

Synthesis, Biological and In Silico Studies of Griseofulvin and usnic acid sulfonamide derivatives as Carbonic Anhydrase Inhibitors

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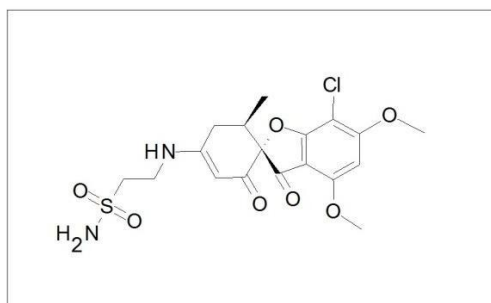
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¹H-NMR and ¹³C-NMR of compounds

Compound 1a

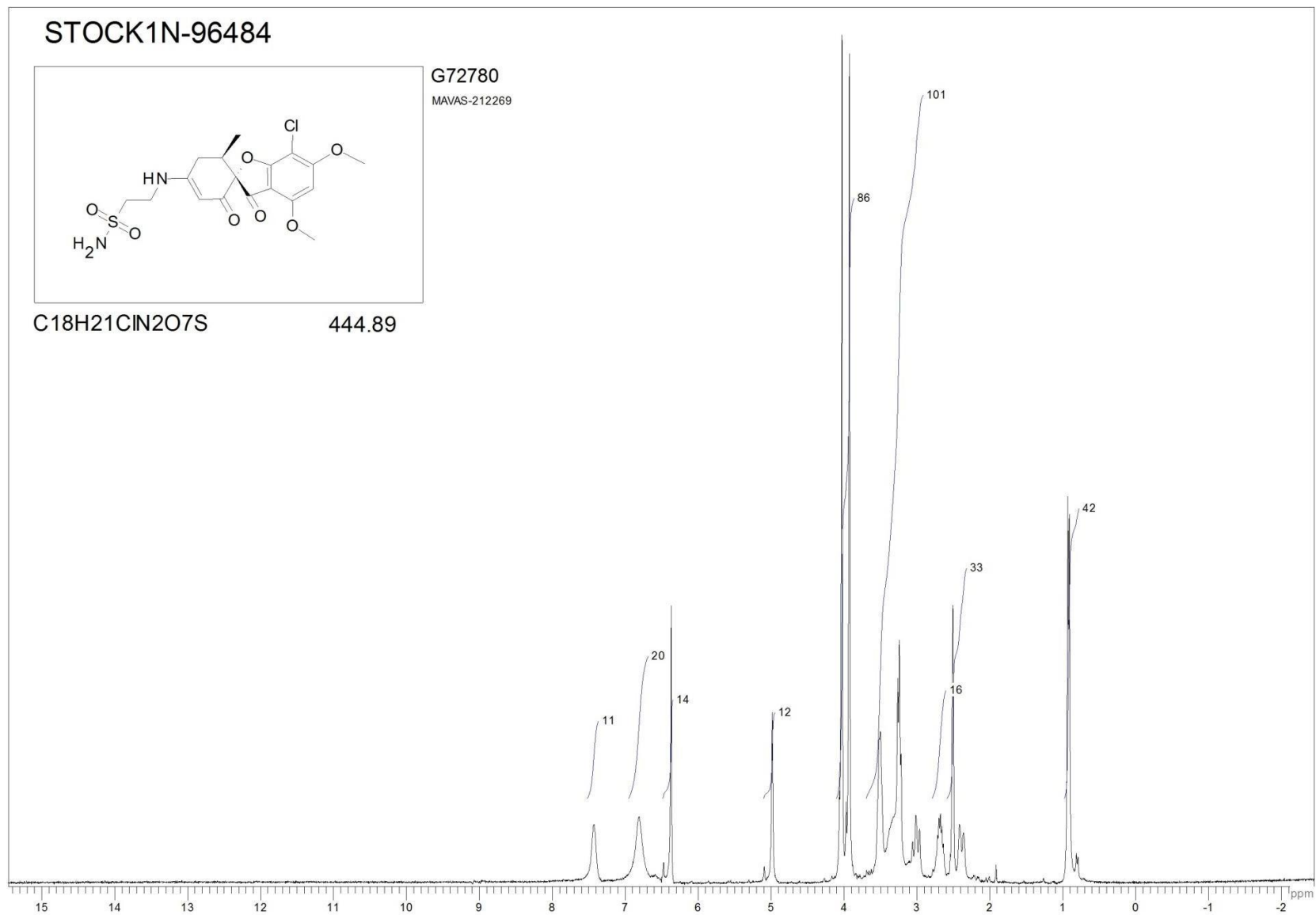
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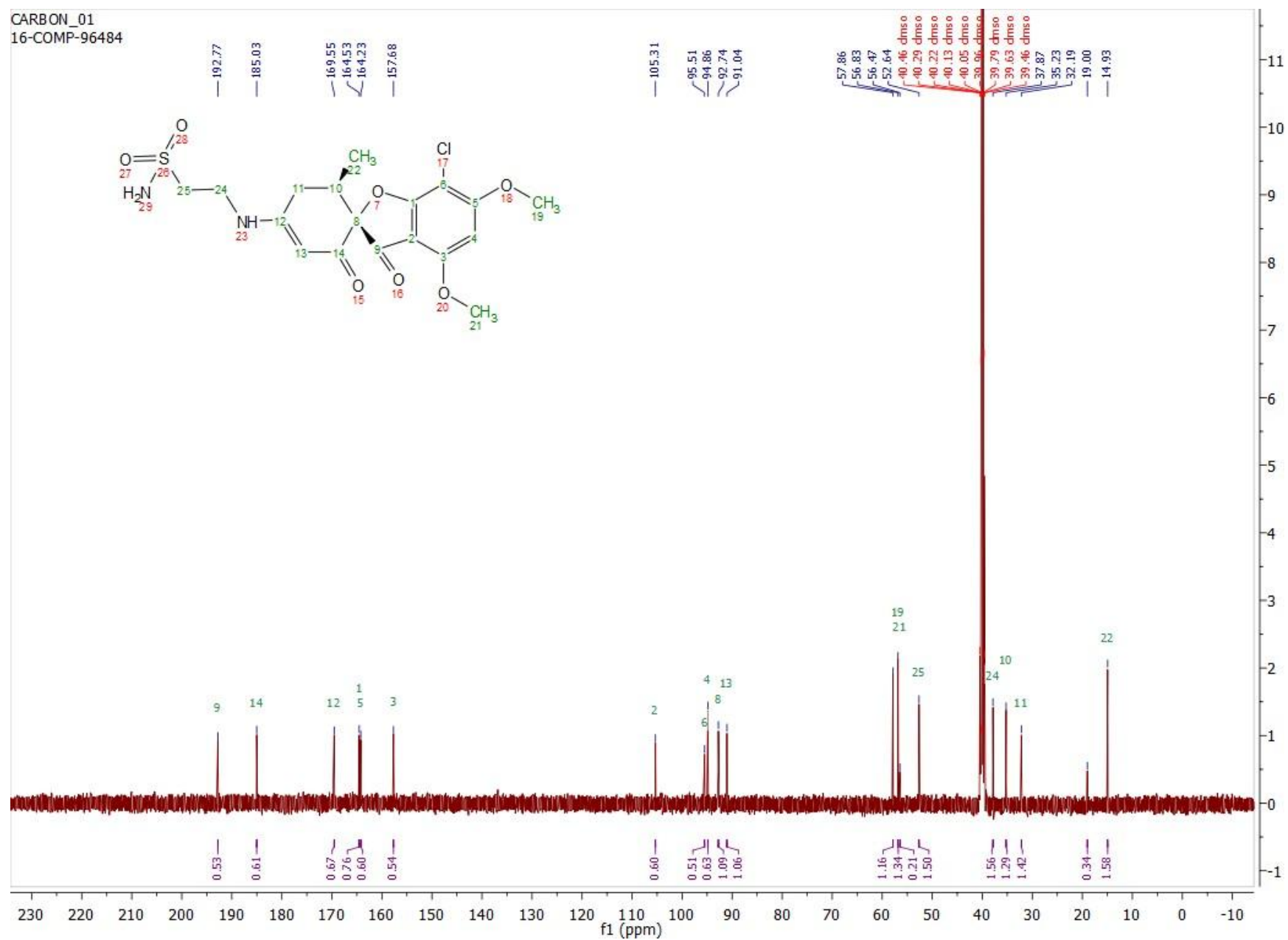
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C18H21ClN2O7S

444.89

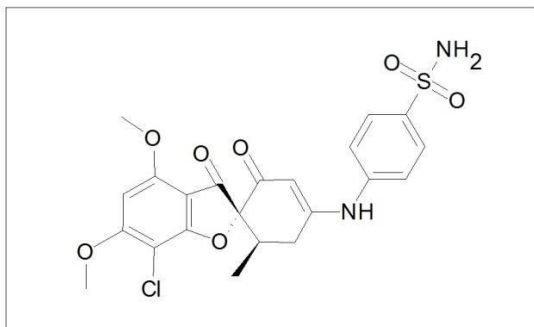


CARBON_01
16-COMP-96484



Compound 1b

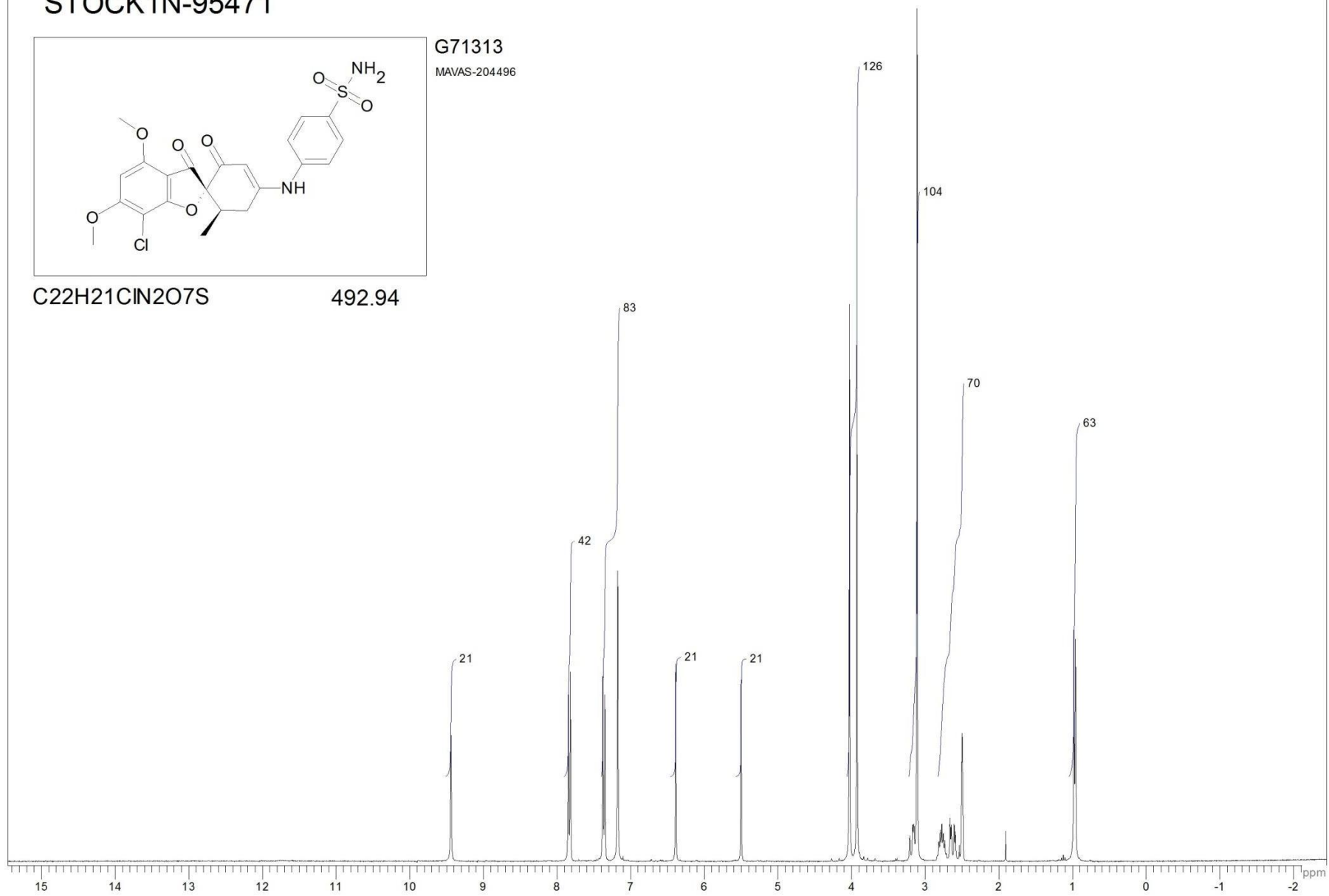
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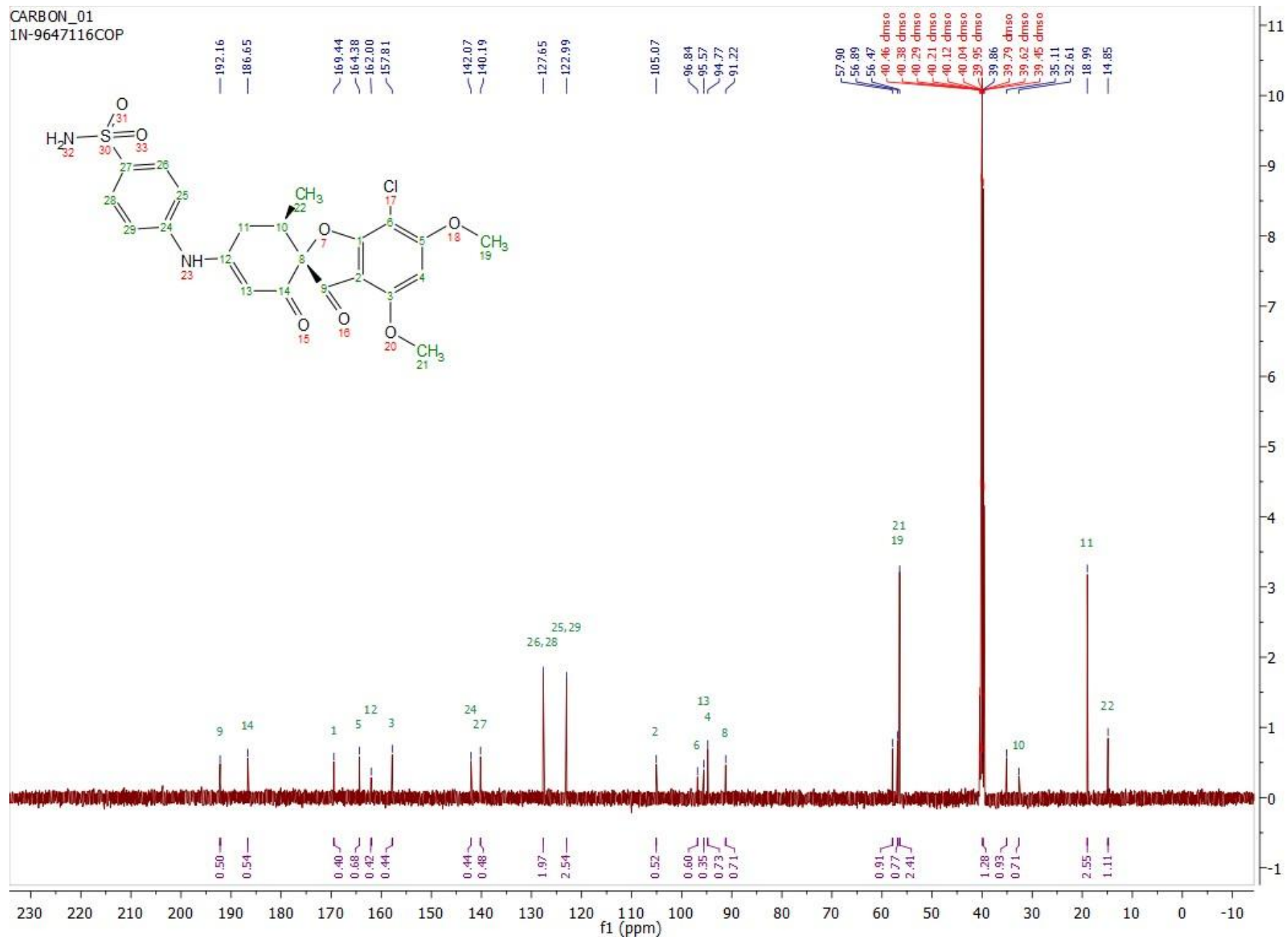
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C22H21ClN2O7S

492.94

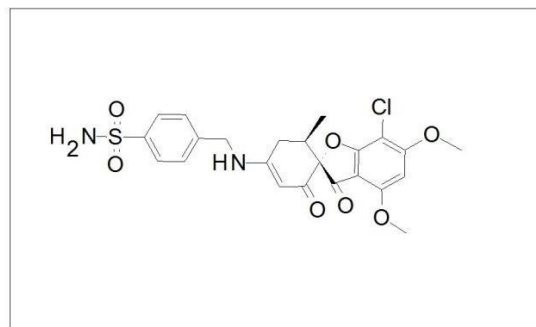


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Compound 1c

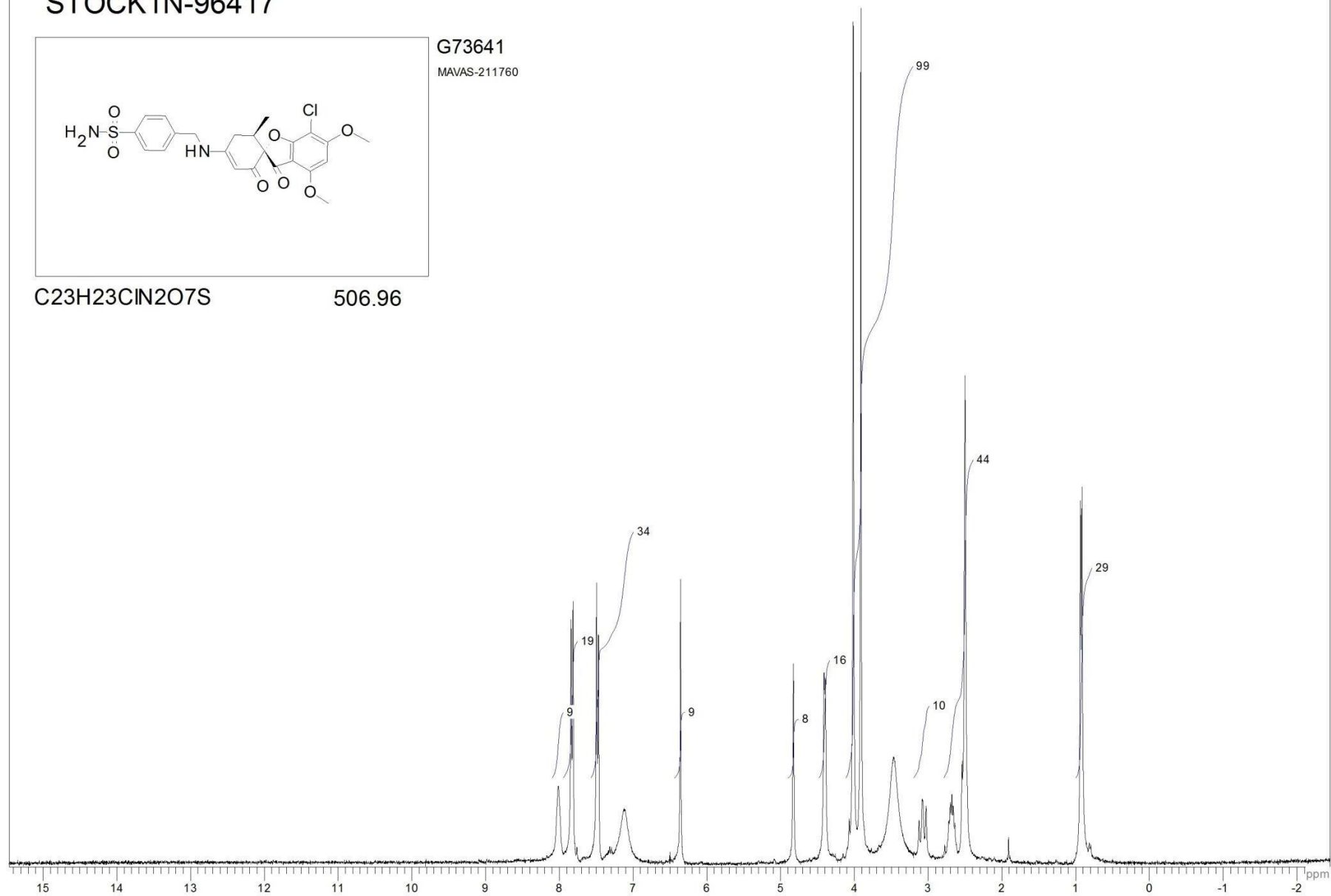
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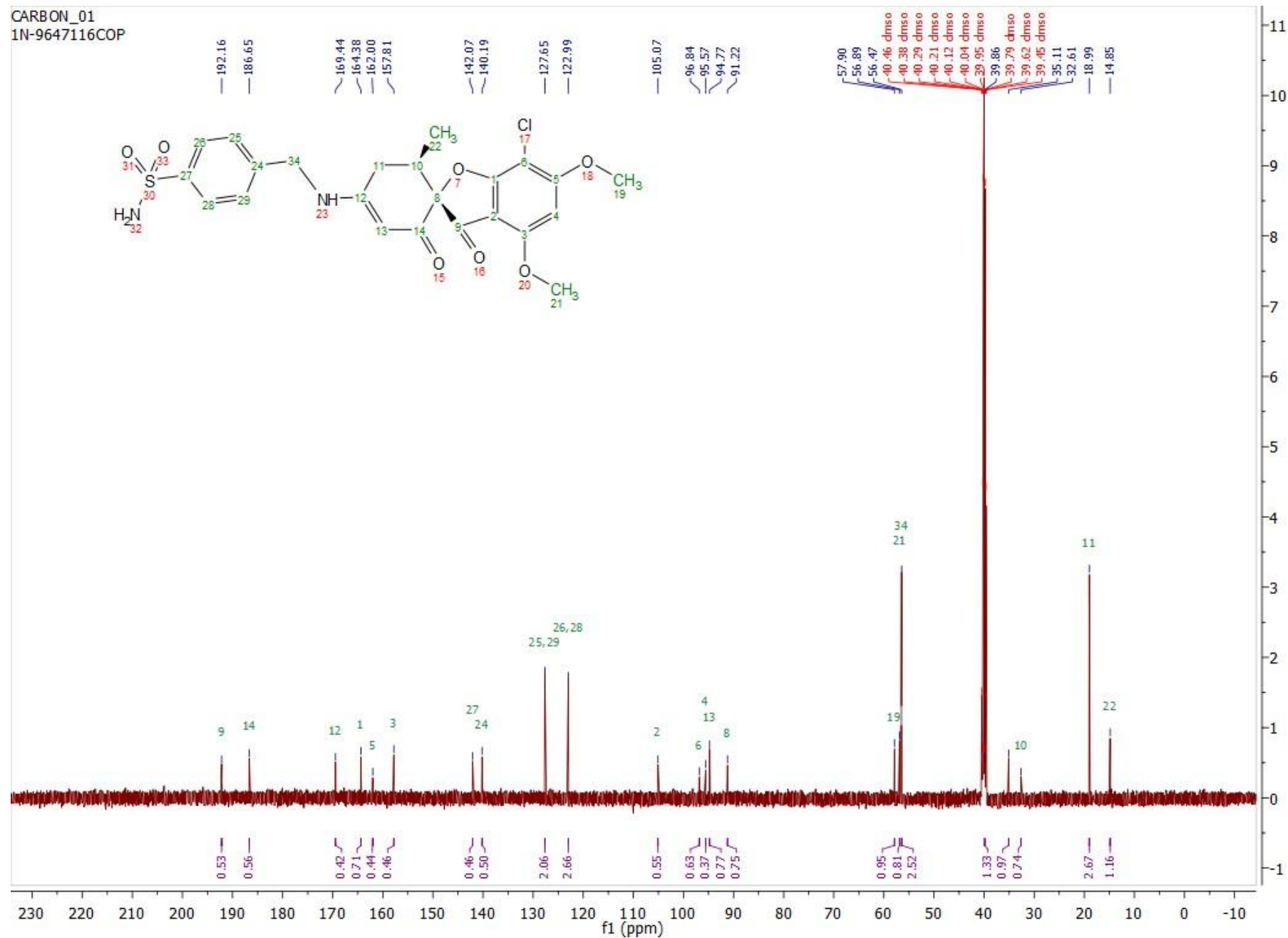
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C23H23ClN2O7S

506.96

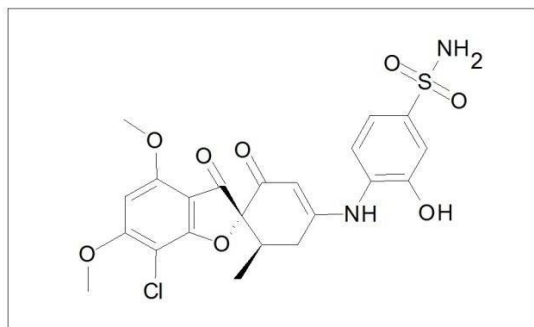


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Compound 1d

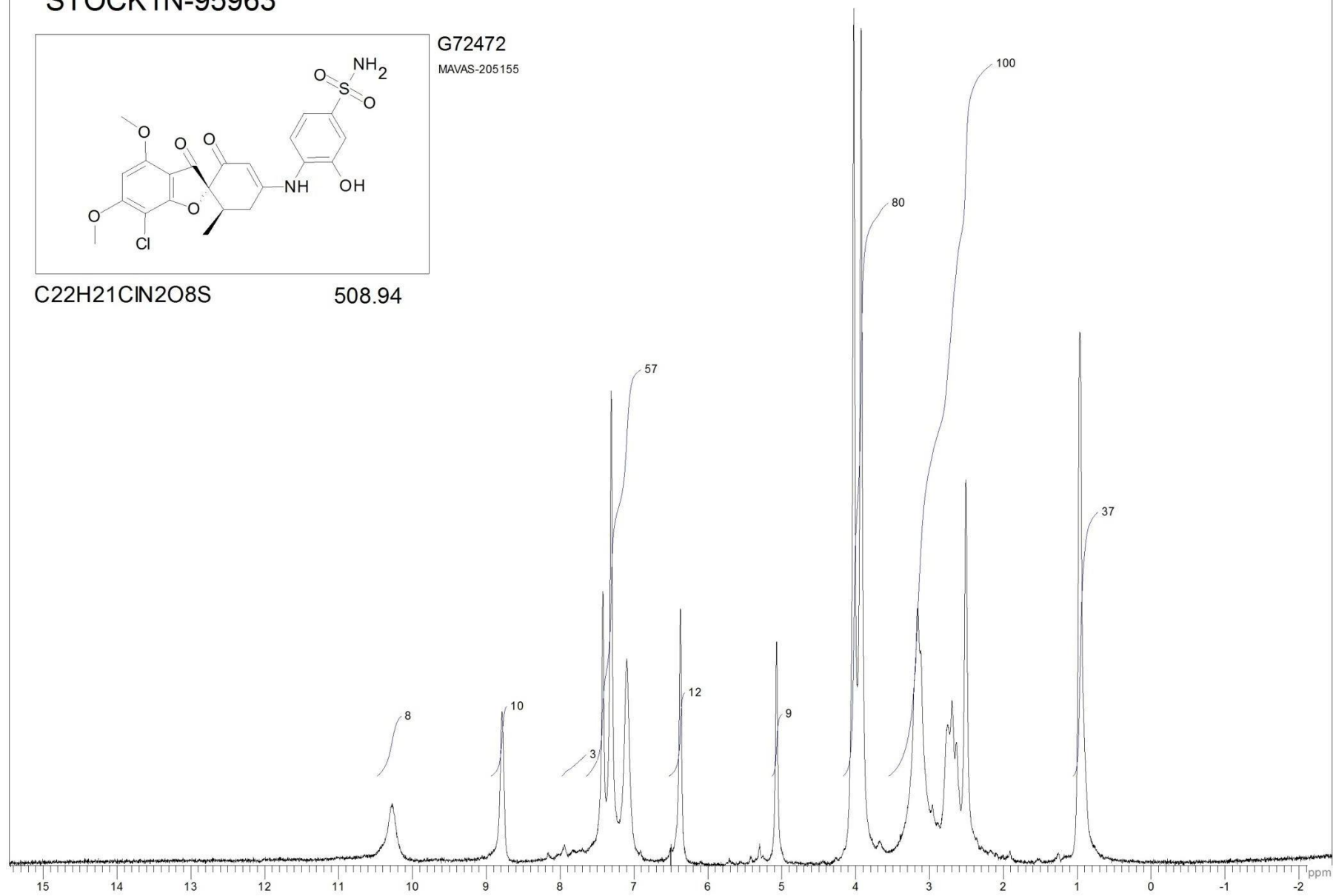
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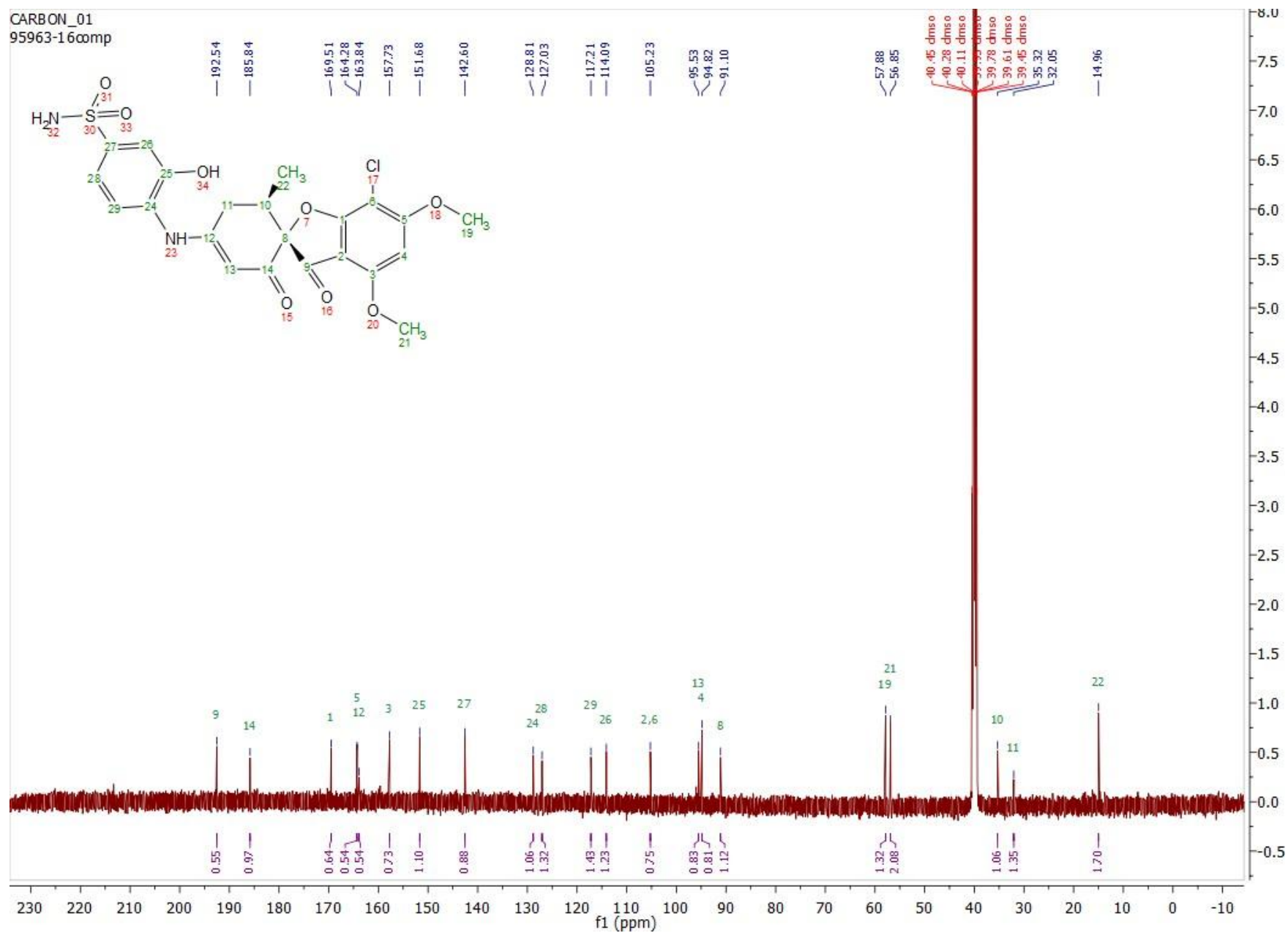
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C₂₂H₂₁ClN₂O₈S

508.94

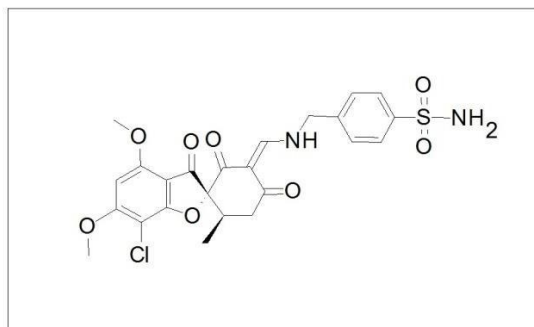


CARBON_01
95963-16amp



Compound 1e

STOCK1N-98537

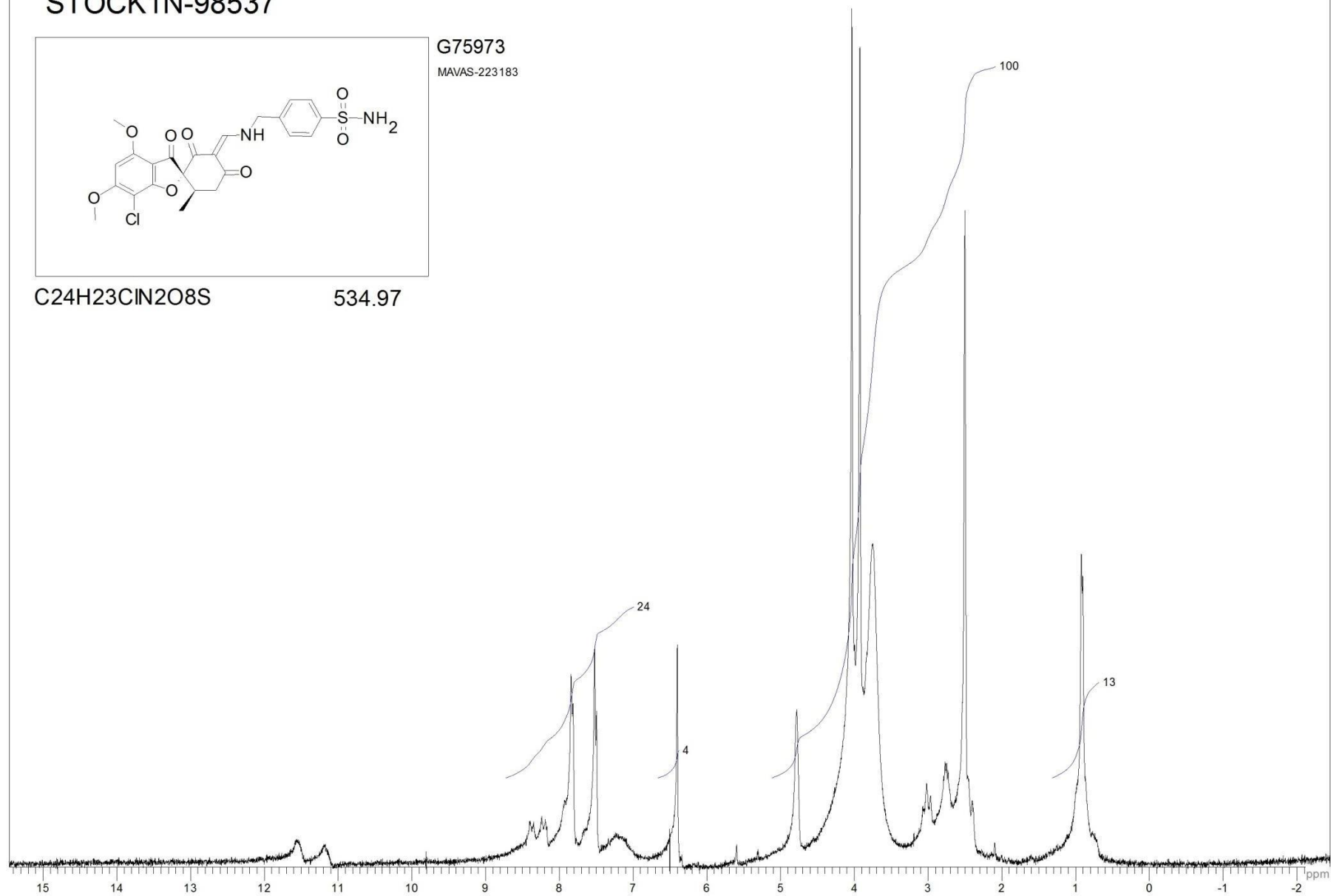


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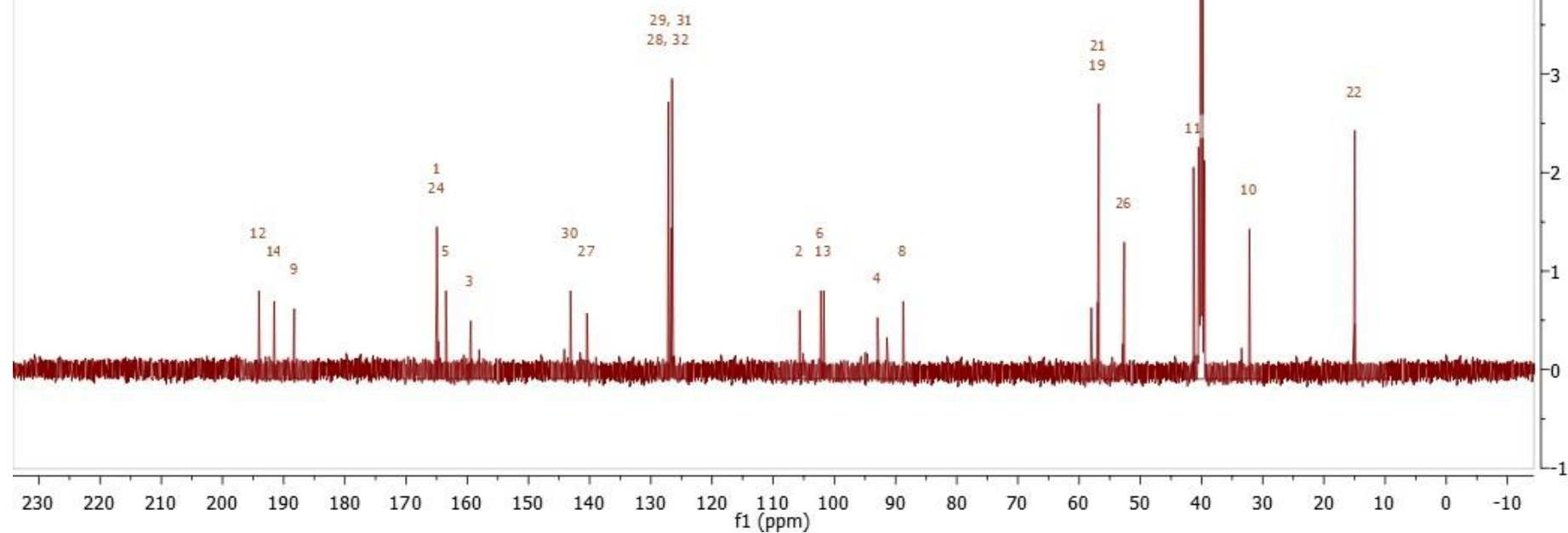
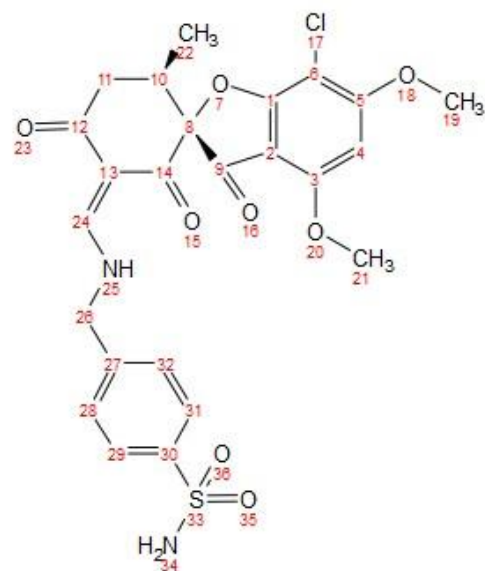
MAVAS-223183

C₂₄H₂₃ClN₂O₈S

534.97

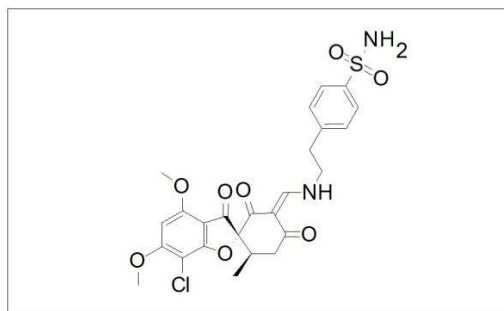


CARBON_01
in-98537



Compound 1f

STOCK1N-97860

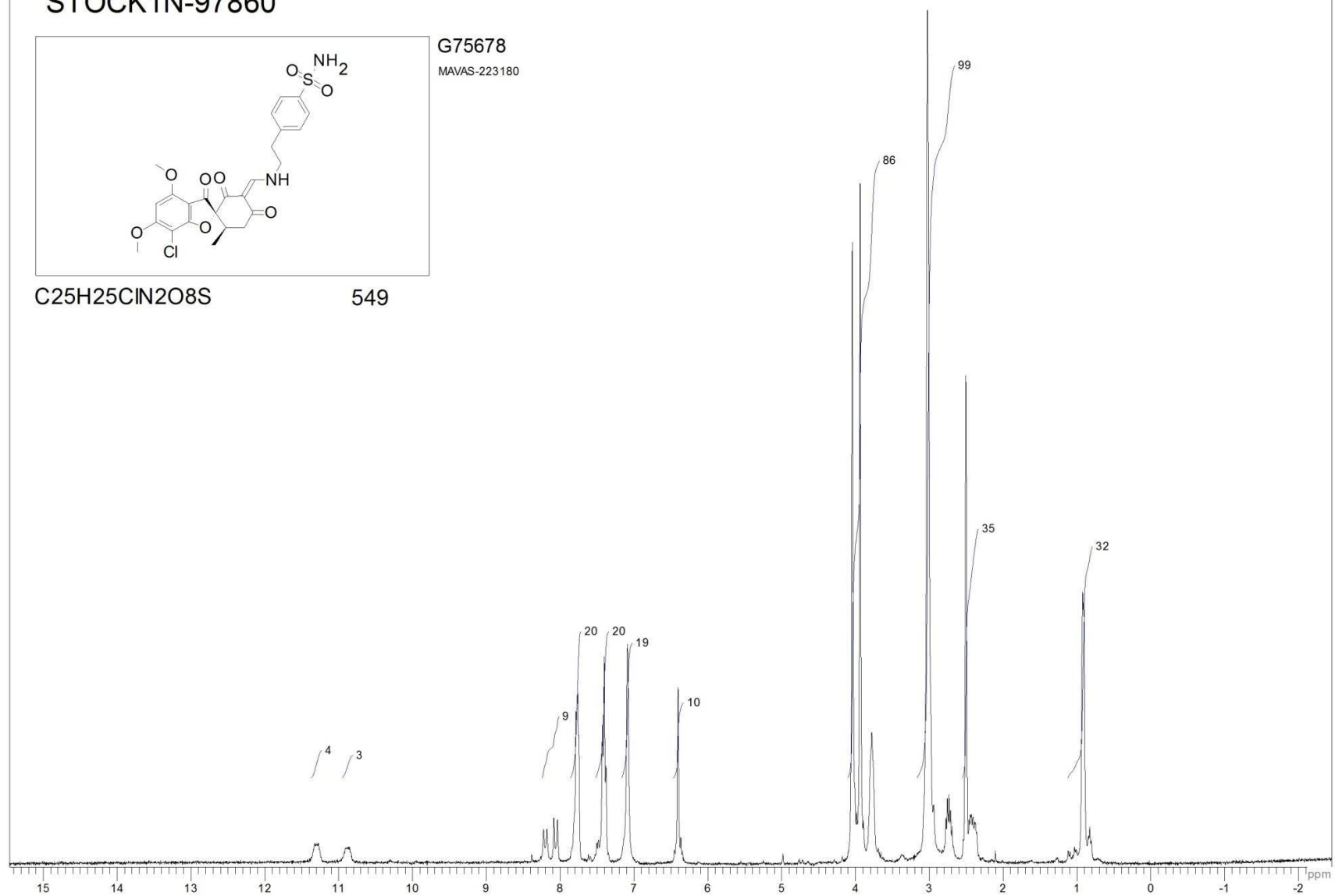


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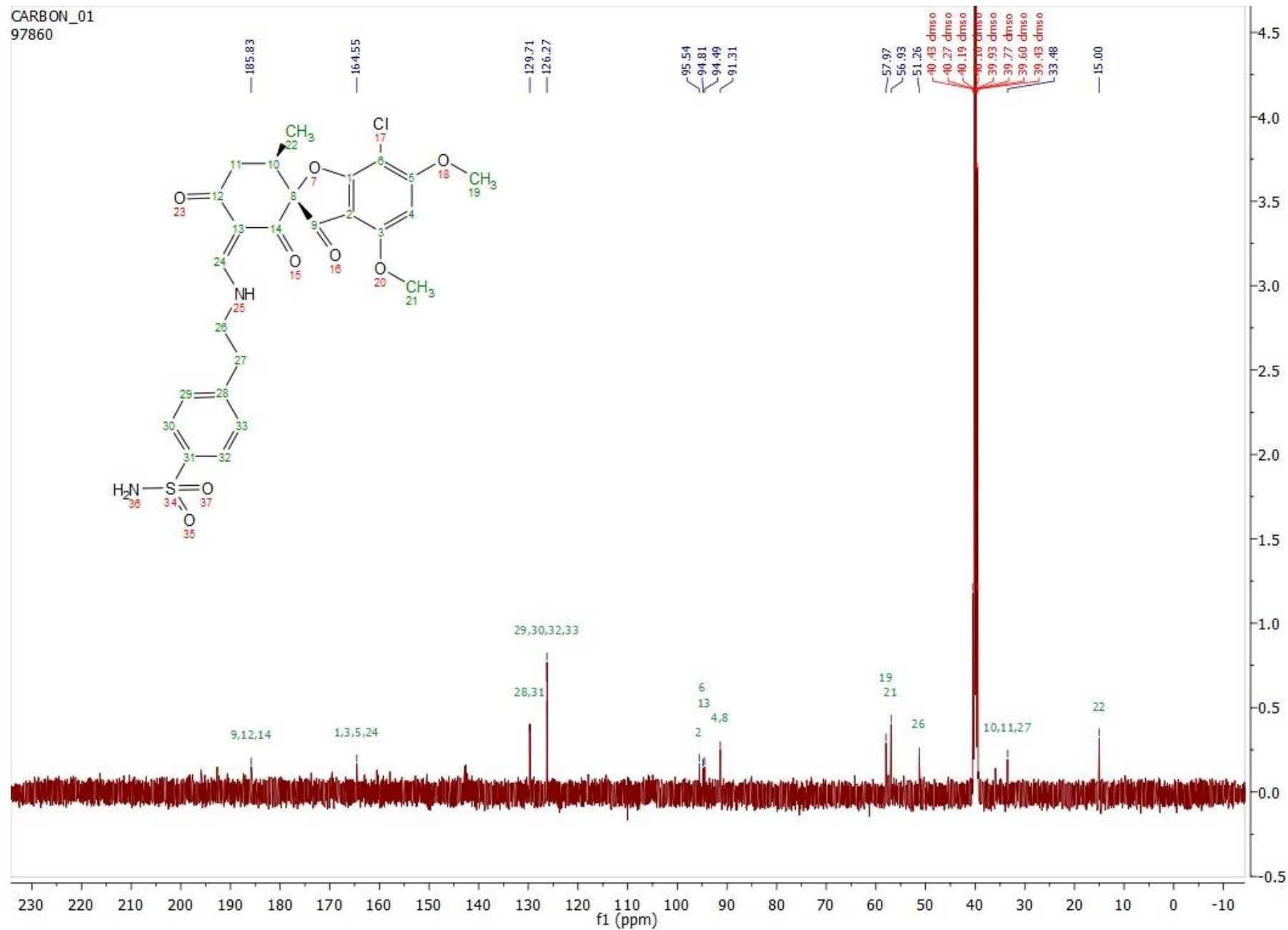
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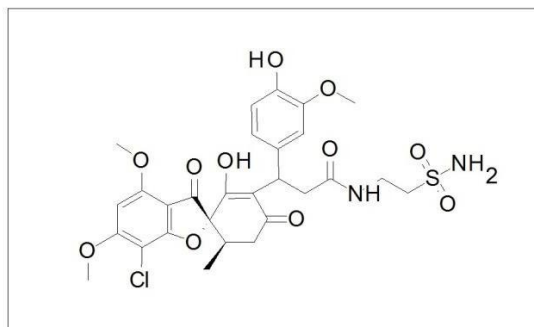


CARBON_01
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Compound 1g

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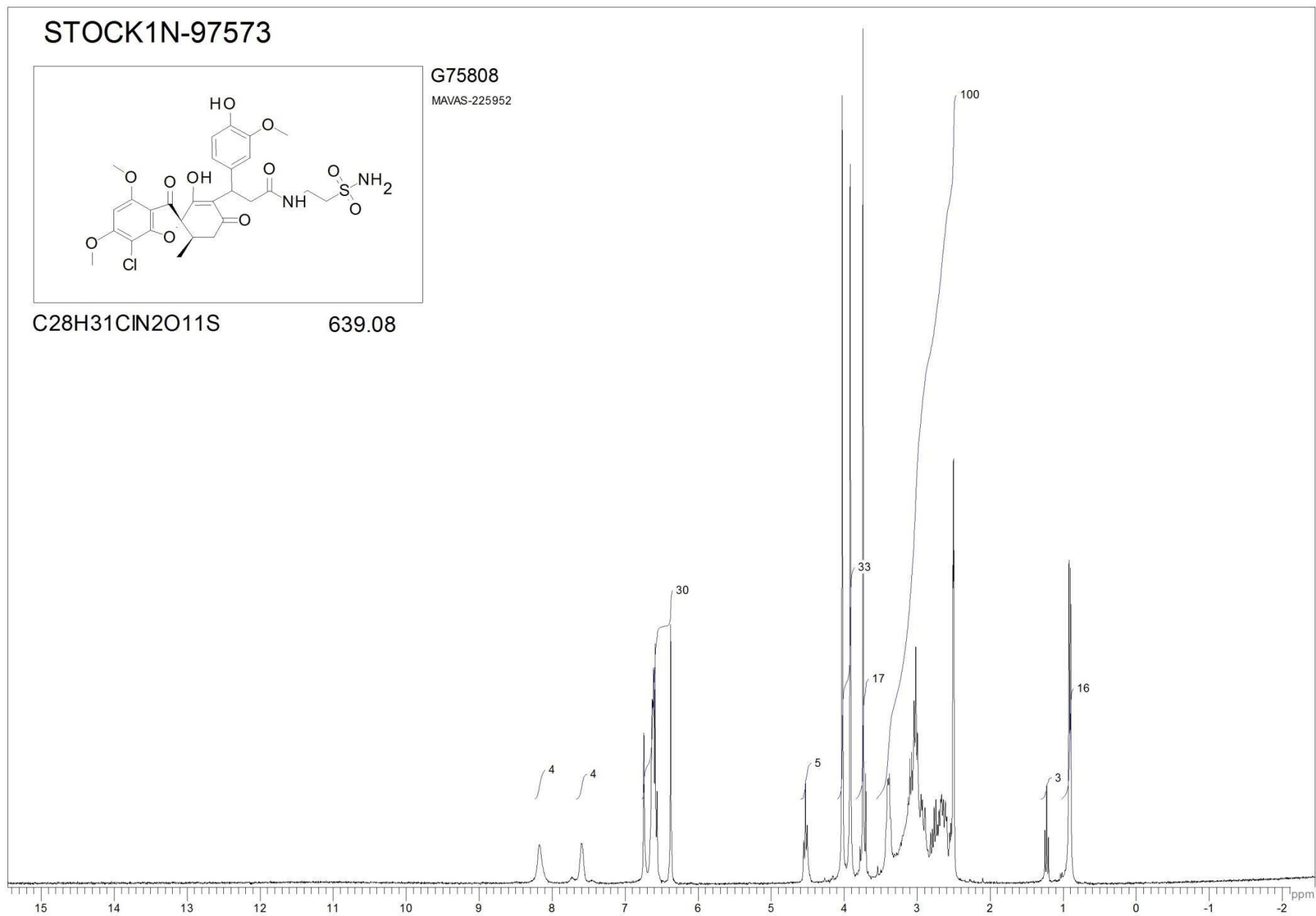


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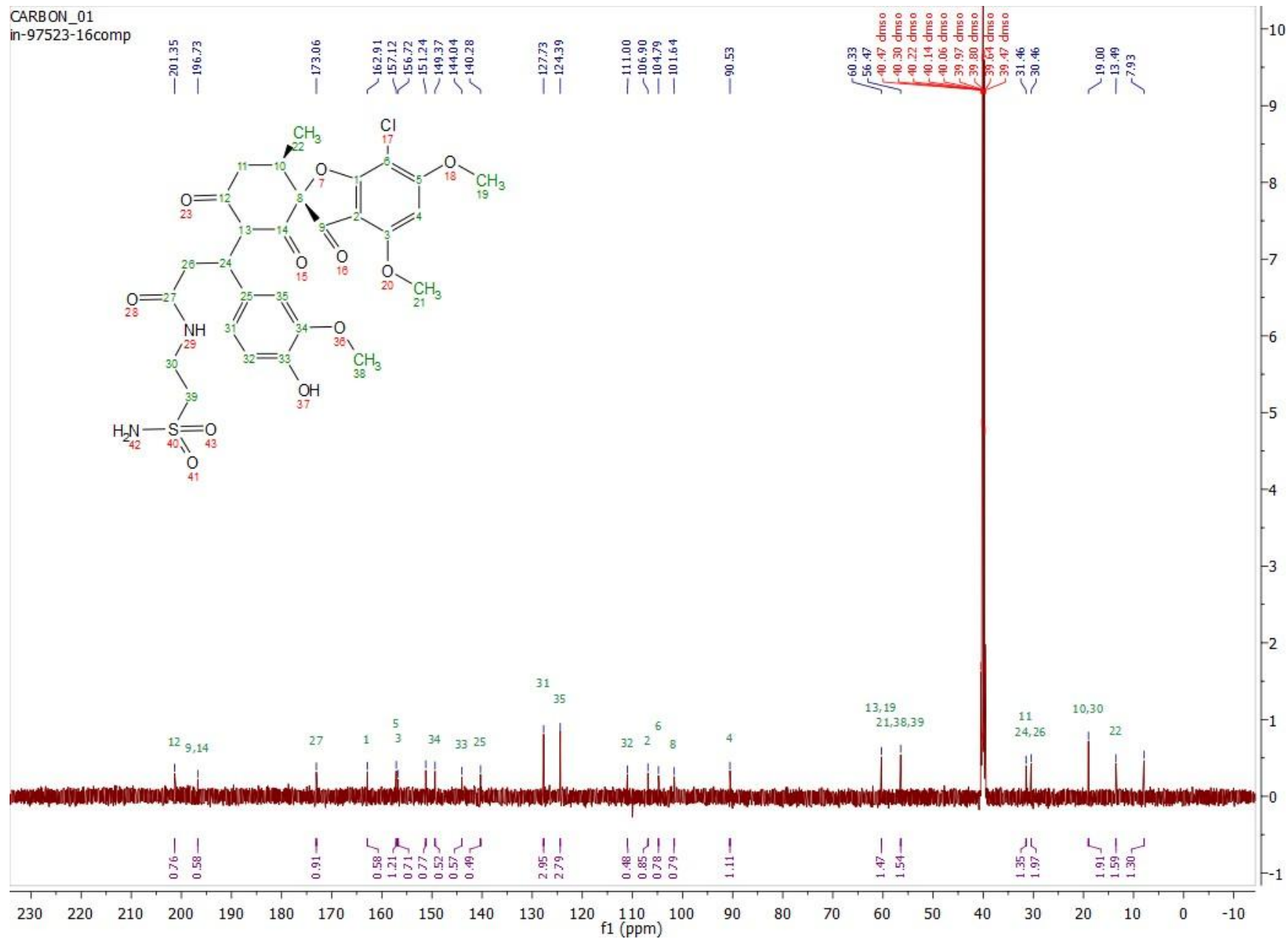
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639.08

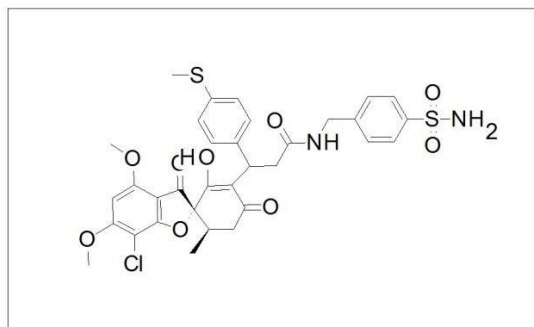


CARBON_01
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Compound 1h

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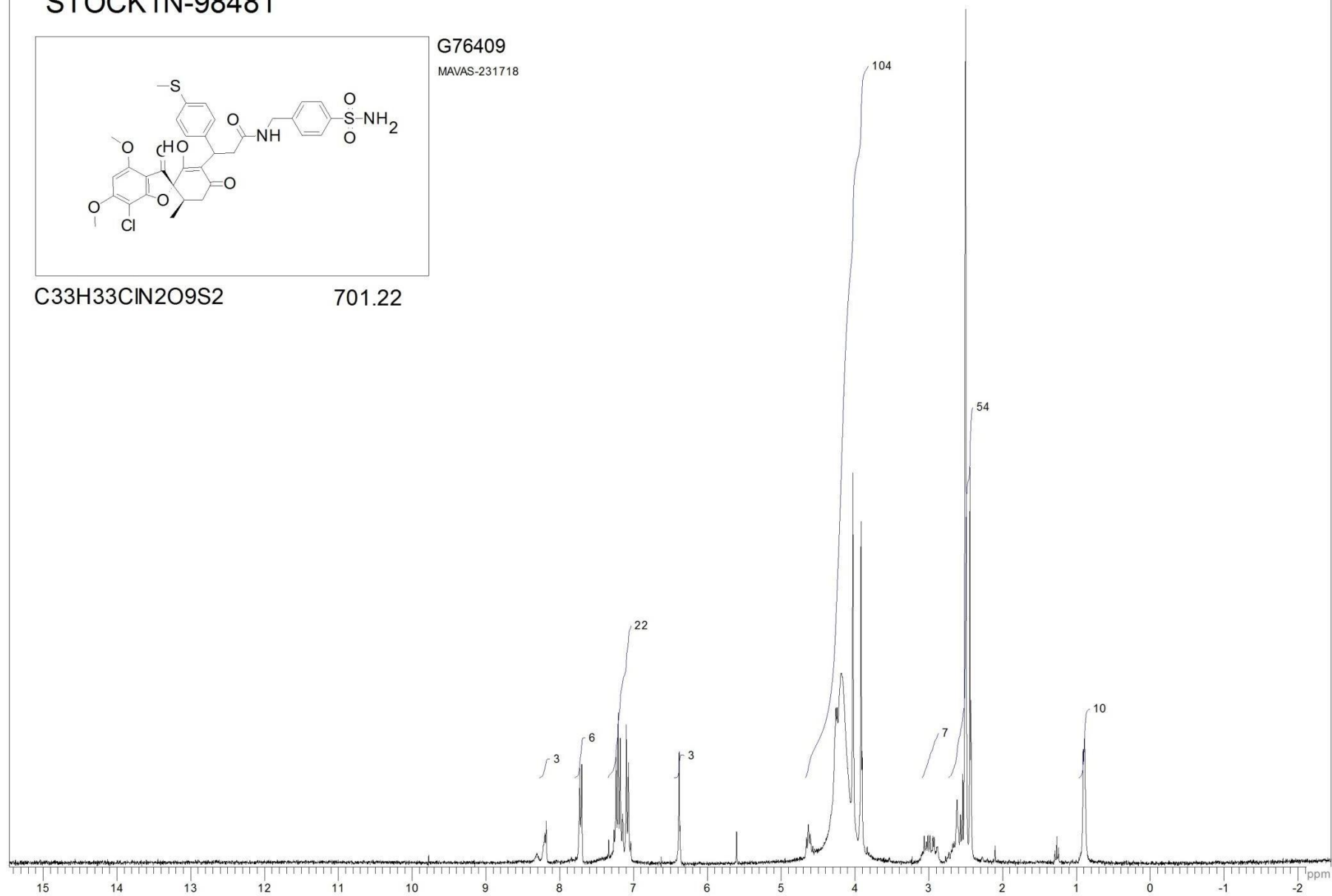


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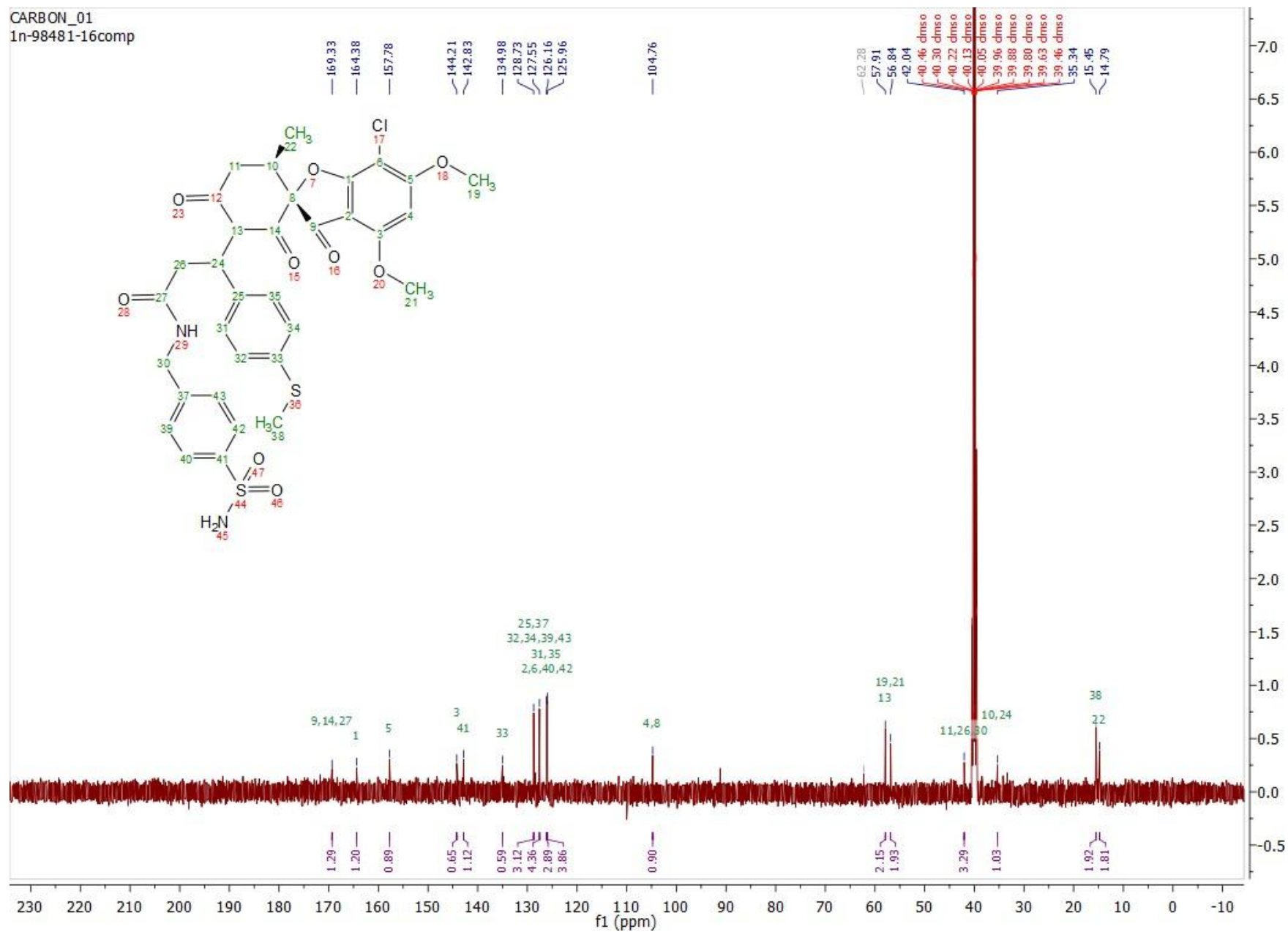
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C₃₃H₃₃ClN₂O₉S₂

701.22

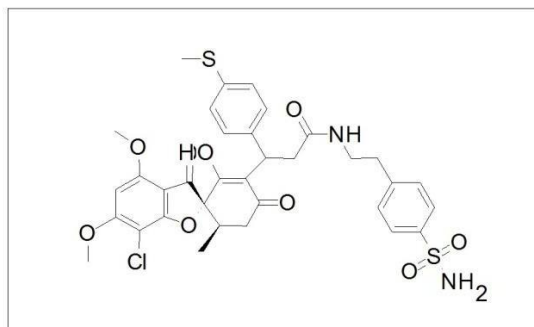


CARBON_01
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Compound 1i

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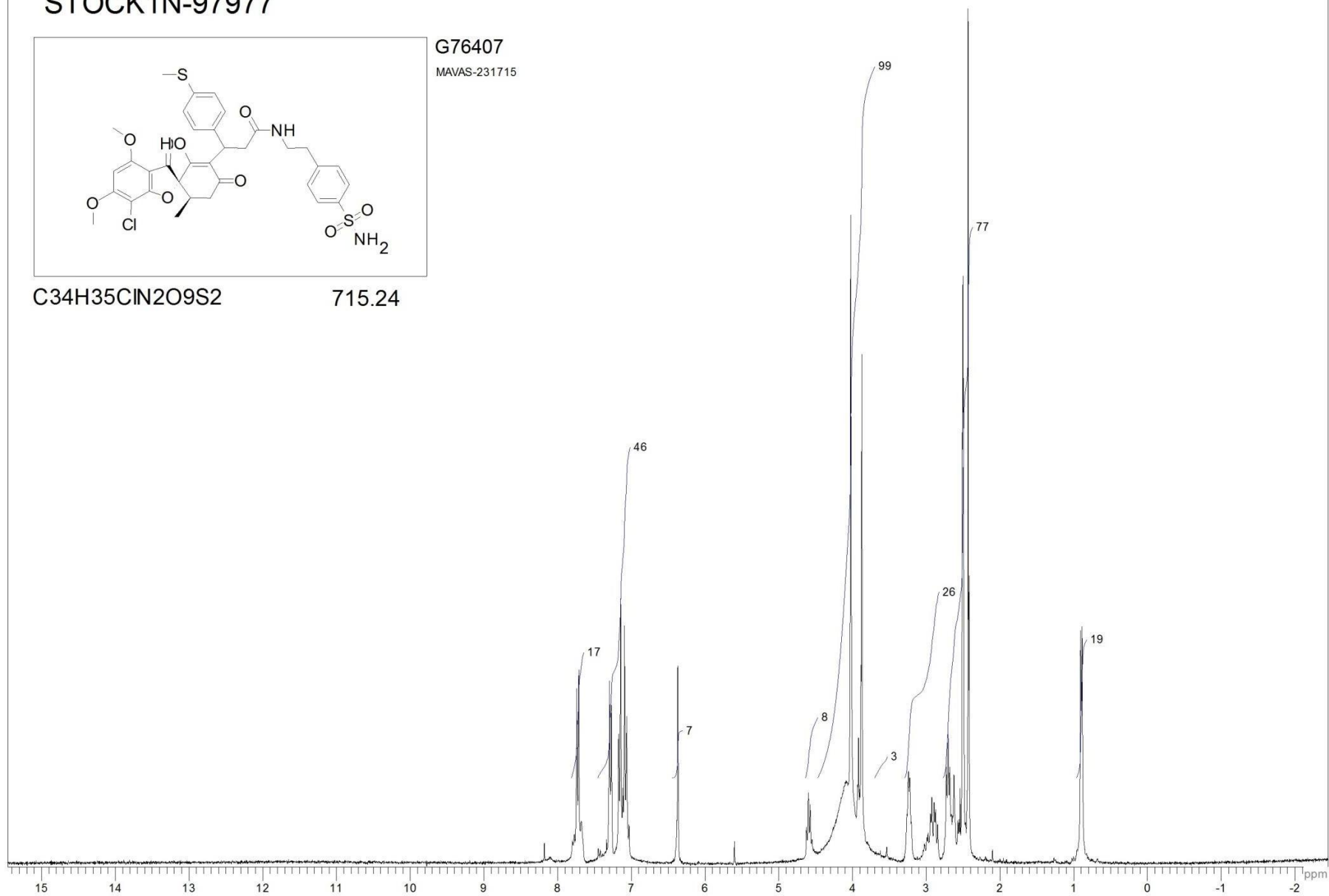


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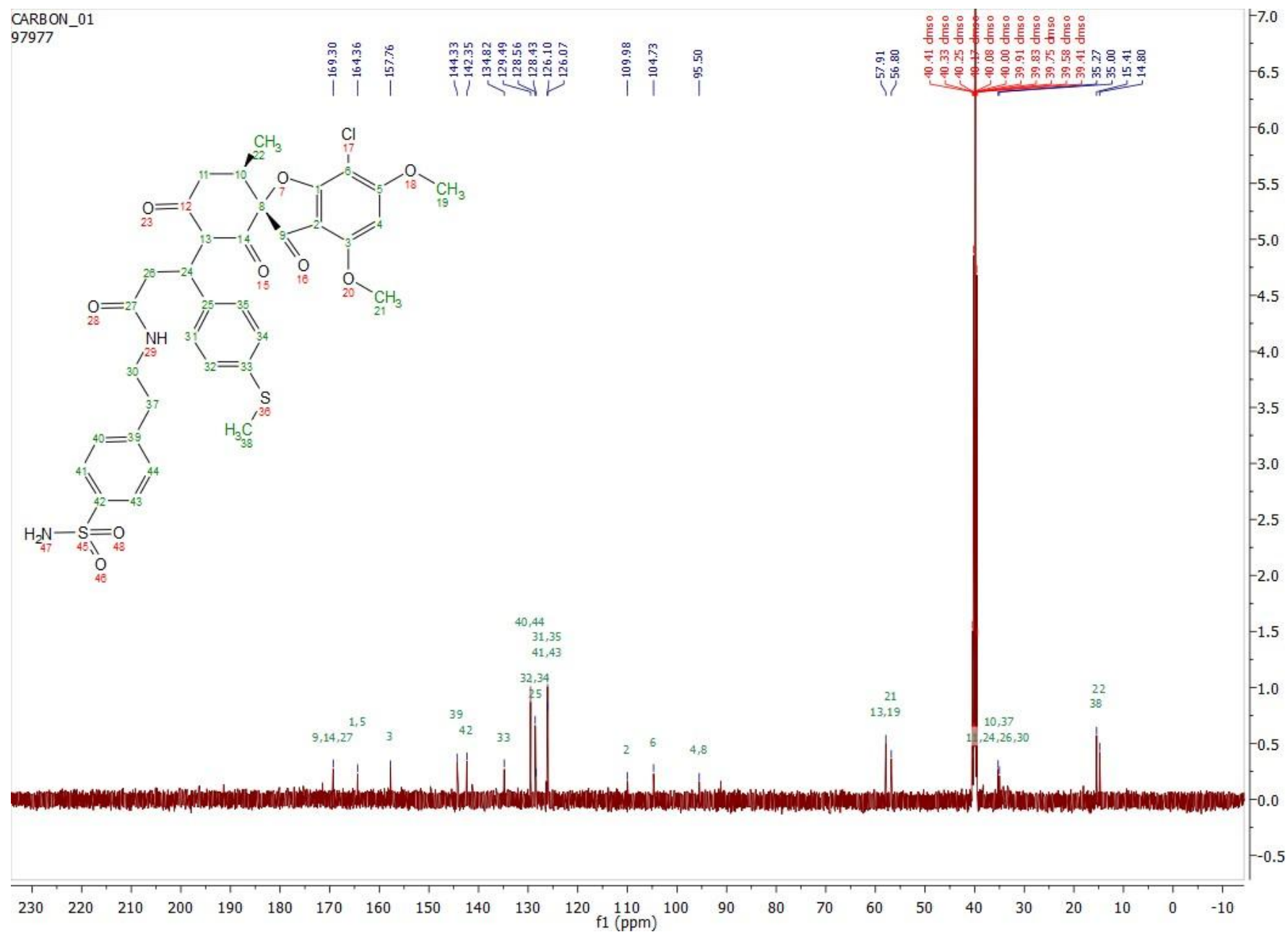
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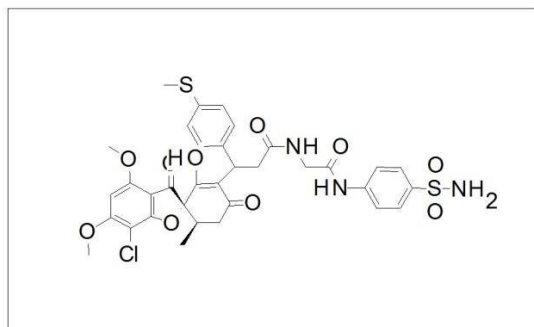


CARBON_01
97977



Compound 1j

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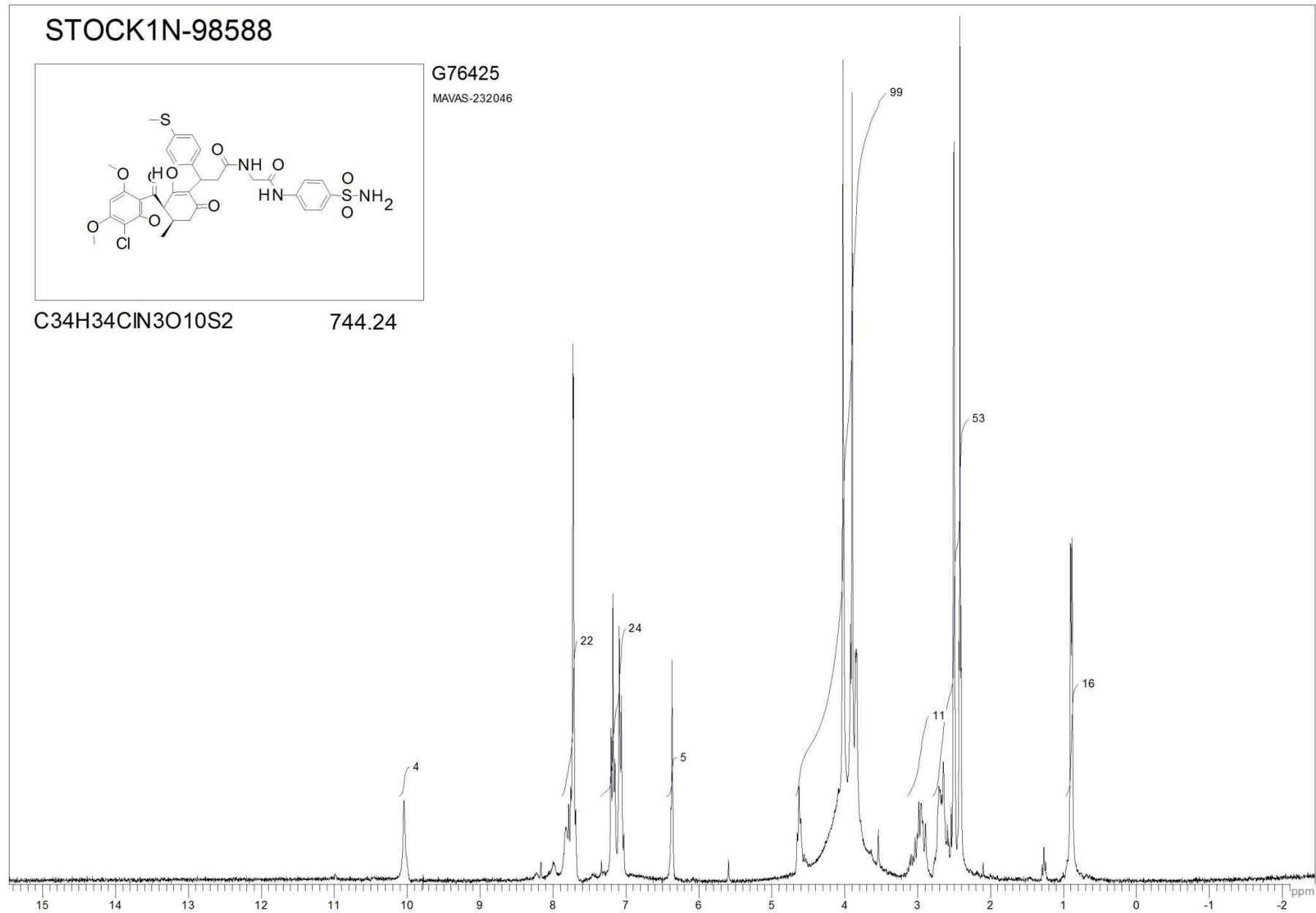


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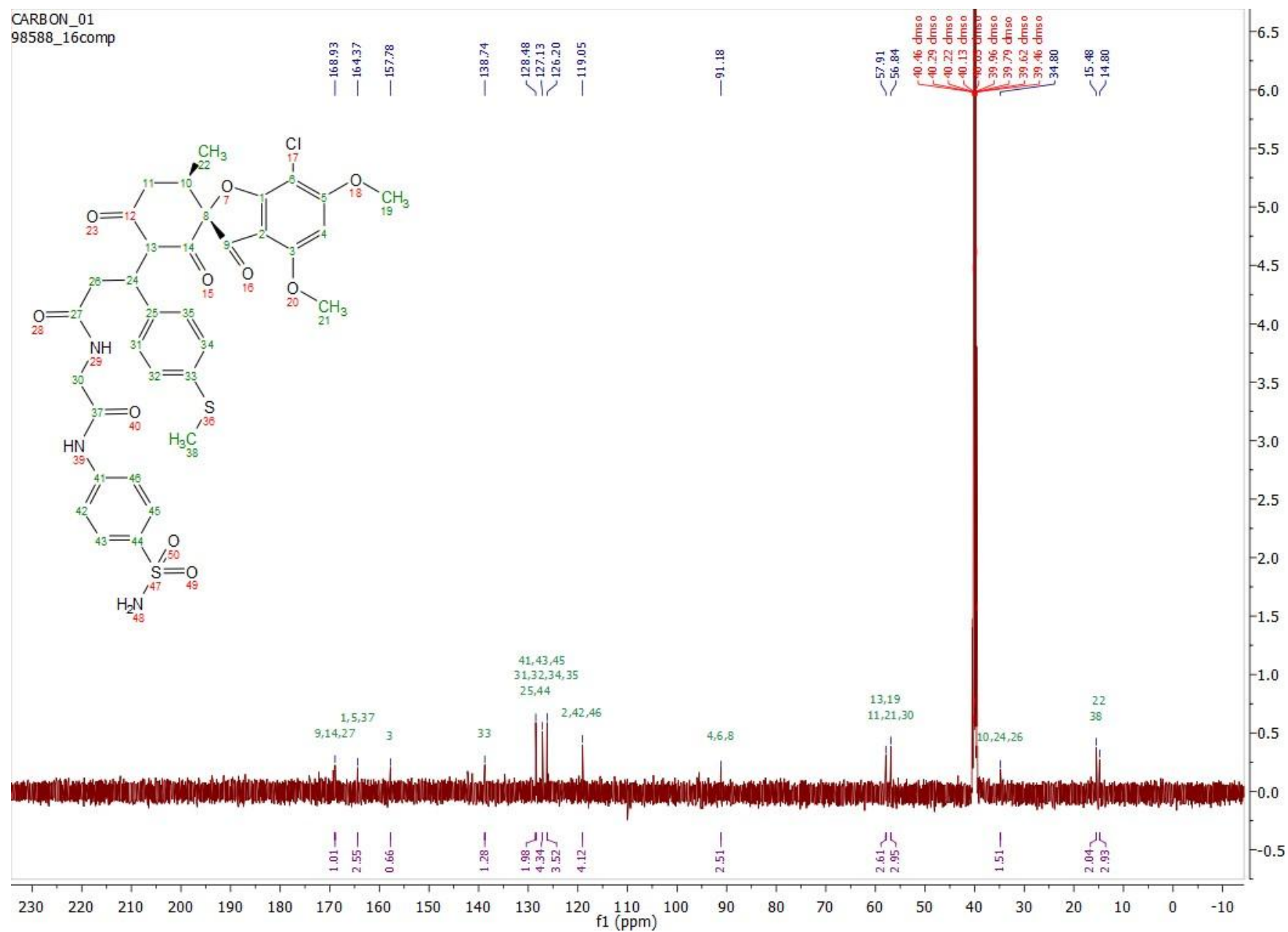
MAVAS-232046

C₃₄H₃₄ClN₃O₁₀S₂

744.24

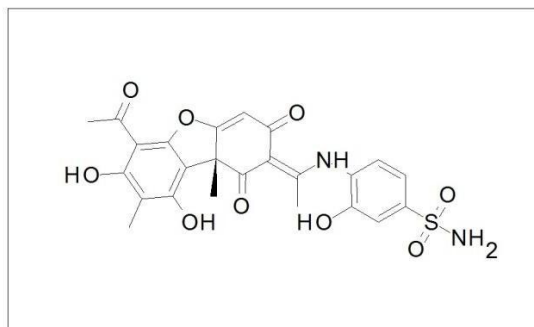


CARBON_01
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Compound 4a

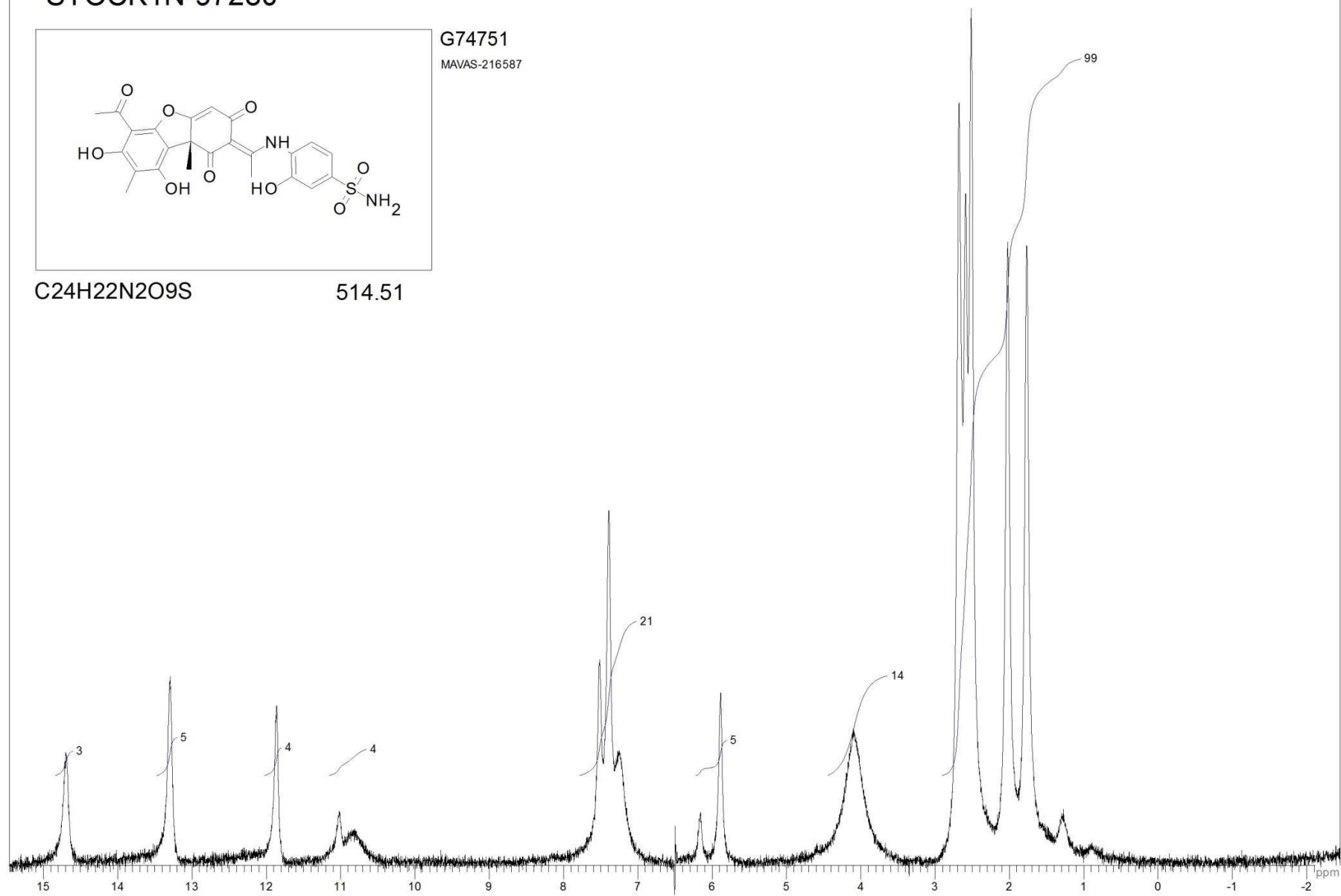
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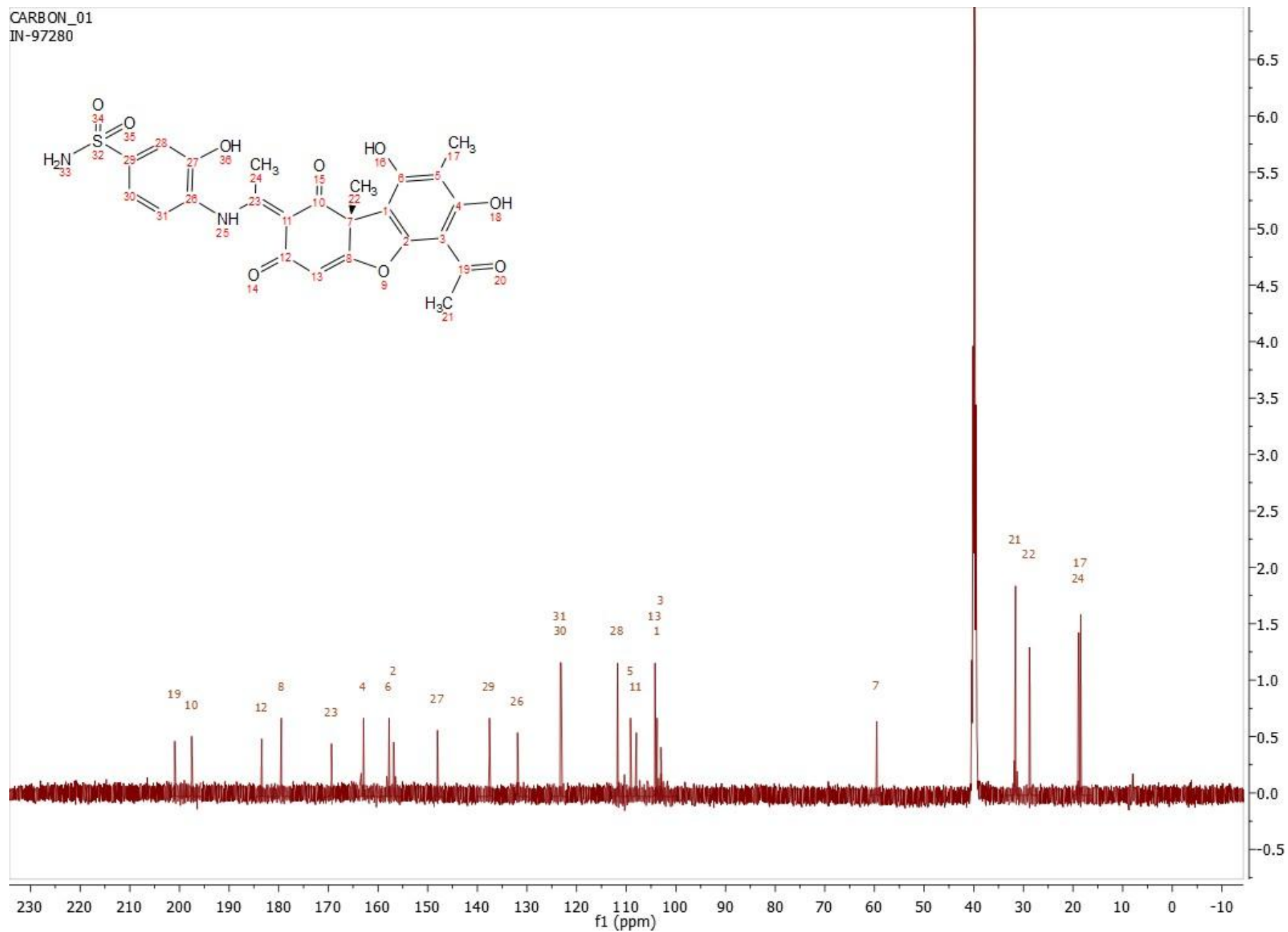
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MAVAS-216587

C₂₄H₂₂N₂O₉S

514.51

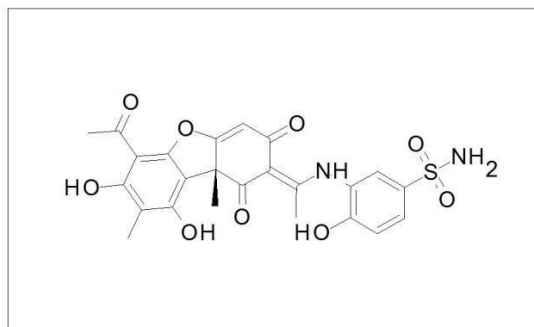


CARBON_01
IN-97280



Compound 4b

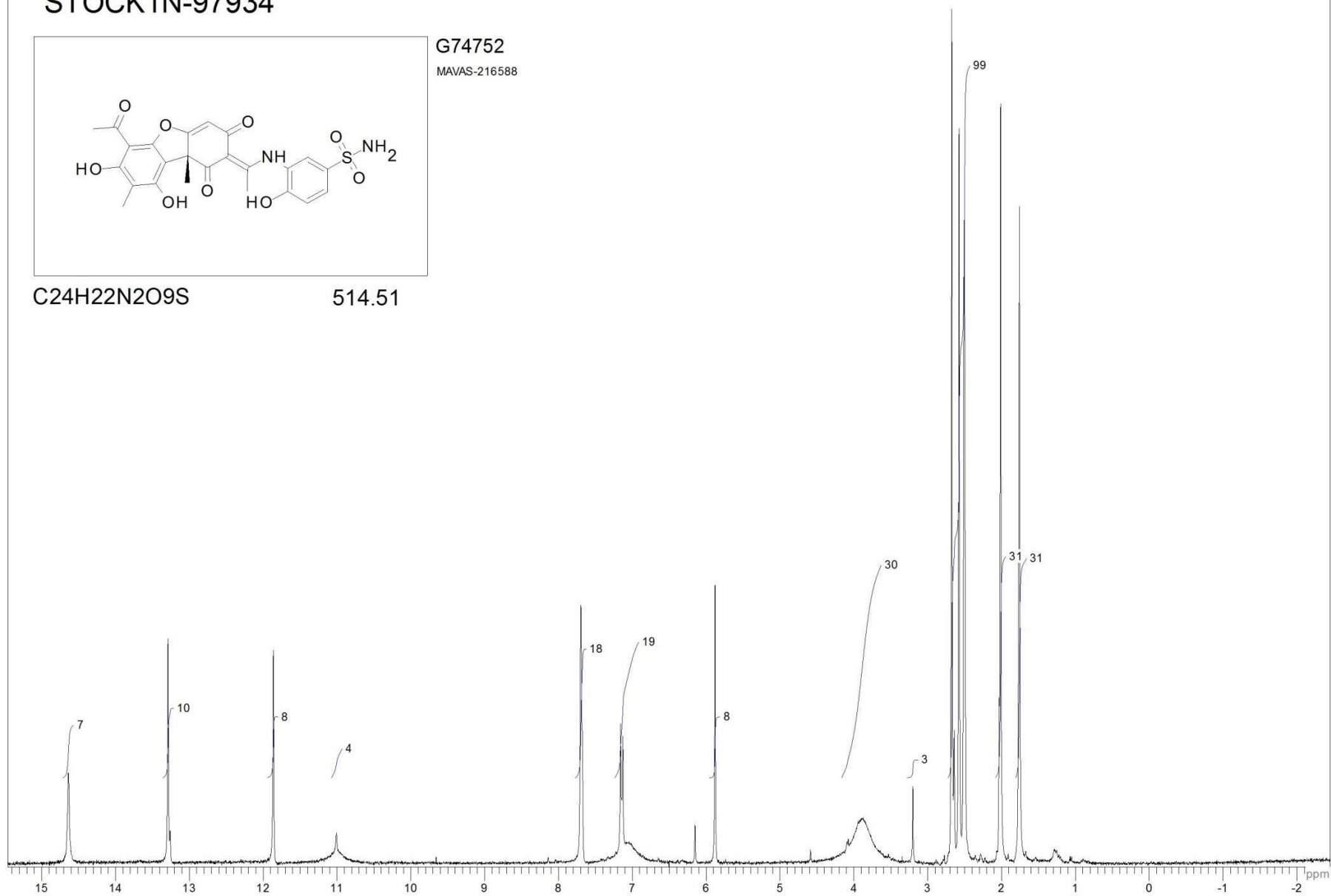
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G74752
MAVAS-216588

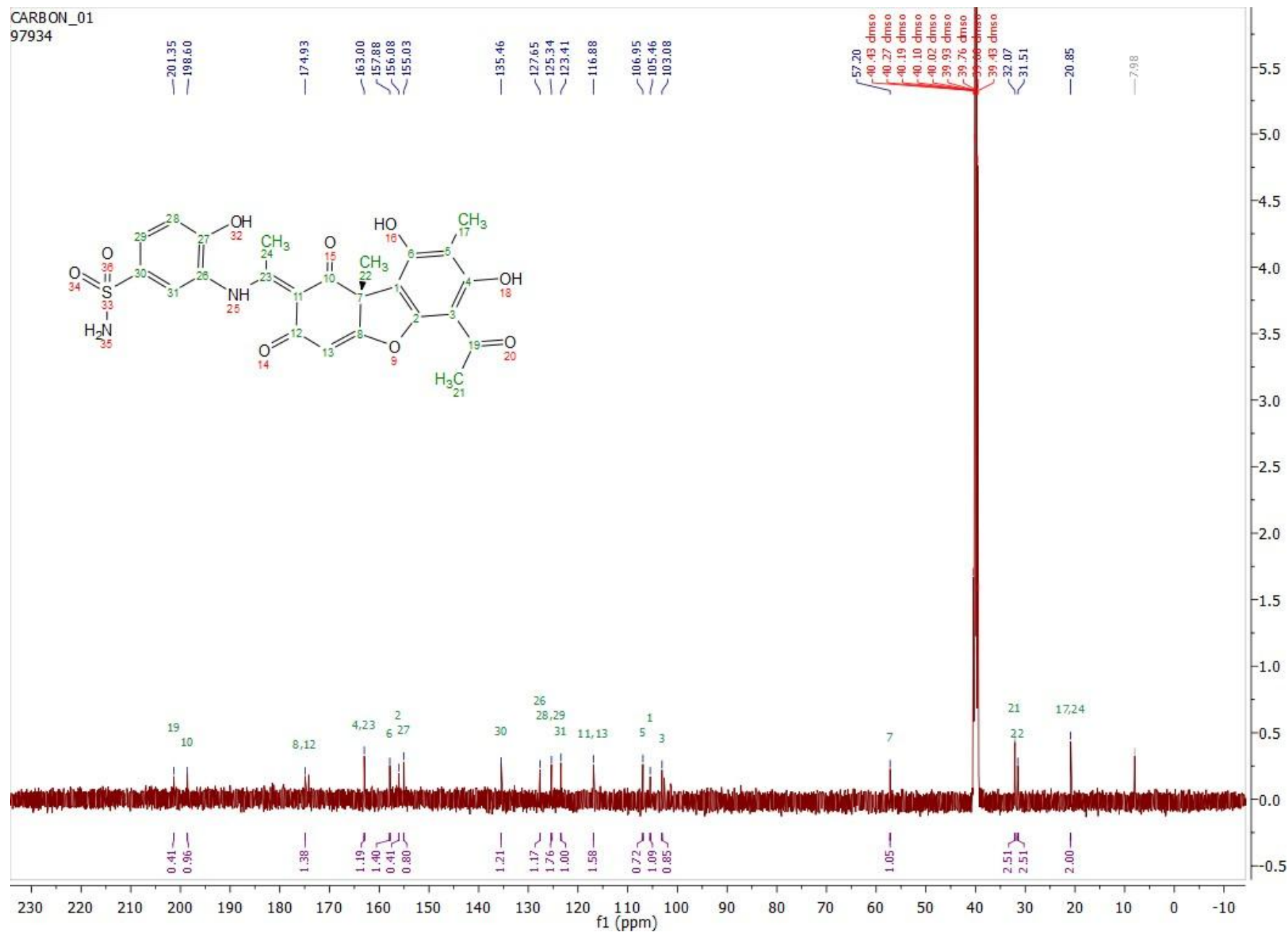
C24H22N2O9S

514.51



CARBON_01

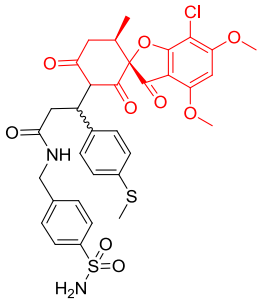
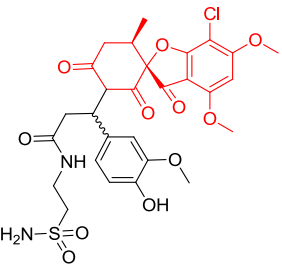
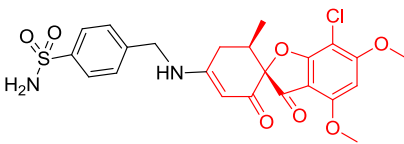
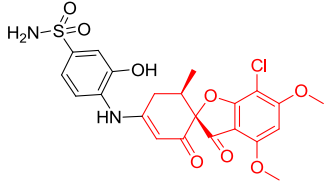
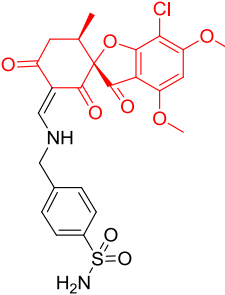
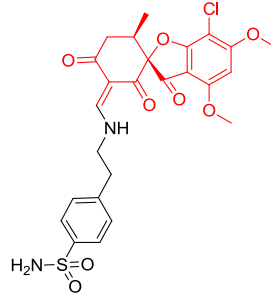
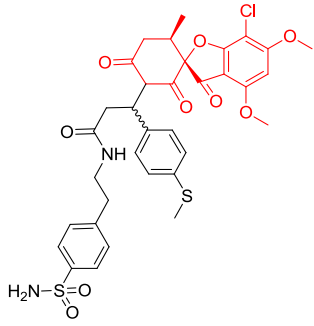
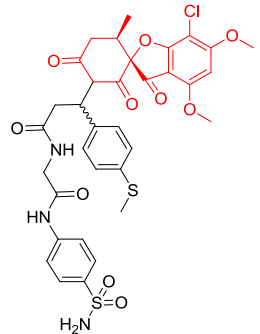
97934



2. Preliminary docking studies

Table S1. Molecular docking free binding energies (kcal/mol) and interactions of designed compounds on hCA I, II and IX isoforms.

Structure	hCA isoform	Estimated free binding energy (Kcal/mol)	Structure	hCA isoform	Estimated free binding energy (Kcal/mol)
	hCA I	-3.20		hCA I	-2.67
	hCA II	-4.64		hCA II	-2.85
	hCA IX	-3.16		hCA IX	-4.32
	hCA I	-3.71		hCA I	-3.60
	hCA II	-4.50		hCA II	-4.16
	hCA IX	-3.62		hCA IX	-5.30
	hCA I	-4.88		hCA I	-4.95
	hCA II	-4.13		hCA II	-3.24
	hCA IX	-5.57		hCA IX	-3.45
	hCA I	-4.29		hCA I	-3.87
	hCA II	-5.74		hCA II	-4.11
	hCA IX	-5.18		hCA IX	-5.46
	hCA I	-6.72		hCA I	-6.25
	hCA II	-10.23		hCA II	-6.24
	hCA IX	-6.57		hCA IX	-8.38
	hCA I	-5.11		hCA I	-5.49
	hCA II	-4.89		hCA II	-7.38

	hCA IX	-6.26		hCA IX	-6.30
	hCA I	-6.13		hCA I	-9.67
	hCA II	-7.39		hCA II	-10.38
	hCA IX	-6.88		hCA IX	-7.70
	hCA I	-9.67		hCA I	-10.35
	hCA II	-9.85		hCA II	-12.02
	hCA IX	-7.32		hCA IX	-8.53
	hCA I	-6.28		hCA I	-5.76
	hCA II	-5.89		hCA II	-6.78
	hCA IX	-8.52		hCA IX	-8.36
	hCA I	-12.45		hCA I	-8.28
	hCA II	-11.64		hCA II	-12.87
	hCA IX	-12.74		hCA IX	-12.02
AAZ	hCA I	-8.28			

hCA II	-8.87
hCA IX	-9.02

3. Prediction of toxicity

Table S2. Prediction of toxicity.

No	Predicted LD50 (mg/kg)	Predicted Toxicity Class	Hepatotoxicity	Carcinogenicity	Mutagenicity	Cytotoxicity
1a	1100	IV	Inactive (0.55)	Inactive (0.58)	Inactive (0.57)	Inactive (0.60)
1b	2000	IV	Inactive (0.54)	Inactive (0.61)	Inactive (0.67)	Inactive (0.74)
1c	1100	IV	Inactive (0.53)	Inactive (0.62)	Inactive (0.64)	Inactive (0.66)
1d	2000	IV	Inactive (0.54)	Inactive (0.60)	Inactive (0.68)	Inactive (0.73)
1e	3500	V	Inactive (0.55)	Inactive (0.59)	Inactive (0.67)	Inactive (0.64)
1f	3500	V	Inactive (0.55)	Inactive (0.61)	Inactive (0.66)	Inactive (0.65)
1g	1100	IV	Inactive (0.55)	Inactive (0.60)	Inactive (0.63)	Inactive (0.60)
1h	5000	V	Inactive (0.52)	Inactive (0.62)	Inactive (0.69)	Inactive (0.64)
1i	5000	V	Inactive (0.53)	Inactive (0.63)	Inactive (0.68)	Inactive (0.64)
1j	5000	V	Inactive (0.56)	Inactive (0.63)	Inactive (0.68)	Inactive (0.64)
4a	838	IV	Inactive (0.54)	Inactive (0.58)	Inactive (0.67)	Inactive (0.74)
4b	838	IV	Inactive (0.54)	Inactive (0.58)	Inactive (0.67)	Inactive (0.74)

Number in brackets indicate possibilities.

4. Molecular Docking Studies

Software Used

The ligand preparations done by using chemdraw 12.0, geometries were optimized using LigandScout 4.4.5. The "Build/check/repair model" to the session "Prepare PDB file for docking programs" module was used for proteins preparation. For the finally preparation of both ligands and protein preparation Wizard of AutoDock tools 1.5.6 are used. Autodock 4 (ver. 4.2.6) was employed for docking simulations and Autogrid4 for affinity grid maps preparation [1]. The resulting poses and potential interactions were visualized using the LigandScout 4.4.5 program.

Protein and ligand preparation

X-ray crystal structures of *hCA I* (PDB code 3W6H) and *hCA II* (PDB code 3HS4) cytosolic isoforms as well as *hCA IX* (PDB code 3IAI) transmembrane tumor-associated isoform were retrieved from Brookhaven Protein Data Bank (PDB) [2]. The pdb files of proteins were submitted to "Build/check/repair model" to the session "Prepare PDB file for docking programs" and missing side chains were modeled in, water positions and symmetry were corrected, and hydrogen atoms were added. Only chain A of each enzyme of the repaired pdb file was evaluated and passed to AutodockTools (ADT ver.1.5.6) for pdbqt file preparation. ADT assigned polar hydrogens, water molecules and non-standard residues were removed, only polar hydrogen was maintained, and Gasteiger charges were computed for protein atoms. AutoDock saved the prepared file in PDBQT format.

All the molecules were sketched in chemdraw 12.0 program. The geometry of built compounds was optimized using the molecular mechanical force fields 94 (MMFF94) energy via program LigandScout, partial charges were also calculated, conformers of each ligand were generated and the one with the best conformation was maintained and saved as mol2 files that were passed, as usual, to ADT for pdbqt file preparation. There, polar hydrogen was added to each structure, followed by computing Gasteiger and Kollman charges, and the torsions.

Docking Procedure

Autodock4 (ver. 4.2.6) was employed for docking simulations. A computationally (relatively) 'hybrid' force field that contains terms based on molecular, mechanics and empirical terms is used by AutoDock. The evaluation step in a nutshell includes: first, calculation of the energy of protein and ligand in the unbound state. Second, calculation of the energy of the ligand-protein complex and take the difference between first and second step.

$$\Delta G = (V_{\text{bound}}^{L-L} - V_{\text{unbound}}^{L-L}) + (V_{\text{bound}}^{P-P} - V_{\text{unbound}}^{P-P}) + (V_{\text{bound}}^{P-L} - V_{\text{unbound}}^{P-L} + \Delta S_{\text{conf}})$$

where P refers to the protein, L refers to the ligand, V are the pair-wise evaluations mentioned above and ΔS_{conf} denotes the loss of conformational entropy upon binding [3]. The ligand molecule is in an arbitrary conformation, orientation and position and this molecular docking program finds favorable poses in a protein-binding site using Lamarckian genetic algorithms implemented therein to search for the best conformers.

Lamarckian genetic algorithm was used as search engine, with a total of 100 runs. The region of interest, used by Autodock4 for docking runs and by Autogrid4 for affinity grid maps preparation, was defined in such a way to comprise the whole catalytic binding site using a grid of 50 x 50 x 50 points with a grid space of 0.375 Å. All parameters used in docking were default. The translation, quaternion and torsions steps were taken from default values in AutoDock. The Lamarckian genetic algorithm and the pseudo-Solis and Wets methods were applied for minimization using default parameters. The number of docking runs was 100. After docking, the 100 solutions were clustered into groups with RMS lower than 1.0 E. The clusters were ranked by the lowest energy representative of each cluster. Upon completion of docking, the best poses were screened by examination of binding energy ($\Delta G_{\text{binding}}$, kcal/mol) and number in cluster. In order to describe the ligand-binding pocket interactions, the top ranked binding mode found by AutoDock in complex with the binding pocket of enzyme was selected. The resulting poses and potential interactions were visualized using the LigandScout 4.4.5. Finally, the docking protocol was verified by re-docking of the co-crystallized ligand acetazolamide (AAZ) in the vicinity of the active sites of each enzyme with RMSD values 0.885, 0.966 and 1.034 Å for *hCA I*, *II* and *IX*, respectively.

1. Morris, G. M., Huey, R., Lindstrom, W., Sanner, M. F., Belew, R. K., Goodsell, D. S. and Olson, A. J. (2009) Autodock4 and AutoDockTools4: automated docking with selective receptor flexibility. *J. Computational Chemistry* **2009**,16: 2785-91.
2. <http://www.rcsb.org/>. (Accessed 19/6/2021).
3. R. Huey, G.M. Morris, A.J. Olson, and D.S. Goodsell, A semiempirical free energy force field with charge-based desolvation, *J. Comp. Chem.* **28** (2007), pp. 1145–1152.