

Supplementary material

Figure S1. Summary of lipid metabolism.

List S1. Main abbreviations used for lipid species.

Table S1. List of detected lipids in leaves of both *ndufs4* and Col0 (Arabidopsis) with loadings of multivariate statistics (p_{corr}) and P -value from univariate statistics.

Table S2. List of detected lipids in leaves of both *ndufs8* and Col0 (Arabidopsis) with loadings of multivariate statistics (p_{corr}) and P -value from univariate statistics.

Table S3. List of detected lipids in leaves of both *CMSII* and WT (tobacco) with loadings of multivariate statistics (p_{corr}) and P -value from univariate statistics.

Table S4. List of detected lipids in seeds of both *ndufs4* and Col0 (Arabidopsis) with loadings of multivariate statistics (p_{corr}) and P -value from univariate statistics.

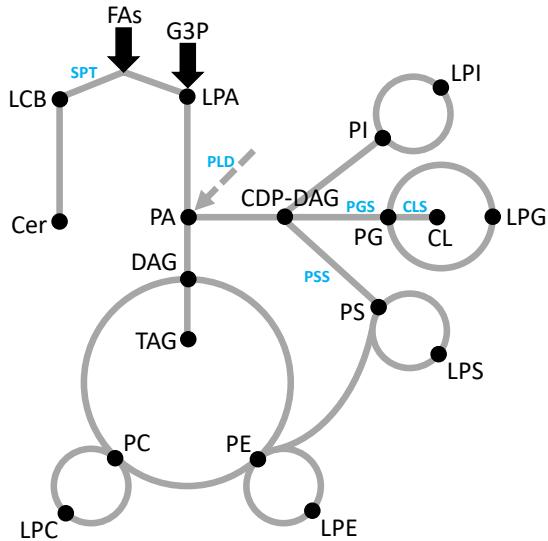
Table S5. List of detected lipids in seeds of both *ndufs8* and Col0 (Arabidopsis) with loadings of multivariate statistics (p_{corr}) and P -value from univariate statistics.

Table S6. List of detected lipids in seeds of both *CMSII* and WT (tobacco) with loadings of multivariate statistics (p_{corr}) and P -value from univariate statistics.

Table S7. List of detected lipids in pollen of both *CMSII* and WT (tobacco) with loadings of multivariate statistics (p_{corr}) and P -value from univariate statistics.

(a) Pathways and enzymes mentioned in main text (section 3.3)

CLS – cardiolipin synthase
PGS – phosphatidylglycerol synthase
PLD – phospholipase D
PSS – phosphatidylserine synthase
SPT – serine palmitoyl-transferase



(b) Pathways changed in complex I mutants (summary)

In leaves
● Increase
● Decrease
In seeds
● Increase
● Decrease

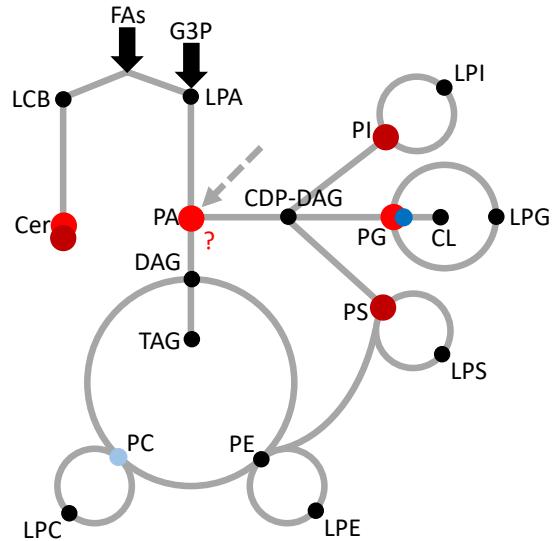


Figure S1. Summary of lipid biosynthetic pathways and observed changes in complex I mutants. (a) pathways with enzymes mentioned in text (in blue). (b) overview of changes observed in mutants with increases (red) and decreases (blue). For simplicity, the biosynthesis of galactolipids (chloroplast) is not shown here. Cer, ceramides; CL, cardiolipins; DAG, diacylglycerol; FAs, fatty acids; G3P, glycerol 3-phosphate; LCB, long chain base; PA, phosphatidic acid; PC, phosphatidyl choline; PE, phosphatidyl ethanolamine; PG, phosphatidyl glycerol; PI, phosphatidyl inositol; PS, phosphatidyl serine; TAG, triglycerides (triacylglycerol). Upper case ‘L’ stands for “lyso”. Grey circles indicate the possibility of cycling between species shown. Phospholipid cleavage and recycling can generate PA, and it is shown with a dashed grey arrow. The link between PE and PS represents the interconversion between PE and PS. In (b), the question mark symbolises uncertainty about the increase in PA (see main text). Changes in pollen (which are limited to data collected in tobacco) are not shown here.

List S1. Main abbreviations used for lipid species in the paper.

Abbreviation	Meaning
LPC	Lyso phosphatidyl-choline
PC	Phosphatidyl-choline
LPG	Lyso phosphatidyl-glycerol
PG	Phosphatidyl-glycerol
LPE	Lyso phosphatidyl-ethanolamine
PE	Phosphatidyl-ethanolamine
dMePE	Dimethyl phosphatidyl-ethanolamine
LPI	Lyso phosphatidyl-inositol
PI	Phosphatidyl-inositol
PL	Phospholipids
PS	Phosphatidyl-serine
DG	Diacylglycerol (diglycerides)
TG	Triacylglycerol (triglycerides)
PA	Phosphatidic acid
PMe	Phosphatidyl-methanol
PEt	Phosphatidyl-ethanol
CL	Cardiolipin
Cer	Ceramide
MGDG	Monogalactosyl-diacylglycerol
DGDG	Digalactosyl-diacylglycerol
StE	Stigmasterol ester
PX(n:i, m:j)	Phosphatidyl-X esterified with C _n and C _m fatty acid chains, with i and j unsaturations, respectively. Example, PC(18:3, 18:3): phosphatidyl-choline esterified with two C ₁₈ chains carrying 3 unsaturations (linolenic type fatty acid).

MGDG(18:0_18:3)	GL	MGDG	8.08E-01	3.37E-03
Cer(d18:1+hO_24:0)	Cer	Cer	8.18E-01	3.36E-03
Cer(d18:2_24:0+O)	Cer	Cer	8.27E-01	2.34E-03
PC(18:0_18:2)	PL	PC	8.36E-01	2.08E-03
PS(39:1)	PL	PS	8.42E-01	1.82E-03
TG(16:0_14:0_16:0)	TG	TG	8.71E-01	1.11E-03
AGlcSiE(16:0)	AGlcSiE	AGlcSiE	8.73E-01	7.11E-04
TG(16:0_16:0_16:0)	TG	TG	8.84E-01	6.64E-04
DGDG(18:0_18:1)	GL	DGDG	9.17E-01	1.16E-04
PC(17:1_17:1)	PL	PC	9.25E-01	1.65E-04
MGDG(18:3_20:1)	GL	MGDG	9.26E-01	8.50E-05
PE(16:0_18:1)	PL	PE	9.27E-01	9.55E-05
PC(38:3)	PL	PC	9.31E-01	8.95E-05
PC(18:1_18:3)	PL	PC	9.31E-01	1.31E-04

PG(16:1_18:2)	PL	PG	9.18E-01	1.62E-04
AGlcSiE(18:0)	AGlcSiE	AGlcSiE	6.34E-01	5.24E-02
MGDG(18:3_18:3)	GL	MGDG	8.70E-01	1.07E-03
DGDG(18:0_18:3)	GL	DGDG	1.27E-01	7.27E-01
PG(18:3_18:2)	PL	PG	8.54E-01	1.74E-03
DG(18:4_18:3)	DG	DG	4.26E-01	2.20E-01
MGDG(18:0_18:3)	GL	MGDG	-4.79E-01	1.58E-01
Cer(d18:1+hO_24:0)	Cer	Cer	8.06E-01	5.31E-03
Cer(d18:2_24:0+O)	Cer	Cer	9.11E-01	2.67E-04
PC(18:0_18:2)	PL	PC	6.25E-02	8.84E-01
PS(39:1)	PL	PS	-8.01E-02	8.06E-01
TG(16:0_14:0_16:0)	TG	TG	-3.33E-01	3.61E-01
AGlcSiE(16:0)	AGlcSiE	AGlcSiE	8.34E-01	2.99E-03
TG(16:0_16:0_16:0)	TG	TG	-3.75E-01	3.00E-01
DGDG(18:0_18:1)	GL	DGDG	5.76E-02	8.89E-01
PC(17:1_17:1)	PL	PC	7.44E-01	1.46E-02
MGDG(18:3_20:1)	GL	MGDG	9.17E-01	1.73E-04
PE(16:0_18:1)	PL	PE	5.28E-01	1.24E-01
PC(38:3)	PL	PC	5.60E-01	1.01E-01
PC(18:1_18:3)	PL	PC	-8.03E-02	8.01E-01

