

Recombinant peptide production softens *Escherichia coli* cells and increases their size during C-limited fed-batch cultivation

Supplementary materials

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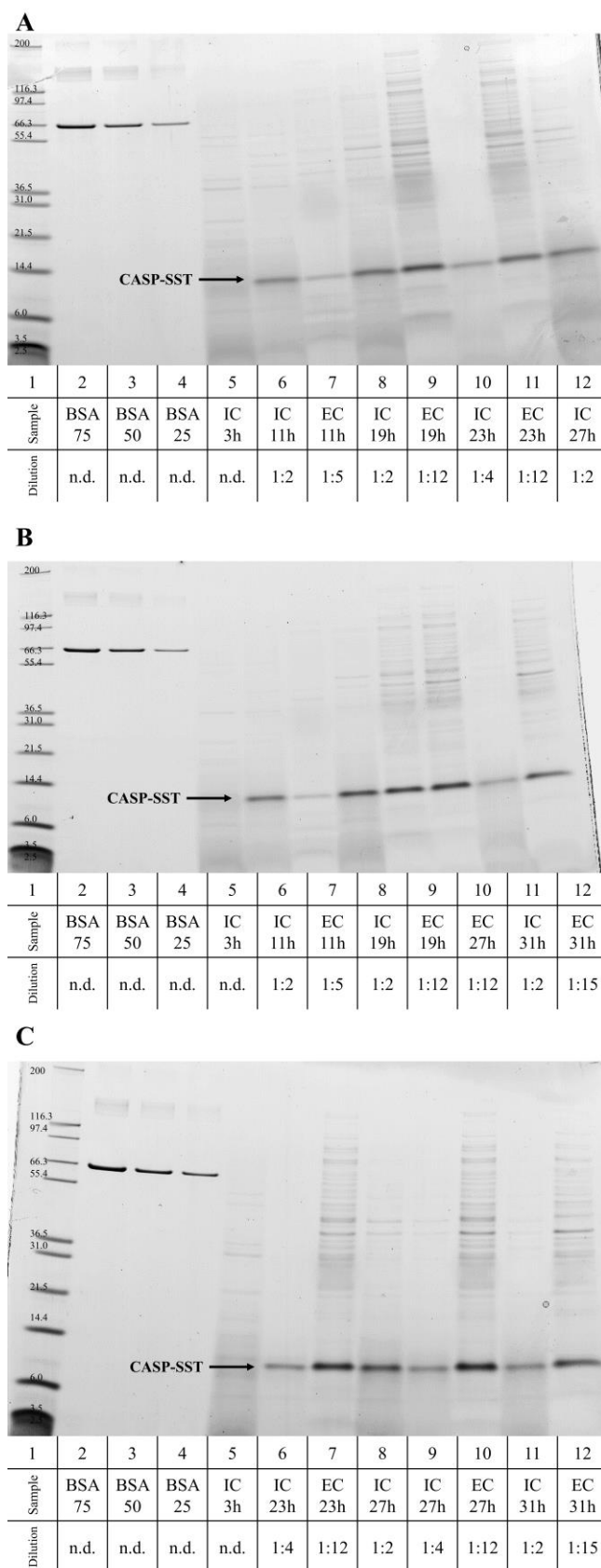


Figure S1. SDS-PAGE gels (panels A–C) for quantification of intracellular (IC) and extracellular (EC) peptide content. All samples were analysed at least twice. 3 h samples represent uninduced reference samples. CASP-SST bands (black arrow) were quantified via ImageQuant™ using BSA standards (lanes

2-4, 75-25 $\mu\text{g mL}^{-1}$). Samples were diluted to fit the linear range of the standard, and non-diluted samples are indicated as n.d.

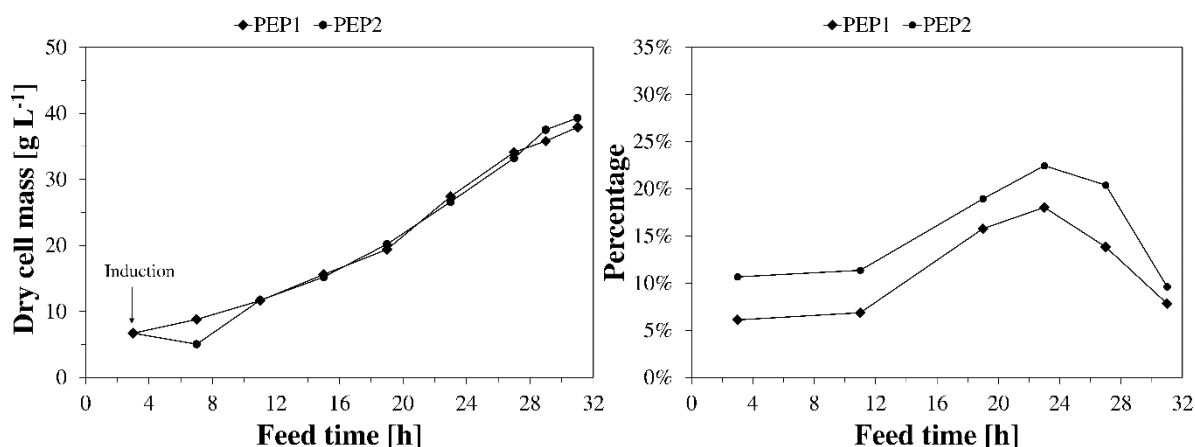


Figure S2: Growth curves (A) and cell lysis curves (B) of duplicate cultivations with PEP. Minor deviations in DCM (e.g. after 7 h) likely arise from errors during sample preparation. Cell lysis variation might arise from differences in timepoint of sample preparation and DNA measurement (DNA instability).

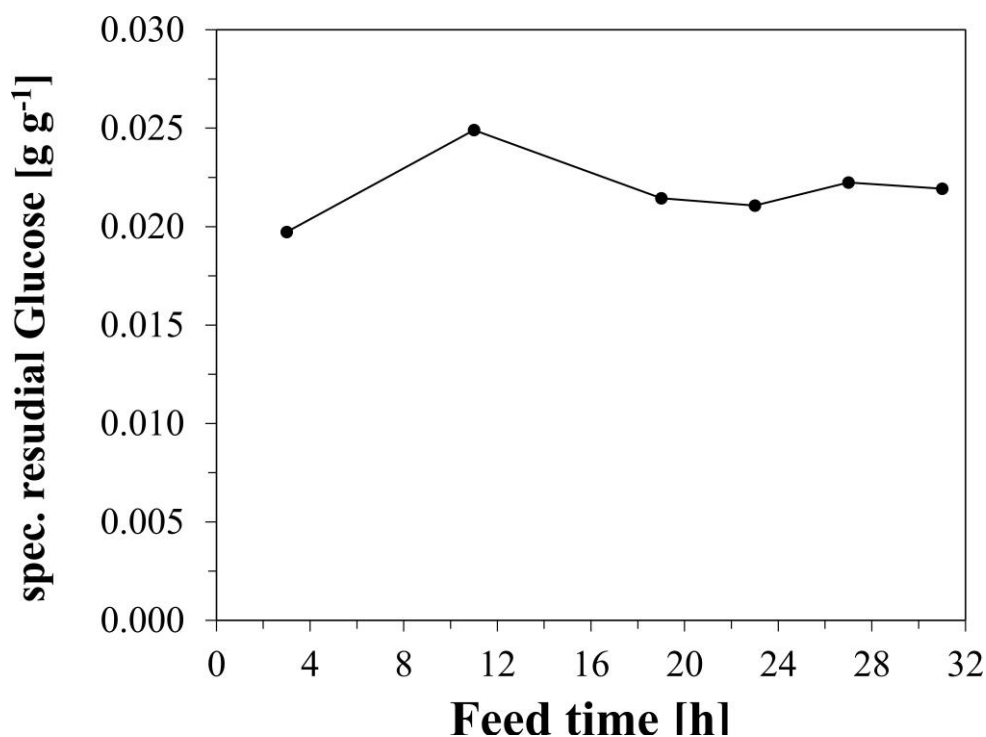


Figure S3. Specific residual glucose concentration in the cell-free cultivation medium of PEP. Glucose concentrations were analysed using HPLC as previously described by Marisch et al., 2013 (<https://doi.org/10.1186/1475-2859-12-58>).

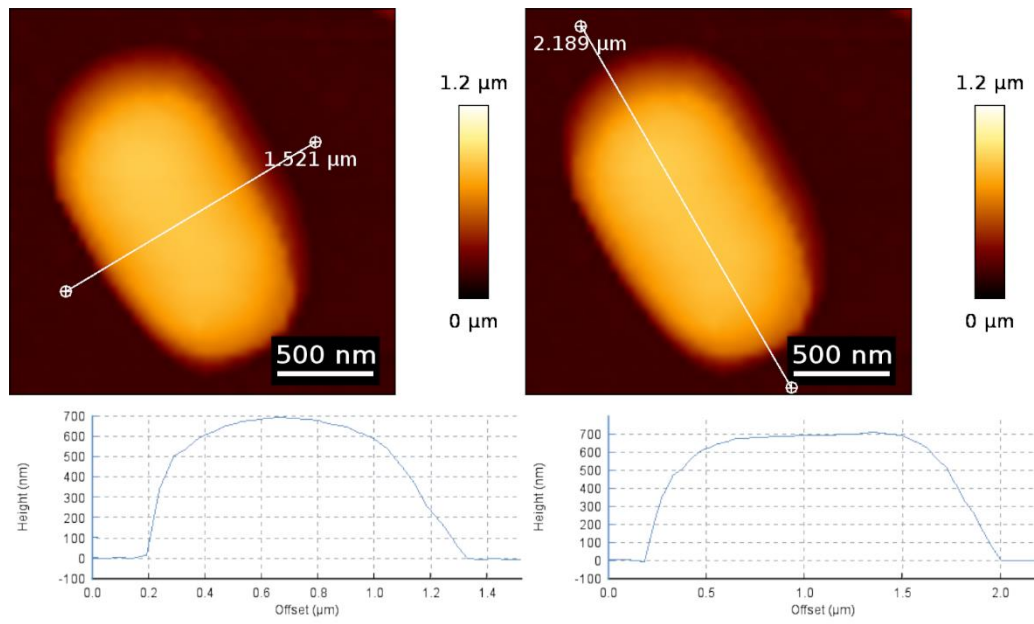


Figure S4. Analysis of cell width (**left**) and cell length (**right**) from AFM images.

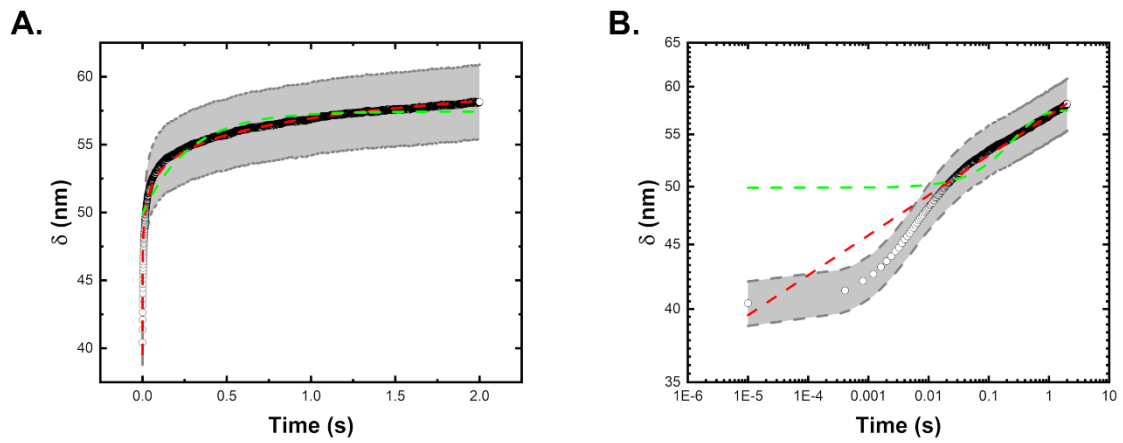


Figure S5. Performance of fitting procedures for creep curves (red power law, green standard linear solid) in normal representation (**A**) and logarithmic representation (**B**).

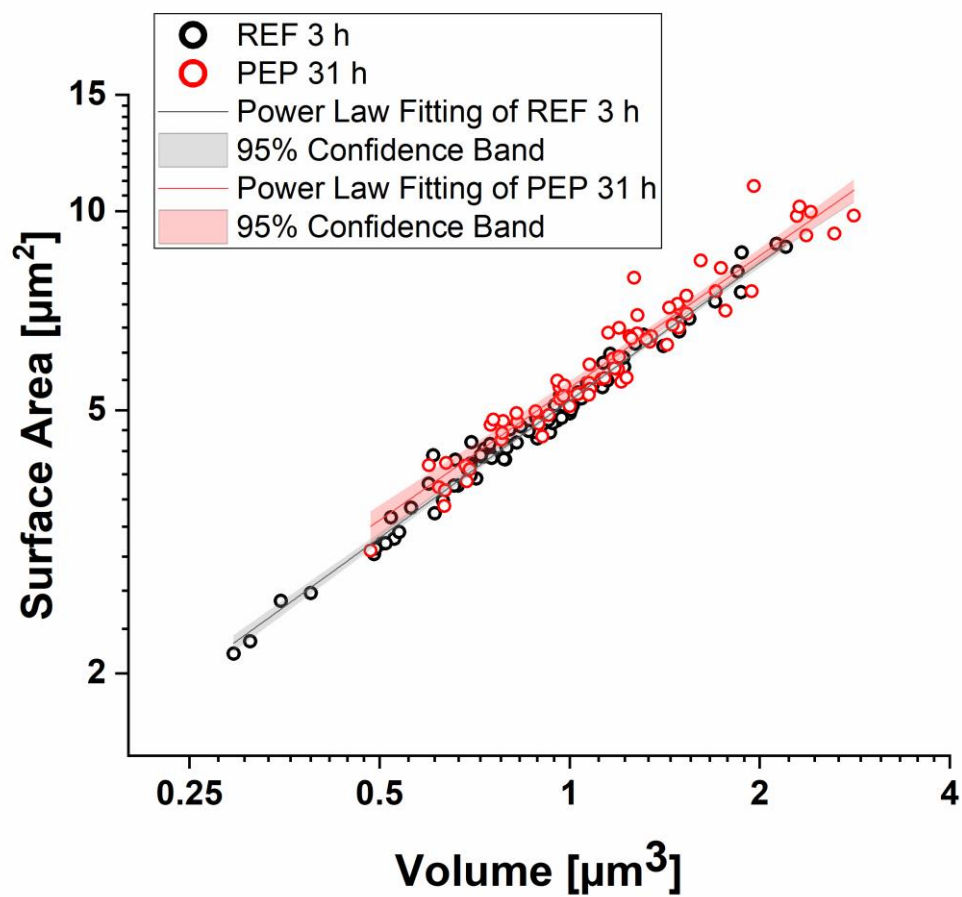


Figure S6. Surface area to volume scaling is conserved for the different samples. Here shown is REF 3h and PEP 31 h.

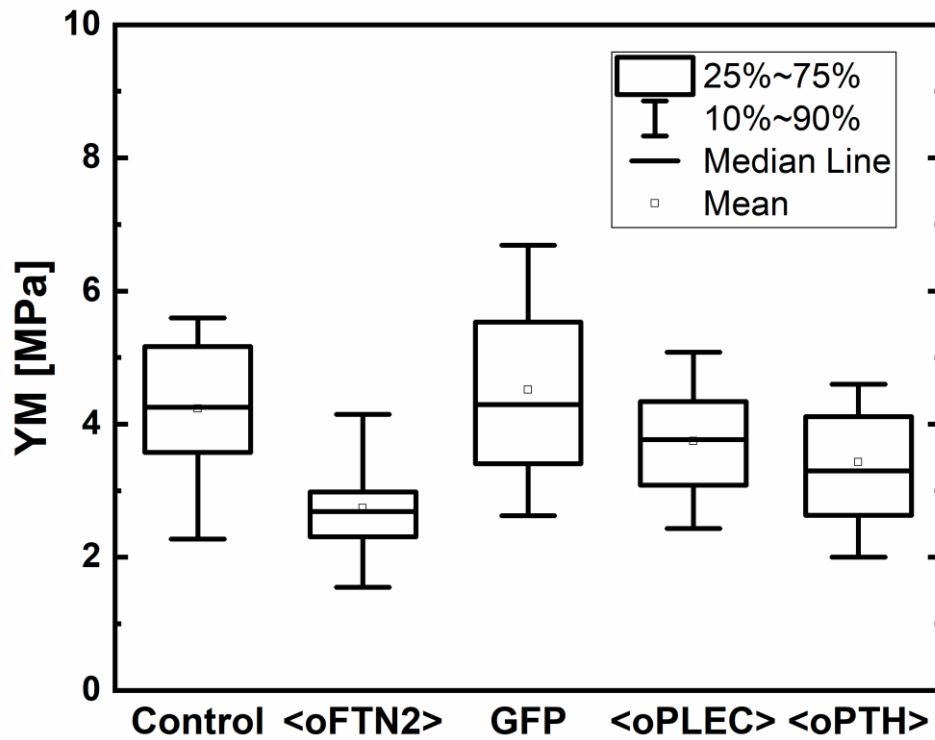


Figure S7. Apparent Young's Modulus of BL21(DE3) cells expressing recombinant Fab fragments (FTN2), green fluorescent protein (GFP), antimicrobial peptide plectasin (PLEC), and parathyroid hormone (PTH). Measurements of BL21(DE3) cells without any protein/peptide production are indicated as control. Brackets (< >) indicate genomic integration of the recombinant gene into the host. Translocation into the periplasmic space via the OmpA signal sequence of the respective protein/peptide is indicated as "o". Respective cultivations were carried out as described in the material and method section.