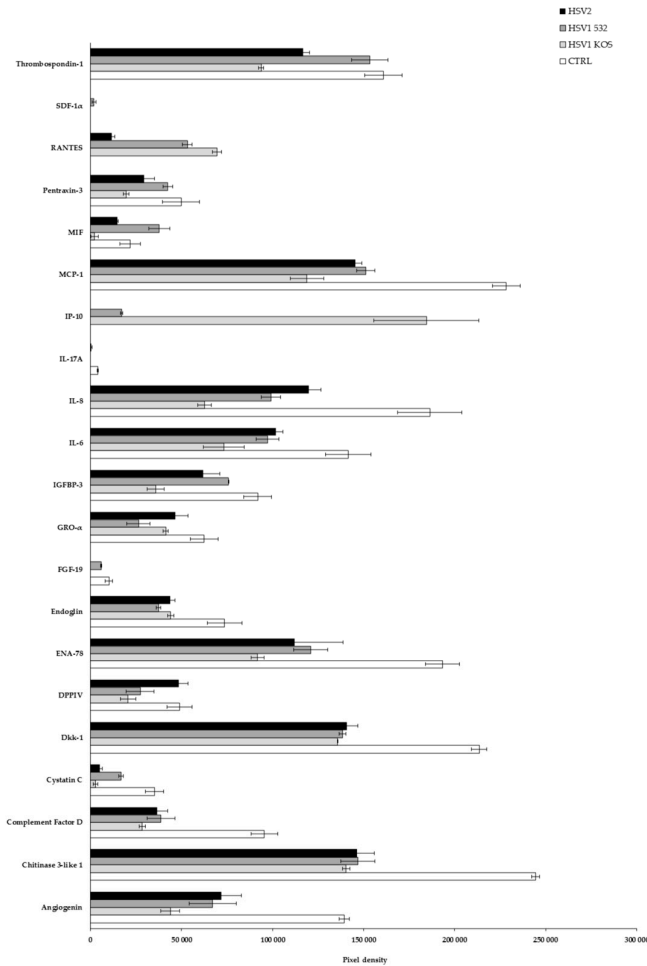


Figure S7. TOX pathways in HSV-1 532 and HSV-2 infected ADMSC
Bar chart of TOX pathways activated by in vitro HSV-1 532 or HSV-2 infection in AD-MSC. Pathways were determined by IPA.

A



B



C

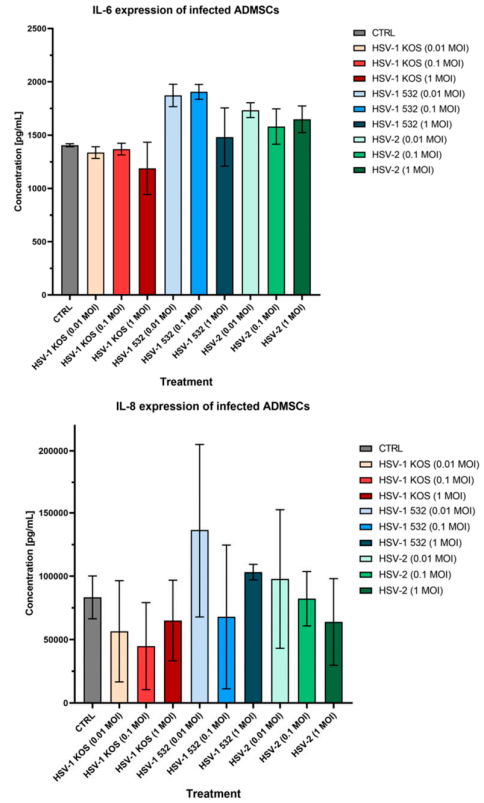


Figure S8. Expression of cytokines released by HSV-infected and uninfected AD-MSC

The setup of the Human XL Cytokine Proteome Profiler to identify targets in the cell culture supernatants (A). All spots were analyzed and semi-quantified by image analysis. (B) IL-6, IL-8 and IP-10 secretion were validated by ELISA. (N=3) (C)