

Supplementary Information

A new phospholipase D from *Moritella* sp. JT01: Biochemical characterization, crystallization and application in the synthesis of phosphatidic acid

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Supplementary Table S1 Summary on purification steps of wild type MsPLD.

Purification steps	Total activity (U) ^a	Total protein (mg)	Specific activity (U/mg)	Purification (fold)	Yield (%)
Crude extract	4111	1242.9	3.31	1.00	100.00
Ni ²⁺ -NTA	3311	261.8	12.65	3.82	80.54
Desalting column	2682	200.6	13.37	4.04	65.23

^a The specific activity and protein concentration were determined as described in methods section.

Supplementary Table S2 Effects of different metal ions on the activity of MsPLD.

Metal ion	Concentration (mM)	Relative activity (%)
None	0	100±0.26
Mn ²⁺	5	106.26±0.22
	10	109.99±0.85
Ca ²⁺	5	100.09±0.35
	10	104.12±1.93
Mg ²⁺	5	96.01±2.93
	10	96.50±1.05
Ni ²⁺	5	106.18±0.13
	10	99.28±6.16
Zn ²⁺	5	62.37±0.74
	10	66.40±2.85
Fe ²⁺	5	79.54±0.48
	10	7.14±4.64
Fe ³⁺	5	10.91±0.96
	10	3.83±2.83
Cu ²⁺	5	16.95±0.70
	10	8.94±1.10

Note: Enzyme activity without metal ions adding was defined as 100%.

Supplementary Table S3 Data collection and refinement statistics of MsPLD.

MsPLD	
Wavelength	0.97919
Resolution range	23.36-2.3 (2.38- 2.3)
Space group	P2 ₁ 2 ₁ 2 ₁
Unit cell	a =93.90 Å, b =216.50Å, c=277.45Å $\alpha = 90^\circ$, $\beta =90^\circ$, $\gamma = 90^\circ$
Unique reflections	250032 (24724)
Multiplicity	12.3(6.8)
Completeness (%)	99.69 (99.38)
Mean I/sigma(I)	0.6
Wilson B-factor	49.48
R-meas	0.178 (2.892)
R-pim	0.050 (0.889)
CC1/2	0.995 (0.304)
Reflections used in refinement	249785 (24611)
Reflections used for R-free	1742 (171)
R-work	0.1805 (0.2957)
R-free	0.2018 (0.3025)
Number of non-hydrogen atoms	26491
macromolecules	25452
ligands	55
solvent	984
Protein residues	3254
RMS(bonds)	0.010
RMS(angles)	0.94
Ramachandran favored (%)	93.72
Ramachandran allowed (%)	5.50
Ramachandran outliers (%)	0.77
Rotamer outliers (%)	1.01
Clashscore	5.30
Average B-factor	61.54
macromolecules	61.82
ligands	57.29
solvent	54.48
Number of TLS groups	25

Supplementary Table S4 Primers used in this study.

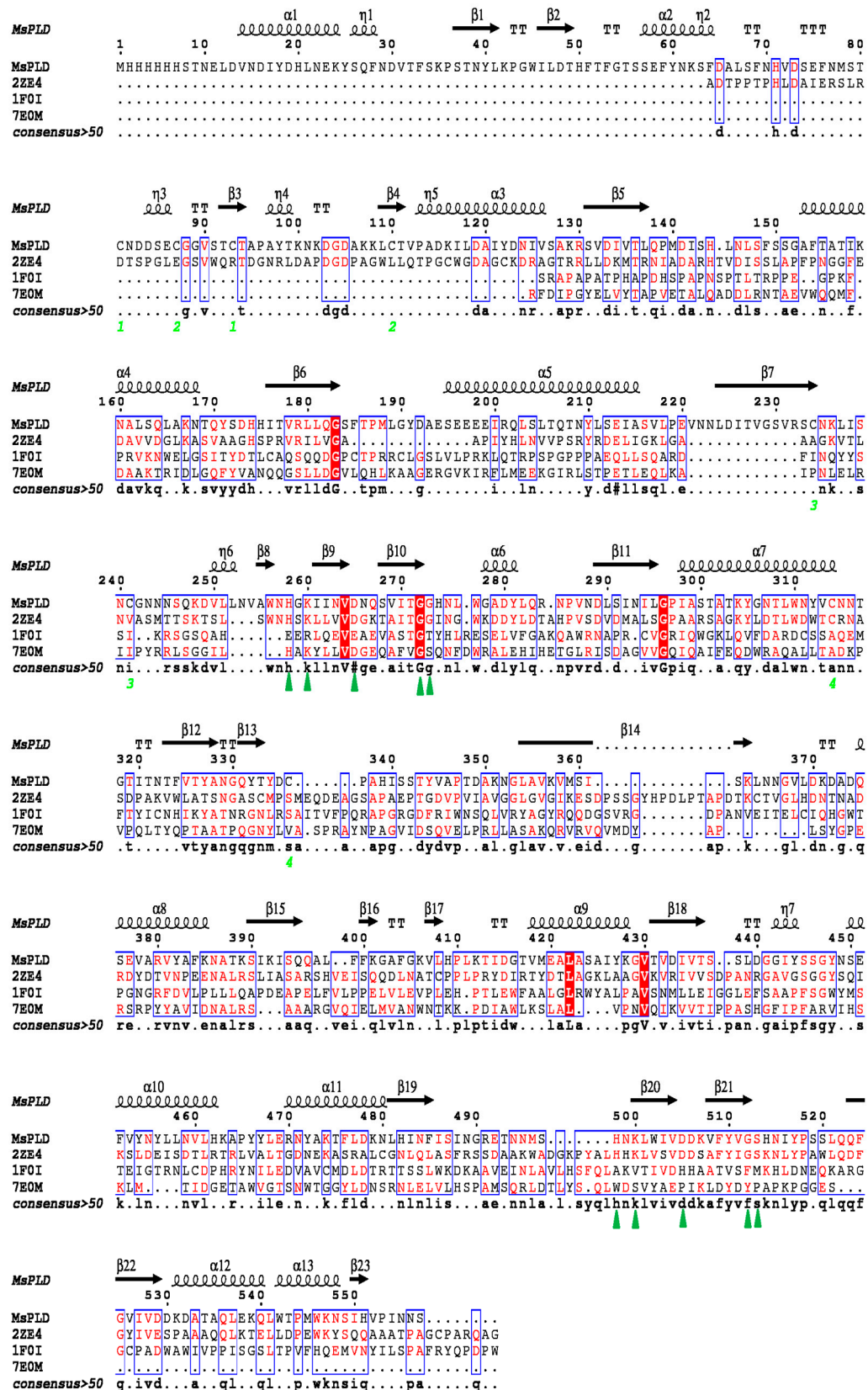
Primer	Sequence (5'to3')*
N61C-F	CACTTCCAGCGAATTTTACT <u>TGCAAATCCTTCGACGCGCTG</u>
N61C-R	CAGCGCGTCGAAGGATT <u>TGCAGT</u> AAAATTCGCTGGAAGTG
E75C-F	GAGCTTCAACCACGTTGACTCT <u>TGCTTCAACATGTCTACCTGTAACG</u>
E75C-R	CGTTACAGGTAGACATGTTGAAG <u>CAAGAGTCAACGTGGTTGAAGCTC</u>
K102C-F	CACAGCACCGGCGTACACCAAAAAC <u>TGCGATGGT</u> GATGCTAAAAAAC
K102C-R	GTTTTTTAGCATCACCATCGCAGTTTTTTGGTGTACGCCGGTGCTGTG
A156C-F	TTTCTAGCGGTGCTTTCACCT <u>TGCACC</u> ATTAAAAACGCGCTGAG
A156C-R	CTCAGCGCGTTTTTAATGGT <u>GCA</u> GGTGAAAGCACCGCTAGAAA
S163C-F	CCATTAAAAACGCGCTGT <u>TGCC</u> CAGCTGGCGAAAAAC
S163C-R	GTTTTTCGCCAGCTGG <u>CA</u> CAGCGCGTTTTTAATGG
R284C-F	GCGGATTATCTGCAGT <u>TGTA</u> ACCCGGTGAACG
R284C-R	CGTTCACCGGGTT <u>ACACT</u> GCAGATAATCCGC
H258A-F	TGAACGTTGCTTGGAAC <u>GCCG</u> GTAAAATTATCAACGTTGACAACC
H258A-R	TTTACC <u>GGC</u> GTTCCAAGCAACGTTTCAGCAGAACATCTTTCTGGCTG TT
MsPLD-1F0I-1-F	CCCATTTCACCTTCGGCACT <u>CCGGGTTGCTGGGGTGACGATGGTGA</u> TGCTAAAAAACT
MsPLD-1F0I-1-R	AGTTTTTTAGCATCACCAT <u>TCGTCACCCCAGCAACCCGGAGTGCCGA</u> AGGTGAAGTGGG
MsPLD-1F0I-2-F	ACTCCGGGTTGCTGGGGTGACGAC <u>CAAATGCGCTGACGATGGT</u> GAT GCTAAAAAACTG
MsPLD-1F0I-2-R	CAGTTTTTTAGCATCACCATC <u>GTCAGCGCATTGTCGTCACCCCAGC</u> AACCCGGAGT
Del57-113-F	GGATACCCACTTCACCTTCGGCACTCCGGCTGATAAAATTCTGGAT GCGA
Del57-113-R	TCGCATCCAGAATTTTATCAGCCGGAGTGCCGAAGGTGAAGTGGGT ATCC
Del65-113-F	TTCCAGCGAATTTTACAACAAATCCCCGGCTGATAAAATTCTGGAT GCGA
Del65-113-R	TCGCATCCAGAATTTTATCAGCCGGGGATTGTTGTAAAATTCGCTG GAA

* Underlined sequences were miss-match regions for site directed mutagenesis.

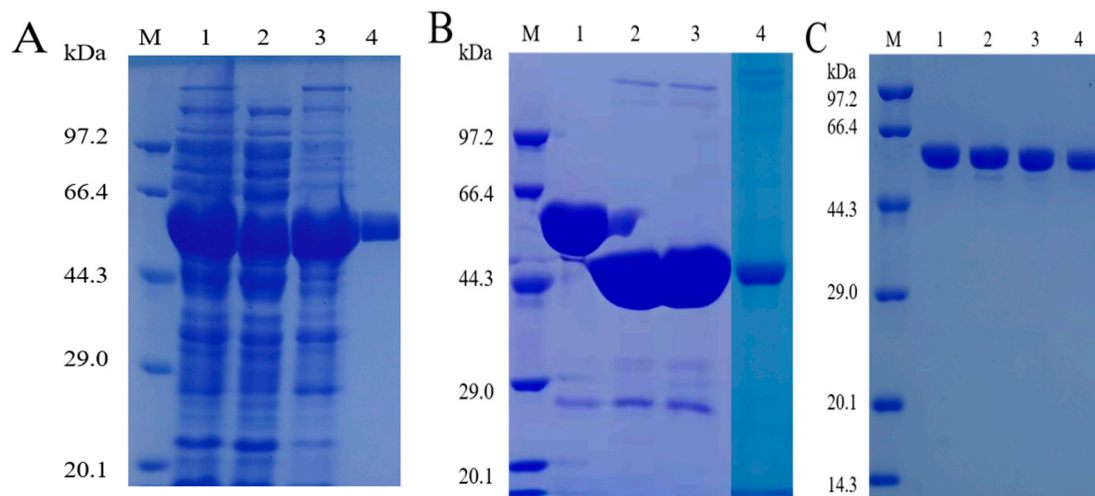
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10	20	30	40	50	60
MKIQNKHAMI	ILKSAASAFV	LVSASASAST	NELDVNDIYD	HLNEKYSQFN	DVTFSKPSTN
70	80	90	100	110	120
YLKPGWILDT	HFTFGTSSEF	YNKSFDAISF	NHVDSEFNMS	TCNDDSECGG	VSTCTAPAYT
130	140	150	160	170	180
KNKDGDAKKL	CTVPADKILD	AIYDNIVSAK	RSVDIVTLQP	MDISHLNLSF	SSGAFTATIK
190	200	210	220	230	240
NALSQIAKNT	QYSDHHITVR	LLQGSFTPML	GYDAESEEEE	IRQLSLTQTN	YLSEIASVLP
250	260	270	280	290	300
EVNNLDITVG	SVRSCNKLIS	NCGNNSQKD	VLLNVAWNHG	KIINVDNQSV	ITGGHNLWGA
310	320	330	340	350	360
DYLQRNPVND	LSINILGPIA	STATKYGNTL	WNYVCNNTGT	ITNTFVTYAN	GQYTYDCPAH
370	380	390	400	410	420
ISSTYVAPTD	AKNGLAVKVM	SISKLNNGVL	DKDADQSEVA	RVYAFKNATK	SIKISQQALF
430	440	450	460	470	480
FKGAFGKVLH	PLKTIDGTVM	EALASAIYKG	VTVDIVTSSL	DGGIYSSGYN	SEFVYNYLLN
490	500	510	520	530	540
VLHKAPYYLE	RNYAKTFLDK	NLHINFISIN	GRETNMNSHN	KLWIVDDKVF	YVGS
550	560	570			
SLQQFGVIVD	DKDATAQLEK	QLWTPMWKNS	IHVPINNS		

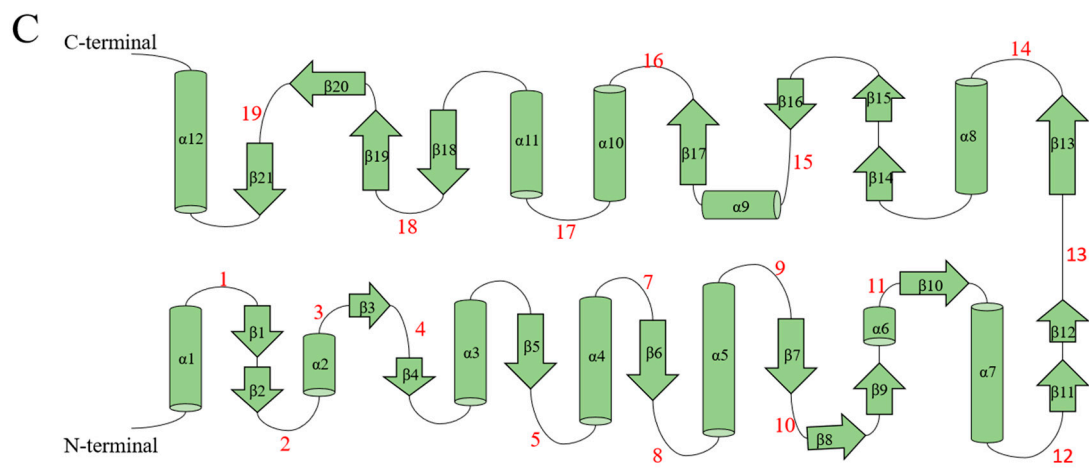
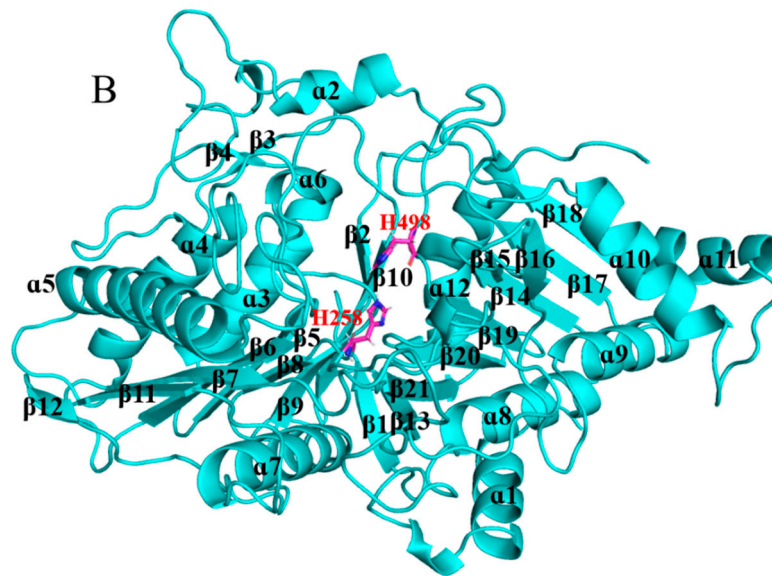
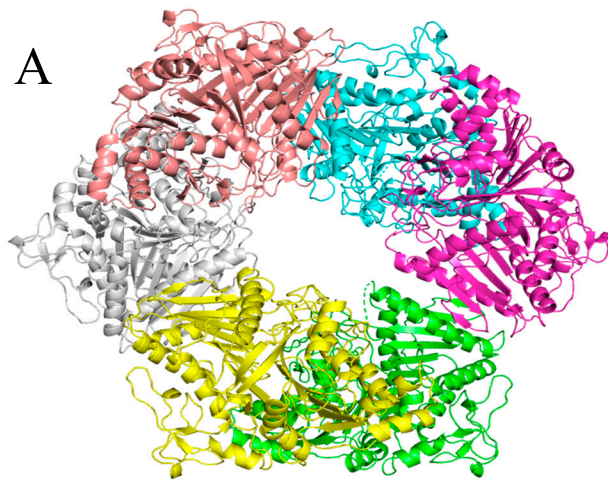
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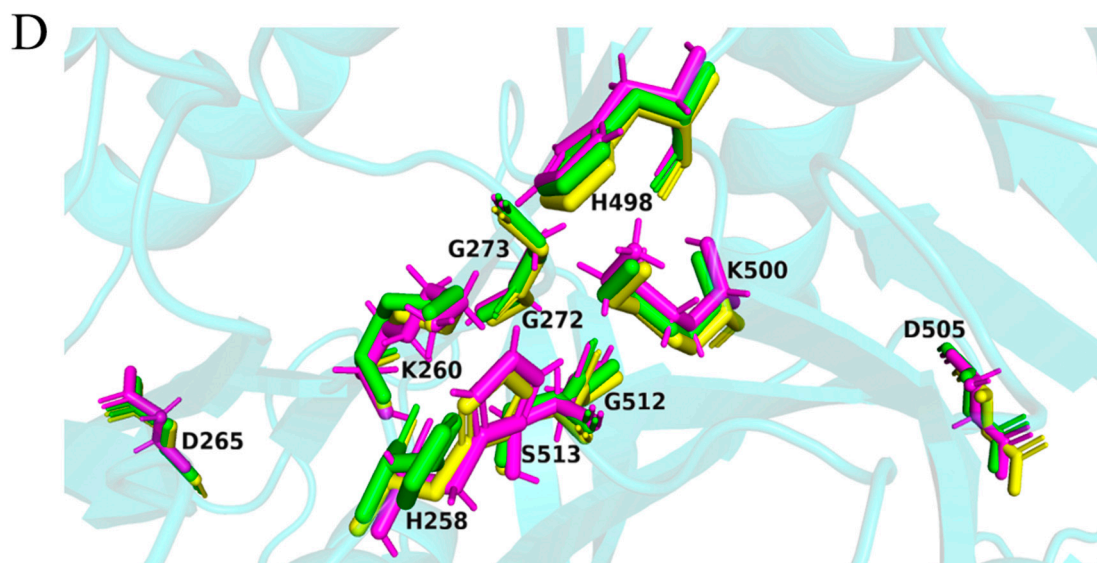


Supplementary Figure S1. Bioinformatic analysis of MsPLD. (A) Full length amino acid sequence of MsPLD. The deduced signal peptide that predicted with SignalP 5.0 server was labeled in purple. Two conserved HXX(X)₄D(X)₆GG/S (HKD) motifs of the PLD superfamily were shown in the red box and key residues were labeled in red. (B) Structural-based amino acid sequence alignment of the mature MsPLD with several other PLDs in the PDB. SpPLD (PDB ID: 7E0M), Streptomyces sp. PMF PLD (PDB ID: 1F0I), SaPLD (PDB ID: 2ZE4). Residues highlighted in blue box were identical among the protein compared. Four pairs of disulfide bonds were numbered.



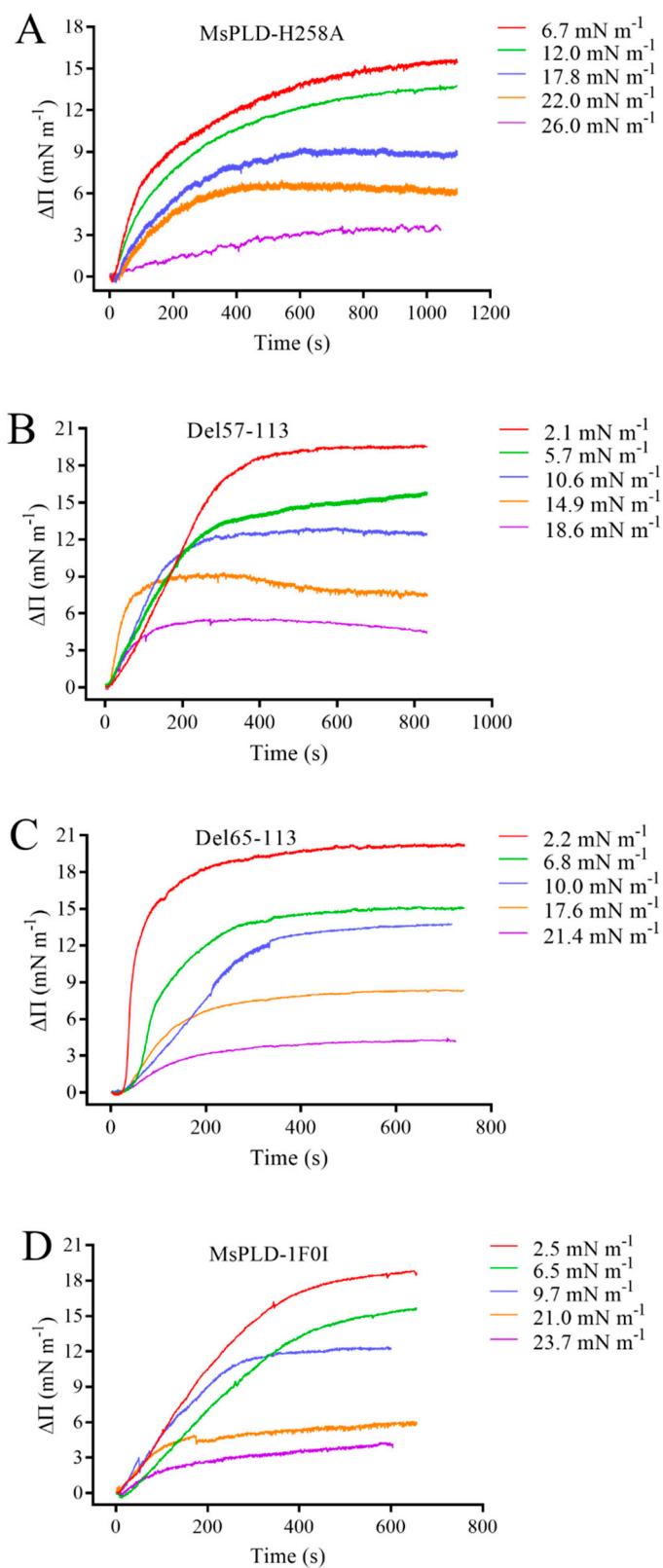
Supplementary Figure S2. Sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) analysis of the recombinant wild type MsPLD and its corresponding mutants. (A) Wild type MsPLD. Lane M, The protein marker; Lane 1, total cell extract; Lane 2, supernatant of total cell extract; Lane 3, pellet of total cell extract; Lane 4, purified MsPLD by using nickel column chromatography. (B) Purified protein of various mutants. Lane M, The protein marker; Lane 1, MsPLD-H258A; Lane 2, Del57-113; Lane 3, Del65-113; Lane 4, MsPLD-1F0I; (C) Purified protein of wild type and various mutants. Lane M, The protein marker; Lane 1, MsPLD; Lane 2, N61C-R284C; Lane 3, E75C-A156C; Lane 4, K102C-S163C.





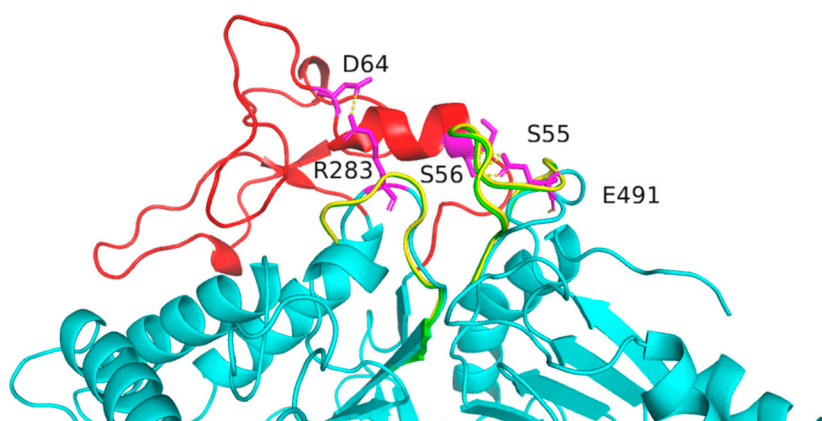
Supplementary Figure S3. The three-dimensional (3D) structure of wild type MsPLD.

(A) Six molecules in the asymmetric unit were shown with different colors. (B) Overall structure of MsPLD. MsPLD was composed by 12 α helixs and 21 β sheets connected by various loops. Two His258 and H498 located at the active site of the protein were shown in sticks. (C) Overall topology of MsPLD. (D) Two classical HxKxxxxD (HKD) motifs were found in MsPLD and showed high structural consistence with several PLDs within the PDB. MsPLD (magentas, PDB ID: 7WU1), *Streptomyces* sp. PMF PLD (yellow, PDB ID: 1F0I), SaPLD (green, PDB ID: 2ZE4). The 3D molecular visualization was performed with PyMOL software.



Supplementary Figure S4. Effects of various initial surface pressure (Π_i) on the adsorption of wild type MsPLD or its mutants onto PC monolayer. (A) Effects of various initial surface pressure (Π_i) on the adsorption of MsPLD-H258A. (B) Effects

of various initial surface pressure (Π_i) on the adsorption of Del57-113. (C) Effects of various initial surface pressure (Π_i) on the adsorption of Del65-113. (D) Effects of various initial surface pressure (Π_i) on the adsorption of MsPLD-1F0I. MsPLD and its corresponding mutants (5.0 μ M final concentration) were injected into the aqueous sub-phase of teflon trough, respectively. Surface pressure increase due to the adsorption of the protein onto phospholipid monolayer that spread at initial surface pressures was continuously monitored.



Supplementary Figure S5. Partial superimposition and comparison of the three typical loop segments of MsPLD (cyans, PDB ID: 7WU1) with other *Streptomyces* PLDs. *Streptomyces* sp. PMF PLD (yellow, PDB ID: 1F0I), SaPLD (green, PDB ID: 2ZE4). The extra loop segment solely exist on MsPLD was shown in red. The hydrogen bonds formed with related residues between the adjacent two loops were marked by dashed line in orange. Amino acid residues with interactions on the loops were shown in magentas sticks.