



Supplementary Materials

Discovery of Chalcone-Based Hybrid Structures as High Affinity and Site-Specific Inhibitors against SARS-CoV-2: A Comprehensive Structural Analysis Based on Various Host-Based and Viral Targets

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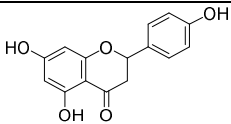
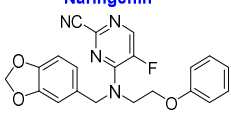
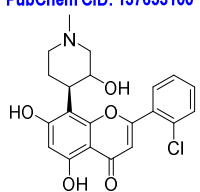
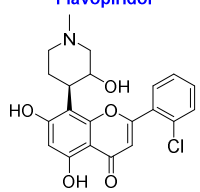
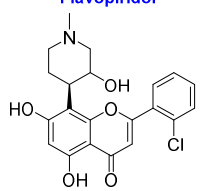
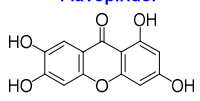
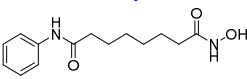
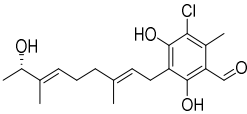
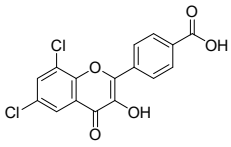
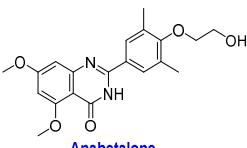
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Table S1. The most important host and virus-based targets selected to evaluate the antiviral activity of the 757 chalcone-based compound library, along with PDB codes, chemical structure of their co-crystal ligands, and the authoritative studies proposing these targets as promising strategies for the treatment of viral diseases such as COVID-19.

N.	HBATs	PDB Code	Co-crystallized ligand	HBATs-References
1	p38 MAPK	4EH3	 Naringenin	[1-8]
2	Cathepsin L	5MQY	 PubChem CID: 137653100	[9-16]
3	CDK1	6GU2	 Flavopiridol	[17-22]
4	CDK2/CyclinA	6GUB	 Flavopiridol	[23-25]
5	CDK9/cyclinT1	3BLR	 Flavopiridol	[26-29]
6	ERK2	3SA0	 Norathyriol	[30-34]
7	HDAC2	4LXZ	 Vorinostat	[35-39]
8	DHODH	5ZF7	 3vva	[40-46]
9	CK2 alpha'	5M4U	 PubChem CID: 38997316	[47-52]
10	BRD2	4J1P	 Apabetalone	[53-55]

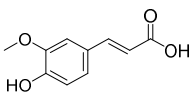
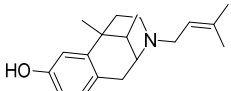
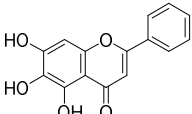
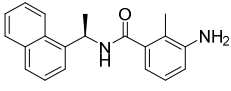
11	BRD4	6HOV	 Ferulic acid	[53, 55-57]
12	Sigma-1 receptor	6DK1	 Pentazocine	[58-64]
13	3CLpro	6M2N	 Baicalein	[65-68]
14	PLpro	7JN2	 PubChem CID: 153835436	[69-72]

Table S2. Some important druglikeness, ADME, and toxicity parameters of the best identified **CHA-12**, **CHA-37**, **CHA-378**, **CHA-384**, and standards quercetin and resveratrol predicted by ad-metSAR, preADME, and swissADME online servers.

Parameters	CHA-12	CHA-37	CHA-378	CHA-384	Quercetin	Resveratrol
Formula	C ₂₆ H ₁₉ N ₅ O ₃	C ₂₁ H ₁₅ N ₃ O	C ₂₂ H ₁₉ NO ₅ S	C ₂₃ H ₁₇ ClN ₂ O ₄	C ₁₅ H ₁₀ O ₇	C ₁₄ H ₁₂ O ₃
Molecular weight (g/mol)	449.46	325.36	409.45	420.85	302.24	228.24
Lipinski rule of Five	Suitable	Suitable	Suitable	Suitable	Suitable	Suitable
Rule of Five Violations a	0	0	0	0	0	0
BBB permeant	No	Yes	No	No	No	Yes
GI absorption	High	High	High	High	High	High
P-glycoprotein Inhibitor	No	Inhibitor	No	No	No	No
Acute Oral Toxicity	III	III	III	III	II	III
Carcinogens	No	No	No	No	No	No

a: Violations of Lipinski rule of 5 (logP < 5, MW < 500, nHBA < 10, and nHBD < 5).

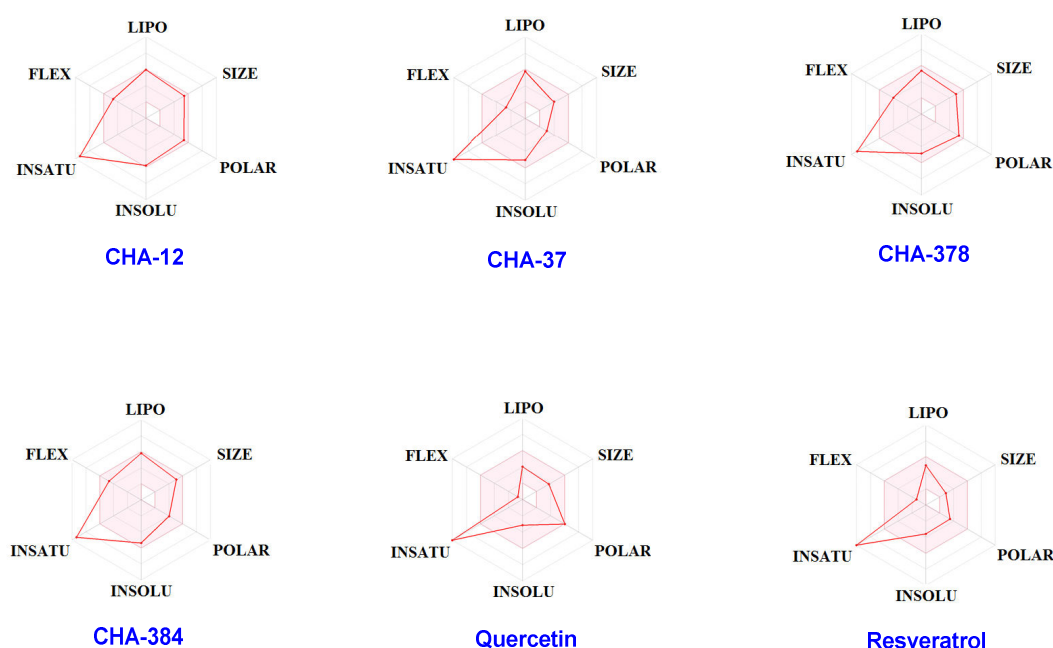


Figure S1. Radar map of the physicochemical properties for oral bioavailability of **CHA-12**, **CHA-37**, **CHA-378**, **CHA-384**, and standards quercetin and resveratrol predicted by SwissADME online server (colored zone is the suitable physicochemical space for oral bioavailability). LIPO (Lipophilicity): $-0.7 < \text{XLOGP3} < +5.0$; SIZE: $150 \text{ g/mol} < \text{mw} < 500 \text{ g/mol}$; POLAR (polarity): $20 \text{ \AA}^2 < \text{TPSA} < 130 \text{ \AA}^2$; INSOLU (insolubility): $0 < \text{Log S (ESOL)} < 6$; INSATU (insaturation): $0.25 < \text{Fraction Csp3} < 1$; FLEX (Flexibility): $0 < \text{Numb. rotatable bonds} < 9$.

Table S3. Predicted inhibitory effects of **CHA-12**, **CHA-37**, **CHA-378**, **CHA-384**, resveratrol, and quercetin on cytochrome P450 enzymes (predicted by preADME online server).

Compounds	CYP2C19 inhibition	CYP2C9 inhibition	CYP2D6 inhibition	CYP2D6 Substrate	CYP3A4 inhibition	CYP3A4 Substrate
CHA-12	Non	Inhibitor	Non	Non	Non	Weakly
CHA-37	Non	Inhibitor	Non	Non	Non	Weakly
CHA-378	Inhibitor	Inhibitor	Non	Non	Inhibitor	Non
CHA-384	Non	Inhibitor	Non	Non	Non	Non
Resveratrol	Inhibitor	Inhibitor	Non	Non	Inhibitor	Non
Quercetin	Inhibitor	Inhibitor	Non	Non	Inhibitor	Non

Table S4. Important interactions of most active **CHA-12**, **CHA-378**, **CHA-384**, and baicalein as standard in the SARS-CoV-2 3CLpro active site.

Interactions	CHA-12		CHA-378		CHA-384		Baicalein	
	Residues	Interacting groups	Residues	Interacting groups	Residues	Interacting groups	Residues	Interacting groups
Conventional H-bond	Glu166	NH-Tetrazole	Asn142	3-OH ring	Arg188	1-NH (Urea-moiety)	Gly143	6-OH Ring (A-Ring)
	Thr190	OH (A-Phenyl ring)			Arg188	2-NH (Urea-moiety)	Ser144	6-OH Ring (A-Ring)
	Gln192	Carbonyl group						
	His41	O (Etheric linker)						
	His41	N-Quinoline						
Pi-donor	Gln189	B-Phenyl ring	Asn142	B-Phenyl ring				
Pi-Alkyl	Met49	B-Phenyl ring	Met49	Phenyl ring (Sulf-moiety)	Met49	A-Phenyl ring	Met49	B-Ring
	Cys145	Quinoline (Benzene ring)	Met165	A-Phenyl ring	Met165	A-Phenyl ring	Cys44	B-Ring
			Cys145	B-Phenyl ring	Pro168	Phenyl ring (Urea-moiety)	Cys145	C-Ring

			His41	Methyl moiety)	(Sulf-			
Pi-Sulfur	Met49	Quinoline (Pyridine ring)					Cys145	A-Ring
Pi-Sigma	Thr25	Quinoline (Pyridine ring)	Gln189	A-Phenyl ring		Thr25	B-Phenyl ring	
			Gln189	A-Phenyl ring				
Acceptor-Acceptor			Asn142	4-OH ring)	(B-Phenyl			
			Ser144	4-OH ring)	(B-Phenyl			
Donor-Donor							Gly143	5-OH (A-Ring)
Pi-Cation			His41	Phenyl ring (Sulf-moiety)				
Pi-Pi Stacked			His41	Phenyl ring (Sulf-moiety)	His41	A-Phenyl ring	His41	B-Ring
Alkyl					Pro168 Ala191	Cl (Urea-moiety) Cl (Urea-moiety)		
Carbon							Met165	Carbonyl (C-Ring)

Table S5. Important interactions of the most active **CHA-12**, **CHA-37**, **CHA-378** and standard compound GRL0617 in the active site of SARS-CoV-2 PLpro.

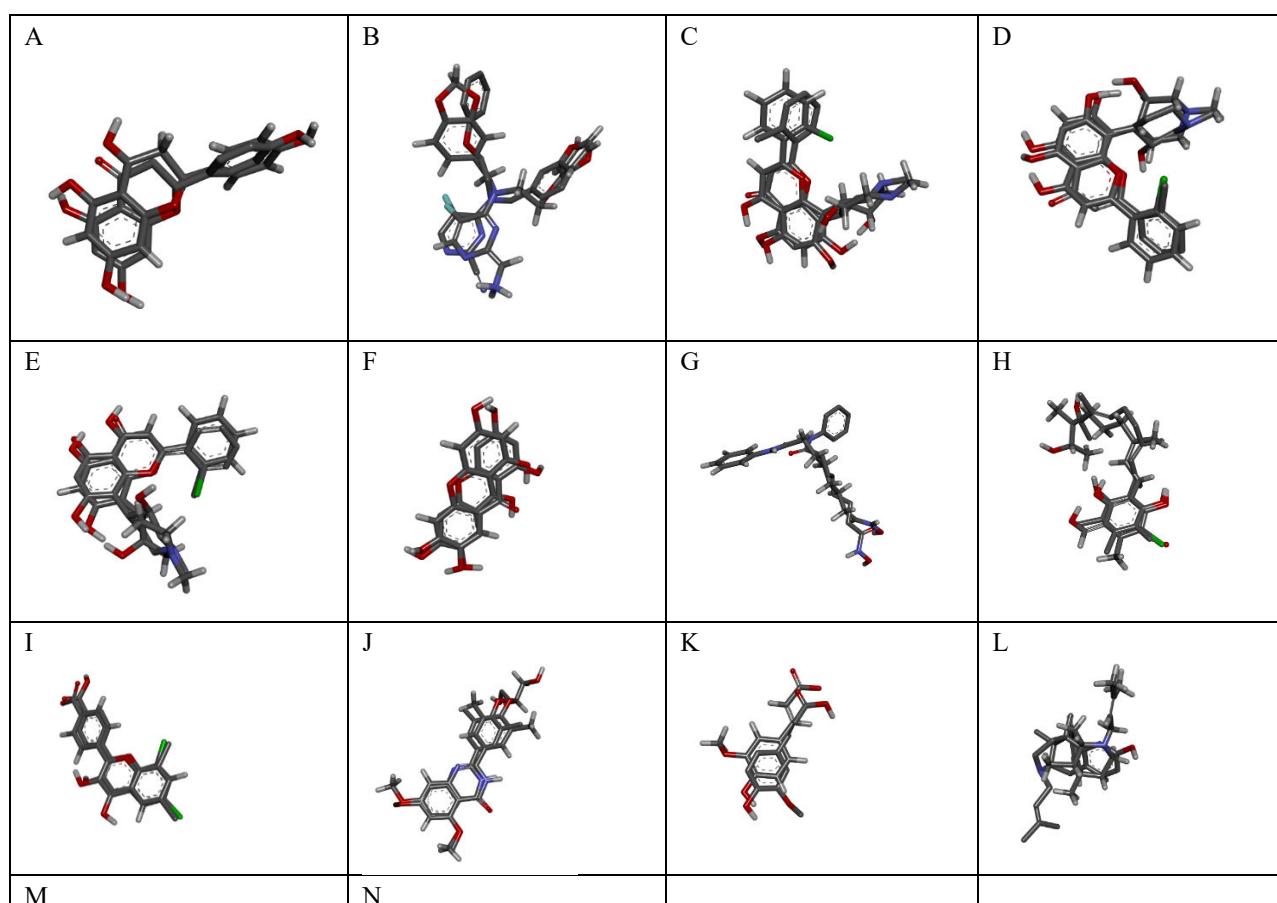
Interactions	CHA-12		CHA-37		CHA-384		GRL0617	
	Residues	Interacting groups	Residues	Interacting groups	Residues	Interacting groups	Residues	Interacting groups
Conventional H-bond	Lys157	N1 Tetrazole			Tyr264	Carbonyl group	Gln269	Carbonyl group
	Gln269	N4 Tetrazole			Glu167	NH (Sulf-moiety)	Asp164	NH (Amide)
	Tyr264	OH (A-Phenyl ring)					Gly163	NH2 (A-Phenyl ring)
Pi-donor	Gln269	Tetrazole ring						
Pi-Alkyl	Pro248	B-Phenyl ring	Pro248	B-Phenyl ring	Pro248	B-Phenyl ring	Tyr268	CH ₃ (A-Phenyl ring)
			Pro248	Triazole (Benzotriazole)	Leu162	A-Phenyl ring	Pro247	Naphthyl (Closer ring)
			Pro248	Benzene (Benzotriazole)			Pro248	Naphthyl (Closer ring)
			Pro247	Benzene (Benzotriazole)			Pro248	Naphthyl (Farther ring)
Amide-Pi Stacked	Gly163	A-Phenyl ring	Gly163	A-Phenyl ring	Gly163	A-Phenyl ring		
			Asn267	Triazole (Benzotriazole)				
Pi-Sigma					Gln269	A-Phenyl ring	Asp164	A-Phenyl ring
Acceptor-Acceptor	Tyr273	Carbonyl group						
Pi-Cation			Lys157	A-Phenyl ring				
Pi-Anion	Glu167	Tetrazole ring			Asp164	B-Phenyl ring	Asp164	A-Phenyl ring
					Glu167	Phenyl ring (Sulf-moiety)		

Pi-Pi Stacked	Tyr268	Quinoline (Pyridine ring)	Tyr264	B-Phenyl ring					
	Tyr268	Quinoline (Benzene ring)							
Pi-Pi T-Shaped	Tyr268	B-Phenyl ring	Tyr268	Benzene (Benzotriazole)	Tyr264	B-Phenyl ring	Tyr268	Naphthyl (Closer ring)	(Closer ring)
							Tyr268	Naphthyl (Farther ring)	(Farther ring)
Carbon					Pro248	4-OH (B-Phenyl ring)			

Table S6. The list of host-based and virus-based targets along with their associated PDB codes as well as the essential parameters for the docking protocol used in this study.

Entry	Target	PDB Code	X	Y	Z	Grid box	Docking score ^a
1	p38 MAPK	4EH3	-2.668	0.007	-20836	20,20,20	- 8.40
2	Cathepsin L	5MQY	54.336	48.381	17.63	20,20,20	-6.80
3	CDK1	6GU2	329.218	212.541	192.325	20,20,20	-9.60
4	CDK2/CyclinA	6GUB	-8.411	-21.607	22.226	20,20,20	-10.10
5	CDK9/cyclinT1	3BLR	52.813	-16.066	-12.998	20,20,20	-10.30
6	ERK2	3SA0	-12.826	10.831	40.627	26,22,20	-7.40
7	HDAC2	4LXZ	25.00	-17.005	-0.024	20,20,20	-7.50
8	DHODH	5ZF7	-33.443	14.428	-21.466	20,22,22	-9.60
9	CK2 alpha'	5M4U	10.536	-19.326	-8.976	20,20,20	-11.40
10	RBD2	4J1P	10.105	20.989	-6.705	20,20,26	-7.30
11	RBD4	6HOV	10.887	5.487	-0.524	20,20,20	-6.50
12	Sigma-1 receptor	6DK1	12.009	37.935	-33.62	20,20,22	-9.0
13	SARS-CoV-2 3CLpro	6M2N	-32.488	-63.85	40.962	22,20,20	-7.80
14	SARS-CoV-2 PLpro	7JN2	51.608	30.592	0.79	20,20,20	-8.80

a: Docking score of the re-dock co-crystal ligand for the method validation.



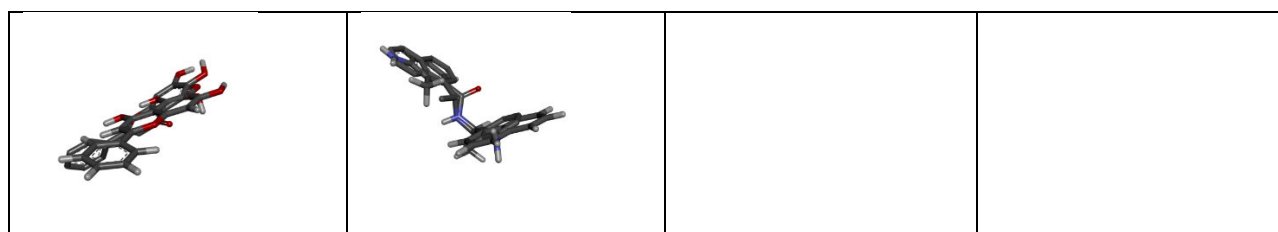


Figure S2. Coordinates of the co-crystal ligand and the re-docked one in the validation of the docking methods with (A) p38 MAPK, (B) Cathepsin L., (C) cyclin-dependent Kinase-1, (D) CDK2/CyclinA, (E) CDK9/cyclinT1, (F) ERK2, (G) HDAC2, (H) DHODH, (I) CK2 alpha, (J) RBD2, (K) RBD4, (L) sigma-1 receptor, (M) 3CLpro and (N) PLpro.

References

1. Kopecky-Bromberg, S. A.; Martinez-Sobrido, L.; Palese, P., 7a protein of severe acute respiratory syndrome coronavirus inhibits cellular protein synthesis and activates p38 mitogen-activated protein kinase. *J. Virol.* **2006**, *80*, 785-793.
2. Marchant, D.; Singhera, G. K.; Utokaparch, S.; Hackett, T. L.; Boyd, J. H.; Luo, Z.; Si, X.; Dorscheid, D. R.; McManus, B. M.; Hegele, R. G., Toll-like receptor 4-mediated activation of p38 mitogen-activated protein kinase is a determinant of respiratory virus entry and tropism. *J. Virol.* **2010**, *84*, 11359-11373.
3. Mizutani, T.; Fukushima, S.; Saijo, M.; Kurane, I.; Morikawa, S., Phosphorylation of p38 MAPK and its downstream targets in SARS coronavirus-infected cells. *Biochem. Biophys. Res. Commun.* **2004**, *319*, 1228-1234.
4. Grimes, J. M.; Grimes, K. V., p38 MAPK inhibition: A promising therapeutic approach for COVID-19. *J. Mol. Cell. Cardiol.* **2020**, *144*, 63-65.
5. Kono, M.; Tatsumi, K.; Imai, A. M.; Saito, K.; Kuriyama, T.; Shirasawa, H., Inhibition of human coronavirus 229E infection in human epithelial lung cells (L132) by chloroquine: involvement of p38 MAPK and ERK. *Antivir. Res.* **2008**, *77*, 150-152.
6. Banerjee, S.; Narayanan, K.; Mizutani, T.; Makino, S., Murine coronavirus replication-induced p38 mitogen-activated protein kinase activation promotes interleukin-6 production and virus replication in cultured cells. *J. Virol.* **2002**, *76*, 5937-5948.
7. Padhan, K.; Minakshi, R.; Towheed, M. A. B.; Jameel, S., Severe acute respiratory syndrome coronavirus 3a protein activates the mitochondrial death pathway through p38 MAP kinase activation. *J. Gen. Virol.* **2008**, *89*, (8), 1960-1969.
8. McCaskill, J. L.; Ressel, S.; Alber, A.; Redford, J.; Power, U. F.; Schwarze, J.; Dutia, B. M.; Buck, A. H., Broad-spectrum inhibition of respiratory virus infection by microRNA mimics targeting p38 MAPK signaling. *Mol. Ther. -Nucleic Acids* **2017**, *7*, 256-266.
9. Liu, T.; Luo, S.; Libby, P.; Shi, G.-P., Cathepsin L-selective inhibitors: A potentially promising treatment for COVID-19 patients. *Pharmacol. Ther.* **2020**, 107587.
10. Huang, I.-C.; Bosch, B. J.; Li, F.; Li, W.; Lee, K. H.; Ghiran, S.; Vasilieva, N.; Dermody, T. S.; Harrison, S. C.; Dormitzer, P. R., SARS coronavirus, but not human coronavirus NL63, utilizes cathepsin L to infect ACE2-expressing cells. *J. Biol. Chem.* **2006**, *281*, 3198-3203.
11. Simmons, G.; Gosalia, D. N.; Rennekamp, A. J.; Reeves, J. D.; Diamond, S. L.; Bates, P., Inhibitors of cathepsin L prevent severe acute respiratory syndrome coronavirus entry. *Proc. Natl. Acad. Sci.* **2005**, *102*, 11876-11881.
12. Zhao, M.-M.; Yang, W.-L.; Yang, F.-Y.; Zhang, L.; Huang, W.-J.; Hou, W.; Fan, C.-F.; Jin, R.-H.; Feng, Y.-M.; Wang, Y.-C., Cathepsin L plays a key role in SARS-CoV-2 infection in humans and humanized mice and is a promising target for new drug development. *Signal Transduct. Target. Ther.* **2021**, *6*, 1-12.

13. Sacco, M. D.; Ma, C.; Lagarias, P.; Gao, A.; Townsend, J. A.; Meng, X.; Dube, P.; Zhang, X.; Hu, Y.; Kitamura, N., Structure and inhibition of the SARS-CoV-2 main protease reveal strategy for developing dual inhibitors against Mpro and cathepsin L. *Sci. Adv.* **2020**, *6*, eabe0751.
14. Bosch, B. J.; Bartelink, W.; Rottier, P. J., Cathepsin L functionally cleaves the severe acute respiratory syndrome coronavirus class I fusion protein upstream of rather than adjacent to the fusion peptide. *J. Virol.* **2008**, *82*, 8887-8890.
15. Ferraro, F.; Merlino, A.; dell' Oca, N.; Gil, J.; Tort, J. F.; Gonzalez, M.; Cerecetto, H.; Cabrera, M.; Corvo, I., Identification of chalcones as *Fasciola hepatica* cathepsin L inhibitors using a comprehensive experimental and computational approach. *PLoS Negl. Trop. Dis.* **2016**, *10*, e0004834.
16. Garg, S.; Raghav, N., Inhibitory potential of some chalcones on cathepsins B, H and L. *RSC Adv.* **2015**, *5*, 72937-72949.
17. Surjit, M.; Liu, B.; Chow, V. T.; Lal, S. K., The nucleocapsid protein of severe acute respiratory syndrome-coronavirus inhibits the activity of cyclin-cyclin-dependent kinase complex and blocks S phase progression in mammalian cells. *J. Biol. Chem.* **2006**, *281*, 10669-10681.
18. Gutierrez-Chamorro, L.; Felip, E.; Ezeonwumelu, I. J.; Margelí, M.; Ballana, E., Cyclin-dependent Kinases as Emerging Targets for Developing Novel Antiviral Therapeutics. *Trends Microbiol.* **2021**.
19. Schang, L. M., Cyclin-dependent kinases as cellular targets for antiviral drugs. *J. Antimicrob. Chemother.* **2002**, *50*, 779-792.
20. Gargouri, M.; Alzwi, A.; Abobaker, A., Cyclin dependent kinase inhibitors as a new potential therapeutic option in management of COVID-19. *Med. Hypotheses* **2021**, *146*, 110380.
21. Heaton, B. E.; Trimarco, J. D.; Hamele, C. E.; Harding, A. T.; Tata, A.; Zhu, X.; Tata, P. R.; Smith, C. M.; Heaton, N. S., SRSF protein kinases 1 and 2 are essential host factors for human coronaviruses including SARS-CoV-2. *BioRxiv* **2020**.
22. Cribier, A.; Descours, B.; Valadão, A. L. C.; Laguette, N.; Benkirane, M., Phosphorylation of SAMHD1 by cyclin A2/CDK1 regulates its restriction activity toward HIV-1. *Cell Rep.* **2013**, *3*, 1036-1043.
23. Ludgate, L.; Ning, X.; Nguyen, D. H.; Adams, C.; Mentzer, L.; Hu, J., Cyclin-dependent kinase 2 phosphorylates s/tp sites in the hepadnavirus core protein C-terminal domain and is incorporated into viral capsids. *J. Virol.* **2012**, *86*, 12237-12250.
24. He, W.; Staples, D.; Smith, C.; Fisher, C., Direct activation of cyclin-dependent kinase 2 by human papillomavirus E7. *J. Virol.* **2003**, *77*, 10566-10574.
25. Bouchard, M.; Giannakopoulos, S.; Wang, E. H.; Tanese, N.; Schneider, R. J., Hepatitis B virus HBx protein activation of cyclin A-cyclin-dependent kinase 2 complexes and G1 transit via a Src kinase pathway. *J. Virol.* **2001**, *75*, 4247-4257.
26. Wang, S.; Fischer, P. M., Cyclin-dependent kinase 9: a key transcriptional regulator and potential drug target in oncology, virology and cardiology. *Trends Pharmacol. Sci.* **2008**, *29*, 302-313.
27. Khan, S. Z.; Mitra, D., Cyclin K inhibits HIV-1 gene expression and replication by interfering with cyclin-dependent kinase 9 (CDK9)-cyclin T1 interaction in Nef-dependent manner. *J. Biol. Chem.* **2011**, *286*, 22943-22954.
28. Zhang, J.; Li, G.; Ye, X., Cyclin T1/CDK9 interacts with influenza A virus polymerase and facilitates its association with cellular RNA polymerase II. *J. Virol.* **2010**, *84*, 12619-12627.
29. Klebl, B. M.; Choidas, A., CDK9/cyclin T1: a host cell target for antiretroviral therapy. **2006**.

30. Kindrachuk, J.; Ork, B.; Hart, B. J.; Mazur, S.; Holbrook, M. R.; Frieman, M. B.; Traynor, D.; Johnson, R. F.; Dyllal, J.; Kuhn, J. H., Antiviral potential of ERK/MAPK and PI3K/AKT/mTOR signaling modulation for Middle East respiratory syndrome coronavirus infection as identified by temporal kinome analysis. *Antimicrob. Agents Chemother.* **2015**, *59*, 1088-1099.
31. Varshney, B.; Lal, S. K., SARS-CoV accessory protein 3b induces AP-1 transcriptional activity through activation of JNK and ERK pathways. *Biochemistry* **2011**, *50*, 5419-5425.
32. Johnson, T. R.; McLellan, J. S.; Graham, B. S., Respiratory syncytial virus glycoprotein G interacts with DC-SIGN and L-SIGN to activate ERK1 and ERK2. *J. Virol.* **2012**, *86*, 1339-1347.
33. Chen, W.; Monick, M. M.; Carter, A. B.; Hunninghake, G. W., Activation of ERK2 by respiratory syncytial virus in A549 cells is linked to the production of interleukin 8. *Exp. Lung Res.* **2000**, *26*, 13-26.
34. Martin, S.; Harris, D. T.; Shisler, J., The C11R gene, which encodes the vaccinia virus growth factor, is partially responsible for MVA-induced NF- κ B and ERK2 activation. *J. Virol.* **2012**, *86*, 9629-9639.
35. Takahashi, Y.; Hayakawa, A.; Sano, R.; Fukuda, H.; Harada, M.; Kubo, R.; Okawa, T.; Kominato, Y., Histone deacetylase inhibitors suppress ACE2 and ABO simultaneously, suggesting a preventive potential against COVID-19. *Sci. Rep.* **2021**, *11*, 1-9.
36. Murthy, P. K.; Sivashanmugam, K.; Kandasamy, M.; Subbiah, R.; Ravikumar, V., Repurposing of histone deacetylase inhibitors: A promising strategy to combat pulmonary fibrosis promoted by TGF- β signalling in COVID-19 survivors. *Life Sci.* **2020**, 118883.
37. Sivashanmugam, K.; Kandasamy, M.; Subbiah, R.; Ravikumar, V., Repurposing of histone deacetylase inhibitors: A promising strategy to combat pulmonary fibrosis promoted by TGF- β signalling in COVID-19 survivors. *Life Sci* **2021**, 118883-118883.
38. Mohamed, M. F.; Shaykoon, M. S. A.; Abdelrahman, M. H.; Elsadek, B. E.; Aboraia, A. S.; Abuo-Rahma, G. E.-D. A., Design, synthesis, docking studies and biological evaluation of novel chalcone derivatives as potential histone deacetylase inhibitors. *Bioorganic Chem.* **2017**, *72*, 32-41.
39. Orlikova, B.; Schneckeburger, M.; Zloh, M.; Golais, F.; Diederich, M.; Tasdemir, D., Natural chalcones as dual inhibitors of HDACs and NF- κ B. *Oncol. Rep.* **2012**, *28*, 797-805.
40. Coelho, A. R.; Oliveira, P. J. *Dihydroorotate Dehydrogenase Inhibitors in SARS-CoV-2 Infection*; Wiley: Hoboken, NJ, USA, 2020.
41. Berber, B.; Doluca, O., A comprehensive drug repurposing study for COVID19 treatment: novel putative dihydroorotate dehydrogenase inhibitors show association to serotonin–dopamine receptors. *Brief. Bioinform.* **2021**.
42. Luban, J.; Sattler, R. A.; Mühlberger, E.; Graci, J. D.; Cao, L.; Weetall, M.; Trotta, C.; Colacino, J. M.; Bavari, S.; Strambio-De-Castillia, C., The DHODH inhibitor PTC299 arrests SARS-CoV-2 replication and suppresses induction of inflammatory cytokines. *Virus Res.* **2021**, *292*, 198246.
43. Xiong, R.; Zhang, L.; Li, S.; Sun, Y.; Ding, M.; Wang, Y.; Zhao, Y.; Wu, Y.; Shang, W.; Jiang, X., Novel and potent inhibitors targeting DHODH are broad-spectrum antivirals against RNA viruses including newly-emerged coronavirus SARS-CoV-2. *Protein Cell* **2020**, *11*, 723-739.
44. Xiong, R.; Zhang, L.; Li, S.; Sun, Y.; Ding, M.; Wang, Y.; Zhao, Y.; Wu, Y.; Shang, W.; Jiang, X., Novel and potent inhibitors targeting DHODH, a rate-limiting enzyme in de novo pyrimidine biosynthesis, are broad-spectrum antiviral against RNA viruses including newly emerged coronavirus SARS-CoV-2. *BioRxiv* **2020**.

-
45. Hahn, F.; Wangen, C.; Häge, S.; Peter, A. S.; Dobler, G.; Hurst, B.; Julander, J.; Fuchs, J.; Ruzsics, Z.; Überla, K., IMU-838, a Developmental DHODH Inhibitor in Phase II for Autoimmune Disease, Shows Anti-SARS-CoV-2 and Broad-Spectrum Antiviral Efficacy In Vitro. *Viruses* **2020**, *12*, 1394.
46. Calistri, A.; Luganini, A.; Conciatori, V.; Del Vecchio, C.; Sainas, S.; Boschi, D.; Lolli, M. L.; Gribaudo, G.; Parolin, C., The new generation hDHODH inhibitor MEDS433 hinders the in vitro replication of SARS-CoV-2. *bioRxiv* **2020**.
47. Xia, C.; Wolf, J. J.; Vijayan, M.; Studstill, C. J.; Ma, W.; Hahm, B., Casein kinase 1 α mediates the degradation of receptors for type I and type II interferons caused by hemagglutinin of influenza A virus. *J. Virol.* **2018**, *92*.
48. Quintavalle, M.; Sambucini, S.; Summa, V.; Orsatti, L.; Talamo, F.; De Francesco, R.; Neddermann, P., Hepatitis C virus NS5A is a direct substrate of casein kinase I- α , a cellular kinase identified by inhibitor affinity chromatography using specific NS5A hyperphosphorylation inhibitors. *J. Biol. Chem.* **2007**, *282*, 5536-5544.
49. Bhattacharya, D.; Ansari, I. H.; Striker, R., The flaviviral methyltransferase is a substrate of Casein Kinase 1. *Virus Res.* **2009**, *141*, 101-104.
50. Bhargavan, B.; Kanmogne, G. D., Epigenetics, N-myristoyltransferase-1 and casein kinase-2-alpha modulates the increased replication of HIV-1 CRF02_AG, compared to subtype-B viruses. *Sci. Rep.* **2019**, *9*, 1-19.
51. Zhang, L.; Li, H.; Chen, Y.; Gao, X.; Lu, Z.; Gao, L.; Wang, Y.; Gao, Y.; Gao, H.; Liu, C., The down-regulation of casein kinase 1 alpha as a host defense response against infectious bursal disease virus infection. *Virology* **2017**, *512*, 211-221.
52. Campagna, M.; Budini, M.; Arnoldi, F.; Desselberger, U.; Allende, J. E.; Burrone, O. R., Impaired hyperphosphorylation of rotavirus NSP5 in cells depleted of casein kinase 1 α is associated with the formation of viroplasms with altered morphology and a moderate decrease in virus replication. *J. Gen. Virol.* **2007**, *88*, 2800-2810.
53. Chlamydas, S.; Papavassiliou, A. G.; Piperi, C., Epigenetic mechanisms regulating COVID-19 infection. *Epigenetics* **2021**, *16*, 263-270.
54. Tian, R.; Samelson, A. J.; Rezelj, V. V.; Chen, M.; Ramadoss, G. N.; Guo, X.; Mac Kain, A.; Tran, Q. D.; Lim, S. A.; Lui, I., BRD2 inhibition blocks SARS-CoV-2 infection in vitro by reducing transcription of the host cell receptor ACE2. *bioRxiv* **2021**.
55. O'Meara, M. J.; Guo, J. Z.; Swaney, D. L.; Tummino, T. A.; Hüttenhain, R., A SARS-CoV-2-human protein-protein interaction map reveals drug targets and potential drug-repurposing. *BioRxiv* **2020**.
56. Acharya, A.; Pandey, K.; Thurman, M.; Challagundala, K. B.; Vann, K. R.; Kutateladze, T. G.; Morales, G. A.; Durden, D. L.; Byrareddy, S. N., Highly potent PI3K- α /mTOR/BRD4 inhibitor for the targeted inhibition of SARS-CoV-2. *bioRxiv* **2021**.
57. Acharya, A.; Pandey, K.; Thurman, M.; Challagundla, K. B.; Vann, K. R.; Kutateladze, T. G.; Morales, G. A.; Durden, D. L.; Byrareddy, S. N., Blockade of SARS-CoV-2 infection in-vitro by highly potent PI3K- α /mTOR/BRD4 inhibitor. *bioRxiv* **2021**.
58. Hashimoto, K., Repurposing of CNS drugs to treat COVID-19 infection: targeting the sigma-1 receptor. *Eur. Arch. Psychiatry Clin. Neurosci.* **2021**, 1-10.
59. Vela, J. M., Repurposing sigma-1 receptor ligands for Covid-19 therapy? *Front. Pharmacol.* **2020**, *11*, 1716.
60. Abate, C.; Niso, M.; Abatematteo, F. S.; Contino, M.; Colabufo, N. A.; Berardi, F., PB28, the Sigma-1 and Sigma-2 Receptors Modulator With Potent Anti-SARS-CoV-2 Activity: A Review About Its Pharmacological Properties and Structure Affinity Relationships. *Front. Pharmacol.* **2020**, *11*.

61. Yesilkaya, U. H.; Balcioglu, Y. H.; Sahin, S., Reissuing the sigma receptors for SARS-CoV-2. *J. Clin. Neurosci.* **2020**, *80*, 72-73.
62. Salerno, J. A.; Torquato, T.; Temerozo, J. R.; Goto-Silva, L.; Mendes, M.; Sacramento, C. Q.; Fintelman-Rodrigues, N.; Vitoria, G.; Souza, L.; Ornelas, I., Inhibition of SARS-CoV-2 infection in human cardiomyocytes by targeting the Sigma-1 receptor disrupts cytoskeleton architecture and contractility. *bioRxiv* **2021**.
63. Hashimoto, K., Review 2:" Inhibition of SARS-CoV-2 infection in human cardiomyocytes by targeting the Sigma-1 receptor disrupts cytoskeleton architecture and contractility". *Rapid Rev. COVID-19* **2021**.
64. Gordon, D. E.; Jang, G. M.; Bouhaddou, M.; Xu, J.; Obernier, K.; White, K. M.; O'Meara, M. J.; Rezelj, V. V.; Guo, J. Z.; Swaney, D. L., A SARS-CoV-2 protein interaction map reveals targets for drug repurposing. *Nature* **2020**, *583*, 459-468.
65. Su, H.; Yao, S.; Zhao, W.; Li, M.; Liu, J.; Shang, W.; Xie, H.; Ke, C.; Gao, M.; Yu, K., Discovery of baicalin and baicalein as novel, natural product inhibitors of SARS-CoV-2 3CL protease in vitro. *bioRxiv* **2020**.
66. Abdallah, H. M.; El-Halawany, A. M.; Sirwi, A.; El-Araby, A. M.; Mohamed, G. A.; Ibrahim, S. R.; Koshak, A. E.; Asfour, H. Z.; Awan, Z. A.; Elfaky, M. A., Repurposing of Some Natural Product Isolates as SARS-COV-2 Main Protease Inhibitors via In Vitro Cell Free and Cell-Based Antiviral Assessments and Molecular Modeling Approaches. *Pharmaceuticals* **2021**, *14*, 213.
67. Ullrich, S.; Nitsche, C., The SARS-CoV-2 main protease as drug target. *Bioorganic Med. Chem. Lett.* **2020**, 127377.
68. ul Qamar, M. T.; Alqahtani, S. M.; Alamri, M. A.; Chen, L.-L., Structural basis of SARS-CoV-2 3CLpro and anti-COVID-19 drug discovery from medicinal plants. *J. Pharm. Anal.* **2020**, *10*, 313-319.
69. Shin, D.; Mukherjee, R.; Grewe, D.; Bojkova, D.; Baek, K.; Bhattacharya, A.; Schulz, L.; Widera, M.; Mehdipour, A. R.; Tascher, G., Papain-like protease regulates SARS-CoV-2 viral spread and innate immunity. *Nature* **2020**, *587*, 657-662.
70. McClain, C. B.; Vabret, N., SARS-CoV-2: the many pros of targeting PLpro. *Signal Transduct. Target. Ther.* **2020**, *5*, 1-2.
71. Klemm, T.; Ebert, G.; Calleja, D. J.; Allison, C. C.; Richardson, L. W.; Bernardini, J. P.; Lu, B. G.; Kuchel, N. W.; Grohmann, C.; Shibata, Y., Mechanism and inhibition of the papain-like protease, PLpro, of SARS-CoV-2. *EMBO J.* **2020**, *39*, e106275.
72. Rut, W.; Lv, Z.; Zmudzinski, M.; Patchett, S.; Nayak, D.; Snipas, S. J.; El Oualid, F.; Huang, T. T.; Bekes, M.; Drag, M., Activity profiling and crystal structures of inhibitor-bound SARS-CoV-2 papain-like protease: A framework for anti-COVID-19 drug design. *Sci. Adv.* **2020**, *6*, eabd4596.

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