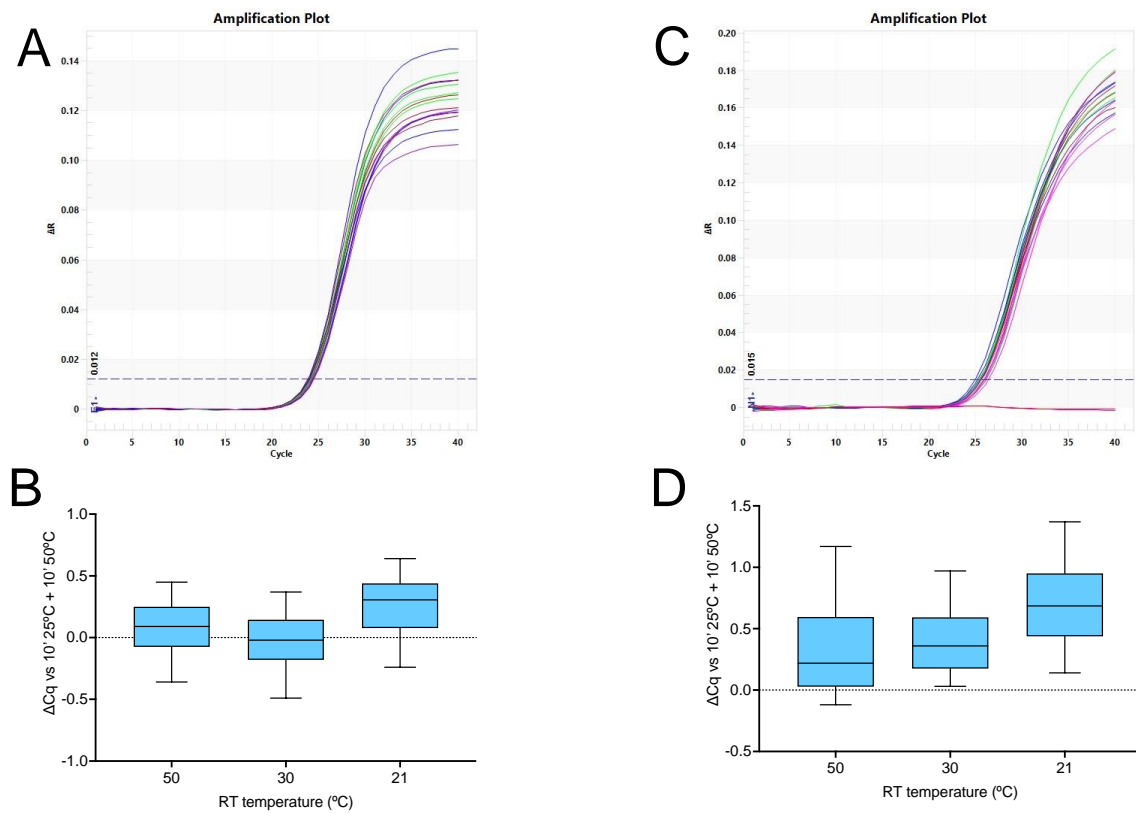
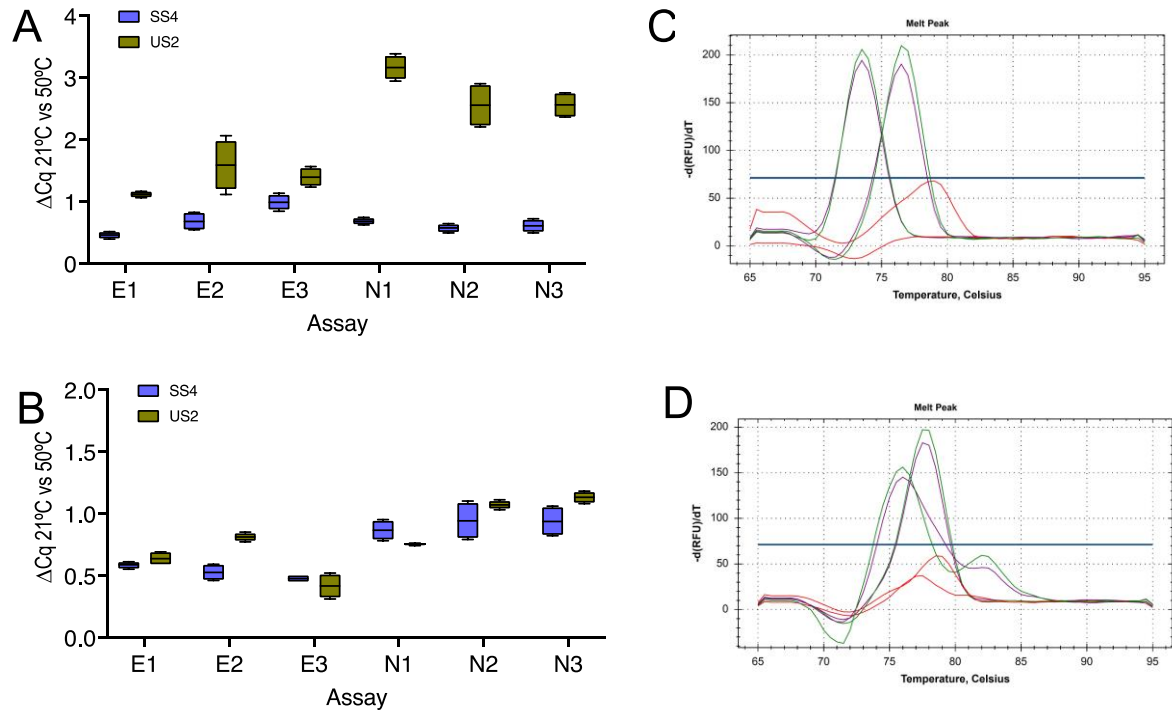


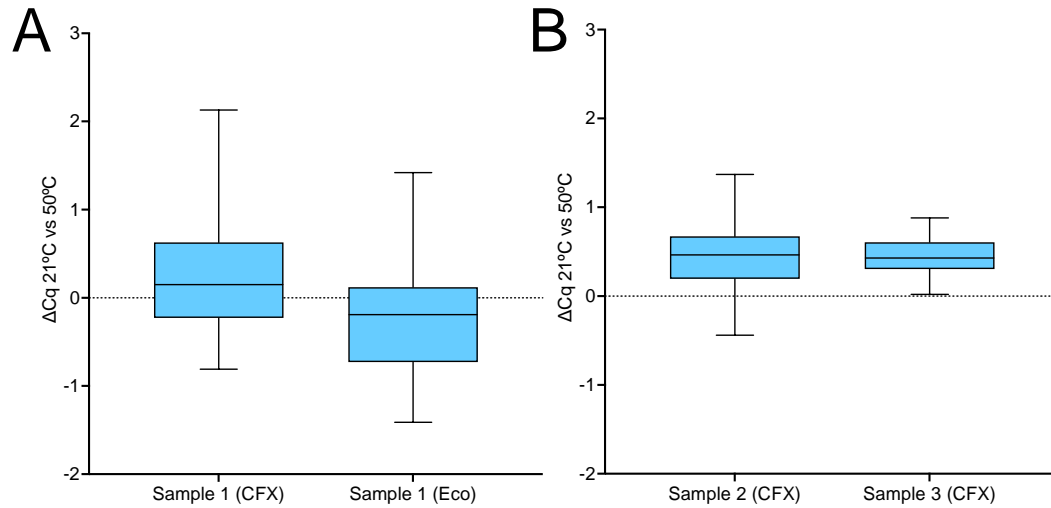
**Figure S1.** Amplification of RNA reverse transcribed by SS4, amplified using primers targeting the viral N-gene (assay N1) and detected using a hydrolysis probe or SYBR Green. **A.** Amplification plots (probe) and Cqs from duplicate qPCR reactions: 10 minutes at 25°C and 50°C (protocol i, blue), 5 minutes at 50°C (protocol ii, green), 30°C (protocol iii, brown) or 21°C (protocol iv, purple). **B.** Box and whiskers plot showing the  $\Delta Cq$  for the three modified conditions relative to the recommended protocol, with the whiskers delineating the minimum and maximum values. **C.** Amplification plots (SYBR Green) and melt curves (inset) from single qPCR reactions: 5 minutes at 50°C (protocol ii, green) or 21°C (protocol iv, purple). The NTC is shown in red. **D.**  $Cq$  values of RNA samples left on the bench for the indicated times relative to control sample kept frozen at -80°C.



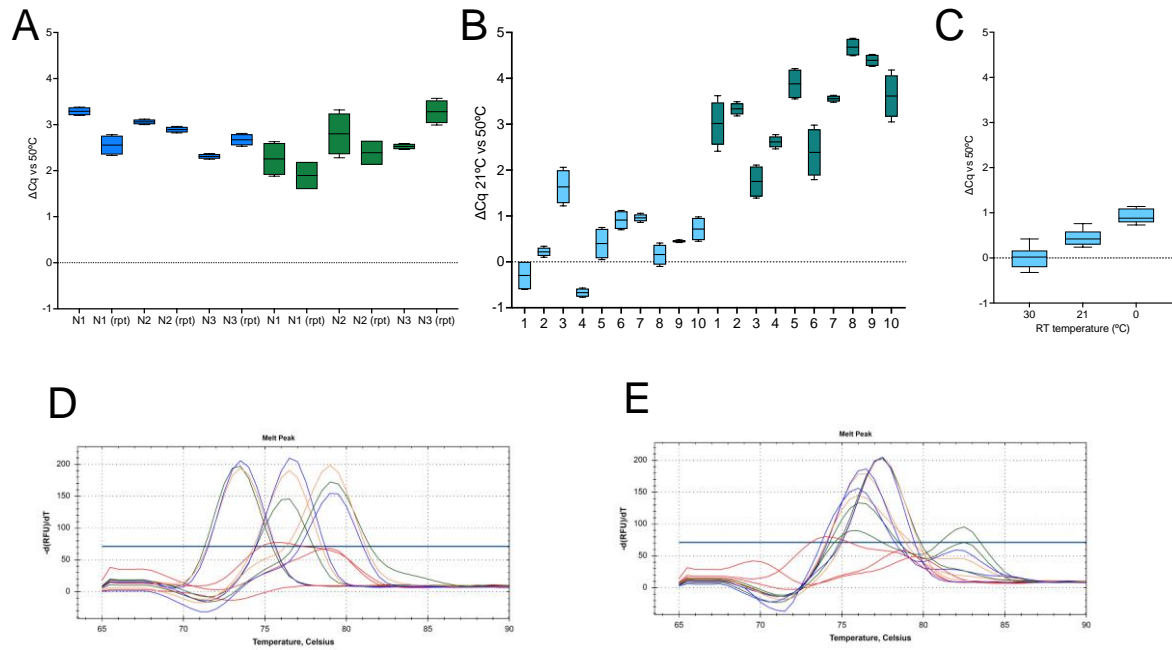
**Figure S2.** Amplification on the PCRMax Eco instrument of RNA reverse transcribed by SS4, amplified using primers targeting the viral E (assay E1) and N-genes (assay N1) and detected using a hydrolysis probe. **A.** Amplification plots (probe) and Cqs from qPCR reactions targeting E1: 10 minutes at 25°C and 50°C (protocol i, blue), 5 minutes at 50°C (protocol ii, green), 30°C (protocol iii, brown) or 21°C (protocol iv, purple). **B.** Box and whiskers plot showing the  $\Delta Cq$  for the three modified conditions relative to the recommended protocol, with the whiskers delineating the minimum and maximum values. **C.** Amplification plots (probe) and Cqs from qPCR reactions targeting N1: 10 minutes at 25°C and 50°C (protocol i, blue), 5 minutes at 50°C (protocol ii, green), 30°C (protocol iii, brown) or 21°C (protocol iv, purple). **D.** Box and whiskers plot showing the  $\Delta Cq$  for the three modified conditions relative to the recommended protocol, with the whiskers delineating the minimum and maximum values.



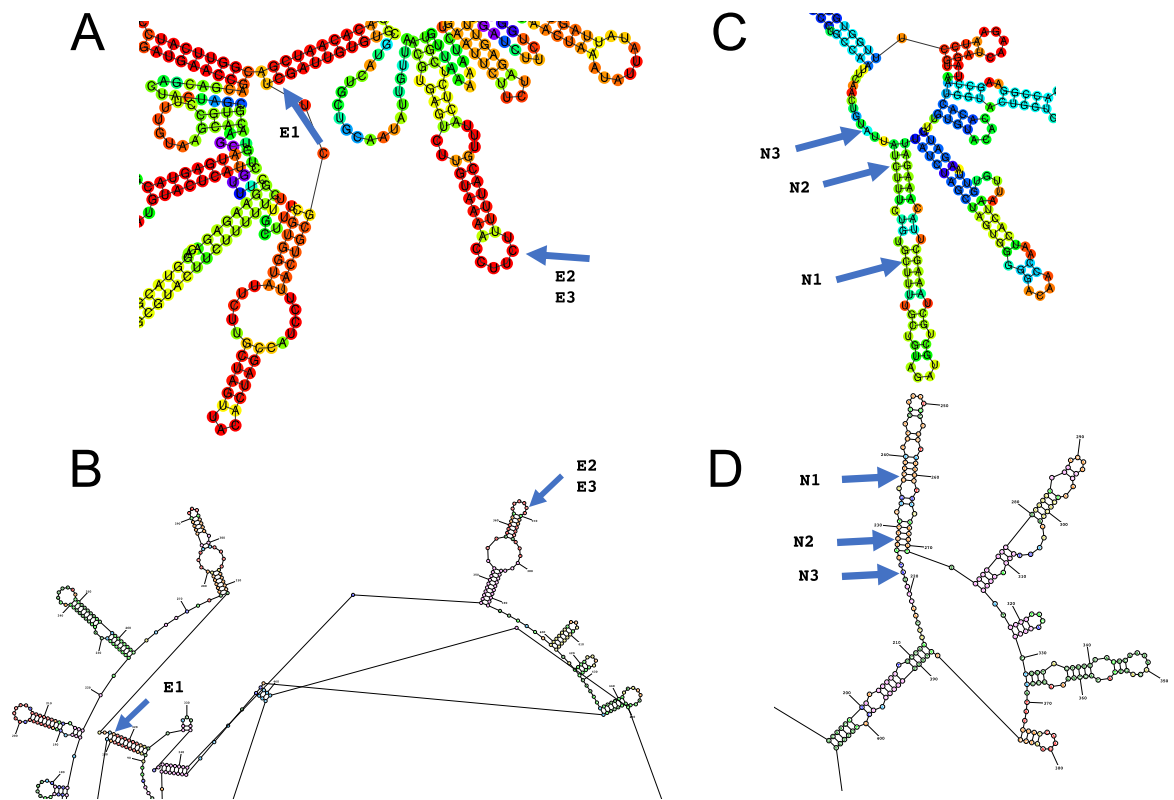
**Figure S3.** Comparison of two further RNA sample reverse transcribed by SS4 and US2 amplified using assays E1 to E3 and N1 to N3. **A.** Box and whiskers plot comparing the  $\Delta Cq$  (21°C vs 50°C) for the six assays with SS4 (blue) and US2 (olive) cDNAs, respectively, carried out with the first RNA sample. **B.** The same plot carried out with the results recorded for the second RNA sample. **C.** Melt curves for E2 and E3 PCR amplicons recorded from RT reactions carried out at 50°C (brown) or 21°C (green). The red melt profiles were obtained from the NTC controls. **D.** Melt curves for N2 and N3 PCR amplicons recorded from RT reactions carried out at 50°C (brown) or 21°C (green). The red melt profiles were obtained from the NTC controls.



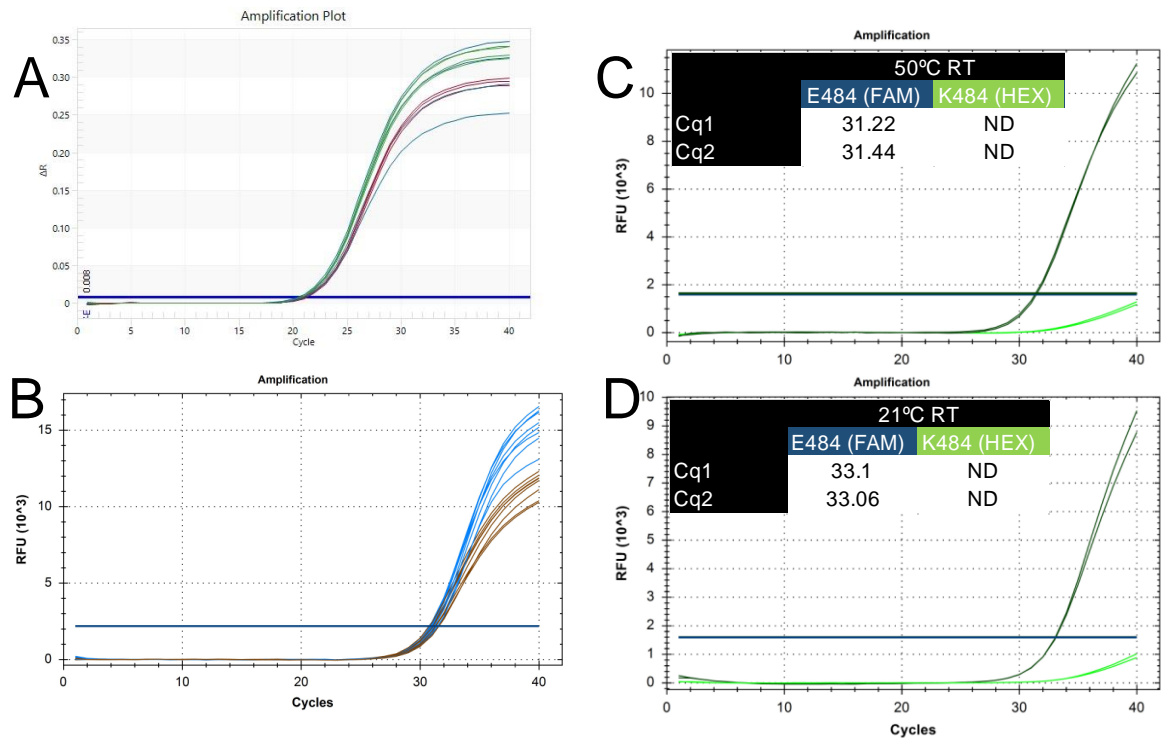
**Figure S4.** Reproducibility of low temperature RT relative to the standard 5-minute 50°C protocol and amplified with the E1 assay. **A.** Aliquots of the same sample were transcribed by SS4 at 50°C or 21°C for 5 minutes and amplified on CFX or Eco instruments. **B.** RT reactions were repeated with a second and third RNA sample transcribed by SS4 at 50°C or 21°C for 5 minutes and amplified on the CFX. All box and whiskers plots show the  $\Delta Cq$  for the reactions carried out at 21°C relative to 50°C, with the whiskers delineating the minimum and maximum  $\Delta Cq$  values. The scales of the y-axes were adjusted to a common scale to simplify the comparison between the runs and instruments.



**Figure S5.** RT efficiency at 0°C. **A.** ΔCqs against control RNA transcribed at 50°C of RNA transcribed in duplicate on ice by SS4 (blue) or US2 (green) and amplified with assays N1, N2 and N3. **B.** ΔCqs against control RNA transcribed at 50°C of RNA transcribed in ten independent RT reactions for 5 minutes at 21°C (blue) or ice (green) with US2 and amplified with assay E1 on the CFX. **C.** ΔCqs against control RNA transcribed at 50°C of RNA transcribed for 5 minutes at 30°C, 21°C or ice with SS4 and amplified with assay E1 on the CFX. **D.** Melt curves of PCR amplicons for assays E1, E2 and E3 obtained at 30°C (orange), 21°C (blue) and 4°C (green). The red melt curve is from the NTC. **E.** Melt curves of PCR amplicons for assays N1, N2 and N3 obtained at 30°C (orange), 21°C (blue) and 4°C (green). The red melt curve is from the NTC. All box and whiskers plots show the ΔCq for the reactions carried out at 21°C relative to 50°C, with the whiskers delineating the minimum and maximum ΔCq values. The scales of the y-axes were adjusted to a common scale to simplify the comparison between the runs and instruments.



**Figure S6.** Mapping of reverse primers to SARS-CoV-2 secondary and tertiary genomic RNA structure. **A.** The locations of primers initiating reverse transcription of assays E1, E2 and E3 are indicated by arrows on one of the predicted structures [43]. The E1 primer maps into a stem structure, whereas the 3'-ends of both E2 and E3 primers terminate in a small loop, **B.** The second model [44] results in a similar prediction for the three primers. **C.** The locations of primers initiating reverse transcription of assays N1, N2 and N3 are indicated by arrows on one of the predicted structures [43]. N1 and N2 primers map into a stem structure, whereas the 3'-end of the N3 primer terminates in a small loop, **D.** The second model [44] results in a similar prediction for the three primers.



**Figure S7.** RT reactions carried out with high and low copy number targets. **A.** Two-step assay (SS4/Bioline SensiFast) with E1 and a high viral load sample. RT reactions were carried out at 50°C (blue), 30°C (green) and 21°C (brown). **B.** One step gradient RT-qPCR (PCRBio 1-step) with E1 (blue) and N1 (brown) and a low viral load. **C.** One-step RT-qPCR (PS3) detection of the WT E484 sequence in variant B.1.1.7 with a 50°C RT reaction. **D.** One-step RT-qPCR (PS3) detection of the WT E484 sequence in variant B.1.1.7 with a 21°C RT reaction.