

Supplementary tables:

Table S1: Primary and secondary antibodies used in Western Blot assays.

Antibody	Supplier	Reference
Anti-PCSK9	Abcam	Ab181142
Anti-TLR4	Santa Cruz Biotech.	Sc-293072
Anti-P-p38 (Thr180/Tyr182)	Cell Signaling	#4511
Anti-p38	Cell Signaling	#8690
Anti-vinculin	Cell signaling	#18799
Anti-caspase-1 p10	Cell Signaling	#3866 AF4022
Anti-cleaved caspase-3	Cell Signaling	#9664
Anti-I κ B- α	Cell Signaling	#9242
Anti-P-I κ B- α (Ser32)	Cell Signaling	#2859
Anti-P-p65 (Ser536)	Cell Signaling	#3033
Anti-p65	Cell Signaling	#8242
Anti- β -actin	Sigma	A5441
Anti-rabbit IgG	Bio-Rad	#1706515
Anti-mouse IgG	Sigma	A-4416
Anti-goat IgG	Sigma	A-5420

Table S2. Specific primer sequences used in qRT-PCR assay (sequences; 5'–3').

Gen	Forward	Reverse
Human primer sequences		
<i>ABCA1</i>	ACCCACCCTATGAACAACATGA	GAGTCGGGTAACGGAAACAGG
<i>ABCG1</i>	CGTGCGCTTTGTGCTGTTT	CCACTGTAGGTACGTGGGGAT
<i>CD36</i>	CTTTGGCTTAATGAGACTGGGAC	GCAACAAACATCACCACACCA
<i>CYBB</i>	ACCGGGTTTATGATATTCCACCT	GATTTCGACAGACTGGCAAGA
<i>FASN</i>	AAGGACCTGTCTAGGTTTGATGC	TGGCTTCATAGGTGACTTCCA
<i>HMGCR</i>	TGATTGACCTTTCCAGAGCAAG	CTAAAATTCGCATTCCACGAGC
<i>IL1B</i>	TCGAGGCACAAGGCACAACAGG	GCCATGGCTGCTTCAGACACTTGA
<i>ITGAM</i>	GCCTTGACCTTATGTCATGGG	CCTGTGCTGTAGTCGCACT
<i>LDLR</i>	TCTGCAACATGGCTAGAGACT	TCCAAGCATTGTTGGTCCC
<i>NLRP3</i>	GATCTTCGCTGCCATCAACAG	CGTGCATTATCTGAACCCAC
<i>PCSK9</i>	GGACTCCTCTGTCTTTGCCC	TCCCGGTGGTCACTCTGTAT
<i>SREBP1</i>	CGCAACCATCTTGGCAACAGT	CGCTTCTCAATGGCGTTGT
<i>TNF</i>	GAGGCCAAGCCCTGGTATG	CGGGCCGATTGATCTCAGC
<i>36B4</i>	GGATTACACCTTCCCACTTGC	GCCACAAAGGCAGATGGATCA
Mouse primer sequences		
<i>Il1b</i>	AGAAGCTGTGGCAGCTACCTG	GGAAAAGAAGGTGCTCATGTCC
<i>Pcsk9</i>	GCCCATCGGGAGATTGAGG	TTCCCTTGACAGTTGAGCACA
<i>Tnf</i>	CATCTTCTCAAAATTCGAGTGACAA	TGGGAGTAGACAAGGTACAACCC
<i>36b4</i>	ACTGGTCTAGGACCCGAGAAG	TCCACCTTGTCTCCAGTCT

Supplementary figures:

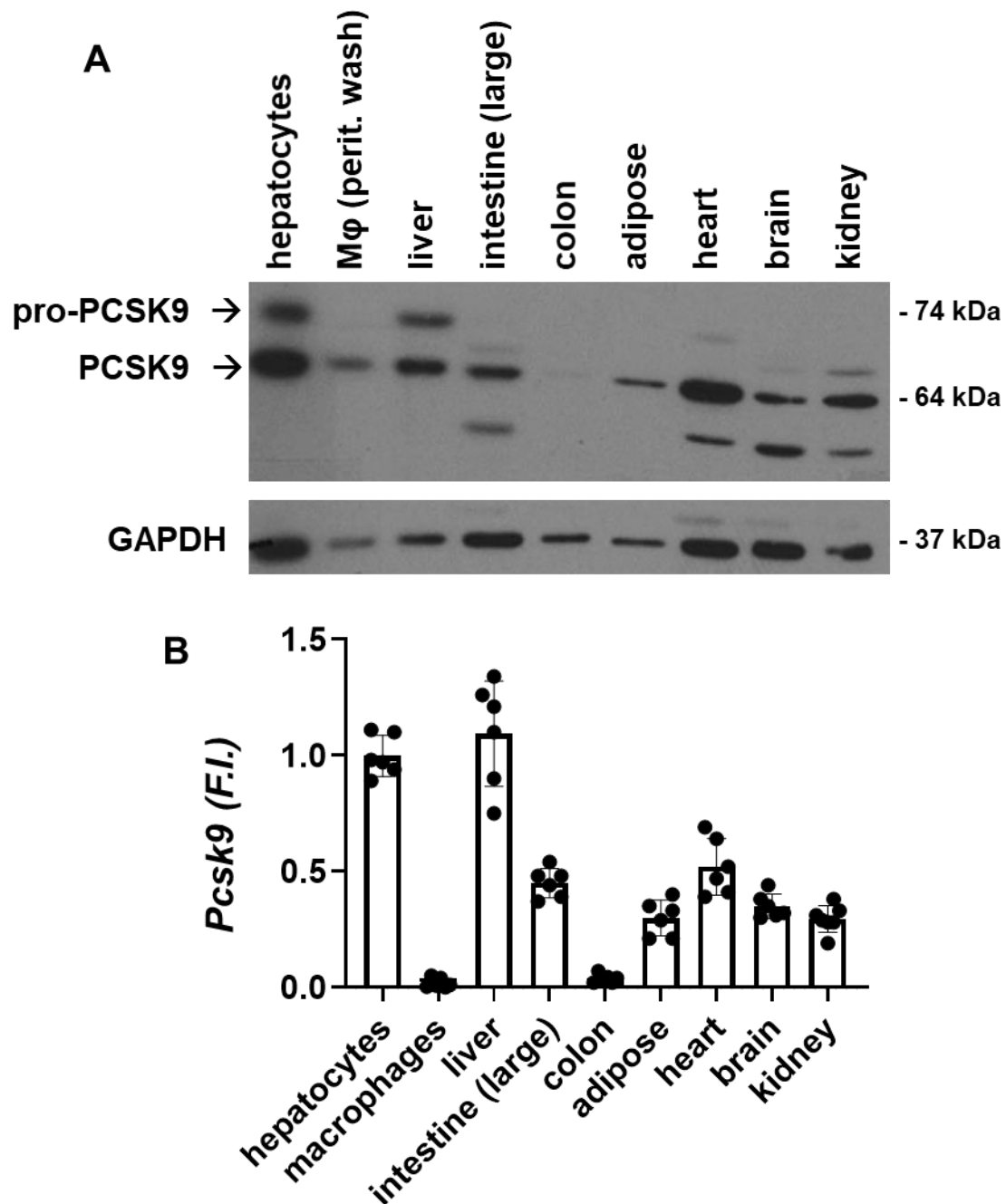


Figure S1. Intracellular content of PCSK9 and mRNA levels of *Pcsk9* in murine tissues. (A) Quantification of PCSK9 in different murine tissues. Mice were fed ad libitum and tissues and cells were prepared, homogenized and the protein levels were determined by Western blot. (B) Determination of mRNA levels of *Pcsk9* in the corresponding samples analyzed in panel A. Results show a representative blot (A) or the mean \pm SD of the corresponding mRNA levels of 6 independent preparations, using *36b4* as internal control, and referencing hepatocyte content.

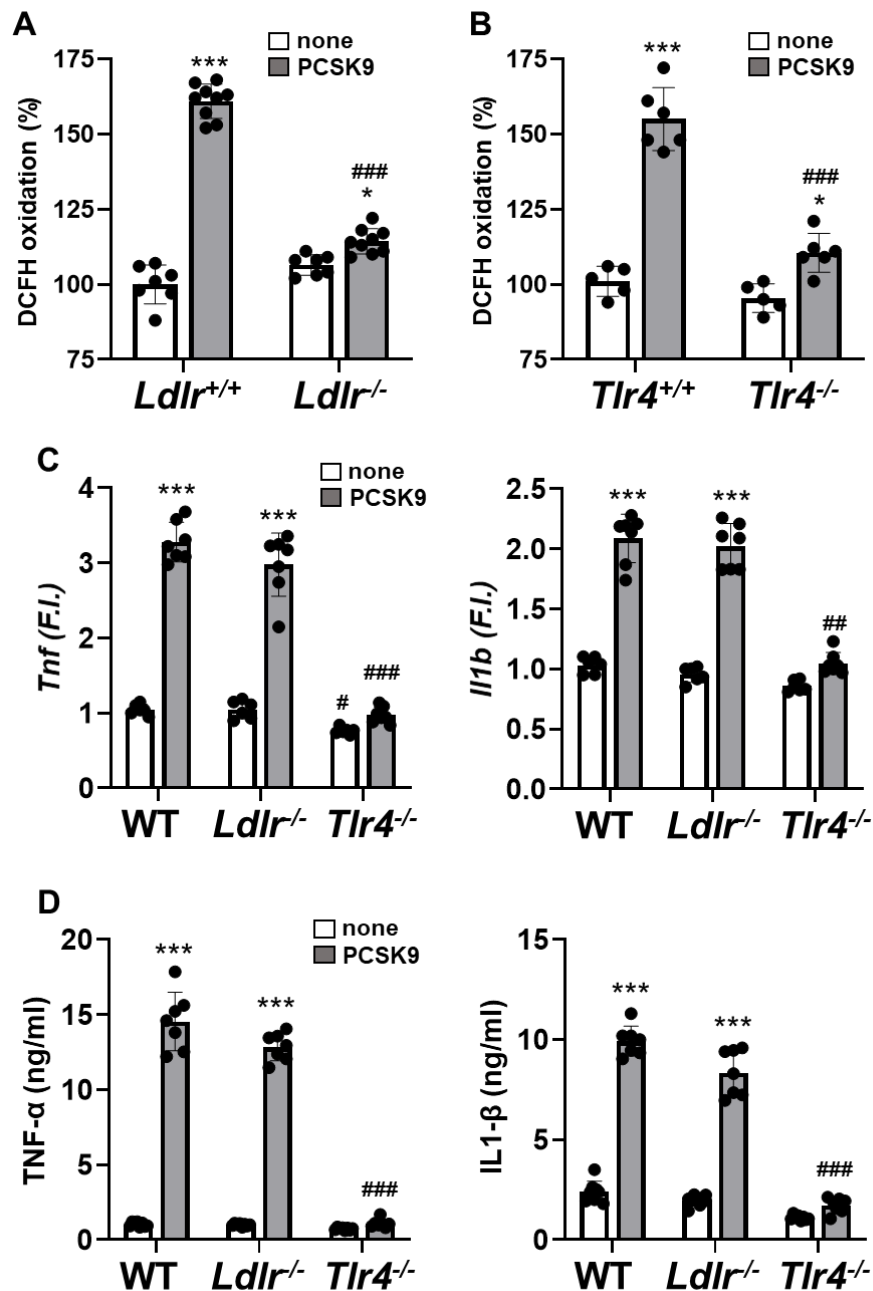


Figure S2. Effect of PCSK9 on the production of ROS and proinflammatory cytokines in elicited peritoneal macrophages from wild-type (WT), *Ldlr*^{-/-} and *Tlr4*^{-/-} mice. (A-B) Peritoneal macrophages from the indicated mice genotypes were treated for 18h with 1 μ g/ml of recombinant PCSK9. The oxidation of DCFH was measured as an indication of ROS synthesis. (C) The mRNA levels of *Tnf* and *Il1b*, and (D) the release of TNF- α and IL-1 β to the culture medium were determined in peritoneal macrophages from mice of the indicated genotype treated for 4h with 1 μ g/ml of recombinant PCSK9. Data show the mean \pm SD of macrophages from 5 mice of each genotype, assayed per triplicate. * P <0.05; *** P <0.001 vs. the corresponding condition in the absence of PCSK9. # P <0.05; ## P <0.005; ### P <0.001 vs. the corresponding WT counterparts.