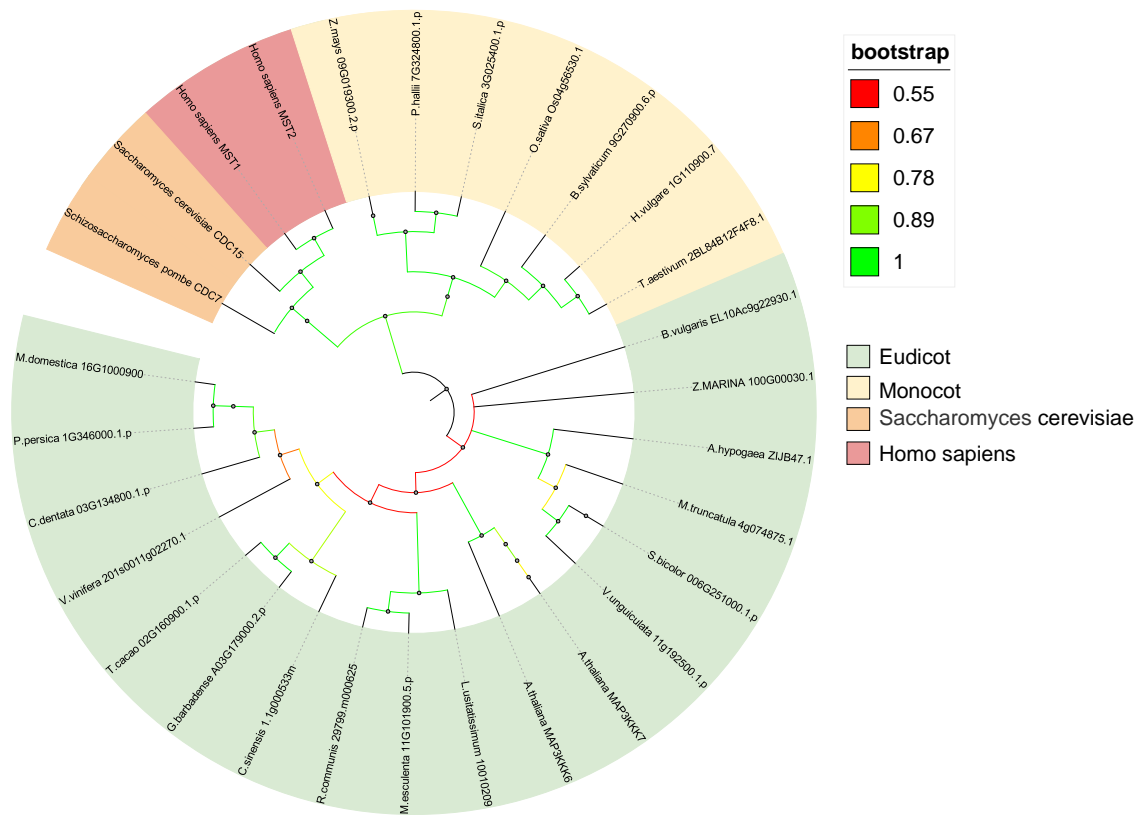


Figure. S1

A



B

		STKc_Cdc7_like											
		#####	#	#	#	#####	##	#	#####	#			
MAP3Kε1 Arabidopsis	22	LGDEIGKGAYGRV----	AIK	---	NIVKYL	GSS-TKTHLHI	LEYV	ENGSLANI	---	HRDIKGANIL	151		
MAP3Kε2 Arabidopsis	22	LGDEIGKGAYGRV----	AIK	---	NIVKYL	GSL-TKTHLHI	LEYV	ENGSLANI	---	HRDIKGANIL	151		
ZmPHJ40.09G019300.2.p Z.mays PHJ	22	LGDEIGKGAYGRV----	AIK	---	NIVKYL	GSL-TKSHLHI	LEYV	ENGSLANI	---	HRDIKGANIL	151		
LOC_Os04g56530.1 O.sativa	22	LGDEIGKGAYGRV----	AIK	---	NIVKYL	GSL-TRSHLHI	LEYV	ENGSLANI	---	HRDIKGANIL	151		
Glyma.11G101700.4.p G.maxWm	22	LGDEIGKGAYGRV----	AIK	---	NIVKYL	GSS-KSHLHIV	LEYV	ENGSLANI	---	HRDIKGANIL	151		
CDC15 Saccharomyces cerevisiae	27	LKQVIGRGSYGVV----	AIK	---	NIVKYH	GFI-KSYELYI	LEYC	ANGSLRRL	---	HRDIKAA NIL	153		
CDC7 Schizosaccharomyces pombe	11	LGDCLGKGAFGAV----	AVK	---	NIVKYR	GSY--NDSLCII	LEYC	ENGSLRSI	---	HRDIKGANIL	138		
		##	#		#####	#							
MAP3Kε1 Arabidopsis	152	TTKEGLVKLADFGVATKL---	NTHS	VVGT	PYWMAPE----	TVIELLTC	VPPY	YDLQPMP			226		
MAP3Kε2 Arabidopsis	152	TTKEGLVKLADFGVATKL---	NTHS	VVGT	PYWMAPE----	TVIELLTC	VPPY	YDLQPMP			226		
ZmPHJ40.09G019300.2.p Z.mays PHJ	152	TTKEGLVKLADFGVATKL---	NTHS	VVGT	PYWMAPE----	TVIELLTC	VPPY	YDLQPMP			226		
LOC_Os04g56530.1 O.sativa	152	TTKEGLVKLADFGVATKL---	NTHS	VVGT	PYWMAPE----	TVIELLTC	VPPY	YDLQPMP			226		
Glyma.11G101700.4.p G.maxWm	152	TTKEGLVKLADFGVATKL---	NTHS	VVGT	PYWMAPE----	TVIELLTC	VPPY	YDLQPMP			226		
CDC15 Saccharomyces cerevisiae	154	LSADNTVKLADFGVSTIVNSSALT	LAGT	LN	WMAPE---	TVVEMLT	KNPPY	HNLT	DAN		225		
CDC7 Schizosaccharomyces pombe	139	TTKDGTIKLADFGVATKI---	EDHS	VVGS	PYWMAPE----	TVIELLDG	NPPY	YDL	DPTS		211		

Figure S1. A phylogenetic analysis of the MAP3Kε family. **A:** Phylogenetic relationships of members of the MAP3Kε family. Rooted circular phylogram representation of the consensus tree generated by the neighbor-joining method (500 bootstraps) on the conserved regions of proteins from the MAP3Kε family. Triangle represents NJ bootstrap values > 50. numbers after the abbreviated species names are internal. The aqua represents eudicot, the buff represents monocot, salmon pink represents *Saccharomyces cerevisiae*, the rose represents *Homo sapiens*. The bootstrap represents similarity, rendered with a color gradient. **B:** The conserved regions of MAP3Kε from Arabidopsis thaliana and examples from other representative taxa.

Figure. S2

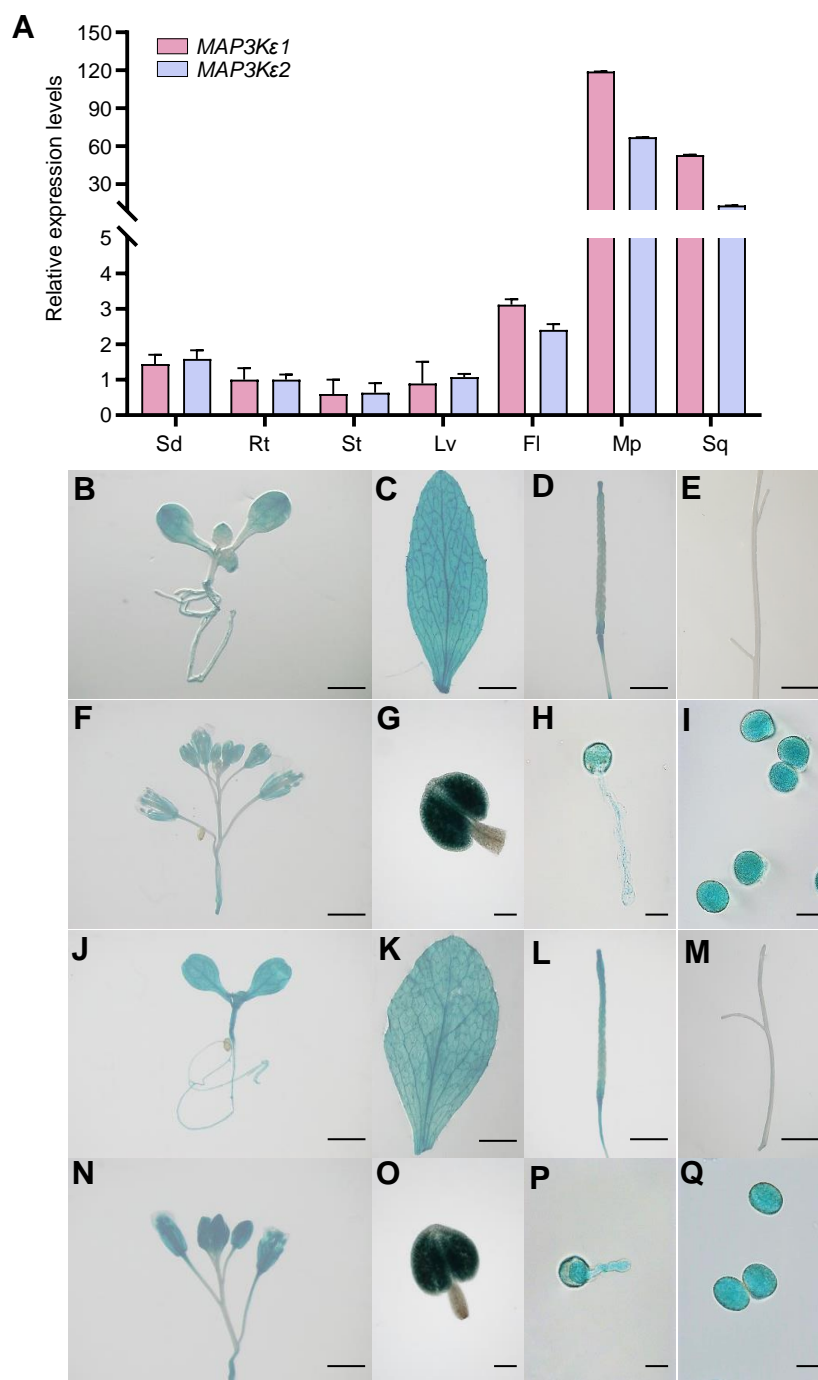


Figure S2. Expression patterns and levels of *MAP3Kε1* and *MAP3Kε2*. **A**: Quantitative real-time PCR showing the expression levels of the *MAP3Kε1* and *MAP3Kε2* in different tissues from Arabidopsis. Sd, seedlings; Rt, root; St, steam; Lv, leaves; Fl, flower; Mp, mature; Sq, silique. Each bar represents the mean \pm SEM of three independent experiments. **B-I**: The *GUS* expression patterns of pMAP3Kε1::GUS transgenic lines. **J-Q**: The *GUS* expression patterns of pMAP3Kε2::GUS transgenic lines. Seedlings (**B**, **J**); leaves (**C**, **K**); siliques (**D**, **L**); stems (**E**, **M**); inflorescences (**F**, **N**); anthers (**G**, **O**); pollen tubes (**H**, **P**); mature pollens (**I**, **Q**). The scale bars : 2 mm for (**B-F**) and (**J-N**), 100 μm for (**G-I**) and (**P-Q**).

Figure. S3

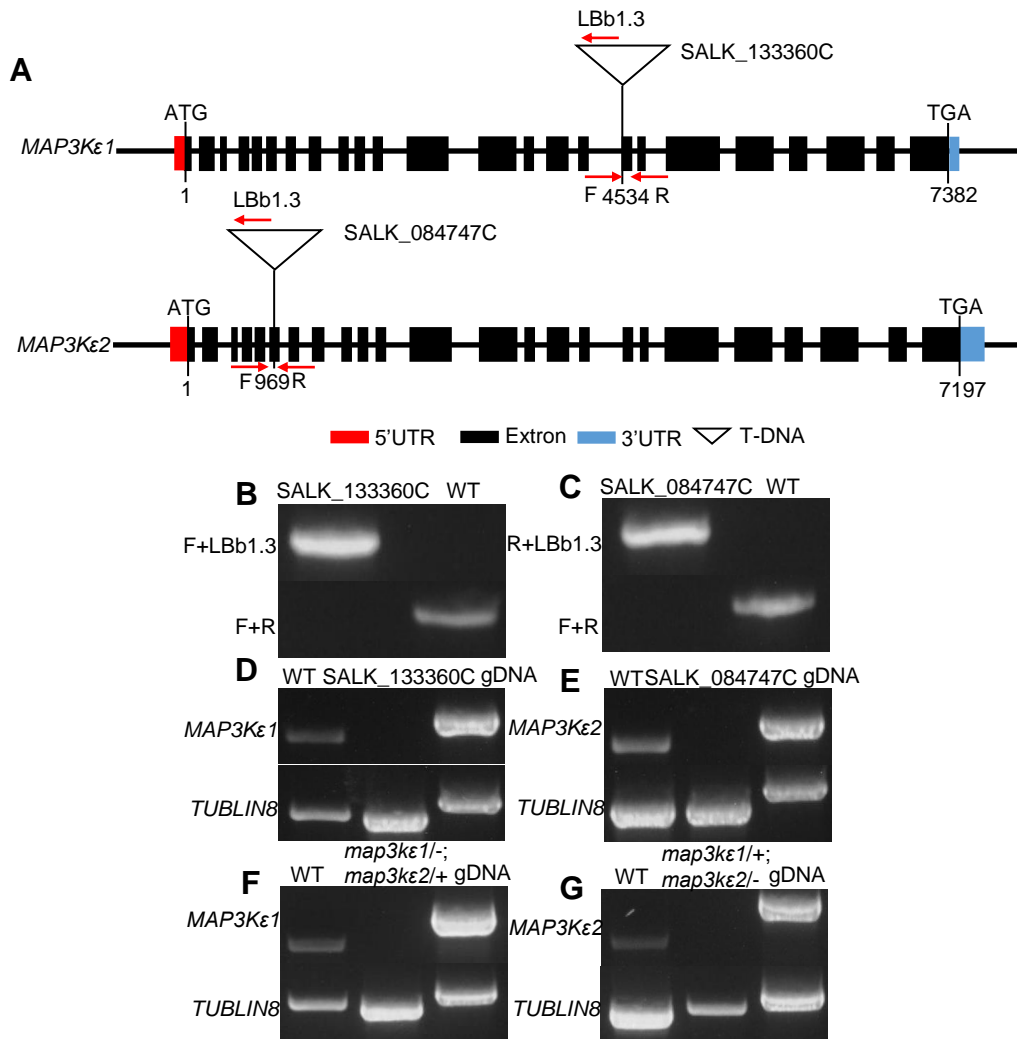


Figure S3. Molecular characterization of *map3kε1*, *map3kε2* mutants. **A:** The schematic diagrams of the *MAP3Kε1*, *MAP3Kε2* structures and T-DNA insertion sites in the mutants. Red arrows represent primers. **B:** Confirmation of the T-DNA insertion in *map3kε1* by PCR. **C:** Confirmation of the T-DNA insertion in *map3kε2* by PCR. **D:** The relative expression levels of *MAP3Kε1* genes in wild-type (WT) and SALK_133360C mutant by RT-PCR. **E:** The relative expression levels of *MAP3Kε2* genes in wild-type (WT) and SALK_084747C mutant by RT-PCR. **F:** The relative expression levels of *MAP3Kε1* genes in wild-type (WT) and *map3kε1*^{-/-}; *map3kε2*^{+/+} mutant by RT-PCR. **G:** The relative expression levels of *MAP3Kε2* genes in wild-type (WT) and *map3kε1*^{+/+}; *map3kε2*^{-/-} mutant by RT-PCR. *TUBLIN8* used for control.

Figure. S4

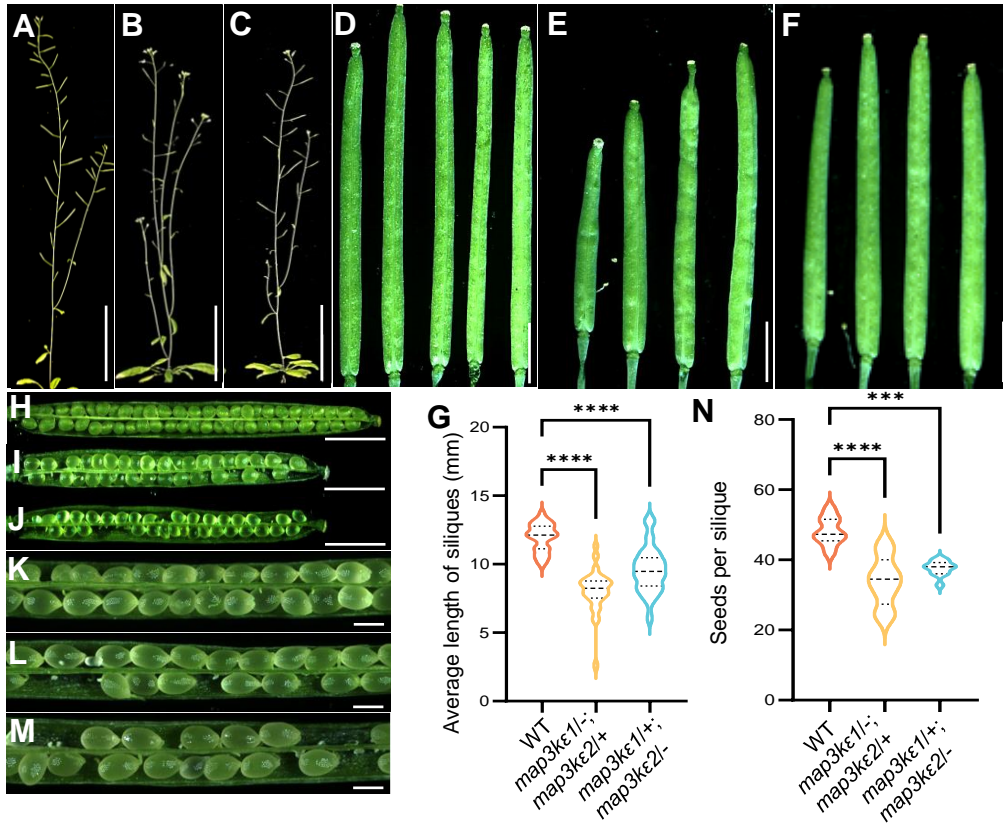


Figure S4. The *map3kε1*^{-/-}; *map3kε2*^{+/+} and *map3kε1*^{+/+}; *map3kε2*^{-/-} mutants reduced fertility. **A-C:** The vegetative and reproductive growth patterns of wild-type (WT) (A) and the *map3kε1*^{-/-}; *map3kε2*^{+/+} (B) and *map3kε1*^{+/+}; *map3kε2*^{-/-} (C) mutant plants. **D-F:** The siliques length from wild-type (WT) plants (D), *map3kε1*^{-/-}; *map3kε2*^{+/+} (E) plants and *map3kε1*^{+/+}; *map3kε2*^{-/-} (F) plants. **G:** Comparison silique length in wild-type (WT) , *map3kε1*^{-/-}; *map3kε2*^{+/+} and *map3kε1*^{+/+}; *map3kε2*^{-/-}. **H-M:** The silique with seed set from wild-type (WT) (H, K) and the *map3kε1*^{-/-}; *map3kε2*^{+/+} (I, L) and *map3kε1*^{+/+}; *map3kε2*^{-/-} (J, M) mutant plants. **N:** Comparison seeds per silique in wild-type (WT) , *map3kε1*^{-/-}; *map3kε2*^{+/+} and *map3kε1*^{+/+}; *map3kε2*^{-/-}, each bar represents the mean \pm SD of three independent experiments. Different letters represent significant difference at *** $p < 0.001$, **** $p < 0.0001$ (one-way ANOVA, Tukey post-test). Bars = 1mm in (K-M), Bars = 2 mm in (D-J), Bar=5 cm in (A-C).

Figure. S5

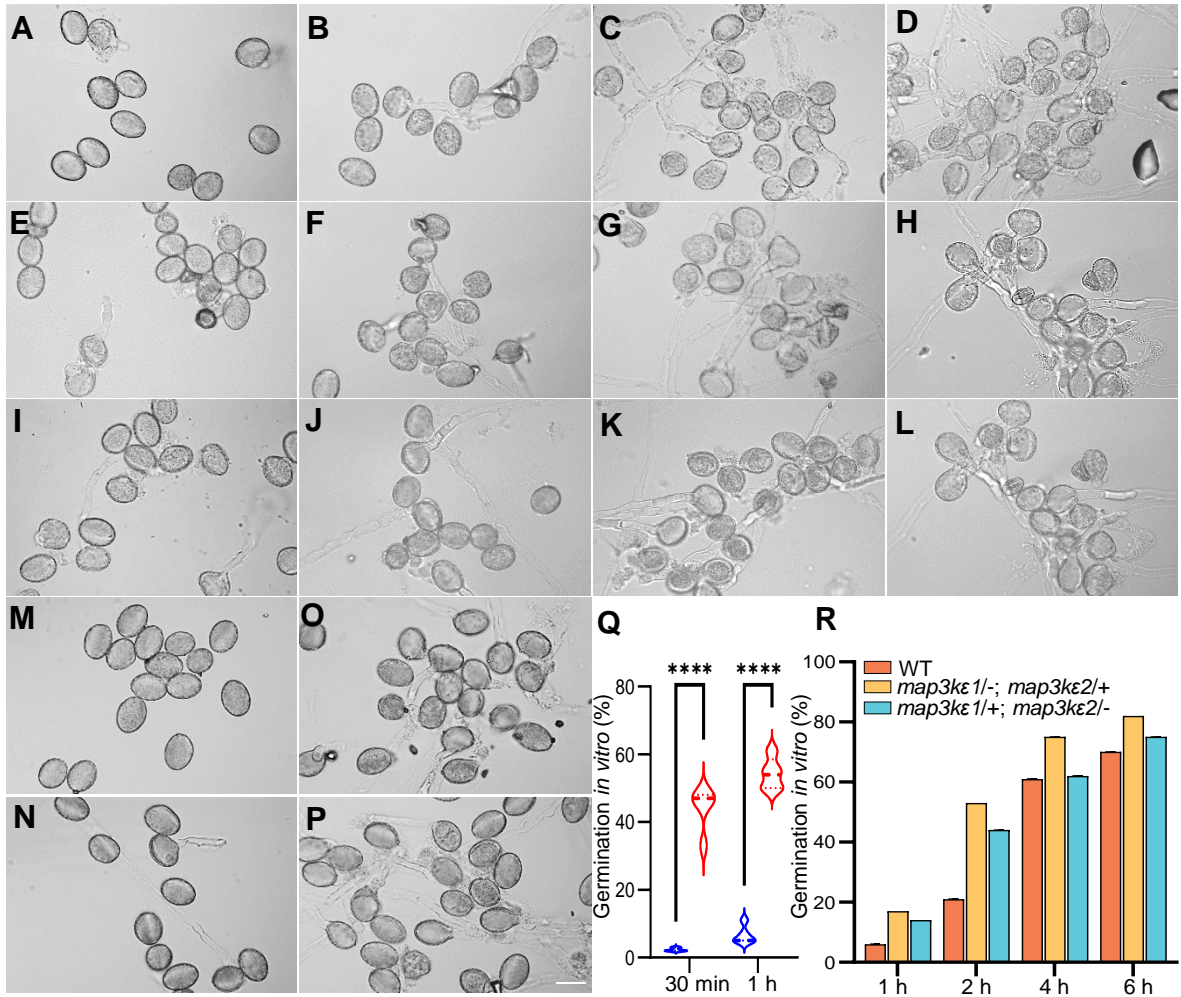


Figure S5. Arabidopsis pollen germinated *in vitro*. **A-D:** The wild-type (WT) pollen grains germinated *in vitro* at 1h, 2h, 4h, 6h respectively. **E-H:** The *map3kε1/-; map3kε2/+* pollen grains germinated *in vitro* at 1h, 2h, 4h, 6h respectively. **I-L:** The *map3kε1/+; map3kε2/-* pollen grains germinated *in vitro* at 1h, 2h, 4h, 6h respectively. **M-N:** The wild-type (WT) pollen grains germinated *in vitro* with the most germination on the medium supplemented with 0 μM MeJA at 30 min and 1h, respectively. **O-P:** The wild-type (WT) pollen grains germinated *in vitro* with the most germination on the medium supplemented with 100 μM MeJA at 30 min and 1h, respectively. **Q:** Germination of wild-type (WT) pollen in standard germination medium supplemented with MeJA. **R:** Comparison pollen germination in wild-type (WT), *map3kε1/-; map3kε2/+* and *map3kε1/+; map3kε2/-*, each bar represents the mean ± SD of three independent experiments. Different letters represent significant difference at **** $p < 0.0001$ (one-way ANOVA, Tukey post-test). Bars = 20 μm (**A-P**).

Figure. S6

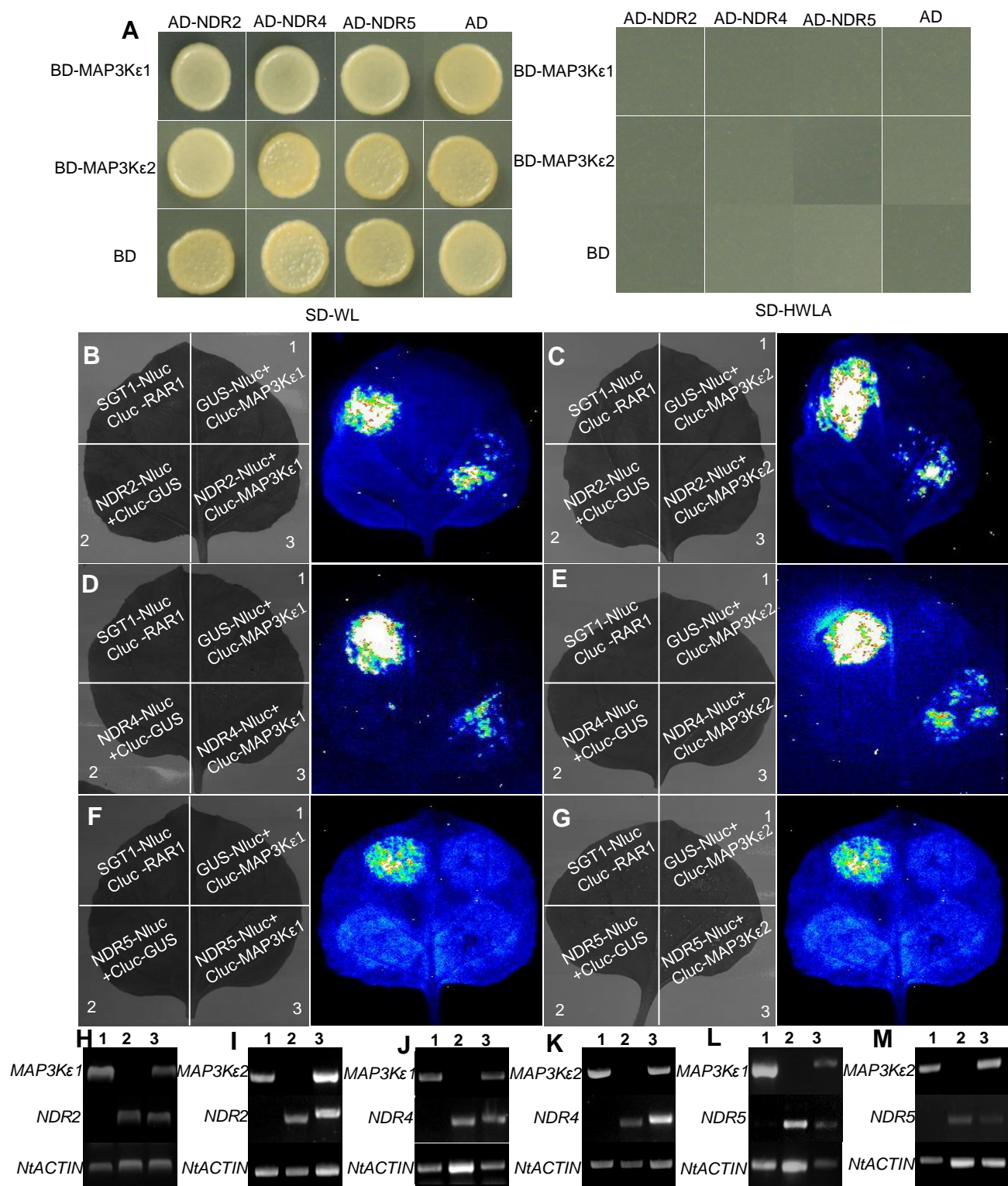


Figure S6. Yeast two-hybrid assay and LCI assay for MAP3K ϵ 1/2 protein and NDR2/4/5. **A:** MAP3K ϵ 1/ 2 can't interact with MOB in the yeast two-hybrid assay. Interaction was determined by yeast growth on the medium lacking His (H), Trp (W) , Leu (L) and Ade(A). **B:** MAP3K ϵ 1 interacts with NDR2 in the luciferase complementation image (LCI) assays. **C:** MAP3K ϵ 2 interacts with NDR2 in the LCI assay. **D:** MAP3K ϵ 1 interacts with NDR4 in the LCI assay. **E:** MAP3K ϵ 2 interacts with NDR4 in the LCI assay. **F:** MAP3K ϵ 1 can't interact with NDR5 in the LCI assay. **G:** MAP3K ϵ 2 can't interact with NDR5 in the LCI assay. **H:** RT-PCR analysis for gene expression in the LCI assay of (**B**). The numbers represent different experimental combinations marked in (**B**). **I:** RT-PCR analysis for gene expression in the LCI assay of (**C**). The numbers represent different experimental combinations marked in (**C**). **J:** RT-PCR analysis for gene expression in the LCI assay of (**D**). The numbers represent different experimental combinations marked in (**D**). **K:** RT-PCR analysis for gene expression in the LCI assay of (**E**). The numbers represent different experimental combinations marked in (**E**). **L:** RT-PCR analysis for gene expression in the LCI assay of (**F**). The numbers represent different experimental combinations marked in (**F**). **M:** RT-PCR analysis for gene expression in the LCI assay of (**G**). The numbers represent different experimental combinations marked in (**G**). The expression of *Nicotiana benthamiana* gene *ACTIN* as a control for the internal standard.

Figure. S7

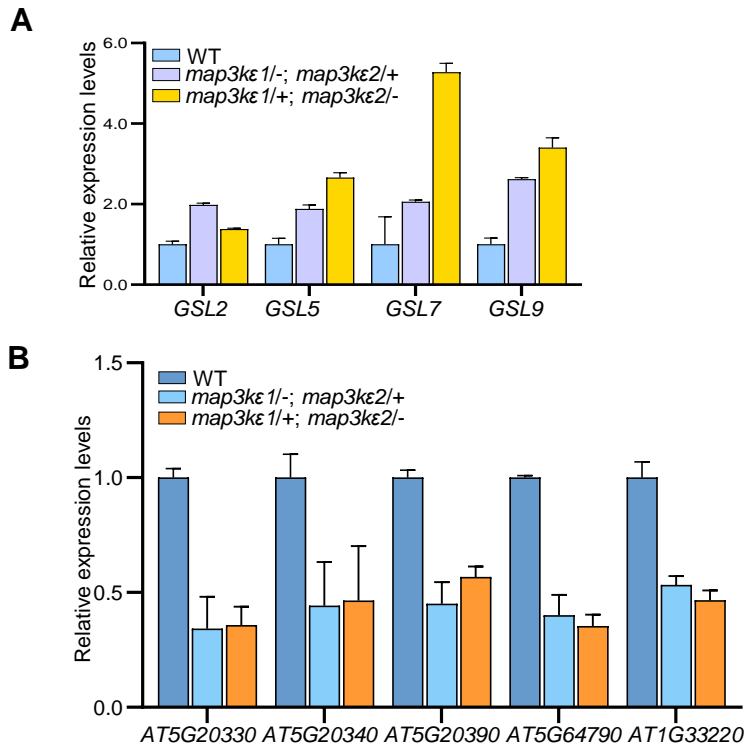


Figure S8. Expression of callose synthase and 1,3-glucanase genes were altered in the *map3kε1/-; map3kε2/+* and *map3kε1/+; map3kε2/-* mutant. **A:** Relative expression of *GSL2*, *GSL5*, *GSL7*, *GSL9* in mature pollen of wild-type (WT) and the *map3kε1/-; map3kε2/+* and *map3kε1/+; map3kε2/-* mutant plants. **B:** Relative expression of *AT5G20330* (β -1,3-glucanase), *AT5G20340* (β -1,3-glucanase), *AT5G20390* (glycosyl hydrolase), *AT5G64790* (o-glycosyl hydrolases), *AT1G33220* (glycosyl hydrolase) in mature pollen of wild type (WT) and the *map3kε1/-; map3kε2/+* and *map3kε1/+; map3kε2/-* mutant plants. *ACTIN2* was used as an internal control.

Table S1. The sequences of the primers used in this study

Uses of the primers	Names of the primers	Sequences of the primers (5'-3')
Mutant identification	LBb1.3	ATTTTGCCGATTTTCGGAAC
	GK-LB	ATATTGACCATCATACTCATTGC
	GK-719G04-LP	TGGATTTCAGGTTCATGGTTGA
	GK-719G04-RP	TGCACTCTTCTTAGGTCGGA
	SALK_06207 0-LP-2	TTTGGAGGAAAAGCTGCACC
	SALK_06207 0-RP-2	ACTGCTGGTCCCAAGTACAG
	SALK_13336 0-F	GCAGGATTTTTGTTGTTGTCC
	SALK_13336 0-R	AATCATTTCTTGGGGTGGATC
	SALK_08474 7-F	TGGAAAACATTGGTCAAGAGG
	SALK_08474 7-R	AGGAATAGGAGGGGTATCATC
Complementati on assays	MAP3K ϵ 1-gus-cz-F((PstI)	AAGCTTGCATGCCTGCAGCCGTGTTACCAACCA AAG
	MAP3K ϵ 1-gus-cz-R((BamHI)	CTGACCACCCGGGGATCCctcttctcgtttcttctt
	MAP3K ϵ 2-GUS-F(PstI):	GCTGCAGCTGTTCTCTGATCACCGGTTG
	MAP3K ϵ 2-GUS-R(BamHI)	CGGATCCctcttctcgtcagcaccaaac
	pMAP3K ϵ 1-1300-F (PstI)	agtgccaaagcttCTGCAGCCGTGTTACCAACCAAAG
	pMAP3K ϵ 1-1300-R (BamHI)	CACCATGGTACCGGATCCctcttctcgtttcttctt

	1300-MAP3K ϵ 1-F(Sac I)	GAGCTGTACAAGGAGCTCATGGCGCGGCAAATGACG
	1300-MAP3K ϵ 1-R(Sac I)	GAGCTGTACAAGGAGCTCATGGCGCGGCAAATGACG
	pMAP3K ϵ 2-1300-F (PstI)	agtgccaaagcttCTGCAG CTGTTCTCTGATCACCGG
	pMAP3K ϵ 2-1300-R (BamHI)	CACCATGGTACCGGATCCctctttctcgtcagcacc
	1300-MAP3K ϵ 2-F(Sac I)	GAGCTGTACAAGGAGCTCATGGCGCGACAGATGACG
	1300-MAP3K ϵ 2-R(Sac I)	TCGGGGAAATTCGAGCTCTCACAAGATGGTGTTGAT
Y2H assays	MAP3K ϵ 1-AD-F(NdeI)	CCAGATTACGCTCATATGATGGCGCGGCAAATGACG
	MAP3K ϵ 1-AD-R(EcoRI)	CCCACCCGGGTGGAATTCTCACAATATTGTGTTGAT
	MAP3K ϵ 2-AD-F(NdeI)	CCAGATTACGCTCATATGATGGCGCGACAGATGACG
	MAP3K ϵ 2-AD-R(EcoRI)	CCCACCCGGGTGGAATTCTCACAAGATGGTGTTGAT
	MOB1A-CDS-BD-F(EcoRI_	CCGGAATTCATGAGTCTCTTTGGGTTAGG
	MOB1A-CDS-BD-R(BamHI)	CGCGGATCCTCAATAAGGTGAAATGATAG
	MOB1B-CDS-BD-F(EcoRI_	CCGGAATTCATGAGTCTATTCTGGGCTTGG
	MOB1B-CDS-BD-R(BamHI)	CGCGGATCCTTAGTAAGGTGCAATGATAG
ColP assays	1390-MAP3K ϵ 1-gfp-F ((BamHI)	GGGATCCATGGCGCGGCAAATGACGTC

	1390- MAP3K ϵ 1- gfp-R (KpnI)	GGGTACCCAATATTGTGTTGATGTGAAG
	1390- MAP3K ϵ 2- gfp-F (Sall)	GGTCGACATGGCGCGACAGATGACGTC
	1390- MAP3K ϵ 2- gfp-R (KpnI)	GGGTACCCAAGATGGTGTGTTGATGTGAAG
LCI assays	MAP3K ϵ 1- cluc-F (KpnI)	GGGTACCATGGCGCGGCAAATGACGTC
	MAP3K ϵ 1- cluc-R (BamHI)	GGGATCCTCACAATATTGTGTTGATGTG
	MAP3K ϵ 2- cluc-F(KpnI)	GGGTACCATGGCGCGACAGATGACGTC
	MAP3K ϵ 2- cluc-R (Sall)	GGTCGACTCACAAGATGGTGTGTTGATGT
	MOB1A-nluc- F(KpnI)	GGGTACCATGAGTCTCTTTGGGTTAGG
	MOB1A-nluc- R(Sall)	GGTCGACATAAGGTGAAATGATAGATTCT
	MOB1B-nluc- F (KpnI)	GGGTACCATGAGTCTATTCTGGGCTTGG
	MOB1B-nluc- R (Sall)	GGTCGACGTAAGGTGCAATGATAGATTC
RT-PCR	NtACTIN-F	CCACACAGGTGTGATGGTTG
	NtACTIN-R	GTGGCTAACACCATCACCAG
	MAP3K ϵ 1- RT-F	CTGAGAGGGATCGTTCTCGC
	MAP4K ϵ 1- RT-R	GGGTGCAAAGCATCTGGTTG
	MAP3K ϵ 2- RT-F	TGGAAAACATTGGTCAAGAGG
	MAP3K ϵ 2- RT-R	AGGAATAGGAGGGGTATCATC
	MOB1A-RT-F	AGAGTGCACCTTCTGGAAGC
	MOB1A-RT- R	ATGAGCTCTTGTAGAGGCGC
	MOB1B-RT-F	TTCCGTCCTAAGAAGAGCGC
	MOB1B-RT- R	AGGAGCGAGTTCTTTCTTGTC

qRT-PCR	ACT2-qPCR-FP	GATGCCCAGAAGTCTTGTTCC
	ACT2-qPCR-RP	TGCTCATACGGTCAGCGATAC
	LOX3-qPCR-FP	GAAATCAATGCTTTGGCTAG
	LOX3-qPCR-RP1	AAGACCGTCGTTGGCGTA
	LOX4-qPCR-FP	TGCTTCACTGCTGGTCAATA
	LOX4-qPCR-RP1	ACCGTCGTTGGCGTATGG
	LOX6-qPCR-FP	TCCTGCTGATCTTGTTCCG
	LOX6-qPCR-RP1	GCTCAAGGTCAGATGTTATG
	AOS-qPCR-FP	CGGTTACGGCTCAATACG
	AOS-qPCR-RP1	TCTCCGGCACAACACTCAT
	AOC1-qPCR-FP	AGAACTTGGGAAATACCG
	AOC1-qPCR-RP1	GTTTGTGAATGGGACGAG
	AOC2-qPCR-FP	TCACTGCCAAGAAGAACC
	AOC2-qPCR-RP1	CTCCGAGACCGAACATTA
	AOC3-qPCR-FP	CTCGTAATCATCAATCTCAC
	AOC3-qPCR-RP1	ACTGCTGGACTGTTTCTAT
	OPR3-qPCR-FP	GGCGTTGGCAGAGTATTA
	OPR3-qPCR-RP1	AAACCTCCCTTAGCGTGA
	CYP94B3-qPCR-FP	ATTAGTGCTCGCCGTGCCA
	CYP94B3-qPCR-RP	TTGCTTTGCTTCTGCTCC
	JOX3-qPCR-FP	ATCTCCATTACCAGCCTTCG

JOX3-qPCR-RP	ATCCACCTGATTCCTCTTTACC
JAR1-qPCR-FP	CTCACTGGTCACCCTGTTCC
JAR1-qPCR-RP	TCCCGTTGATATGTA CTGCTTG
JAZ1-qPCR-FP	CGGGCAAGTGATTGTATT
JAZ1-qPCR-RP	GAGGATTGGATTGGCTCT
JAZ2-qPCR-FP	AATGATGAATCTGTTCCCTT
JAZ2-qPCR-RP	CACCATAACTCGACCACC
JAZ5-qPCR-FP	CCTGATCTCAATGAGCCTAC
JAZ5-qPCR-RP	CGATGATGACCTGCGTTT
JAZ9-qPCR-FP	ACACTGCTGCTACCTCATCA
JAZ9-qPCR-RP	CTCATAAGCCTCTCTTTGCG
JAZ10-qPCR-FP	CGATTTCTCGGACTTGA
JAZ10-qPCR-RP	CTTCCCTCGGAGTAGACG
MYC2-qPCR-FP	GAACGAAGATAAAGTTCTATCA
MYC2-qPCR-RP	CAACCGCTCGTAACGCGTAGA
GSL2- qPCR-FP	ACGAGAAGTTTGCAGAGAACTA
GSL2- qPCR-RP	CTCTGCTGCTTTCCCATAAATC
GSL5- qPCR-FP	CAAGCAATGAAGACTTCTACCG
GSL5- qPCR-RP	ATTACTGAAGAAAGCAATCCGC
GSL7- qPCR-FP	CCTTGGTTTCTTGTCAGTCATG
GSL7- qPCR-RP	CTAATCCTTTGAACACGCTACG

GSL9- qPCR-FP	ATTGCATGAGTTGTGCTGTTAG
GSL9- qPCR-RP	TTGCAGCACATATTCAGTTAGC
AT5G20330-qPCR-FP	ATCGAGTCTTACTTAGCTGACG
AT5G20330-qPCR-RP	TAGGTTGGTTAGAGACTGCATG
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AT5G20340-qPCR-RP	CGTTAAGAACCTCAGTGTTTGG
AT5G20390-qPCR-FP	CTATCCTCGTCAACATCTACCC
AT5G20390-qPCR-RP	GTTGGTGTTGAAGATGGCATAG
AT5G64790-qPCR-FP	AAACGTGACCCAGTATATTCGT
AT5G64790-qPCR-RP	TGTGAATGTTCTTTAACGCTGG
AT1G33220-qPCR-FP	AACCTTTACAAGTCCATAGGCA
AT1G33220-qPCR-RP	GTGACTTCAATATTACGGTGGC