

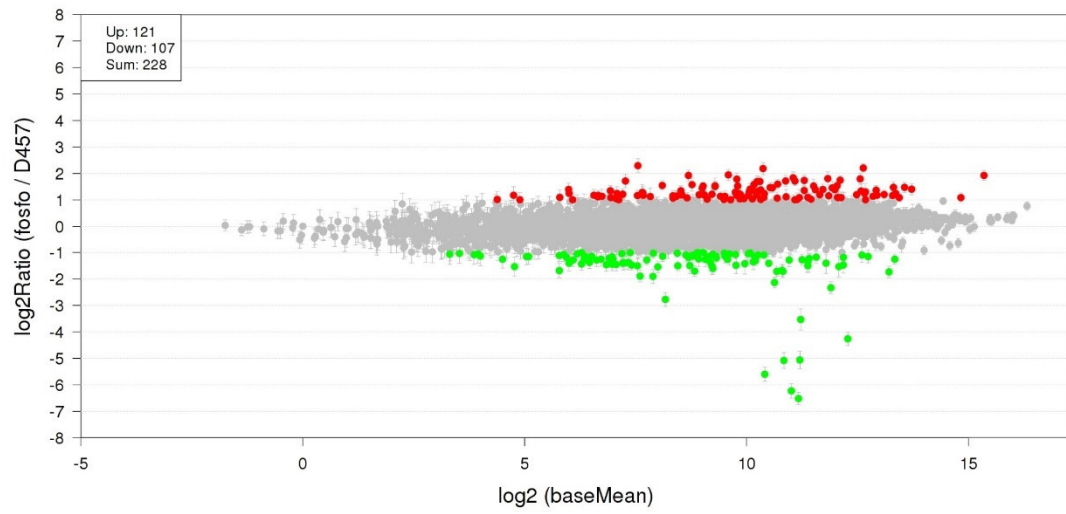
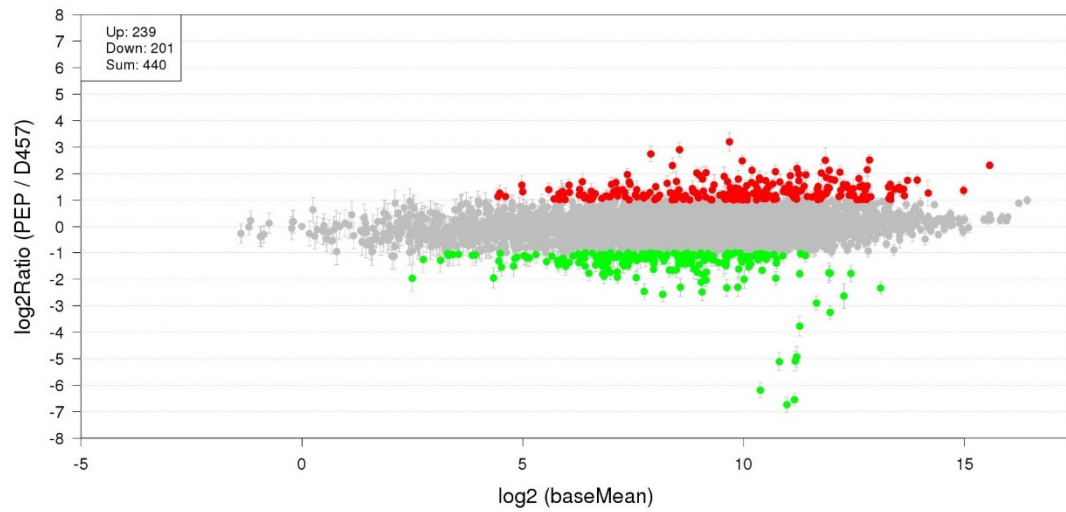
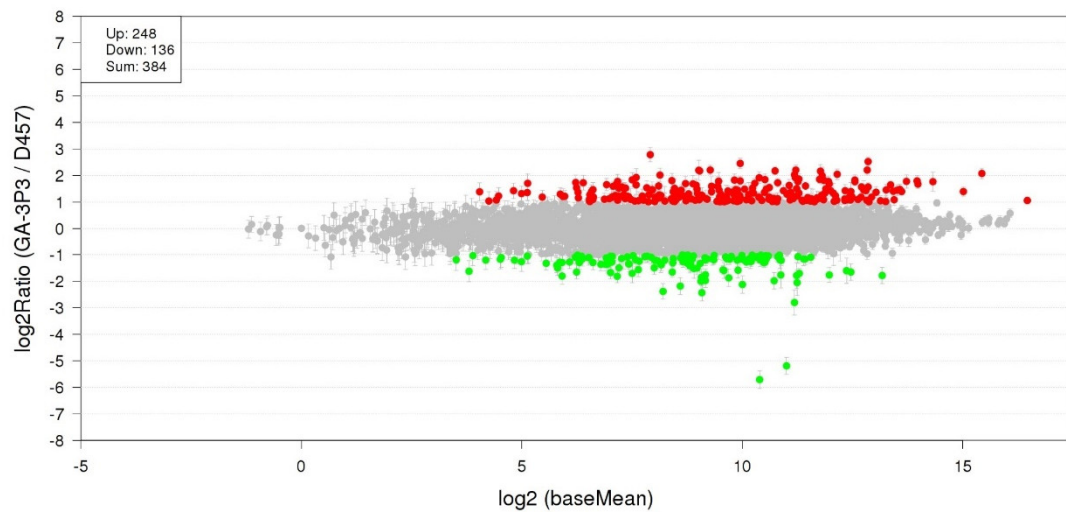
A**fosfo vs D457 (FDR ≤ 0.05 and $|\log\text{Ratio}| \geq 1$)****B****PEP vs D457 (FDR ≤ 0.05 and $|\log\text{Ratio}| \geq 1$)****C****GA-3P3 vs D457 (FDR ≤ 0.05 and $|\log\text{Ratio}| \geq 1$)**

Figure S1. Differential expression between sample groups and their statistical significance. A) Fosfomycin; B) PEP and C) GA-3P. Fold changes were calculated as \log_2 of RPKM after the treatment in comparison with the RPKM of the wild-type D457 strain without treatment. Fold changes <-1 or > 1 were considered significant.

Table S1. List of putative *S. maltophilia* D457 T4SS effectors* identified by BLAST search using XVIPCD domains from *X. citri* effectors.

<i>X. citri</i> XVIPCD	<i>S. maltophilia</i> D457											
Entry	Entry/E-value											
XAC4264 (140-279)	SMD_RS10 655 6,0E-13	SMD_RS13 930 2,0E-09	SMD_RS13 920 5,0E-08									
XAC3634 (189-306)	SMD_RS01 005 3,0E-16	SMD_RS13 735 5,0E-16	SMD_RS01 500 2,0E-15	SMD_RS03 560 3,0E-14	SMD_RS13 725 9,0E-11	SMD_RS10 655 5,0E-10	SMD_RS02 370 4,0E-08	SMD_RS13 930 1,0E-04	SMD_RS00 480 2,0E-04	SMD_RS02 350 4,0E-04	SMD_RS13 920 2,0E-03	SMD_RS12 385 5,6E+00
XAC3266 (735-861)	SMD_RS14 045 5,7E+00											
XAC2885 (271-395)	SMD_RS13 735 2,0E-12	SMD_RS01 005 5,0E-08	SMD_RS13 920 1,0E-06	SMD_RS13 930 3,0E-06	SMD_RS01 500 3,0E-05							
XAC2609 (315-431)	SMD_RS01 500 7,0E-27	SMD_RS03 560 2,0E-21	SMD_RS13 735 1,0E-20	SMD_RS01 005 2,0E-16	SMD_RS13 725 2,0E-11	SMD_RS00 480 2,0E-06	SMD_RS02 350 3,4E-02					
XAC1918 (477-606)	SMD_RS13 920 5,0E-25	SMD_RS13 930 5,0E-25	SMD_RS10 655 3,0E-19	SMD_RS02 370 2,0E-18	SMD_RS02 340 4,0E-16	SMD_RS13 735 3,0E-07	SMD_RS00 480 7,0E-07	SMD_RS02 350 1,0E-06	SMD_RS01 500 8,0E-06	SMD_RS03 560 9,0E-03		
XAC1165 (1-112)	SMD_RS02 370 5,0E-28	SMD_RS13 930 8,0E-26	SMD_RS10 655 1,0E-24	SMD_RS13 920 5,0E-24	SMD_RS02 340 4,0E-18	SMD_RS02 350 3,0E-13	SMD_RS13 725 3,0E-11	SMD_RS00 480 1,0E-10				

XAC0574 (317-440)	SMD_RS13 725 3,0E-44	SMD_RS10 655 5,0E-15	SMD_RS13 930 1,0E-14	SMD_RS13 920 1,0E-12	SMD_RS13 735 1,0E-12	SMD_RS01 005 5,0E-12	SMD_RS02 370 1,0E-11				
XAC0466 (488-584)	SMD_RS14 175 2,0E-18	SMD_RS14 175 9,0E-14									
XAC0323 (16-136)	SMD_RS02 370 1,0E-25	SMD_RS10 655 1,0E-22	SMD_RS02 340 2,0E-22	SMD_RS13 930 1,0E-21	SMD_RS13 920 1,0E-20	SMD_RS00 480 1,0E-11					
XAC0151 (120-254)	SMD_RS02 370 3,0E-36	SMD_RS10 655 8,0E-32	SMD_RS13 930 1,0E-29	SMD_RS13 920 2,0E-29	SMD_RS02 340 9,0E-29	SMD_RS02 350 4,0E-18	SMD_RS00 480 2,0E-15	SMD_RS13 735 3,0E-11	SMD_RS13 725 2,0E-06	SMD_RS03 560 2,0E-05	
XAC0096 (506-646)	SMD_RS10 655 4,0E-21	SMD_RS13 930 5,0E-20	SMD_RS13 920 1,0E-16	SMD_RS02 370 2,0E-16	SMD_RS02 340 6,0E-15	SMD_RS13 725 1,0E-09	SMD_RS13 735 5,0E-09	SMD_RS02 350 1,0E-06	SMD_RS01 500 6,0E-05	SMD_RS03 560 7,0E-03	

*Effectors are color coded according to their accession number and organized according to the e-value.

Table S2 Primers used in this study

Primer	Sequence (5' - 3')	Description
27	TGCCAGCGACAGTGCAAAGGGTC	Amplification of <i>smeT</i> to test DNA contamination
48	CCGTGTTCATGGAAGCAGGC	
<i>gyrA_F</i>	CGCAAGAGXTACCTCGATTA	To amplify the housekeeping gene <i>gyrA</i> by real-time PCR
<i>gyrA_R</i>	GGTGGTACTTACCGATGACG	
<i>virB2_F</i>	AATCTGGTAGCCAGCAAAGA	To amplify <i>virB2</i> by real-time PCR
<i>virB2_R</i>	ACGCTGAAGACTCTGCTGAT	
<i>virB8_F</i>	CCAGCTGGTTGTCAAGGTAG	To amplify <i>virB8</i> by real-time PCR
<i>VirB8_R</i>	GTAACCCTCAGAACCCGTTT	
<i>virB9_F</i>	CAGTACAAGATCGCCTTCGT	To amplify <i>virB9</i> by real-time PCR
<i>virB9_R</i>	GAAACGACCATCGTCATAGG	
<i>virB10_F</i>	GTCCAAAATGTCTCCAATGC	To amplify <i>virB10</i> by real-time PCR
<i>virB10_R</i>	ATAGATCTTGGAGCCCTTCG	
<i>virD4_F</i>	TCAAGCAGGAGAACTTCGAC	To amplify <i>virD4</i> by real-time PCR
<i>virD4_R</i>	TACAGGGTAAAGGCCATGAA	
<i>fruK_F</i>	AACATCAAGCTGGTGAAGA	To amplify <i>fruK</i> by real-time PCR
<i>fruK_R</i>	GCCACTGGTATCCAGCAGTA	
<i>fruA_F</i>	TCGGCCTGTTGATGATGTAT	To amplify <i>fruA</i> by real-time PCR
<i>fruA_R</i>	GGAAATCAATGCGGTAGAG	
<i>rpfN_F</i>	TGATCGAGGATCATTTCCTG	To amplify <i>rpfN</i> by real-time PCR
<i>rpfN_R</i>	CCATAGCTCAGCTCCATCAT	
<i>dld_F</i>	GTTCCACCAGGACTACATCG	To amplify <i>dld</i> by real-time PCR
<i>dld_R</i>	CCTTTTTCCTTCGAGGTCTG	

Primer	Sequence (5' - 3')	Description
<i>lctD_F</i>	GACCTGAGCCTGGAAACC	To amplify <i>lctD</i> by real-time PCR
<i>lctD_R</i>	ACGTACAGCTGGAACCACAT	
<i>lldR_F</i>	GAATACCGTTTTGACGTGCT	To amplify <i>lldR</i> by real-time PCR
<i>lldR_R</i>	CGTCAGCCTCTGCTTCAC	
<i>lctP_F</i>	GTTCGGATACCTCGTCCAAT	To amplify <i>lctP</i> by real-time PCR
<i>lctP_R</i>	GTGAAAATCAGGCTGTGCTT	
<i>smeY_F</i>	CATTGGTGACCGAAGGTG	To amplify <i>smeY</i> by real-time PCR
<i>smeY_R</i>	TTGATACCGGAGAACAGCAG	
<i>smeZ_F</i>	TGTACGAGAGCTGGTCGATT	To amplify <i>smeZ</i> by real-time PCR
<i>smeZ_R</i>	GAACTCGACAATGAGGATCG	
<i>oprO_F</i>	GAACAAGGATGACACCGAG	To amplify <i>oprO</i> by real-time PCR
<i>oprO_R</i>	TCCAGCGAGAAATACTGCTT	