

Table S1. Bacterial strains and plasmids used in this study.

Strain or plasmid	Genotype or relevant phenotype	Source
<i>P. mirabilis</i>		
wt	Wild-type N2; Tet ^r	Clinical isolate
<i>qseE</i>	wt derivative; <i>qseE</i> -knockout mutant; Km ^r	This study
<i>qseF</i>	wt derivative; <i>qseF</i> -knockout mutant; Km ^r	This study
<i>qseG</i>	wt derivative; <i>qseG</i> -knockout mutant; Km ^r	This study
<i>qseEGF</i>	wt derivative; <i>qseEGF</i> -knockout mutant; Km ^r	This study
<i>qseFc</i>	<i>qseF</i> mutant containing pGEM- <i>qseF</i> ; <i>qseF</i> -complemented strain; Amp ^r	This study
<i>glmY</i>	wt derivative; <i>glmY</i> -knockout mutant; Km ^r	This study
<i>glmYc</i>	<i>glmY</i> mutant containing pGEM- <i>glmY</i> ; <i>glmY</i> -complemented strain; Amp ^r	This study
<i>E. coli</i>		
DH5α	<i>fhuA2 lac(del)U169 phoA glnV44 Φ80' lacZ(del)M15 gyrA96 recA1 relA1 endA1 thi-1 hsdR17</i>	Invitrogen
S17-1 λ <i>pir</i>	λ <i>pir</i> lysogen of S17-1 [<i>thi pro hsdR⁻ hsdM⁺ recA</i> RP4 2-Tc::Mu-Km::Tn7 (Tp ^r Sm ^r)]; permissive host able to transfer suicide plasmids requiring the Pir protein by conjugation to recipient cells	Biomedical
Plasmids		
pGEM-T Easy	High-copy TA cloning vector; Amp ^r	Promega
pUT-Km1	Suicide plasmid requiring the Pir protein for replication and containing a mini-Tn5 cassette containing Km ^r gene	
pACYC184	Low-copy cloning vector, P15A replicon; Cm ^r Tet ^r	New England Biolabs
pBAD33	Cloning expression vector, P15A replicon; Cm ^r	ATCC

Tet, tetracycline; Km, kanamycin; Cm, chloramphenicol; Amp, ampicillin; Tp, trimethoprim; Sm, streptomycin.

Table S2. Primers used in this study.

Primers	Sequence (5' to 3')	Description
qseF-upF	CAGGTCACAAAAACAGGTGTC	For <i>qseF</i> knockout. Paired with “XbaI-qseF-upR”.
XbaI-qseF-upR	<u>TCTAGAC</u> AGCCATGCACATTCTCC	For <i>qseF</i> knockout. Paired with “qseF-dnR”.
XbaI-qseF-dnF	<u>TCTAGAC</u> GTTCATCGAGCAATGTGT CG	
qseF-dnR	GTTCTCGCTCATCAAAGAC	For GlmY knockout. Paired with “XbaI-glmY-upR”.
glmY-upF	TTTACGTGATGAGTTTGCCC	
XbaI-glmY-upR	<u>TCTAGAC</u> TTGGCTTATGTGGTATG	For GlmY knockout. Paired with “glmY-dnR”.
glmY-dnF	CATTCTAGAGCCTGTTGC	
glmY-dnR	TCATGGCGTTGCGATTCTA	For <i>qseF</i> complementation. Paired with “qseF-comR”.
qseF-comF	AGCGCAACGGTTAAACCTAC	
qseF-comR	CAGTTTAACTGAGCAAGTGCGAG	For GlmY complementation. Paired with “glmY-comR”.
glmY-comF	CCTTGGATGCGCTTATTCCGT	
glmY-comR	AAGGTGGTGCCTCACTCCA	For <i>flhDC</i> real-time RT-PCR. Paired with “flhDCrt-R”.
flhDCrt-F	CACGAGCATGGACATTAG	
flhDCrt-R	GCAGGATTGGCGGAAAGTT	For <i>glmY</i> real-time RT-PCR. Paired with “glmYrt-R”.
glmYrt-F	GGAACATACCACATAAGCCAAG	
glmYrt-R	CGCTCGTCAATATTCATCTCTT	For <i>rcsB</i> real-time RT-PCR. Paired with “rcsBrt-R”.
rcsBrt-F	GCAGATGCTCTTATCACC	
rcsBrt-R	CAGGCGCACCTTGTTTTA	For <i>cheA</i> real-time RT-PCR. Paired with “cheArt-R”.
cheArt-F	AGATGCGAATGCGTGTACCTT	
cheArt-R	AGGCGATATCATCGGGCTTA	Internal control for real-time RT-PCR. Paired with “gyrB-
gyrBrt-F	GACCCGTACGCTAAACAAC	

		rtR”.
gyrBrT-R flhDC-reF	AGAAATAACCGCAATCAGG GGGTAGATTCGCTTATTAATT CTC	For <i>flhDC</i> reporter assay. Paired with “flhDC-reR”.
flhDC-reR glmY-reF	CTCTTTACATCCCGTCCGAT <u>GCATGCCTTGGATGCGCTTATTCCG</u>	For <i>glmY</i> reporter assay. Paired with “glmY-reR”.
glmY-reR	<u>CTGCAGTTAATATAATGCACAAGGC</u> GT	
BglIII-xylE-F	<u>AGATCTATGAACAAAGGTGTAATGC</u> G	For <i>cheA</i> translational reporter assay. Paired with “xylE-R”.
xylE-R SacI-cheA-5’UTR- F	AAGTCGTACCGGACCATCAG <u>GAGCTCTATCAGGTCATACCGATGA</u> T	For <i>cheA</i> translational reporter assay. Paired with “BglIII-cheA- 5’UTR-R”.
BglIII-cheA- 5’UTR-R	<u>AGATCTTGT</u> TTTGATAAAACTCGG	
cheA-5’UTR 253bp	CACCAGCAATAAAAGAGGCATCAG	For <i>cheA</i> translational reporter assay. Paired with “xylE-orf-R”.
cheA-5’UTR 138bp	GCTTCCCGAATCAAAAAAGGA	For <i>cheA</i> translational reporter assay. Paired with “xylE-orf-R”.
cheA-5’UTR 61bp	CGACATAACAGGAAACAACCTCC	For <i>cheA</i> translational reporter assay. Paired with “xylE-orf-R”.
xylE-orf-R cheA-mut-F	AAGTCGTACCGGACCATCAG CAGCTAATATTTGTCGATTGTG	For site-direct mutagenesis. Paired with “cheA-mut-R”.
cheA-mut-R glmY-mut-F	TATTCAGGAAACAACCTCTTAG AATACAGCGTCCACAATATGGGTGG GAA	For site-direct mutagenesis. Paired with “glmY-mut-R”.
glmY-mut-R qseF-SP-RT	AAAGTTGAATGAGCAACTGA GCTGTATTTCTGCAAATAGC	<i>qseF</i> gene-specific RT primer
PM-qse-us-F	GTTCCCTCGCTCATCAAAGAC	For <i>qseEGF</i> knockout. Paired with “PM-qse-us-R”.
PM-qse-us-R PM-qse-ds-F	TCTAGCGTCATCGAGCAATGTGTCG <u>TCTAGAAAGAGAATGAAGGTGGTG</u> CC	For <i>qseEGF</i> knockout. Paired with “PM-qse-ds-R”.
PM-qse-ds-R	TTTACGTGATGAGTTTGCCC	

PM-qseG-us-F	GCTCAACGTGGGGATACG	For <i>qseG</i> knockout. Paired with “PM-qseG-us-R”.
PM-qseG-us-R	<u>TCTAGAC</u> AGTTAAACCTGAACCAC C	
PM-qseG-ds-F	<u>TCTAGATT</u> GATGCAGGGG	For <i>qseG</i> knockout. Paired with “PM-qseG-ds-R”.
PM-qseG-ds-R	GCCACGTGAATTACGCTC	
PM-qseE-us-F	GACCCAGTCACGTAGCGATAGCGG AGTGTAAGATGGCCGCTCAACAAT C	For <i>qseE</i> knockout. Paired with “PM-qseE-us-R”.
PM-qseE-us-R	TTGAGACACAACGTGTCGGGGAAA AATACGCCAC	
PM-qseE-ds-F	GATGAGTTTTTCTAATCGTATTGAA CTACCGCTAC	For <i>qseE</i> knockout. Paired with “PM-qseE-ds-R”.
PM-qseE-ds-R	CTGCTCTGATGCCGCATAGTTAAGC CAGTAAATAGCATCCACAGAAGC	

The recognition sequences of endonucleases are underlined.