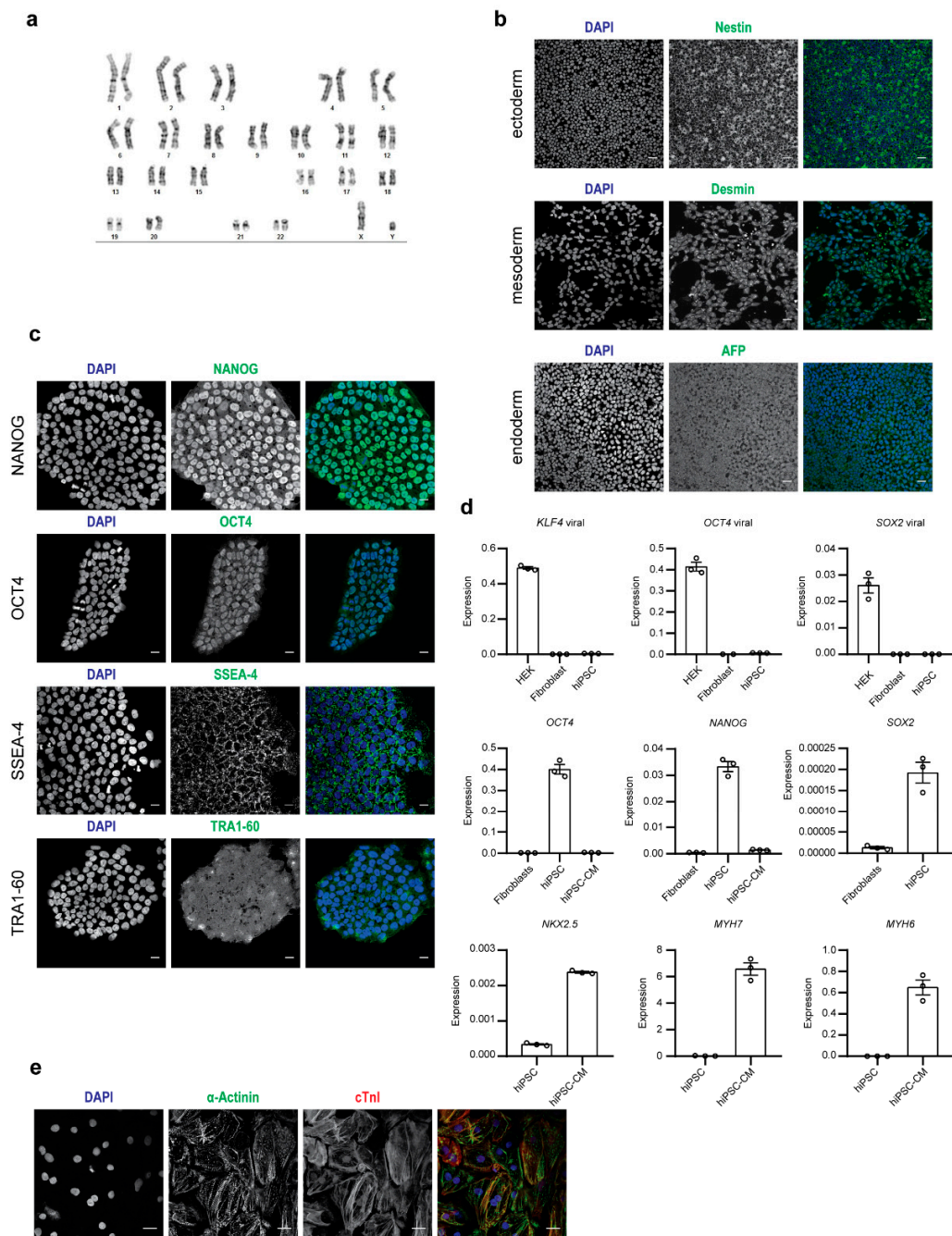


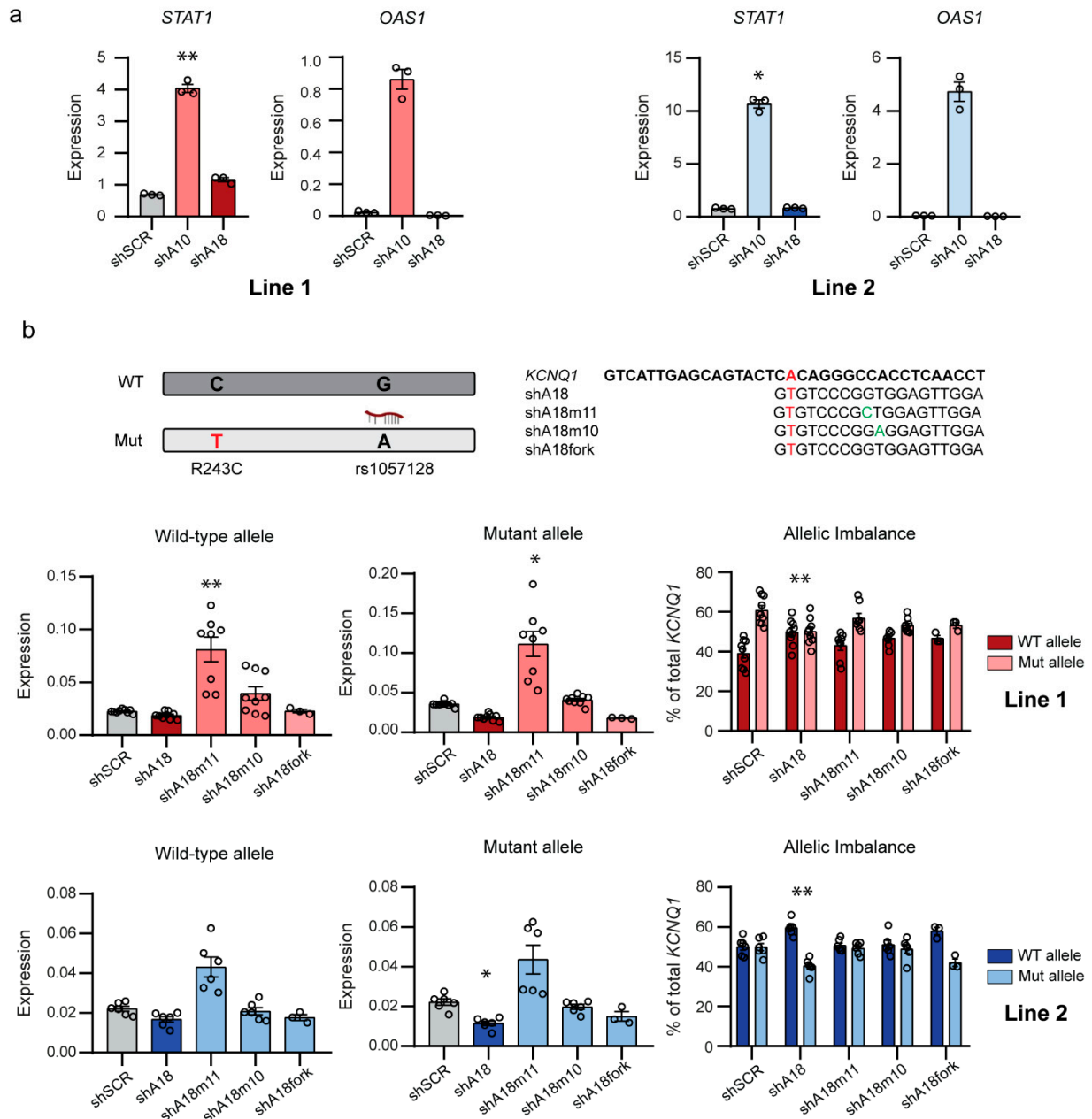
Lucía Cócera-Ortega et al. shRNAs Targeting a Common *KCNQ1* Variant Could Alleviate Long-QT1 Disease Severity by Inhibiting a Mutant Allele. *Int. J. Mol. Sci.* 2022

## **Supplemental tables and figures**



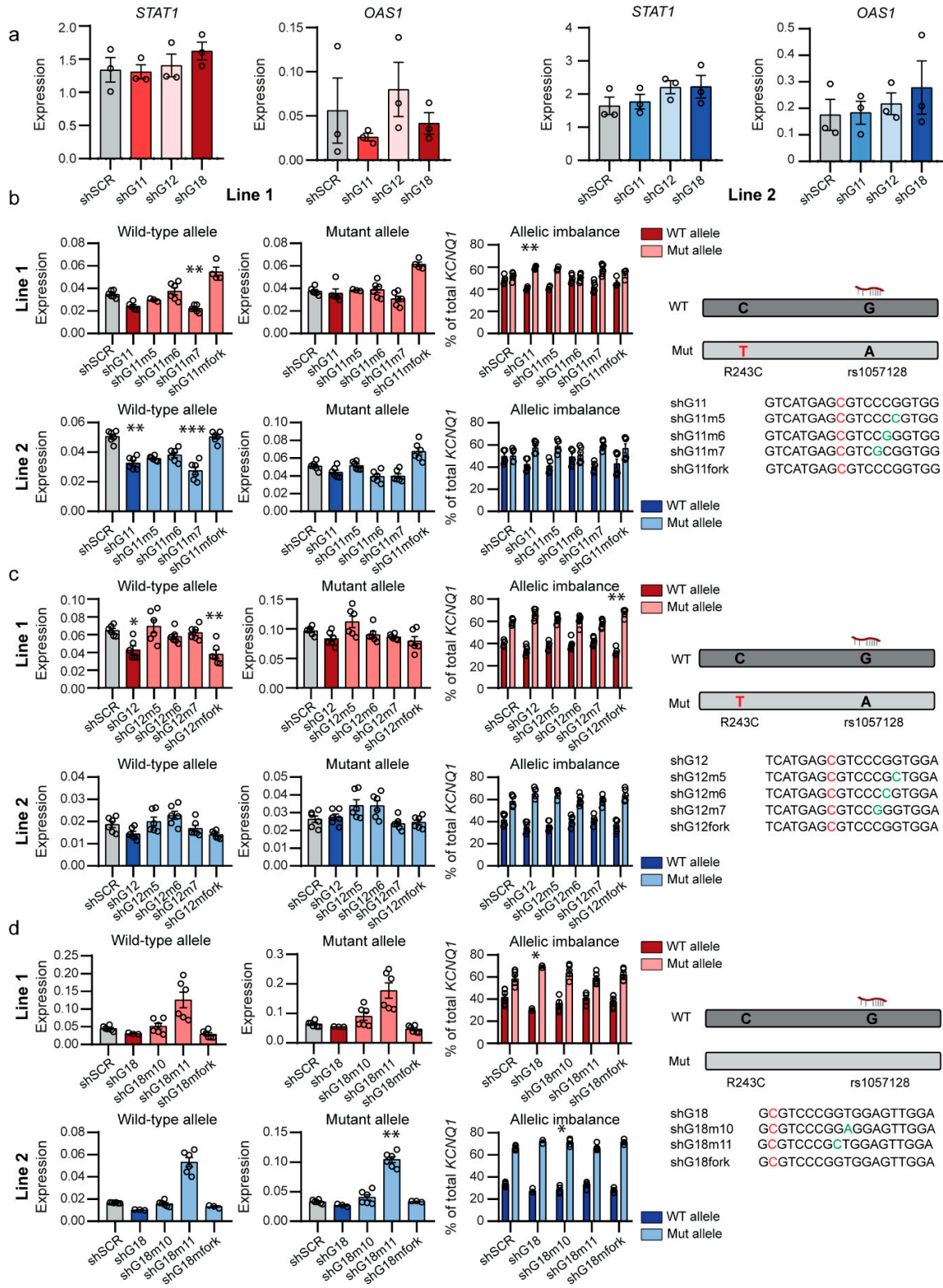
**Figure S1. Characterization of Line 2.** **a)** Karyotype analysis of hiPSCs of Line 2. **b)** Immunofluorescence after in vitro 3 lineage differentiation. Scale bars 20  $\mu$ m. **c)** Immunofluorescence of pluripotency markers in undifferentiated hiPSCs. Scale bars 20  $\mu$ m. **d)** Downregulation of viral genes in hiPSCs after induction of pluripotency with HEK cells transfected with the viral plasmids as positive control, expression of pluripotency markers OCT4, NANOG and SOX2 in undifferentiated hiPSCs and expression of cardiomyocyte markers NKX2.5, MYH7 and MYH6 in hiPSC-CMs measured by RT-qPCR. Error bars indicate SEM. **e)** Immunofluorescence of cardiomyocyte markers in hiPSC-CMs. Scale bars 20  $\mu$ m. For all immunofluorescence pictures the separate panels

on the left are merged into the rightmost panel with colour coding as indicated above the separate panels. Characterization of line 1 is included elsewhere (Tijssen et al. manuscript in draft).

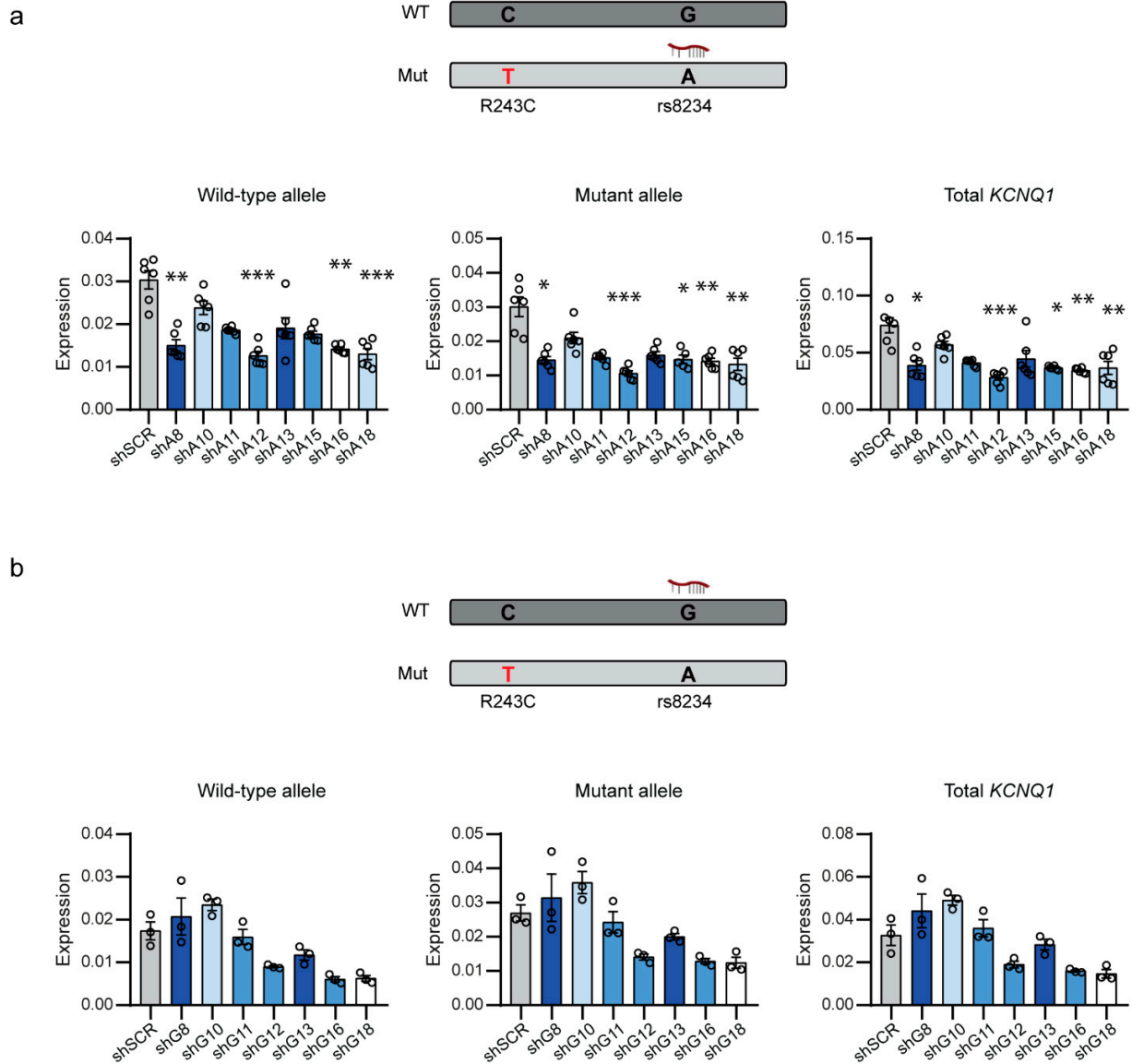


**Figure S2. Introduction of additional mismatches does not improve allele-specificity of shRNAs targeting the A allele of SNP rs1057128 on the mutant *KCNQ1* allele.** **a)** Relative mRNA expression of interferon response genes *STAT1* and *OAS1* for the most efficient shRNAs targeting the A allele of rs1057128 in hiPSC-CMs of Line 1 (red; left) and Line 2 (blue; right). **b)** Top, schematic representation of the shRNAs targeting the A allele of rs1057128 on the mutant *KCNQ1* allele with the mismatch positions indicated (original mismatch in red and additional mismatches in green). Middle and bottom, allele-specific relative mRNA expression of the wild-type and mutant *KCNQ1* alleles

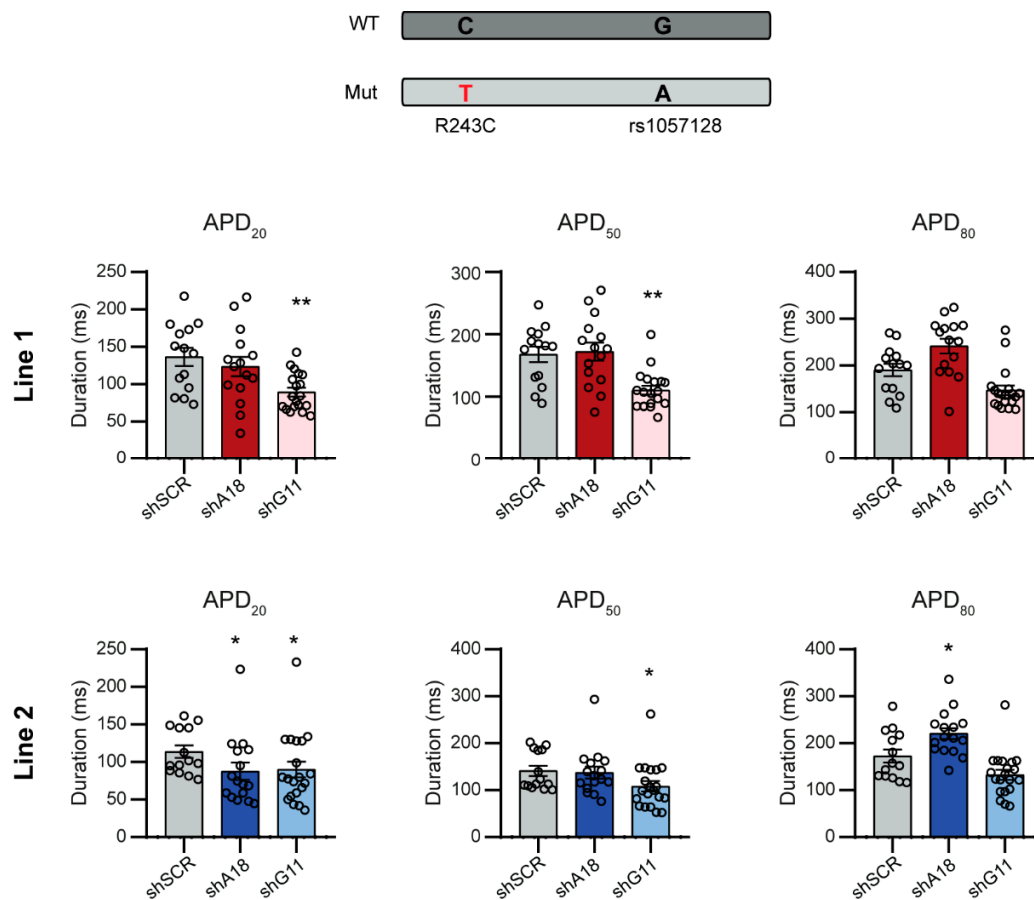
(left two panels) and the allelic expression of the wild-type and mutant *KCNQ1* alleles presented as % of total *KCNQ1* expression (right panel). In red hiPSC-CMs of Line 1 (middle, n=3-9) and in blue hiPSC-CMs of Line 2 (bottom, n=3-6). \* $P < 0.05$ ; \*\* $P < 0.025$  compared to shSCR negative control; error bars indicate SEM.



**Figure S3. Introduction of additional mismatches does not improve allele-specificity of shRNAs targeting the G allele of SNP rs1057128 on the wild-type *KCNQ1* allele.** **a)** Relative mRNA expression of interferon response genes *STAT1* and *OAS1* for the most efficient shRNAs targeting the G allele of rs1057128 in hiPSC-CMs of Line 1 (red; left) and Line 2 (blue; right). **b-d)** Right, schematic representation of the shRNAs targeting the G allele of rs1057128 on the wild-type *KCNQ1* allele with the mismatch positions indicated (original mismatch in red and additional mismatches in green). Left, allele-specific relative mRNA expression of the wild-type and mutant *KCNQ1* alleles and the allelic expression of the wild-type and mutant *KCNQ1* alleles presented as % of total *KCNQ1* expression. In red hiPSC-CMs of Line 1 (top; n=3-9) and in blue hiPSC-CMs of Line 2 (bottom; n=3-6). \* $P < 0.05$ ; \*\* $P < 0.025$ ; \*\*\* $P < 0.001$  compared to shSCR negative control; error bars indicate SEM.

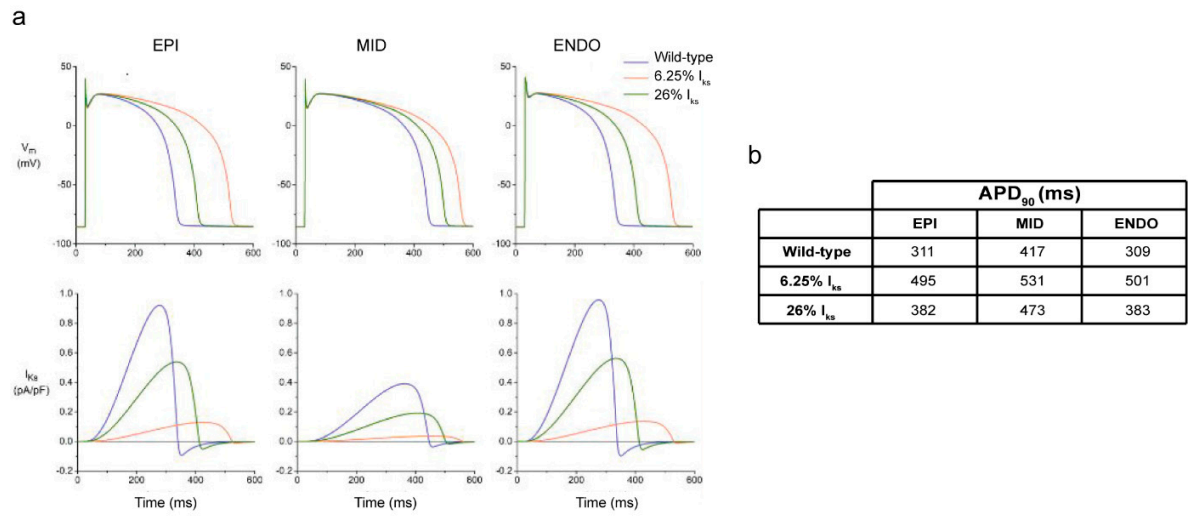


**Figure S4. Allele specific shRNAs targeting SNP rs8234 in the 3'UTR of *KCNQ1*.** **a)** Top, representation of the targeting of the A allele of rs8234 on the mutant *KCNQ1* allele in hiPSC-CMs of Line 2. Bottom, relative allele-specific expression of the wild-type and mutant *KCNQ1* allele and total *KCNQ1* (n=6). **b)** Top, representation of the targeting of the G allele of rs8234 on the wild-type *KCNQ1* allele. Bottom, relative allele-specific expression of the wild-type and mutant *KCNQ1* allele and total *KCNQ1* measured by qRT-PCR (n=3). \* $P < 0.05$ ; \*\* $P < 0.025$ ; \*\*\* $P < 0.001$  compared to shSCR negative control; error bars indicate SEM.



**Figure S5. Action potential duration is affected by shifts in allelic balance.** Schematic representation of the SNP and mutation in *KCNQ1* (top) and action potential duration at 20, 50 or 80% of repolarization (APD<sub>20</sub>, APD<sub>50</sub>, and APD<sub>80</sub>, respectively) of optical action potentials derived from ArcLight fluorescence changes in hiPSC-CMs from Line 1 (red) or Line 2 (blue) treated with either negative control shSCR, shA18 targeting the mutant *KCNQ1* allele or shG11 targeting the wild-type *KCNQ1* allele stimulated at 2 Hz. \* $P < 0.05$ ; \*\* $P < 0.025$  compared to shSCR negative control; error bars indicate SEM.





**Figure S6. Effects of changes in the slow delayed rectifier potassium current ( $I_{Ks}$ ) in a human ventricular cell model. a)** Membrane potential ( $V_m$ ; top) and associated  $I_{Ks}$  (bottom) at 1 Hz stimulation under wild-type conditions, upon reduction of functional  $K_v7.1$  channels to 1/16<sup>th</sup> as expected when both alleles are equally expressed (6.25%) or to 26% as expected when the mutant *KCNQ1* allele is downregulated by 60%. **b)** Associated APD<sub>90</sub> values in the epicardial (EPI), midmyocardial (MID) and endocardial (ENDO) versions of the human cardiomyocyte model.

**Table S1**

Antibody	Supplier	Species	Dilution	Cat. number
<b>OCT-4</b>	STEMCELL Technologies Vancouver, Canada	Mouse	1:200	60093
<b>SSEA-4</b>	STEMCELL Technologies Vancouver, Canada	Mouse	1:100	60062
<b>Tra1-60</b>	STEMCELL Technologies Vancouver, Canada	Mouse	1:100	60064
<b>NANOG</b>	Peprtech Rocky Hill, NJ,USA	Rabbit	1:200	500-P236
<b><math>\alpha</math>-Actinin</b>	Abcam Cambridge, UK	Rabbit	1:500	68167
<b>cTnl</b>	HyTest Turku, Finland	Goat	1:400	4T21/2

<b>Nestin</b>	STEMCELL Technologies Vancouver, Canada	Mouse	1:1000	60091
<b>AFP</b>	ThermoFisher Rockford, IL, USA	Rabbit	1:100	PA5-16658
<b>Desmin</b>	ThermoFisher Rockford, IL, USA	Rabbit	1:100	PA5-16705
<b>Alexa fluor 488 Anti-rabbit</b>	Invitrogen Eugene, OR, USA	Goat	1:250 ICC	A32731
<b>Alexa fluor 488 Anti-mouse</b>	Invitrogen Eugene, OR, USA	Goat	1:250 ICC	A32731
<b>Alexa fluor 488 Anti-rabbit</b>	Invitrogen Eugene, OR, USA	Donkey	1:250 ICC	A10037
<b>Alexa fluor 647 Anti-goat</b>	Invitrogen Eugene, OR, USA	Donkey	1:250 ICC	A32814

Table S2

Targeted SNP	Name shRNA	Oligonucleotides for cloning of shRNAs (upper forward and lower reverse oligonucleotide)
<b>rs1057128</b>	shA18	ccggaaCACAGGGCCACCTCAACCTtcaagacAGGTTGAGGTGGCCCTGTGtttttttg
		aattcaaaaaaaCACAGGGCCACCTCAACCTgtcttgaAGGTTGAGGTGGCCCTGTGtt
<b>rs1057128</b>	shA10	ccggaaGCAGTACTCACAGGGCCACtcaagacGTGGCCCTGTGAGTACTGcttttttg
		aattcaaaaaaaGCAGTACTCACAGGGCCACgtcttgaGTGGCCCTGTGAGTACTGctt
<b>rs1057128</b>	shA13	ccggaaGTACTCACAGGGCCACCTCtcaagacGAGGTGGCCCTGTGAGTACTtttttttg
		aattcaaaaaaaGTACTCACAGGGCCACCTCgtcttgaGAGGTGGCCCTGTGAGTACTt
<b>rs1057128</b>	shA15	ccggaaACTCACAGGGCCACCTCAAtcaagacTTGAGGTGGCCCTGTGAGTtttttttg
		aattcaaaaaaaACTCACAGGGCCACCTCAAgcttgaTTGAGGTGGCCCTGTGAGTtt
<b>rs1057128</b>	shA16	ccggaaCTCACAGGGCCACCTCAACtcaagacGTTGAGGTGGCCCTGTGAGTtttttttg
		aattcaaaaaaaCTCACAGGGCCACCTCAACgtcttgaGTTGAGGTGGCCCTGTGAGtt
<b>rs1057128</b>	shG18	ccggaaCGCAGGGCCACCTCAACCTtcaagacAGGTTGAGGTGGCCCTGCGtttttttg
		aattcaaaaaaaCGCAGGGCCACCTCAACCTgtcttgaAGGTTGAGGTGGCCCTGCGtt
<b>rs1057128</b>	shG10	ccggaaGCAGTACTCGCAGGGCCACtcaagacGTGGCCCTGCGAGTACTGcttttttg

		aattcaaaaaaaGCAGTACTCGCAGGGCCACGtcttgaGTGGCCCTGCGAGTACTGctt
<b>rs1057128</b>	shG11	ccggaaCAGTACTCGCAGGGCCACctcaagacGGTGGCCCTGCGAGTACTGtttttttg
		aattcaaaaaaaCAGTACTCGCAGGGCCACCgtcttgaGGTGGCCCTGCGAGTACTGtt
<b>rs1057128</b>	shG12	ccggaaAGTACTCGCAGGGCCACCTcaagacAGGTGGCCCTGCGAGTACTtttttttg
		aattcaaaaaaaAGTACTCGCAGGGCCACCTgtcttgaAGGTGGCCCTGCGAGTACTtt
<b>rs1057128</b>	shG16	ccggaaCTCGCAGGGCCACCTCAACtcaagacGTTGAGGTGGCCCTGCGAGtttttttg
		aattcaaaaaaaCTCGCAGGGCCACCTCAACgtcttgaGTTGAGGTGGCCCTGCGAGtt
<b>rs1057128</b>	shG11m5	ccggaaCAGTACTCGCAGGGGCACctcaagacGGTGCCCTGCGAGTACTGtttttttg
		aattcaaaaaaaCAGTACTCGCAGGGGCACCgtcttgaGGTGCCCTGCGAGTACTGtt
<b>rs1057128</b>	shG11m6	ccggaaCAGTACTCGCAGGGCCACctcaagacGGTGGCCCTGCGAGTACTGtttttttg
		aattcaaaaaaaCAGTACTCGCAGGGCCACCgtcttgaGGTGGCCCTGCGAGTACTGtt
<b>rs1057128</b>	shG11m7	ccggaaCAGTACTCGCAGGCCACctcaagacGGTGGCGCTGCGAGTACTGtttttttg
		aattcaaaaaaaCAGTACTCGCAGGCCACCgtcttgaGGTGGCGCTGCGAGTACTGtt
<b>rs1057128</b>	shG11fork	ccggaaCAGTACTCGCAGGGCCATatcaagacGGTGGCCCTGCGAGTACTGtttttttg
		aattcaaaaaaaCAGTACTCGCAGGGCCACCgtcttgaTATGGCCCTGCGAGTACTGtt
<b>rs1057128</b>	shG12m5	ccggaaAGTACTCGCAGGGCGACCTcaagacAGGTGCGCCTGCGAGTACTtttttttg
		aattcaaaaaaaAGTACTCGCAGGGCGACCTgtcttgaAGGTGCGCCTGCGAGTACTtt
<b>rs1057128</b>	shG12m6	ccggaaAGTACTCGCAGGGGCACCTcaagacAGGTGCCCTGCGAGTACTtttttttg
		aattcaaaaaaaAGTACTCGCAGGGGCACCTgtcttgaAGGTGCCCTGCGAGTACTtt
<b>rs1057128</b>	shG12m7	ccggaaAGTACTCGCAGGGCCACCTcaagacAGGTGGGCCTGCGAGTACTtttttttg
		aattcaaaaaaaAGTACTCGCAGGGCCACCTgtcttgaAGGTGGGCCTGCGAGTACTtt
<b>rs1057128</b>	shG12fork	ccggaaAGTACTCGCAGGGCCACTatcaagacAGGTGGCCCTGCGAGTACTtttttttg
		aattcaaaaaaaAGTACTCGCAGGGCCACCTgtcttgaTAGTGGCCCTGCGAGTACTtt
<b>rs1057128</b>	shG18m10	ccggaaCGCAGGGCCTCCTCAACCTcaagacAGGTTGAGGAGGCCCTGCGtttttttg
		aattcaaaaaaaCGCAGGGCCTCCTCAACCTgtcttgaAGGTTGAGGAGGCCCTGCGtt

<b>rs1057128</b>	shG18m11	ccggaaCGCAGGGCGACCTCAACCTtcaagacAGGTTGAGGTGCGCCTGCGtttttttg aattcaaaaaaaCGCAGGGCGACCTCAACCTgtcttgaAGGTTGAGGTGCGCCTGCGtt
<b>rs1057128</b>	shG18 fork	ccggaaCGCAGGGCCACCTCAACTAtcaagacAGGTTGAGGTGGCCCTGCGtttttttg aattcaaaaaaaCGCAGGGCCACCTCAACCTgtcttgaTAGTTGAGGTGGCCCTGCGtt
<b>rs1057128</b>	shA18m11	ccggaaCACAGGGCGACCTCAACCTtcaagacAGGTTGAGGTGCGCCTGTGtttttttg aattcaaaaaaaCACAGGGCGACCTCAACCTgtcttgaAGGTTGAGGTGCGCCTGTGtt
<b>rs1057128</b>	shA18m10	ccggaaCACAGGGCCTCCTCAACCTtcaagacAGGTTGAGGAGGCCCTGTGtttttttg aattcaaaaaaaCACAGGGCCTCCTCAACCTgtcttgaAGGTTGAGGAGGCCCTGTGtt
<b>rs1057128</b>	shA18 fork	ccggaaCACAGGGCCACCTCAACTAtcaagacAGGTTGAGGTGGCCCTGTGtttttttg aattcaaaaaaaCACAGGGCCACCTCAACCTgtcttgaTAGTTGAGGTGGCCCTGTGtt
<b>rs8234</b>	shA8	ccggaaCTGGGCATTACATCGCATAtcaagacTATGCGATGTAATGCCAGtttttttg aattcaaaaaaaCTGGGCATTACATCGCATAgcttgaTATGCGATGTAATGCCAGtt
<b>rs8234</b>	shA10	ccggaaGGGCATTACATCGCATAGAtcaagacTCTATGCGATGTAATGCCtttttttg aattcaaaaaaaGGGCATTACATCGCATAGAgcttgaTCTATGCGATGTAATGCCtt
<b>rs8234</b>	shA11	ccggaaGGCATTACATCGCATAGAAAtcaagacTTCTATGCGATGTAATGCCtttttttg aattcaaaaaaaGGCATTACATCGCATAGAAgcttgaTTCTATGCGATGTAATGCCtt
<b>rs8234</b>	shA12	ccggaaGCATTACATCGCATAGAAAtcaagacTTTCTATGCGATGTAATGCTtttttttg aattcaaaaaaaGCATTACATCGCATAGAAAgcttgaTTTCTATGCGATGTAATGCTtt
<b>rs8234</b>	shA13	ccggaaCATTACATCGCATAGAAATtcaagacATTTCTATGCGATGTAATGtttttttg aattcaaaaaaaCATTACATCGCATAGAAATgtcttgaATTTCTATGCGATGTAATGtt
<b>rs8234</b>	shA15	ccggaaTTACATCGCATAGAAATCAAtcaagacTGATTCTATGCGATGTAAtttttttg aattcaaaaaaaTTACATCGCATAGAAATCAgtcttgaTGATTCTATGCGATGTAAtt
<b>rs8234</b>	shA16	ccggaaTACATCGCATAGAAATCAAtcaagacTTGATTCTATGCGATGTAtttttttg aattcaaaaaaaTACATCGCATAGAAATCAAgcttgaTTGATTCTATGCGATGTAtt
<b>rs8234</b>	shA18	ccggaaCATCGCATAGAAATCAATAtcaagacTATTGATTCTATGCGATGtttttttg

		aattcaaaaaaCATCGCATAGAAATCAATAgcttgaTATTGATTCTATGCGATGtt
<b>rs8234</b>	shG8	ccggaaCTGGGCATTACGTGCGATAtcaagacTATGCGACGTAATGCCAGtttttttg
		aattcaaaaaaCTGGGCATTACGTGCGATAgcttgaTATGCGACGTAATGCCAGtt
<b>rs8234</b>	shG10	ccggaaGGGCATTACGTGCGATAGAtcaagacTCTATGCGACGTAATGCCtttttttg
		aattcaaaaaaGGGCATTACGTGCGATAGAgcttgaTCTATGCGACGTAATGCCtt
<b>rs8234</b>	shG11	ccggaaGGCATTACGTGCGATAGAAAtcaagacTTCTATGCGACGTAATGCCtttttttg
		aattcaaaaaaGGCATTACGTGCGATAGAAgcttgaTTCTATGCGACGTAATGCCtt
<b>rs8234</b>	shG12	ccggaaGCATTACGTGCGATAGAAAtcaagacTTTCTATGCGACGTAATGCtttttttg
		aattcaaaaaaGCATTACGTGCGATAGAAAgcttgaTTTCTATGCGACGTAATGCtt
<b>rs8234</b>	shG13	ccggaaCATTACGTGCGATAGAAATtcaagacATTTCTATGCGACGTAATGtttttttg
		aattcaaaaaaCATTACGTGCGATAGAAATgtcttgaATTTCTATGCGACGTAATGtt
<b>rs8234</b>	shG16	ccggaaTACGTGCGATAGAAATCAAtcaagacTTGATTCTATGCGACGTAAttttttg
		aattcaaaaaaTACGTGCGATAGAAATCAAgcttgaTTGATTCTATGCGACGTAtt
<b>rs8234</b>	shG18	ccggaaCGTCGCATAGAAATCAATAtcaagacTATTGATTCTATGCGACGtttttttg
		aattcaaaaaaCGTCGCATAGAAATCAATAgcttgaTATTGATTCTATGCGACGtt

**Table S3**

PCR primers			
Gene	Forward	Reverse	Annealing temperature
<b>KCNQ1_wt</b>	TGAGGATGCTACACGTCGACC	AGCCGATGTACAGGGTGGTTATC	66
<b>KCNQ1_mut</b>	TGAGGATGCTACACGTCGACT	AGCCGATGTACAGGGTGGTTATC	66
<b>KCNQ1_total</b>	TCCTGAGGATGCTACACGTC	AGCCGATGTACAGGGTGGTTATC	66
<b>STAT1</b>	ATCACATTACATGGGTGGAGC	ACAGATACTTCAGGGGATTCTCAGG	60
<b>OAS1</b>	CATCCGCCTAGTCAAGCACT	AAGACCGTCCGAAATCCCTGG	60

<b><i>GAPDH</i></b>	ACCCACTCCTCCACCTTTGAC	ACCCTGTTGCTGTAGCCAAATT	60
<b><i>HPRT</i></b>	TGACACTGGCAAAACAATGCA	GGTCCTTTTCACCAGCAAGCT	60
<b><i>TBP</i></b>	GCTCACCCACCAACAATTTAG	TCTGCTCTGACTTTAGCACCTG	60
<b><i>NANOG</i></b>	AGAATAGCAATGGTGTGACGCAG	TGGATGTTCTGGGTCTGGTTGC	60
<b><i>OCT4</i></b>	TGGTTGGAGGGAAGGTGAAG	TGTCTATCTACTGTGTCCCAG	60
<b><i>OCT4_viral</i></b>	TGTACTCCTCGGTCCCTTTC	CAGGTGGGGTCTTTCATTC	63
<b><i>SOX2</i></b>	ACCAATCCCATCCACACTCAC	TCTATACAAGGTCCATTCCCCC	63
<b><i>SOX2_viral</i></b>	ATCCCAGTGTGGTGGTACG	AAGGCATTCATGGGCCGCTTG	63
<b><i>MLC2V</i></b>	ACAACTGACACCAACACCTGC	AGTCCAAGTTGCCAGTCACGTC	63
<b><i>NKX2.5</i></b>	TCTATCCACGTGCCTACAGC	AGAAAGTCAGGCTGGCTCAAG	63
<b><i>KLF4_viral</i></b>	CTGCGGCAAAACCTACACAAA	TTATCGTCGACCACTGTGCTGG	63
<b><i>MYH6</i></b>	ACCTGTCCAAGTCCGCAAGG	TTACAGGTTGGCAAGAGTGAGG	60
<b><i>MYH7</i></b>	ACCTGTCCAAGTCCGCAAGG	TTTGCTGGCACCTCCAGGG	60