

Superhydrophobic paper-based microfluidic field-effect transistor biosensor functionalized with semiconducting single-walled carbon nanotube and DNAzyme for hypocalcemia diagnosis

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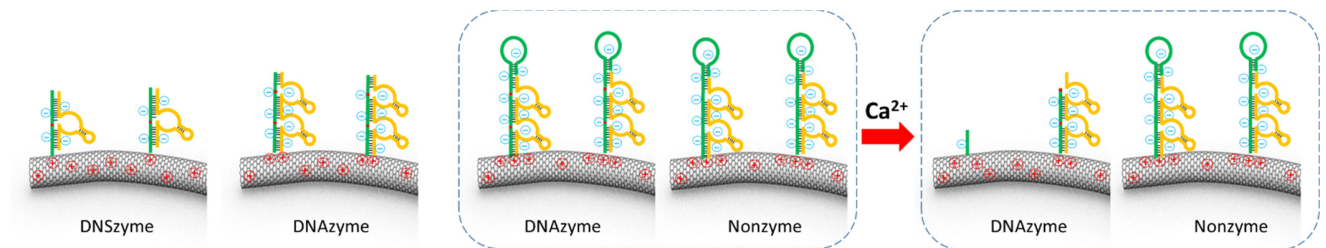


Figure S1. Mechanism of electrochemical field-effect biosensor for Ca^{2+} detection.

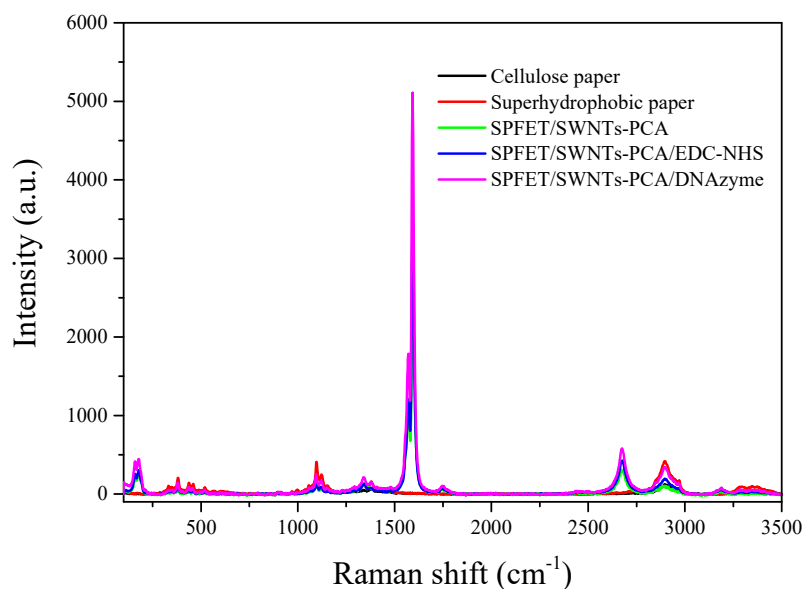


Figure S2. Raman spectra of cellulose paper before and after functionalized different materials: (a)Cellulose paper; (b)Superhydrophobic paper; (c)SPFET/SWNTs-PCA; (d)SPFET/SWNT-PCA/EDC-NHS and (e) SPFET/SWNT-PCA/DNAzyme.

Table S1. The ratios of different bands to G-band

	SWNT-PCA	EDC-NHS	DNAzyme
I_{RBM}/I_G	0.0984	0.0805	0.0814
I_D/I_G	0.0325	0.0286	0.0263
I_G/I_G	0.109621	0.114043	0.105141

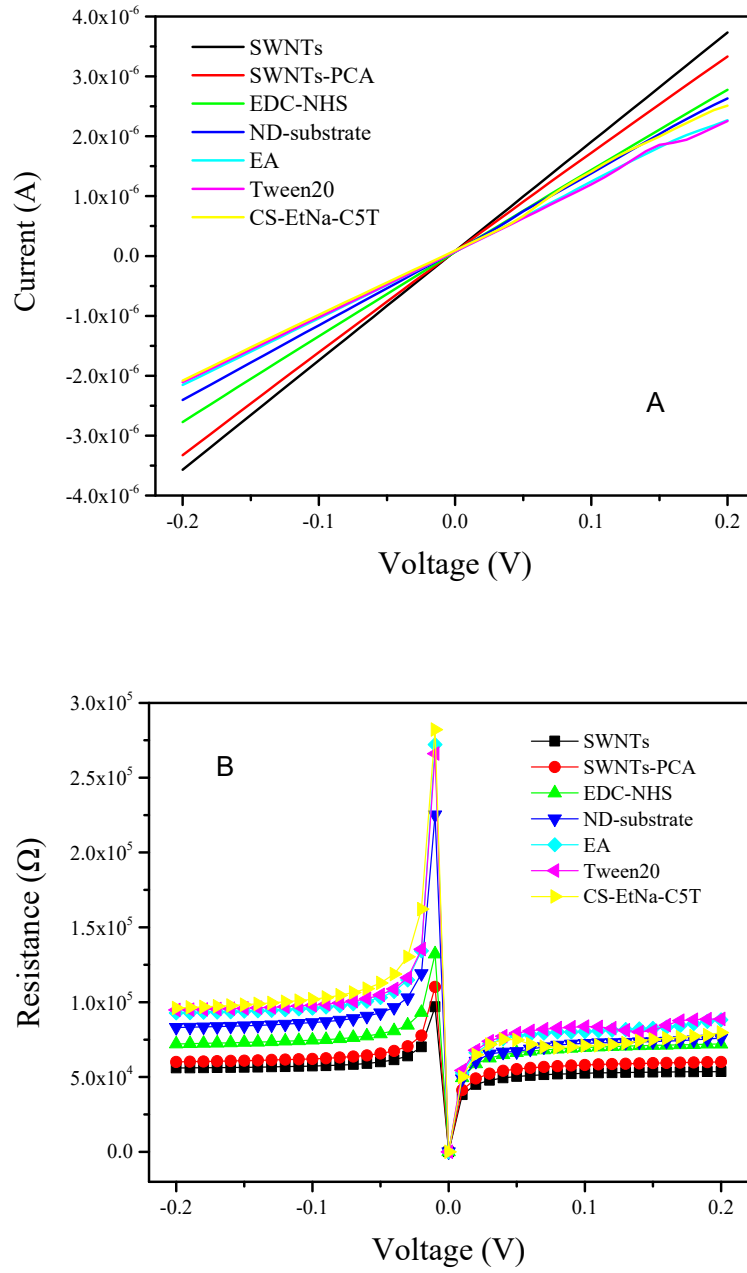


Figure S3. (A) I_{DS} - V_{DS} and (B) Resistances of HPFET modified with different materials (SWNTs, PCA, EDC-NHS, ND-substrate, EA, Tween 20 and CS-EtNa-C5T) at different voltages.

Optimization

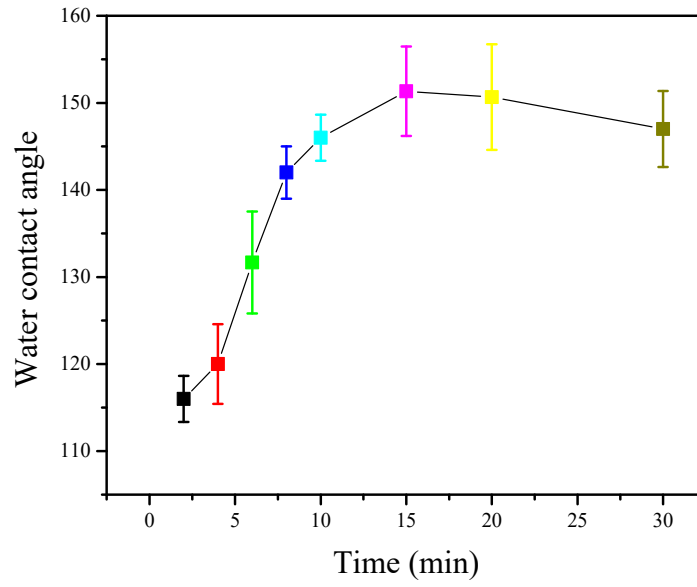


Figure S4. Water contact angle of cellulose paper changing with immersion time from 2 min to 30 min.

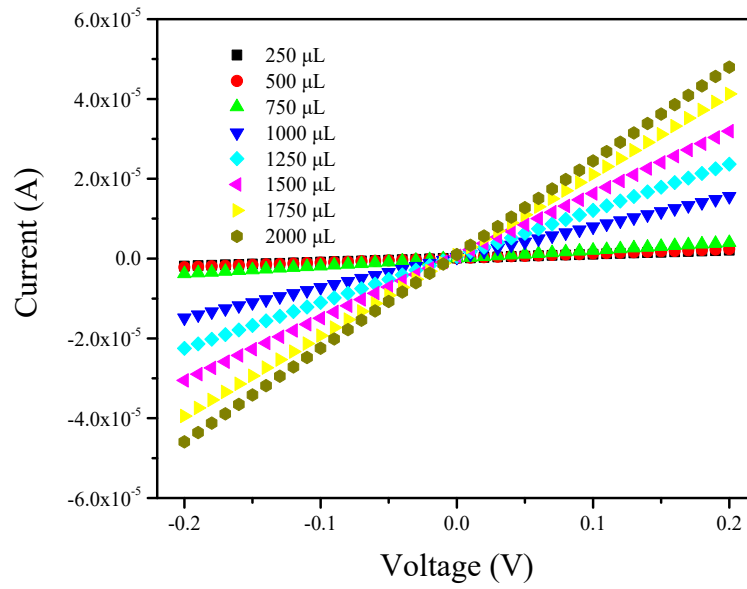
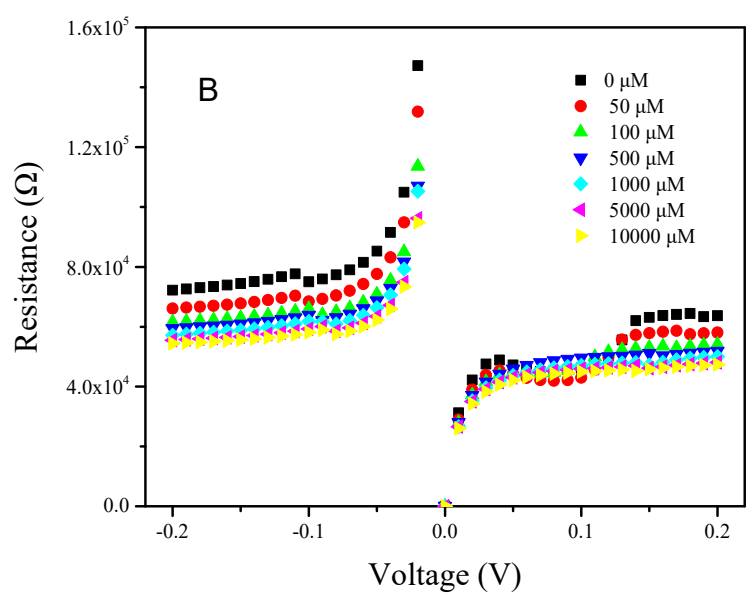
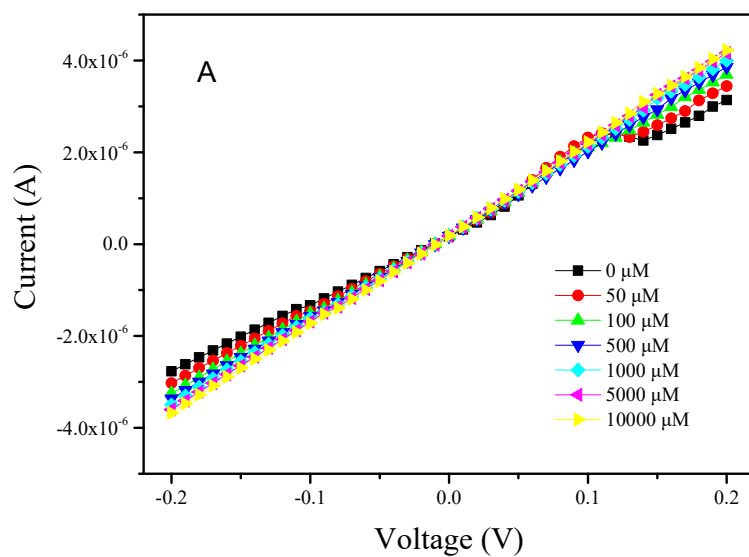


Figure S5. The I_{DS} – V_{DS} of HPFET modified with different volume of SWNTs-PCA.



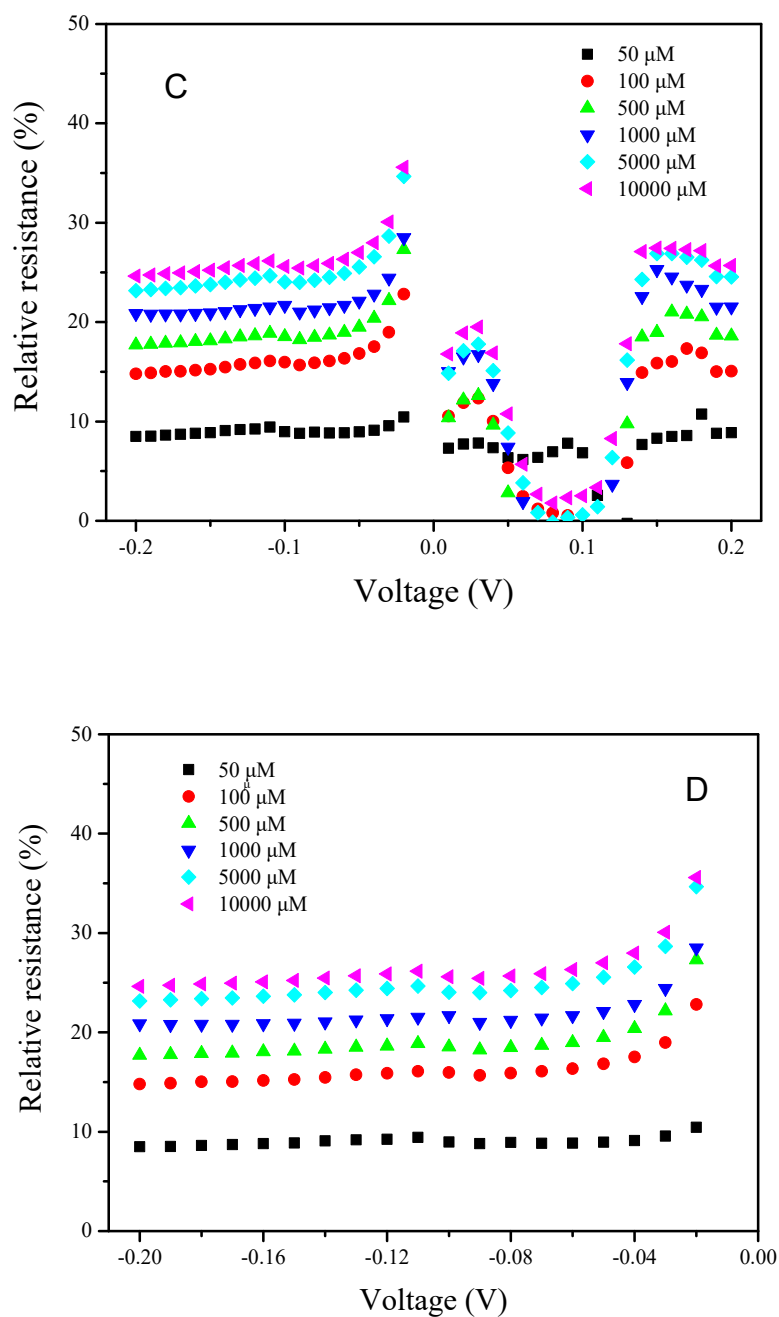


Figure S6. (A)The current-voltage of SPFET/SWNTs-PCA/DNAzyme exposure to different Ca^{2+} concentrations; (B)The resistances at each voltages after SPFET/SWNTs-PCA/DNAzyme exposure to different Ca^{2+} concentrations; (C)The relative resistances

ranging from -0.2 V to 0.2 V after SPFET/SWNTs-PCA/DNAzyme exposure to different Ca^{2+} concentrations. (D) Relative resistances of SPFET/sSWNT-PCA/DNAzyme ranging from -0.2 V to 0 V.

The cleavage efficiency of DNAzyme is associated with Ca^{2+} concentration and incubation time. Three different Ca^{2+} concentrations (100, 1000, and 10000 μM) were measured by SPFET/SWNTs-PCA/DNAzyme with varying incubation times in the range from 0 to 11 min. As shown in Figure S10, the relative resistance of SPFET/SWNTs-PCA/DNAzyme was proportional to Ca^{2+} concentrations and incubation time. For the same Ca^{2+} concentration, the relative resistance increased with incubation time, but the growth rate decreased significantly when the incubation time was higher than 7 min. For this reason, 7 min was chosen as the optimal incubation time for the experiment.

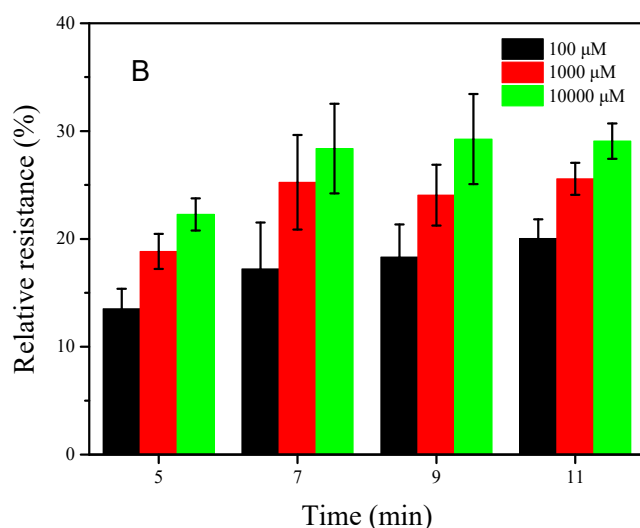


Figure S7. Relative resistances of SPFET/SWNTs-PCA/DNAzyme changing with the

incubation time in the range from 0 to 11 min for three different Ca^{2+} concentrations.

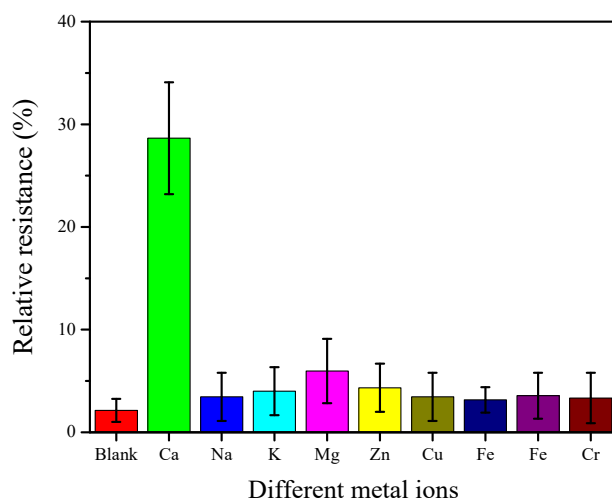


Figure S8. Relative resistances of SPFET/SWNT/DNAzyme before and after incubation in 10 mM solutions of different metal ions including Na^+ , K^+ , Mg^{2+} , Zn^{2+} , Cu^{2+} , Fe^{2+} , Fe^{3+} , and Cr^{3+} .

For SPFET/SWNTs-PCA/DNAzyme, the regression equations were $y_1 = 22.132x + 39.265$ and $y_2 = 5.4265x + 23.577$ with the R-squared of 0.9939 and 0.9782, respectively, which the detection limit was $10.7 \mu\text{M}$ ($N/S=3$). For SPFET/SWNTs-PCA/Nonzyme, the relative resistance revealed excellent relationship with the logarithm of Ca^{2+} concentration in the range of 0.025 mM to 10 mM. The linear regression equation was $y = 2.3384x + 4.5091$ with the linear regression correlation coefficient of 0.987.

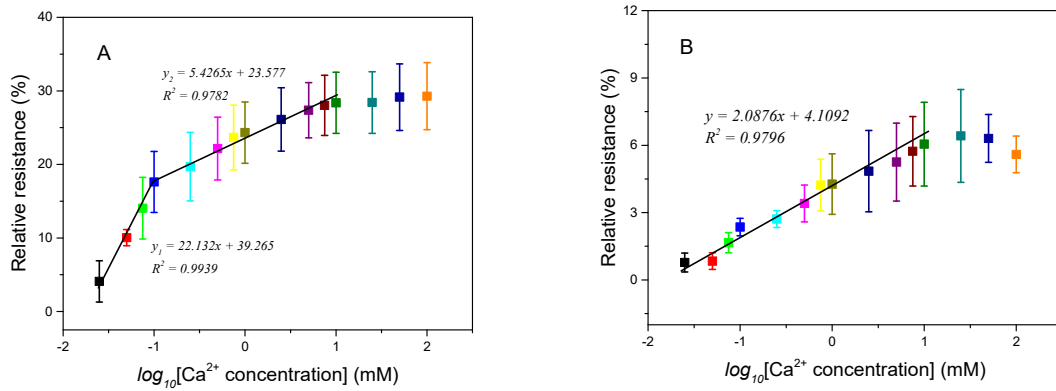


Figure S9. The linear relationship between the relative resistances of SPFET/SWNTs-PCA/DNAzyme(A) and SPFET/SWNTs-PCA/Nonzyme(B) and the logarithm of the Ca^{2+} concentration.

Table S2. Comparison of the performances of Dual-BioFET and other reported sensors for Ca^{2+} determination.

Sensor	Method	pH	Time (min)	Linear range (μM)	LOD (μM)	Ref.
Ca-ISE	Potential	2.0	1.67	$3.1 \times 10^{-3} \sim 3.2 \times 10^4$	-	1
Ca^{2+} -SCISE	Potential	-	1.0	$1.0 \times 10^{-1} \sim 1.0 \times 10^4$	5.00	2
ECL-based ion-selective nano-optodes	Potential	7.4	-	$1.0 \times 10^0 \sim 5.0 \times 10^2$	0.11	3
Fe_2O_3 -ZnO NRs/FET	Curent	7.6	-	$1.0 \times 10^1 \sim 3.0 \times 10^3$	0.05	4
Co_3O_4 CNT	Curent	-	0.84	$1.0 \times 10^2 \sim 1.1 \times 10^4$	3.80	5
IS-OECT	Curent	-	20	$1.0 \times 10^1 \sim 1.0 \times 10^4$		6
CD-EGTA	Fluorescent	7	240	$1.5 \times 10^1 \sim 3.0 \times 10^2$	0.38	7
DLLME	UV-visiable	12.8	30	$1.5 \times 10^0 \sim 3.7 \times 10^1$	0.43	8
SD-ISO	Fluorescence	6.0-8.0	5	$1.0 \times 10^1 \sim 1.0 \times 10^6$	9.30	9
SA-4CO ₂ Et	Fluorescence	4.0-10.0	240	$6.0 \times 10^2 \sim 3.0 \times 10^3$	-	10
DNAzyme/SWNT/FET	Curent	6.9	9	$1.0 \times 10^1 \sim 1.0 \times 10^3$	7.20	11
Dual BioFET	Curent	7.4	7	$2.5 \times 10^1 \sim 5.0 \times 10^3$	10.70	This work

CD-EGTA: carbon dot-ethylenebis(oxyethylenenitrilo)tetraacetic acid; ISO: ion-selective optode; SD: solvatochromic

dye; ISE: potentiometry based on a calcium-selective electrode; SCISE: solid-contact calcium-selective electrode; IS-OECT: ion-selective organic electrochemical transistor; SA: salicyladazine; ECL: electrochemiluminescence;

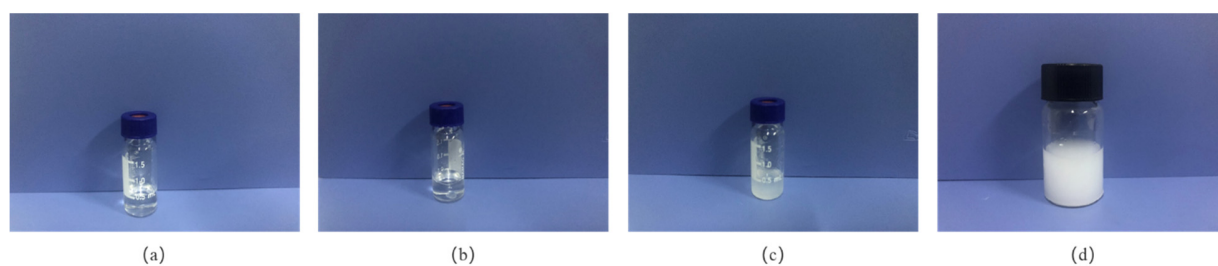


Figure S10. Preparation of superhydrophobic solution: (a) pure OTS; (b) OTS added with water; (c) mixing by vortex and sonication; and (d) incubation for 2 h under ambient conditions and dilution with hexane.

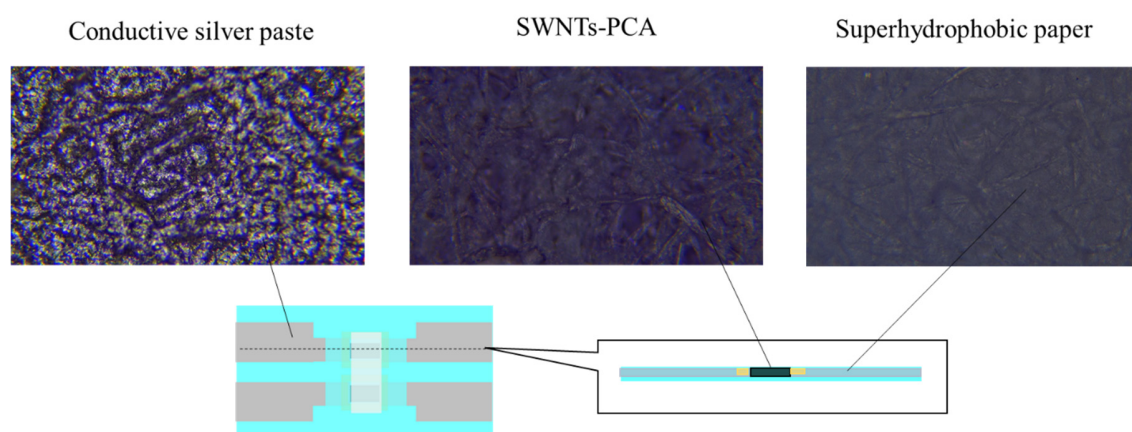
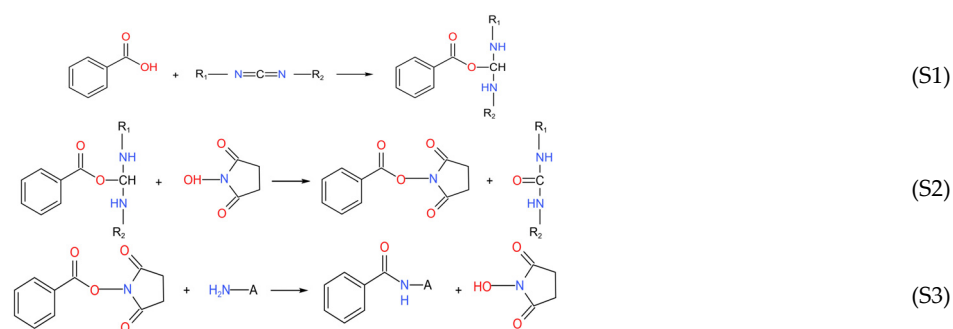


Figure S11. Structural diagram and microscope photographs of SPFET/SWNTs-PCA.



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