

Figure S1. SIBRII T825A affected the cell size of the stems. (A) Cross section (top) and longitudinal section (bottom) of stems from both the transgenic and *cu3^{-abs1}* plants. Scale bars represent 200 μm . (B) The cross-section area of the stem per plant. (C) The cell length of the stem per plant. The data for (B, C) are presented as the mean \pm SD ($n=5$). The different letters indicate significant differences (one-way ANOVA, Tukey's test, $p < 0.05$).

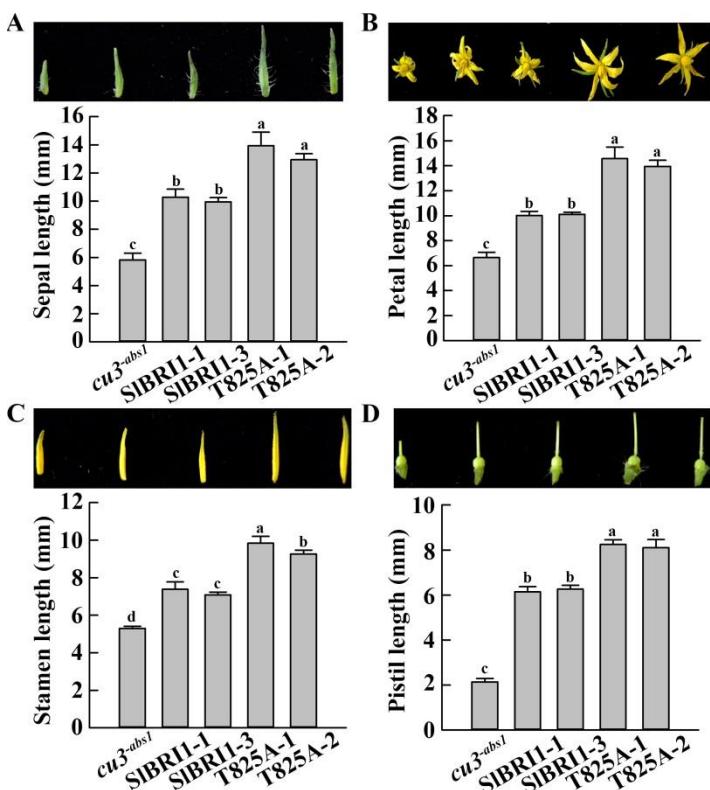


Figure S2. SIBRII T825A affected the development of the floral organs. (A) Top, phenotypes of a single

sepal per plant. Bottom, sepal length per plant. (B) Top, phenotypes of a single petal per plant. Bottom, petal length per plant. (C) Top, phenotypes of a single stamen per plant. Bottom, stamen length per plant. (D) Top, phenotypes of a single pistil per plant. Bottom, pistil length per plant. The floral organs of (A-D) shown on the top from left to right are as follows: *cu3^{abs1}*, SIBRI1-1, SIBRI1-3, T825A-1, and T825A-2. The data for (A-D) are presented as the mean \pm SD (n=6). The different letters indicate significant differences (one-way ANOVA, Tukey's test, $p < 0.05$).

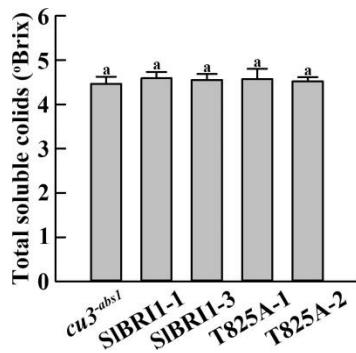


Figure S3. SIBRI1 T825A had no influence on fruit quality during the red ripe period. The data are presented as the mean \pm SD (n=15). The different letters indicate significant differences (one-way ANOVA, Tukey's test, $p < 0.05$).

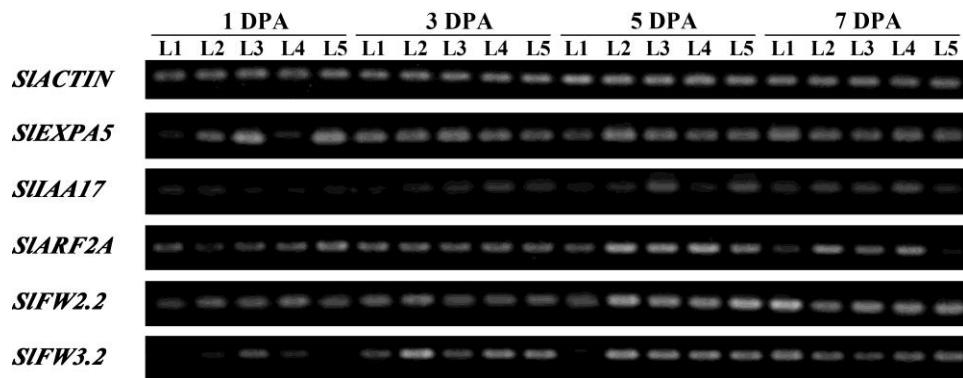


Figure S4. SIBRI1 T825A had no influence on the relative transcript levels of *SIIAA17*, *SIARF2A*, *SIEXPAS5*, *FW2.2* and *FW3.2*. *SIACHTIN* was used as a reference gene in tomato. Total RNA from 1 DPA, 3 DPA, 5 DPA, and 7 DPA ovaries from both the transgenic and *cu3^{abs1}* plants was extracted and transcribed to cDNA. The relative transcript levels were tested by RT-PCR. L1 (Line 1), *cu3^{abs1}*; L2 (Line 2), SIBRI1-1; L3 (Line 3), SIBRI1-3; L4 (Line 4), T825A-1; L5 (Line 5), T825A-2. Each product had 3 biological replications.

Table S1. Primers sequence used in this research

Primer name	Primer sequence (5'→3')
Promoter of <i>SIBRI1</i> -F	GACCATGATTACGCCAAGCTTCT CCATTCAATTATTGCTCAAAGG
Promoter of <i>SIBRI1</i> -R	ACCACCCGGGGATCCTCTAGACT TCAAAGATTGAAACTTATAGCTTAAA
<i>SIBRI1</i> -F for tomato transformation	TCTTGAAAGTCTAGAGCTCGAGAT GAAAGCTCACAAACTGTGTTAAC
<i>SIBRI1</i> -R for tomato transformation	GCCCTTGCTCACCATGGTACCCCTTA TCGTCGTACATCCTGTAATC
T825A-F for site mutation	GTTGCCATAGAGGCAGAAGAAGAGGA
T825A-R for site mutation	TCCTCTTCTCGCCTCTATGGCAAC
T825D-F for site mutation	GTTGCCATAGAGGACAAGAAGAGGA
T825D-R for site mutation	TCCTCTTCTGTGCTCTATGGCAAC
K916E-F for site mutation	GAGTGTGTAGCTATTGAGAAATTGATAC ACG
K916E-R for site mutation	CGTGTATCAATTCTCAATAGCTACAACA CTC
<i>SIBRI1</i> -CD-F for phosphorylation analysis in vitro	GACGATGACAAAGTCAAGCTTGAGA CGAAGAACAGGAGGAGG
<i>SIBRI1</i> -CD-R for phosphorylation analysis in vitro	TCTGCAGGTACCCGGGAATTCAAGG TGTTCGTCAGCTCATTG
<i>SI</i> ACTIN-F for qRT-PCR and RT-PCR	GTGTGGGCTCACCTACGTT
<i>SI</i> ACTIN-R for qRT-PCR and RT-PCR	ACAATCCAAGGGTTGTCAC
<i>SI</i> CPD-F for qRT-PCR and RT-PCR	ATCCAATTAACGTCCAACAT
<i>SI</i> CPD-R for qRT-PCR and RT-PCR	ACCTTCATACACCTCCCTC
<i>SIBRI1</i> -F for qRT-PCR	TTCAATGGCACGATCCGAA
<i>SIBRI1</i> -R for qRT-PCR	TGGGGAGAGGATAACCCACAG
<i>SID</i> WARF-F for qRT-PCR	ATTCAATGCTTGATTGGCA
<i>SID</i> WARF-R for qRT-PCR	GGGAGAGAACATTGCAGGAAG
<i>SI</i> CDKB1-F for RT-PCR	ATGGAGAAATACGAGAAATTGGAG
<i>SI</i> CDKB1-R for RT-PCR	ACGATGTAGAGAGAACATGAGATAGC
<i>SI</i> CDKB2.1-F for RT-PCR	ATGCTGGTAAGAGTGTATCGG
<i>SI</i> CDKB2.1-R for RT-PCR	CGGAGAGTAGTTGGAGGAAC
<i>SIGA20ox1</i> -F for RT-PCR	CTCATTTCTAATGCTCATCGT
<i>SIGA20ox1</i> -R for RT-PCR	R: TGCAGATGATTCTTCTTAGCG
<i>SIGID1</i> -F for RT-PCR	GATCTTGATACACCTCTCAGTACTA
<i>SIGID1</i> -R for RT-PCR	ACAGCCTTACATATACTAACAAAGAC
<i>SIDE</i> LLA-F for RT-PCR	TGATGCGACTATACTTGATATAAG
<i>SIDE</i> LLA-R for RT-PCR	GGGTAACTGTTAATAGAGTTC
<i>SIBZR1</i> -F for RT-PCR	AAGTATGCGTCAAATGTT
<i>SIBZR1</i> -R for RT-PCR	TGCTCTTACCCCTGTCTG
<i>SIE</i> XPAs-F for RT-PCR	AAGGGTTCAAGAACTCAATGGCAAC

<i>SIEXP A5</i> -R for RT-PCR	ACCATGCCTGTAGTGACCTAAAG
<i>SIIA17</i> -F for RT-PCR	TTAAGGCTAGGGTGCCTGG
<i>SIIA17</i> -R for RT-PCR	AGCTGGGTTTGTGTGTTG
<i>SIARF2A</i> -F for RT-PCR	GGATGGCAAAGGGCATAGTA
<i>SIARF2A</i> -R for RT-PCR	TACGCTGCACCTCCTTTT
<i>SIFW2.2</i> -F for RT-PCR	CTGGGATTGACAGGATTGCCT
<i>SIFW2.2</i> -R for RT-PCR	TAGCTTGCCACCTATTCCC
<i>SIFW2.3</i> -F for RT-PCR	ATTACTCTCGTGGGCAAGGC
<i>SIFW2.3</i> -R for RT-PCR	CTGAACCGAATGGTGCAAGC
