

## **Supplementary Material**

### **Wnt/ $\beta$ -Catenin Signaling Pathway Is Strongly Implicated in Cadmium-induced Developmental Neurotoxicity and Neuroinflammation: Clues from Zebrafish Neurobehavior and In Vivo Neuroimaging**

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**Table S1.** Primers used for qRT-PCR analysis.

	Gene name	Sequences of the primers (5' – 3')
Reference gene	<i>β-actin</i>	F: CACTGAGGCTCCCCTGAATCCC
		R: CGTACAGAGAGAGCACAGCCTGG
Neurodevelopment related genes	<i>α1-tubulin</i>	F: AATCACCAATGCTTGCTTCGAGCC
		R: TTCACGTCTTTGGGTACCACGTCA
	<i>syn2a</i>	F: GTGACCATGCCAGCATTTTC
		R: TGGTTCTCCACTTTCACCTT
	<i>elav13</i>	F: AGACAAGATCACAGGCCAGAGCTT
		R: TGGTCTGCAGTTTGAGACCGTTGA
	<i>nestin</i>	F: CTTCAACATCTTCAGGCCCAAG
		R: GTGTTGGTCTGTGATTCTCAG
	<i>gap43</i>	F: GCAGCAGGAAGTGGAGAAGCCA
		R: GGATTCCTCAGCAGCGTCTGGT
Apoptosis related genes	<i>bcl</i>	F: AACCCAAATTCTGCGCAACG
		R: ATCTACCTGGGACGCCATCT
	<i>p53</i>	F: GGGCAATCAGCGAGCAAA
		R: ACTGACCTTCCTGAGTCTCCA
	<i>bax</i>	F: GGCTATTTCAACCAGGGTTCC
		R: TGCGAATCACCAATGCTGT
	<i>caspase-3a</i>	F: GTTCTTTATTCAGGCTTGTCGAG
		R: TTTCTCCAGGAGTAATAACCTGG
	<i>caspase-8</i>	F: TGTCTCAGCCTATAGAAAGATGC
		R: AAATGAAGTAGATCTTCCCAGCT
	<i>irf8</i>	F: CAAAAGCCCAGATTTTGAGG
		R: TCTTTTACGGTGGTGACTGT
	<i>apoeb</i>	F: GCAGATGACGTGAAGAACCG
		R: GTTGCTACGGTGTTGCGGAT
	<i>tnf-a</i>	F: TCACGCTCCATAAGACCCAG
		R: GATGTGCAAAGACACCTGGC
Wnt signaling pathway related genes	<i>Wnt4a</i>	CAATGCGAGCAACTGGCTATAC
		AATGCAGCTTCCCTCGTACCTT
	<i>β-catenin</i>	GGAGCTCACCAGCTCTCTGT
		TAGCTTGGGTCGTCCTGTCT
	<i>gsk3β</i>	TCTGCTCACCGTTTCCTTTC
		CTCCGACCCACTTAACTCCA
	<i>Wnt10b</i>	TCCTGAAACAGGCTCGAAGT
		GCTGCTCACTTGACACA
	<i>ddk1</i>	ATGCCAGAGACACTAAATGAACA
		TATGAAGGAAACCAAGTTGAAAA

	<i>lef1</i>	AATGATCCCGTTCAAAGACG
		CGCTAAGTCTCCCTCCTCCT
	<i>axin2</i>	AGCACTGGAGAAAGGCTACAGGTC
		GGAGTCATCCGTCAGTGCATC
	<i>ccnd</i>	CGTGT CAGGAACAGATCGAG
		TCAAATGTTAATGTCTCTGACGT
	<i>myca</i>	AGAAAGCTGGAGTCCTCGAC
		CTGCTGCAGTGTGTT CAGC
	<i>Sp51</i>	GGAGGTCACGTTGAGGATGG
		GCGACAGCGACGAGTAGAGC

**Table S2.** Detailed statistical information in this study.

Experiment (Figure)	Test method		Significance		N
<b>β-catenin expression (Figure. 1B)</b>		One-way ANOVA	***	F=42.14 P<0.001	3
	CTRL	Dunnett's multiple comparisons			
	100 ug/L vs. CTRL	-	***	<0.001	
	200 ug/L vs. CTRL	-	***	<0.001	
<b>Wnt signaling pathway related genes (Figure. 1C)</b>	<i>wnt4a</i>	One-way ANOVA	*	F=8.252 P=0.019	3
	100 ug/L vs. CTRL	Dunnett's multiple comparisons	ns	0.998	
	200 ug/L vs. CTRL	-	*	0.023	
	<i>wnt10b</i>	One-way ANOVA	**	F=108.4 P=0.002	
	100 ug/L vs. CTRL	Dunnett's multiple comparisons	*	0.017	
	200 ug/L vs. CTRL	-	**	0.005	
	<i>gsk3β</i>	One-way ANOVA	*	F=5.913 P=0.031	
	100 ug/L vs. CTRL	Dunnett's multiple comparisons	ns	0.052	
	200 ug/L vs. CTRL	-	*	0.046	
	<i>β-catenin</i>	One-way ANOVA	**	F=8.848 P=0.005	
	100 ug/L vs. CTRL	Dunnett's multiple comparisons	ns	0.811	
	200 ug/L vs. CTRL	-	**	0.004	
<b>Endogenous protein inhibitor of the Wnt/β-catenin signaling pathway (Figure. 1D)</b>	<i>ddk1</i>	One-way ANOVA	***	F=101.3 P<0.001	3
	100 ug/L vs. CTRL	Dunnett's multiple comparisons	***	<0.001	
	200 ug/L vs. CTRL	-	***	<0.001	

Target genes regulated by $\beta$ -catenin/TCF transcription (Figure. 1E)	<i>lef1</i>	One-way ANOVA	***	F=368.9 P<0.001	
	100 ug/L vs. CTRL	Dunnett's multiple comparisons	***	<0.001	
	200 ug/L vs. CTRL	-	***	<0.001	
	<i>axin2</i>	One-way ANOVA	***	F=40.38 P<0.001	
	100 ug/L vs. CTRL	Dunnett's multiple comparisons	*	0.042	
	200 ug/L vs. CTRL	-	***	<0.001	
	<i>myca</i>	One-way ANOVA	**	F=21.62 P=0.002	
	100 ug/L vs. CTRL	Dunnett's multiple comparisons	*	0.022	
	200 ug/L vs. CTRL	-	**	0.001	
	<i>ccnd</i>	One-way ANOVA	*	F=7.579 P=0.023	
	100 ug/L vs. CTRL	Dunnett's multiple comparisons	*	0.015	
	200 ug/L vs. CTRL	-	*	0.042	
	<i>sp52</i>	One-way ANOVA	***	F=18.80 P<0.001	
	100 ug/L vs. CTRL	Dunnett's multiple comparisons	*	0.018	
	200 ug/L vs. CTRL	-	***	<0.001	
Time to reach different period (Figure. 2C)	30%-epiboly	One-way ANOVA	**	F=3.934 P=0.002	
	Cd				25
	CTRL vs. Cd	Dunnett's multiple comparisons	*	0.023	22
	XAV939 vs. Cd	-	ns	0.116	21
	TWS119 vs. Cd	-	*	0.034	22
	Cd+XAV939 vs. Cd	-	ns	>0.999	23

	Cd+TWS119 vs. Cd	-	*	0.028	22
	<b>bud</b>	One-way ANOVA	***	F=11.85 P<0.001	
	Cd				23
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	21
	XAV939 vs. Cd	-	**	0.01	21
	TWS119 vs. Cd	-	***	<0.001	22
	Cd+XAV939 vs. Cd	-	ns	0.476	21
	Cd+TWS119 vs. Cd	-	*	0.025	22
	<b>15-somite</b>	One-way ANOVA	***	F=10.65 P<0.001	
	Cd				23
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	22
	XAV939 vs. Cd	-	*	0.046	21
	TWS119 vs. Cd	-	***	<0.001	22
	Cd+XAV939 vs. Cd	-	ns	0.223	22
	Cd+TWS119 vs. Cd	-	*	0.022	22
	<b>20-somite</b>	One-way ANOVA	***	F=20.45 P<0.001	
	Cd				24
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	22
	XAV939 vs. Cd	-	***	<0.001	21

	TWS119 vs. Cd	-	***	<0.001	22
	Cd+XAV939 vs. Cd	-	ns	0.244	22
	Cd+TWS119 vs. Cd	-	***	<0.001	22
Heart rate at 5 dpf (Figure. 2D)		One-way ANOVA	***	F=26.14 P<0.001	15
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	
	XAV939 vs. Cd	-	***	<0.001	
	TWS119 vs. Cd	-	***	<0.001	
	Cd+XAV939 vs. Cd	-	*	0.022	
	Cd+TWS119 vs. Cd	-	*	0.015	
Embryo malformation rate at 7 dpf (Figure. 2E)		One-way ANOVA	***	F=37.54 P<0.001	10
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	
	XAV939 vs. Cd	-	***	<0.001	
	TWS119 vs. Cd	-	***	<0.001	
	Cd+XAV939 vs. Cd	-	*	0.034	
	Cd+TWS119 vs. Cd	-	*	0.023	
Body length at 6 dpf (Figure. 2F)		One-way ANOVA	***	F=28.99 P<0.001	
	Cd				10
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	10
	XAV939 vs. Cd	-	***	<0.001	10

	TWS119 vs. Cd	-	***	<0.001	14
	Cd+XAV939 vs. Cd	-	*	0.015	10
	Cd+TWS119 vs. Cd	-	*	0.023	11
Distance moved of spontaneous activity of larvae (Figure. 3C)		One-way ANOVA	***	F=14.35 P<0.001	
	Cd				18
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	15
	XAV939 vs. Cd	-	***	<0.001	18
	TWS119 vs. Cd	-	***	<0.001	18
	Cd+XAV939 vs. Cd	-	ns	>0.999	21
	Cd+TWS119 vs. Cd	-	*	0.015	18
Movement time of spontaneous activity of larvae (Figure. 3D)		One-way ANOVA	***	F=17.42 P<0.001	
	Cd				18
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	15
	XAV939 vs. Cd	-	***	<0.001	17
	TWS119 vs. Cd	-	***	<0.001	12
	Cd+XAV939 vs. Cd	-	ns	>0.999	21
	Cd+TWS119 vs. Cd	-	**	0.007	17
Mean turn angle of spontaneous activity of larvae (Figure. 3E)		One-way ANOVA	***	F=12.40 P<0.001	
	Cd				17
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	15
	XAV939 vs. Cd	-	***	<0.001	18



	TWS119 vs. Cd	-	**	0.004	11
	Cd+XAV939 vs. Cd	-	ns	0.922	21
	Cd+TWS119 vs. Cd	-	**	0.005	17
Distance moved of larvae under light-dark stimulation (Figure. 3H)		One-way ANOVA	***	F=39.43 P<0.001	
	Cd				18
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	24
	XAV939 vs. Cd	-	***	<0.001	24
	TWS119 vs. Cd	-	***	<0.001	21
	Cd+XAV939 vs. Cd	-	ns	0.067	17
	Cd+TWS119 vs. Cd	-	**	0.003	24
Movement time of larvae under light-dark stimulation (Figure. 3I)		One-way ANOVA	***	F=38.91 P<0.001	
	Cd				24
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	24
	XAV939 vs. Cd	-	***	<0.001	24
	TWS119 vs. Cd	-	***	<0.001	21
	Cd+XAV939 vs. Cd	-	ns	0.297	24
	Cd+TWS119 vs. Cd	-	*	0.025	24
Mean turn angle under light-dark stimulation (Figure. 3J)		One-way ANOVA	***	F=23.20 P<0.001	
	Cd				24
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	24
	XAV939 vs. Cd	-	***	<0.001	24

	TWS119 vs. Cd	-	***	<0.001	18
	Cd+XAV939 vs. Cd	-	ns	0.208	24
	Cd+TWS119 vs. Cd	-	***	<0.001	24
<b>Distance moved under vibration stimulation (Figure 3M)</b>		Kruskal-Wallis	***	H=41.33 P<0.001	
	Cd				20
	CTRL vs. Cd	Dunn's multiple comparisons	*	0.01	20
	TWS119 vs. Cd	-	*	0.024	8
	XAV939 vs. Cd	-	*	0.011	24
	Cd+TWS119 vs. Cd	-	**	0.009	21
	Cd+XAV939 vs. Cd	-	ns	0.81	22
<b>Movement duration under vibration stimulation (Figure 3N)</b>		Kruskal-Wallis	***	H=41.53 P<0.001	
	Cd				23
	CTRL vs. Cd	Dunn's multiple comparisons	**	0.005	21
	TWS119 vs. Cd	-	*	0.041	8
	XAV939 vs. Cd	-	*	0.019	23
	Cd+TWS119 vs. Cd	-	*	0.035	19
	Cd+XAV939 vs. Cd	-	ns	0.485	24
<b>Mean turn angle under vibration stimulation (Figure 3O)</b>		Kruskal-Wallis	***	H=32.6 P<0.001	
	Cd				21
	CTRL vs. Cd	Dunn's multiple comparisons	*	0.045	18
	TWS119 vs. Cd	-	*	0.039	8

	XAV939 vs. Cd	-	*	0.019	24
	Cd+TWS119 vs. Cd	-	**	0.003	22
	Cd+XAV939 vs. Cd	-	ns	>.999	17
The mean optical density at 6 dpf (Figure. 4B)		One-way ANOVA	***	F=41.13 P<0.001	
	Cd				8
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	11
	Cd+TWS119 vs. Cd	-	***	<0.001	10
	Cd+XAV939 vs. Cd	-	ns	0.079	9
Telencephalon proportion (Figure. 4D)		One-way ANOVA	***	F=13.51 P<0.001	
	Cd				7
	CTRL vs. Cd	Dunnett's multiple comparisons	*	0.026	
	Cd+TWS119 vs. Cd	-	*	0.019	
	Cd+XAV939 vs. Cd	-	*	0.048	
Neuron number/mm <sup>2</sup> (Figure. 4F)		One-way ANOVA	***	F=69.66 P<0.001	
	Cd				8
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	
	Cd+TWS119 vs. Cd	-	**	0.001	
	Cd+XAV939 vs. Cd	-	**	0.008	
Filament length (Figure. 4H)		One-way ANOVA	F=4.995 P=0.012	*	
	Cd				5

	CTRL vs. Cd	Dunnett's multiple comparisons	0.957	ns	
	Cd+TWS119 vs. Cd	-	0.233	ns	
	Cd+XAV939 vs. Cd	-	0.022	*	
Branch number (Figure. 4I)		One-way ANOVA	F=4.509 P=0.018	*	5
	Cd	Dunnett's multiple comparisons			
	CTRL vs. Cd		0.95	ns	
	Cd+TWS119 vs. Cd	-	0.212	ns	
	Cd+XAV939 vs. Cd	-	0.036	*	
Neurodevelopment related genes (Figure. 4J)	<i>a1-tublin</i>	One-way ANOVA	***	F=27.18 P<0.001	3
	CTRL vs. Cd	Dunnett's multiple comparisons	**	0.003	
	Cd+TWS119 vs. Cd	-	*	0.015	
	Cd+XAV939 vs. Cd	-	*	0.036	
	<i>syn2a</i>	One-way ANOVA	***	F=17.32 P<0.001	
	CTRL vs. Cd	Dunnett's multiple comparisons	*	0.031	
	Cd+TWS119 vs. Cd	-	**	0.001	
	Cd+XAV939 vs. Cd	-	ns	0.748	
	<i>elavl3</i>	One-way ANOVA	***	F=35.02 P<0.001	
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	
	Cd+TWS119 vs. Cd	-	***	<0.001	
	Cd+XAV939 vs. Cd	-	ns	0.818	

	<b><i>nestin</i></b>	One-way ANOVA	*	F=5.954 P=0.013	
	CTRL vs. Cd	Dunnett's multiple comparisons	*	0.049	
	Cd+TWS119 vs. Cd	-	*	0.011	
	Cd+XAV939 vs. Cd	-	ns	0.873	
	<b><i>gap43</i></b>	One-way ANOVA	***	F=61.99 P<0.001	
	CTRL vs. Cd	Dunnett's multiple comparisons	**	0.004	
	Cd+TWS119 vs. Cd	-	ns	0.135	
	Cd+XAV939 vs. Cd	-	**	0.003	
Cell cycle distribution (Figure. 5B)	<b>G1/G0</b>	One-way ANOVA	***	F=22.29 P<0.001	3
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	*	0.025	
	Cd+TWS119 vs. Cd	-	*	0.043	
	Cd+XAV939 vs. Cd	-	*	0.012	
	<b>S</b>	One-way ANOVA	**	F=13.14 P=0.002	
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	*	0.015	
	Cd+TWS119 vs. Cd	-	*	0.028	
	Cd+XAV939 vs. Cd	-	ns	0.329	
	<b>G2/M</b>	One-way ANOVA	***	F=30.26 P<0.001	
	Cd				

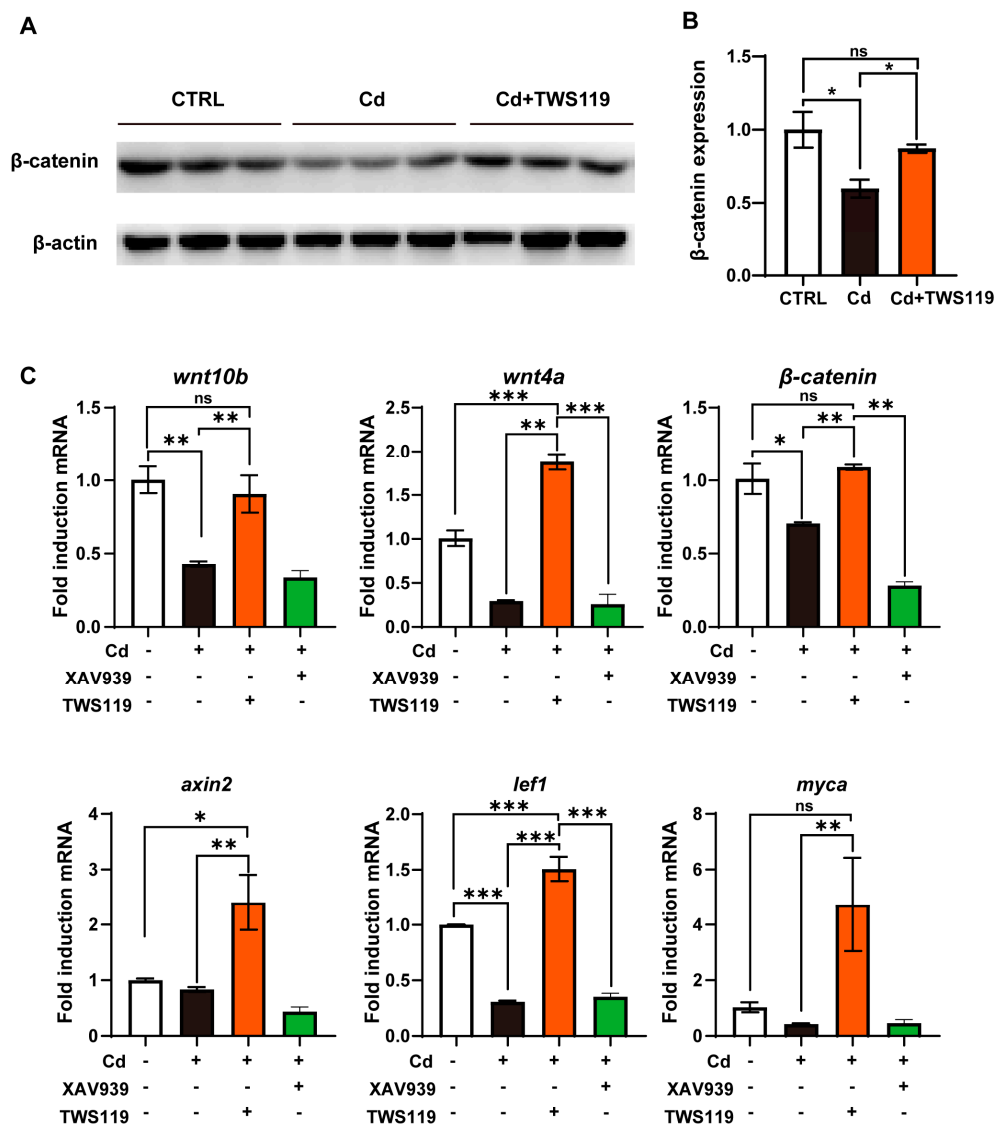
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	
	Cd+TWS119 vs. Cd	-	**	0.002	
	Cd+XAV939 vs. Cd	-	ns	0.204	
The number of AO positive cells (Figure. 5D)		One-way ANOVA	***	F=199.9 P<0.001	10
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	
	Cd+TWS119 vs. Cd	-	***	<0.001	
	Cd+XAV939 vs. Cd	-	**	0.001	
The expression profiles of apoptotic related genes (Figure. 5E)	<i>bcl</i>	One-way ANOVA	***	F=17.71 P<0.001	3
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	*	0.021	
	Cd+TWS119 vs. Cd	-	*	0.014	
	Cd+XAV939 vs. Cd	-	*	0.044	
	<i>p53</i>	One-way ANOVA	***	F=40.71 P<0.001	
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	**	0.003	
	Cd+TWS119 vs. Cd	-	**	0.004	
	Cd+XAV939 vs. Cd	-	***	<0.001	
	<i>bax</i>	One-way ANOVA	***	F=68.49 P<0.001	
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	

	Cd+TWS119 vs. Cd	-	***	<0.001	
	Cd+XAV939 vs. Cd	-	ns	0.368	
	<b>cas8</b>	One-way ANOVA	***	F=89.78 P<0.001	
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	
	Cd+TWS119 vs. Cd	-	***	<0.001	
	Cd+XAV939 vs. Cd	-	ns	0.216	
	<b>cas3a</b>	One-way ANOVA	***	F=27.86 P<0.001	
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	*	0.045	
	Cd+TWS119 vs. Cd	-	**	0.008	
	Cd+XAV939 vs. Cd	-	*	0.018	
<b>Microglia surface (Figure. 6B)</b>		One-way ANOVA	***	F=22.86 P<0.001	
	Cd				8
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	8
	Cd+TWS119 vs. Cd	-	**	0.008	8
	Cd+XAV939 vs. Cd	-	*	0.034	7
<b>Microglia Sphericity (Figure. 6C)</b>		One-way ANOVA	***	F=8.484 P<0.001	
	Cd				8
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	8
	Cd+TWS119 vs. Cd	-	**	0.003	8

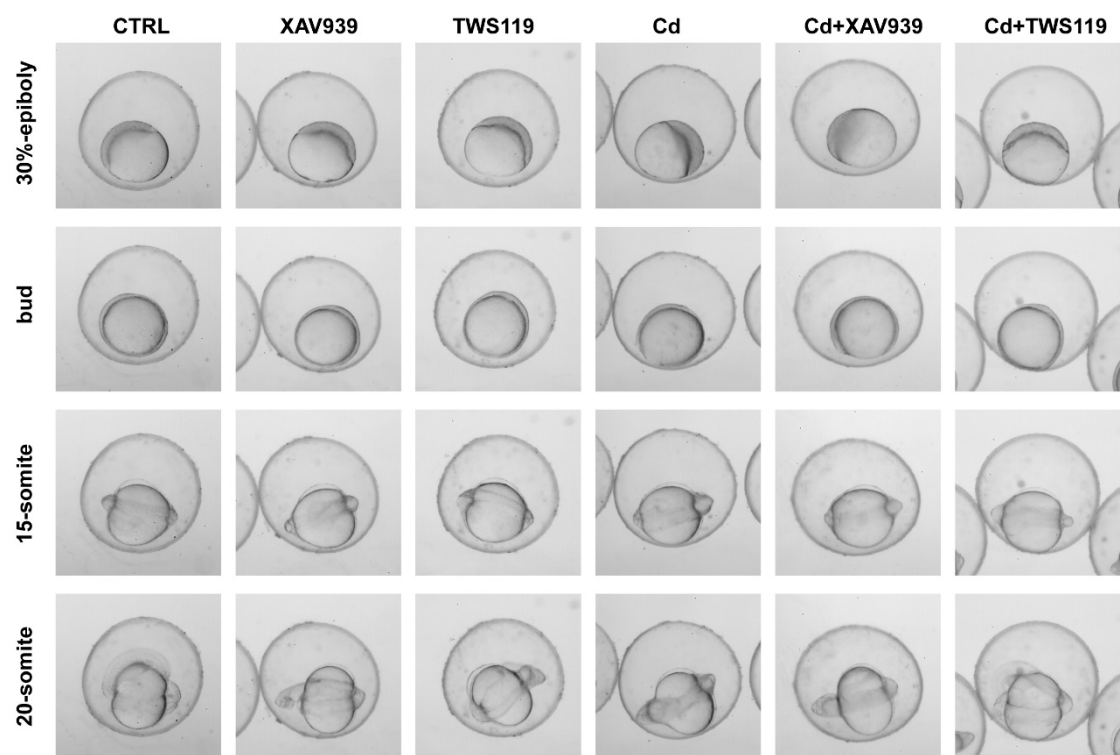
	Cd+XAV939 vs. Cd	-	ns	0.85	7
Microglia Volume (Figure. 6D)		One-way ANOVA	***	F=23.20 P<0.001	
	Cd				8
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	8
	Cd+TWS119 vs. Cd	-	*	0.021	8
	Cd+XAV939 vs. Cd	-	*	0.011	7
Number of microglia in optic tectum (Figure. 6F)		One-way ANOVA	***	F=12.43 P<0.001	
	Cd				11
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	8
	Cd+TWS119 vs. Cd	-	**	0.001	8
	Cd+XAV939 vs. Cd	-	ns	0.997	8
Microglia and inflammation related genes (Figure 6G)	<i>irf8</i>	One-way ANOVA	***	F=24.16 P<0.001	3
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	**	0.002	
	Cd+TWS119 vs. Cd	-	**	0.006	
	Cd+XAV939 vs. Cd	-	ns	0.228	
	<i>apoeb</i>	One-way ANOVA	***	F=77.87 P<0.001	
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	
	Cd+TWS119 vs. Cd	-	***	<0.001	
	Cd+XAV939 vs. Cd	-	ns	0.482	



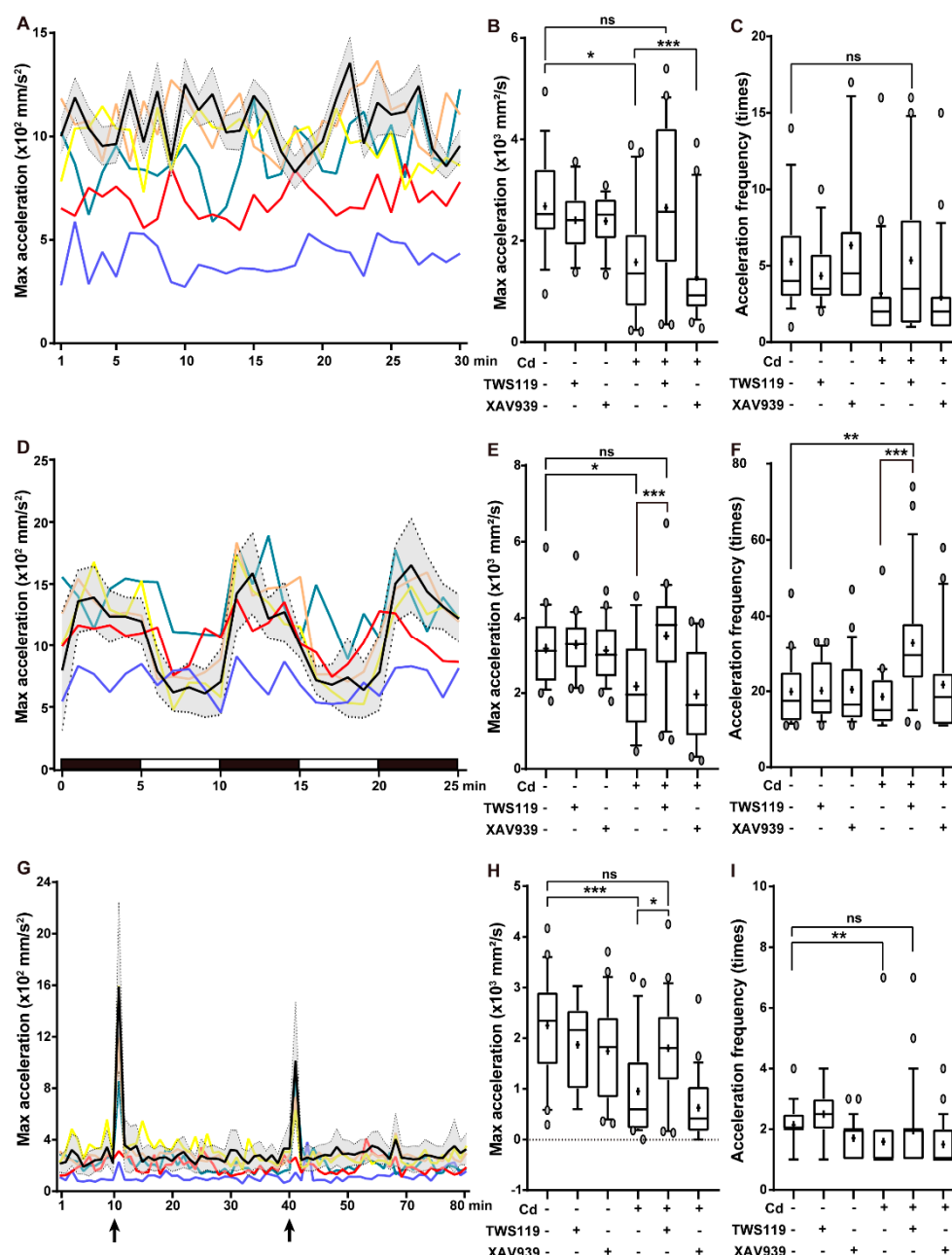
	<i>tnf-a</i>	One-way ANOVA	***	F=33.02 P<0.001	
	Cd				
	CTRL vs. Cd	Dunnett's multiple comparisons	***	<0.001	
	Cd+TWS119 vs. Cd	-	***	<0.001	
	Cd+XAV939 vs. Cd	-	ns	0.062	



**Figure S1.** Effects of TWS119 and XAV939 on Wnt signaling pathway related proteins and genes.



**Figure S2.** Representative images of high-throughput time-lapse tracking of zebrafish early development. See also **Video S1**.



**Figure S3. Zebrafish locomotor activities and reactivity to light-dark/vibration stimulation in different groups.** Swimming distance, velocity of 6 dpf zebrafish larvae in 30 minutes (A-C), subjected to light-dark cycles (D-F) and vibration stimulation in every 30 s (as indicated by the arrows below) (G-I). Left panels were plotted in 5 min or 10 s time bin and right panels were plotted with average values of the whole-tracking. Each experiment was performed independently three times. The values are presented as median (line) and mean (+), while whiskers show 90% confidence levels in boxplots. Nonparametric Kruskal-Wallis test or one-way ANOVA followed by multiple comparisons test results are reported in **Table S2**. Significant differences are indicated by asterisks (\*\*\*p < 0.001, \*\*p < 0.01, \*p < 0.05).

**Video S1.** High-throughput time-lapse imaging to track the early development of zebrafish embryos.