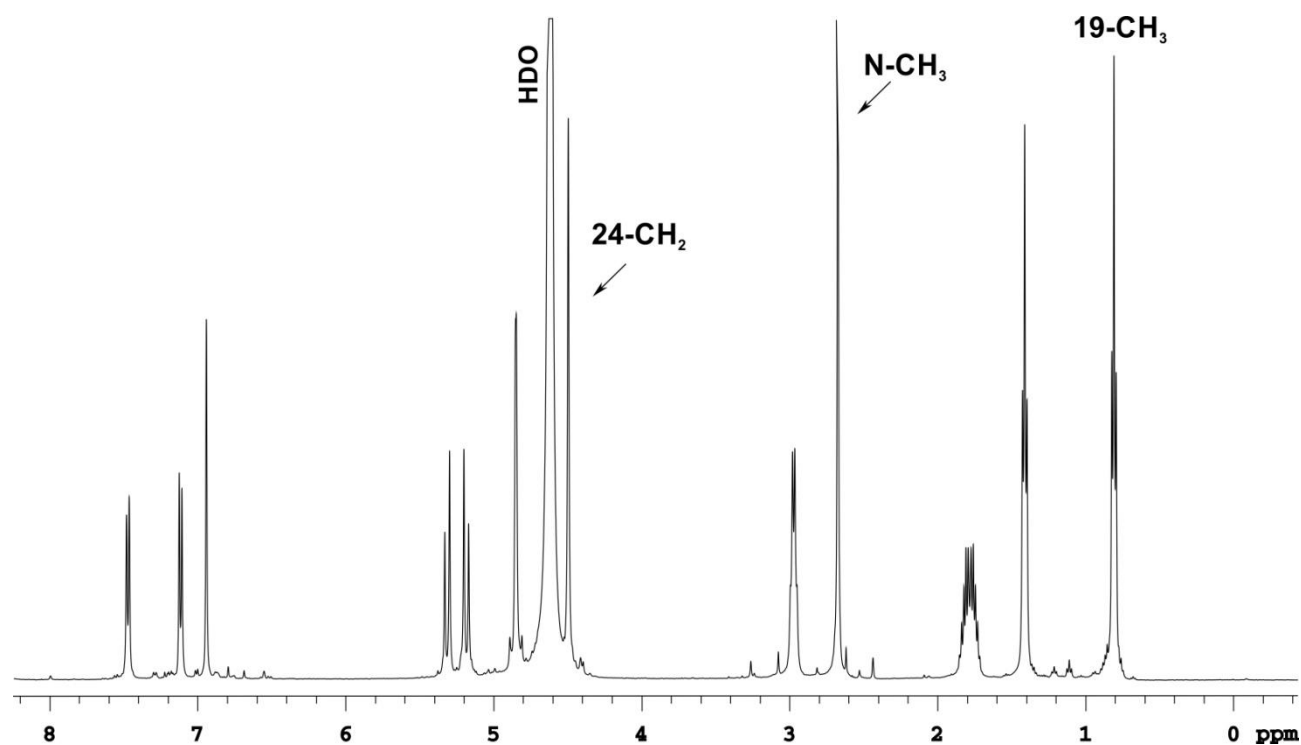
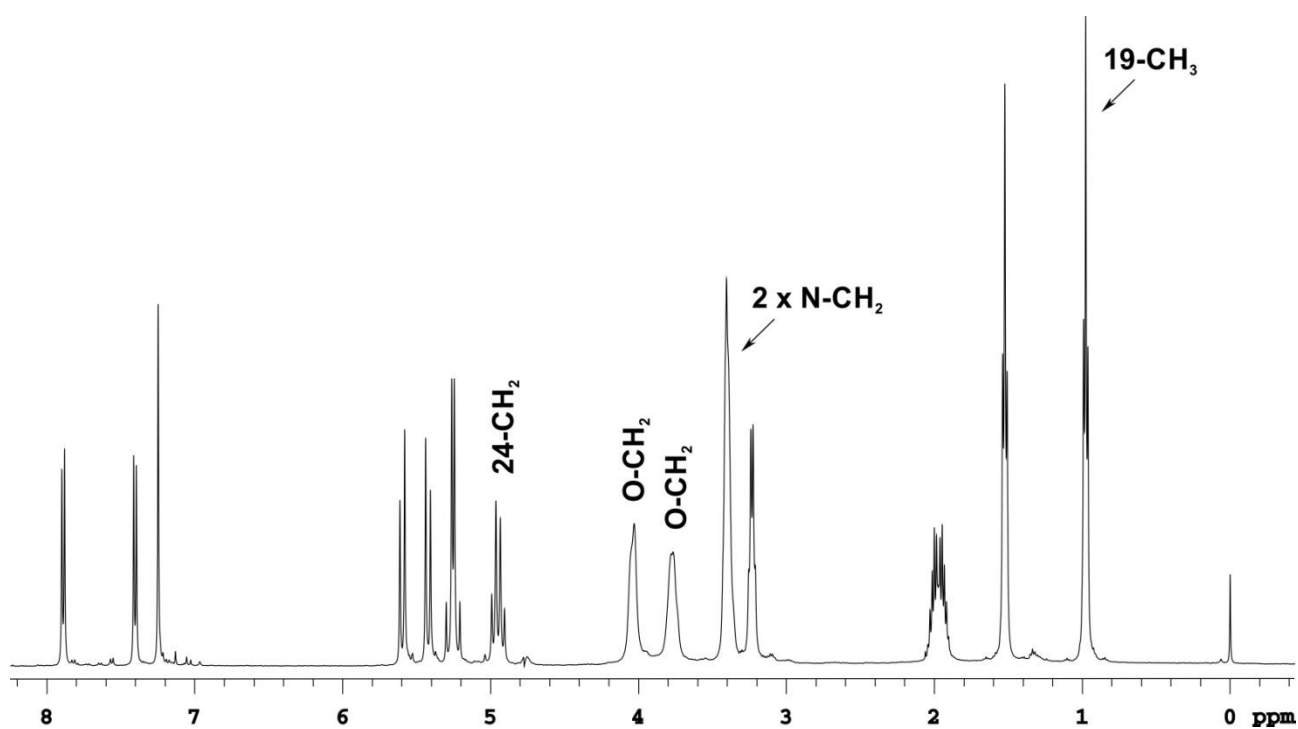


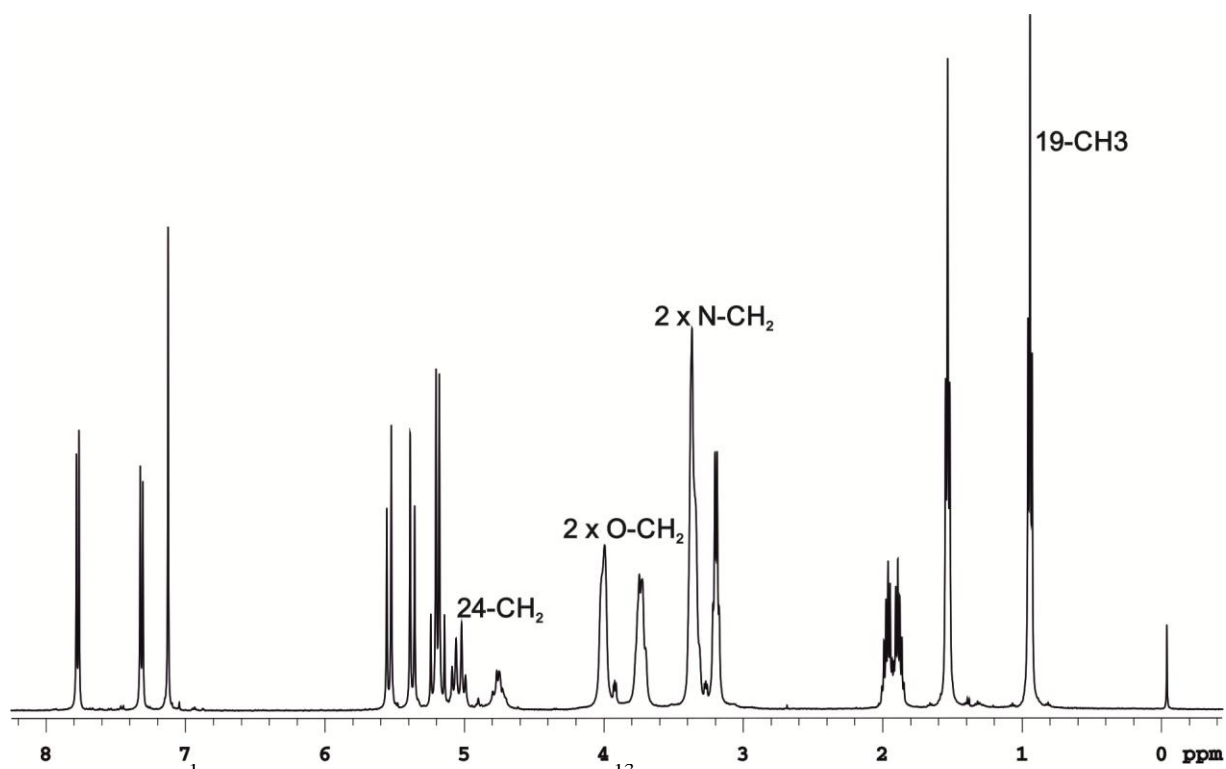
**Fig. S1** The  $^1\text{H}$  NMR spectrum of nicked decamer **1** in buffered  $\text{H}_2\text{O}/\text{D}_2\text{O}$ , (90/10 vol%) 25 mM NaCl /25 mM  $\text{K}_3\text{PO}_4$ , at pH 6. Seven guanosine and 3 thymidine NHs hydrogen bonded, forming a duplex are shown in an inset.



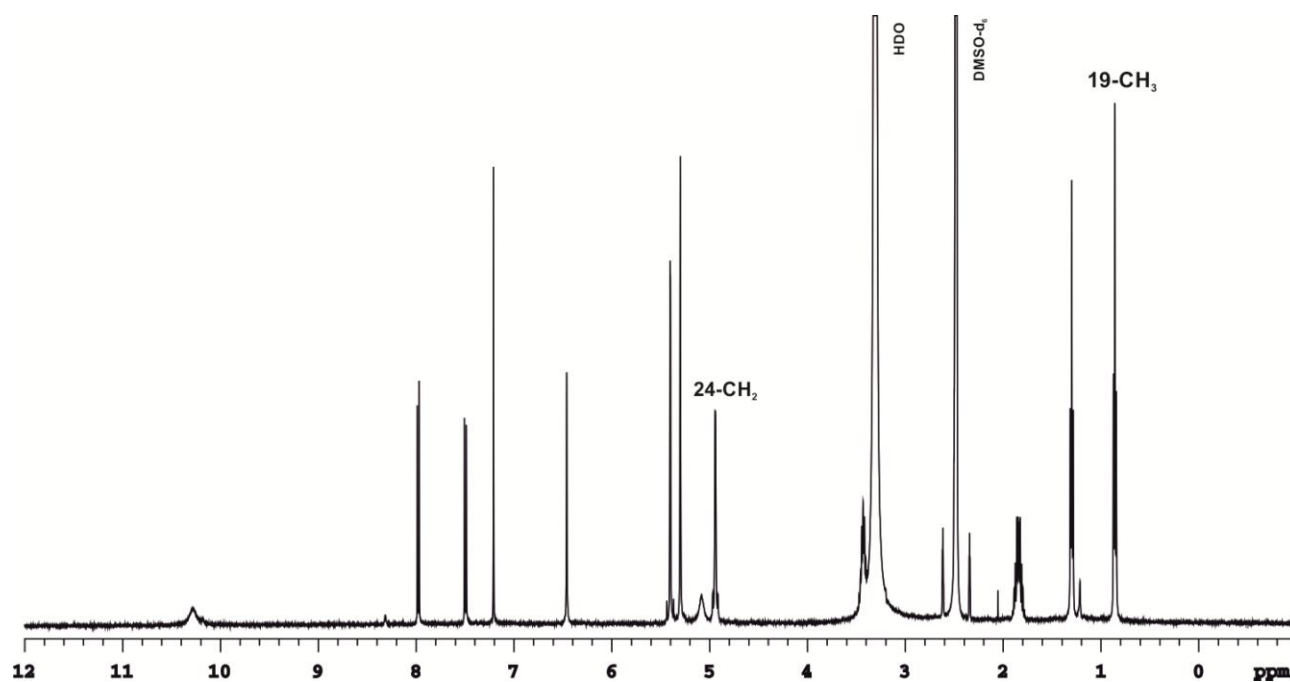
**Fig. S2** The  $^1\text{H}$  NMR spectrum of **2** in buffered  $\text{D}_2\text{O}$ , 25 mM NaCl /25 mM  $\text{K}_3\text{PO}_4$ , at pH 6.



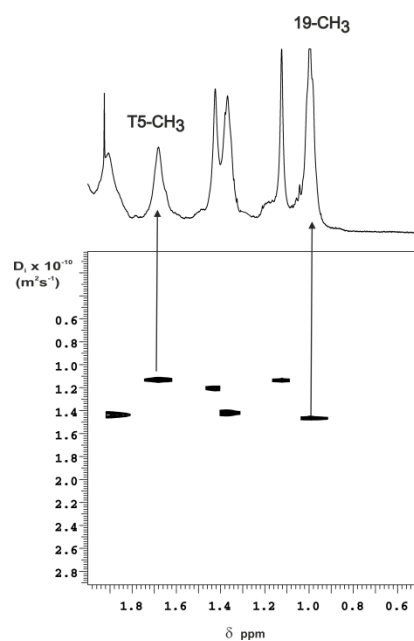
**Fig.S3** The <sup>1</sup>H NMR spectrum of **3** in buffered D<sub>2</sub>O, 25 mM NaCl /25 mM K<sub>3</sub>PO<sub>4</sub>, at pH 6.



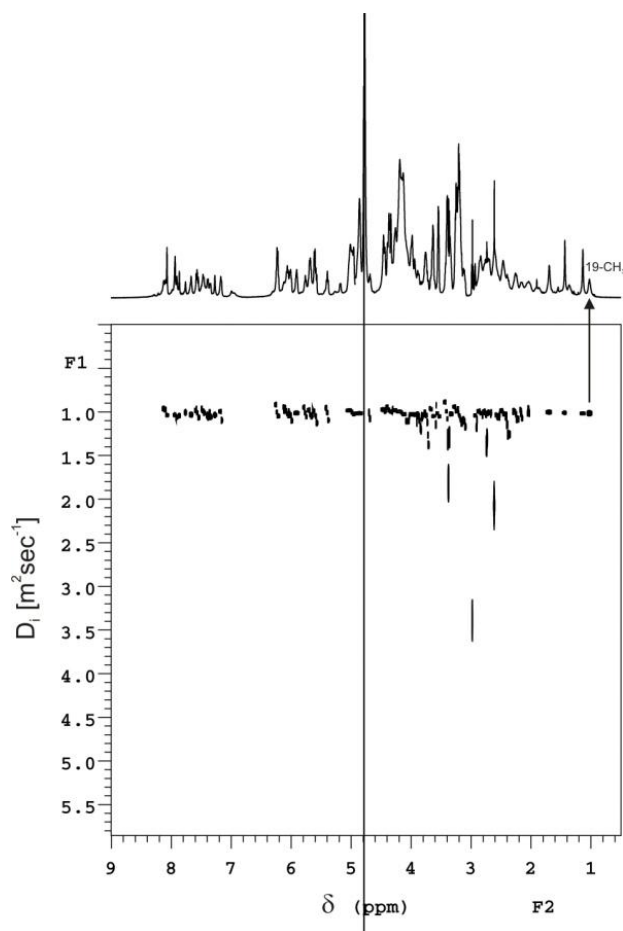
**Fig.S3a** The <sup>1</sup>H NMR spectrum of **3** enriched <sup>13</sup>C-24 in buffered D<sub>2</sub>O, 25 mM NaCl /25 mM K<sub>3</sub>PO<sub>4</sub>, at pH 6.



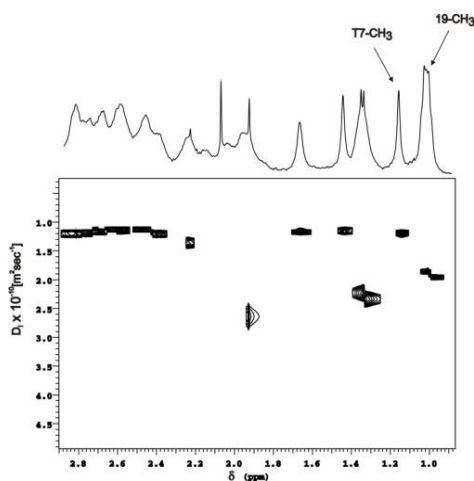
**Fig. S3b** The  $^1\text{H}$  NMR spectrum of metabolite **4** in  $\text{DMSO-d}_6$ .



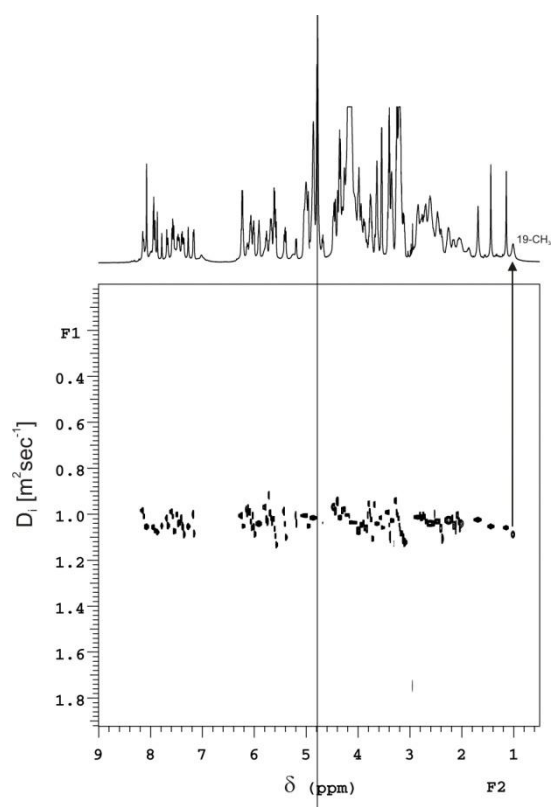
**Fig. S4** The part of the DOSY spectrum presenting result for compounds **1** and **2** after 5 days of incubation in buffered  $\text{D}_2\text{O}$ , 25 mM  $\text{NaCl}$  /25 mM  $\text{K}_3\text{PO}_4$ , at pH 6.



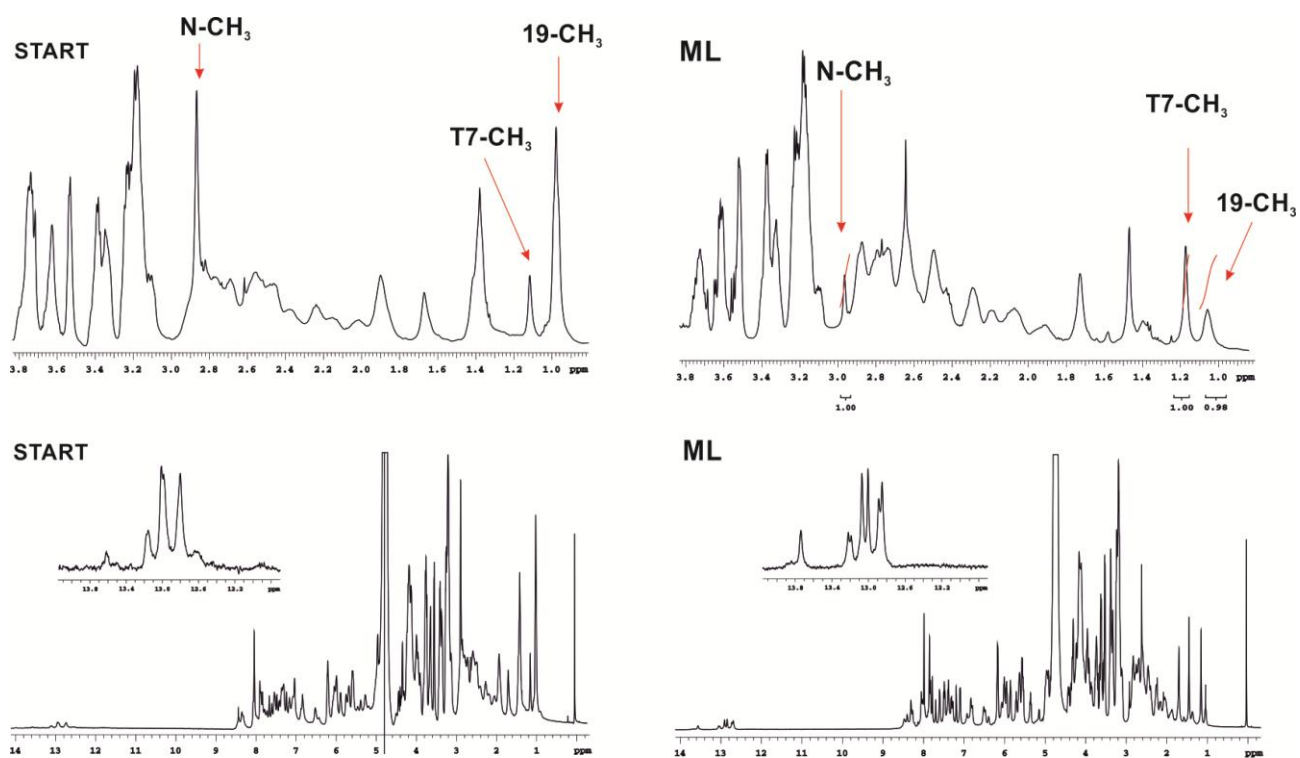
**Fig. S5** The full DOSY spectrum for decamer **1** and derivative **2** after filtering the sample of reaction solution, ML. The DOSY spectrum of a 1:1 complex of **1** and **2** in D<sub>2</sub>O buffer, pH 6, 25°C. The diffusion coefficient  $D_i$ ,  $1.0 \pm 0.1 \times 10^{-10} \text{ [m}^2\text{s}^{-1}\text{]}$  is equal for both components.



**Fig. S6a** The part of the DOSY result of compounds **1** and **3** at the start, in buffered D<sub>2</sub>O, 25 mM NaCl / 25 mM K<sub>3</sub>PO<sub>4</sub>, at pH 6.



**Fig.S6b** The full DOSY spectrum for decamer **1** and derivative **3** after filtering the sample of reaction solution.



**Fig. S7** The shape of 1D NMR spectrum of reaction mixture of **2** with **1** at the start and in a mother liquor ML after filtering of reaction solution. Expansion of low frequency region of a spectrum is given in both cases.

**Table S1.** The  $^1\text{H}$  NMR chemical shifts  $\delta$  [ppm] of DNA **1** and SN38 derivative **2** from NOESY spectrum in buffered  $\text{D}_2\text{O}$ , at  $25^\circ\text{C}$  (**1:2** ratio) acquired after 4 days of incubation the sample at  $25^\circ\text{C}$ . \*

Base	H1'	H2'	H2''	H3'	H4'	H5', H5''	H6/8	H2/5/Me
G1	5.998	2.760	2.833	4.955	4.334	3.883/ 3.991	8.065	-
C2	5.692	2.140	2.439	4.860	4.217	4.161	7.384	5.380
G3	6.059	2.680	2.820	5.008	4.430	4.112/ 4.156	7.923	-
T4	6.055	2.058	2.559	4.885	4.239	na/ 4.310	7.263	1.423
<b>T5</b>	<b>6.222</b>	<b>2.550</b>	<b>2.571</b>	<b>4.984</b>	<b>4.191</b>	<b>4.098/ na</b>	<b>7.492</b>	<b>1.679</b>
nick	-----	-----	-----	-----	-----	-----	-----	-----
<b>G6</b>	<b>5.490</b>	<b>2.358</b>	<b>2.574</b>	<b>4.687</b>	<b>4.170</b>	<b>3.798/ 3.911</b>	<b>7.685</b>	<b>-</b>
T7	6.023	2.158	2.520	4.853	4.235	4.003/ 4.114	7.338	1.122
C8	5.742	2.021	2.391	4.853	4.111	na/ na	7.439	5.590
G9	5.898	2.626	2.695	4.999	4.364	4.075/ 4.133	7.925	-
C10	6.225	2.239	2.466	4.851	4.192	4.139/ 4.249	7.565	5.598

Base	H1'	H2'	H2''	H3'	H4'	H5', H5''	H6/8	H2/5/Me
G11	6.002	2.755	2.828	4.955	4.333	3.880/ 3.991	8.064	-
C12	5.679	2.002	2.376	4.850	4.172	4.168	7.338	5.384
G13	5.673	2.757	2.778	5.032	4.391	4.078/ 4.156	7.894	-
A14	6.203	2.646	2.820	4.913	4.436	na/ na	8.065	7.739
<b>C15</b>	<b>5.733</b>	<b>1.948</b>	<b>2.163</b>	<b>na</b>	<b>4.165</b>	<b>na/ 4.303</b>	<b>7.183</b>	<b>5.045</b>
<b>A16</b>	<b>5.601</b>	<b>2.598</b>	<b>2.715</b>	<b>4.965</b>	<b>4.387</b>	<b>na / na</b>	<b>7.806</b>	<b>6.854</b>
A17	6.093	2.591	2.807	5.011	4.444	na/ 4.289	8.092	7.622
C18	5.584	1.856	2.272	na	4.124	na/ 4.240	7.168	5.176
G19	5.889	2.583	2.686	4.972	4.337	4.038/ 4.118	7.858	-
C20	6.213	2.232	2.458	4.849	4.190	4.127 / 4.235	7.550	5.576

\* The G6-C15 and T5-A16 base pairs, in bold, are flanking both faces of a nick.

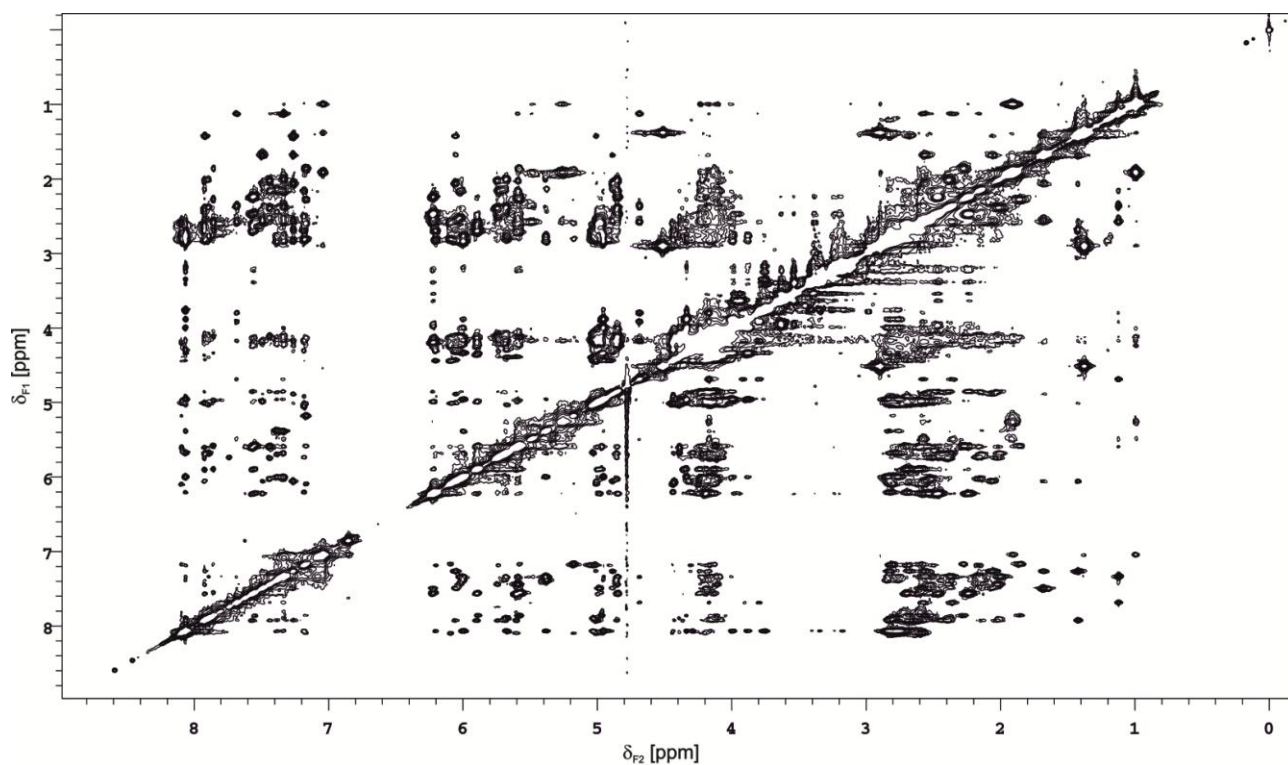
na-not assigned.

**Table S2.** The  $^1\text{H}$  NMR chemical shifts  $\delta$  [ppm] of changes induced in free DNA decamer and **2** after 4 days incubation in  $\text{D}_2\text{O}$  (**1:2** ratio).

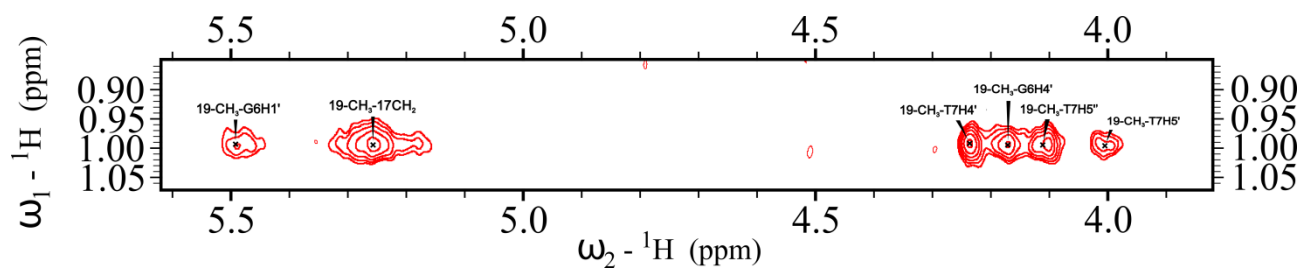
Base	H1'	H2'	H2''	H3'	H4'	H5', H5''	H6/8	H2/5/Me
G1	-0.006	-0.013	-0.006	-0.004	-0.009	0.001 / -0.020	-0.013	-
C2	-0.003	-0.011	-0.020	-0.008	-0.010	0.001	-0.007	-0.027
G3	-0.029	-0.021	-0.034	-0.013	-0.008	-0.004 / -0.009	-0.024	-
T4	-0.008	-0.026	-0.048	0.014	-0.035	- / -0.003	-0.005	-0.023
T5	0.020	<b>0.052</b>	-0.046	-0.006	-0.009	-0.001 / -	<b>0.067</b>	-0.012
nick	-----	-----	-----	-----	-----	-----	-----	-----
G6	<b>-0.342</b>	<b>-0.284</b>	<b>-0.068</b>	0.017	-0.037	<b>-0.076 / -0.222</b>	0.020	-
T7	-0.045	-0.022	-0.037	-0.018	<b>-0.082</b>	- / 0.013	<b>-0.204</b>	-0.025
C8	-0.018	-0.045	-0.027	-0.015	-0.022	- / -	-0.036	-0.046
G9	-0.006	0.012	0.003	0.009	0.010	-0.003 / -0.001	-0.013	-
C10	-0.002	-0.017	0.005	0.005	-0.009	0.001 / -0.003	-0.007	-0.018

Base	H1'	H2'	H2''	H3'	H4'	H5', H5''	H6/8	H2/5/Me
G11	-0.002	-0.018	-0.011	-0.004	-0.010	-0.002 / -0.020	-0.014	-
C12	0.016	-0.013	-0.009	-0.001	-0.009	0.024	-0.029	-0.044
G13	0.012	0.019	-0.037	-0.015	-0.003	0.014 / 0.010	-0.016	-
A14	-0.044	-0.035	<b>-0.128</b>	<b>-0.118</b>	<b>-0.057</b>	- / -	<b>-0.093</b>	<b>-0.058</b>
C15	<b>0.170</b>	0.010	<b>-0.199</b>	?	0.021	- / 0.000	0.027	<b>-0.245</b>
A16	<b>-0.230</b>	0.024	<b>-0.093</b>	-0.043	0.029	- / -	<b>-0.224</b>	<b>-0.204</b>
A17	-0.040	-0.048	-0.031	-0.026	-0.015	- / <b>0.073</b>	<b>-0.069</b>	<b>-0.081</b>
C18	-0.018	0.001	-0.007	?	-0.018	- / -0.017	0.001	-0.024
G19	-0.015	-0.031	-0.006	-0.018	-0.017	0.008 / -0.001	-0.010	-
C20	-0.014	-0.024	-0.003	0.003	-0.011	-0.003 / -0.005	-0.005	-0.014

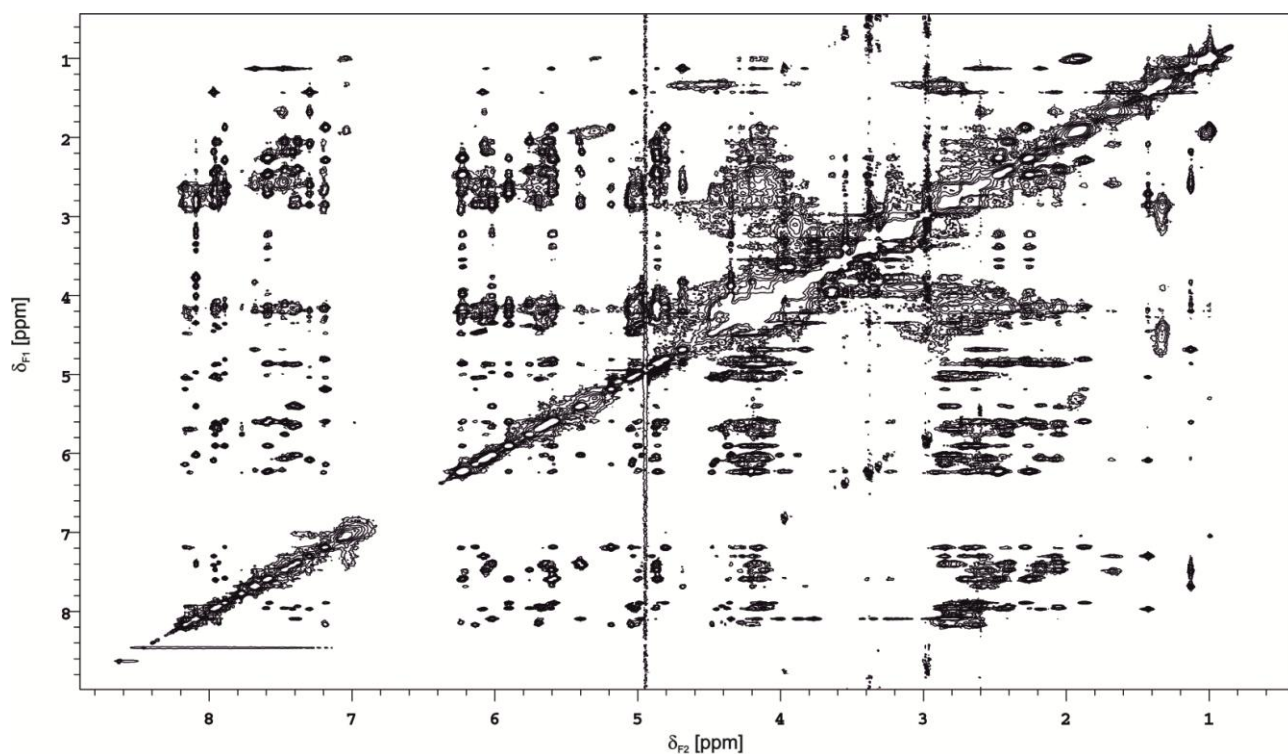




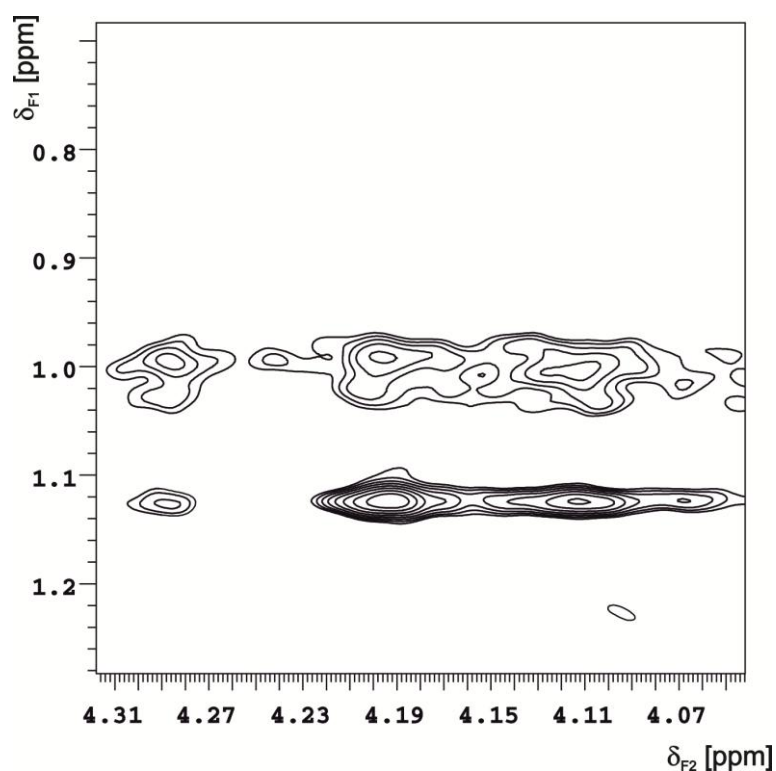
**Fig. S8** The NOESY spectrum of the sample 1+2 after 4 days of incubation (1:2 ratio).



**Fig. S8a** The example of intermolecular cross-peaks in complex 1+2 after 4 days of incubation(1:2 ratio), before filtering the reaction solution ,see Table 1 in manuscript.



**Fig S9** The NOESY spectrum of the sample 1+3 (see experimental section) after 24h.

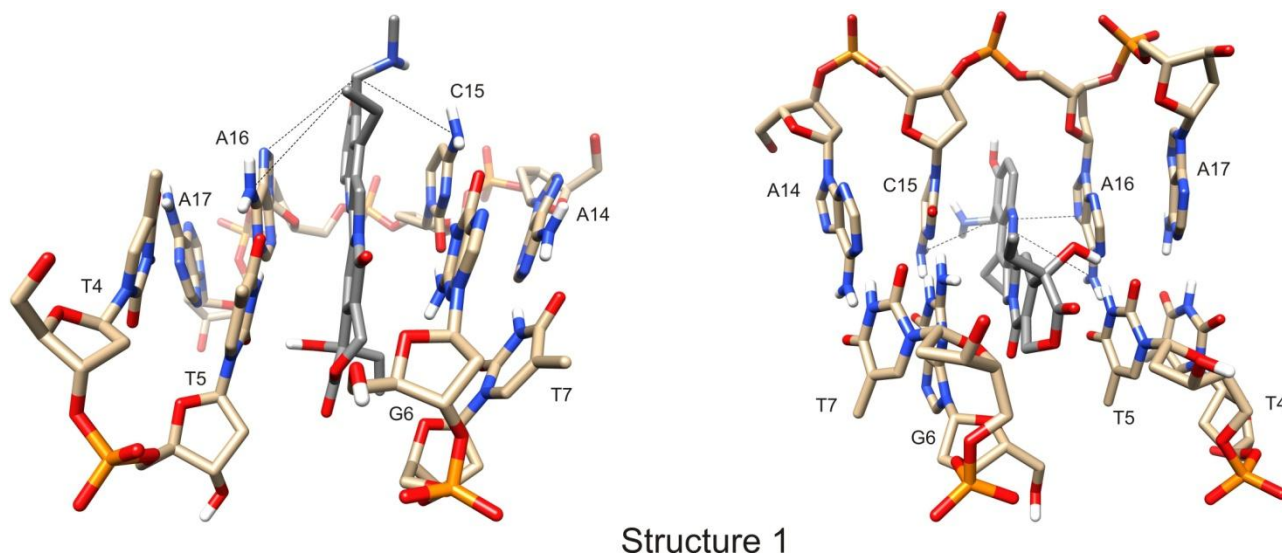


**Fig. S10** The example of the cross-peaks in sample 1+3 (see experimental section). The cross peaks at 1.0 and 1.02 ppm are the intermolecular cross peaks, not assigned, of two different methyl groups 19-CH<sub>3</sub> due to **4** and SN38 and cross peaks at 1.12 ppm are the intramolecular cross peaks of T7-CH<sub>3</sub> in **1**.

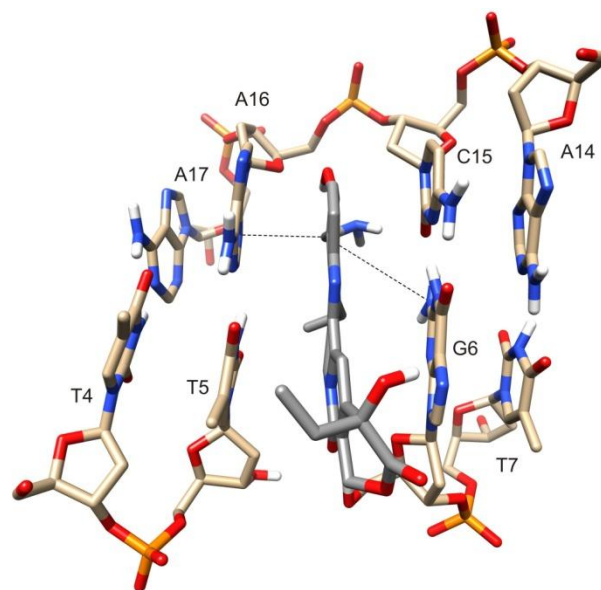
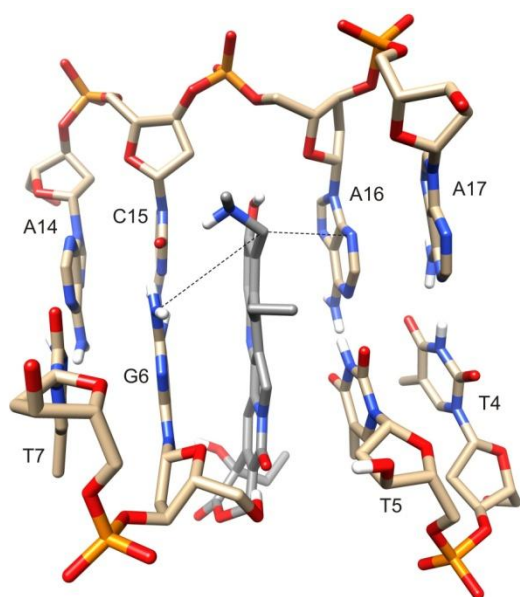
**Table S3.** The most populated cluster energy from PBSA and GBSA analysis.

Structure	Energy [kcal/mol]	
	PBSA	GBSA
NHMe-1	-35.56 +/- 2.97	-35.68 +/- 3.03
NHMe-2	-34.43 +/- 3.19	-33.77 +/- 2.60
NHMe-3	-37.69 +/- 2.80	-34.84 +/- 2.68
NHMe-4	-34.21 +/- 3.33	-35.01 +/- 2.50

The above calculations and NOE effects in Table 4 ( manuscript) point to structures 1 and 3 as the best ones. The HB also favors structure 3.

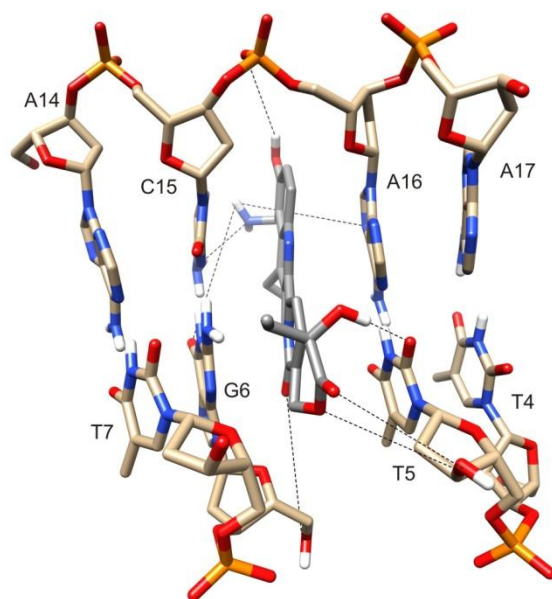
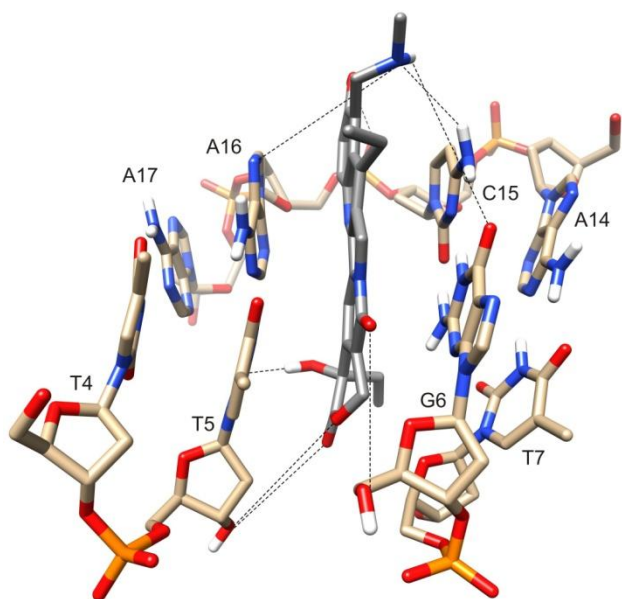


**Fig. S11** The best structure from modeling showing potential sites of hybrid formation in a molecular complex **1+2**

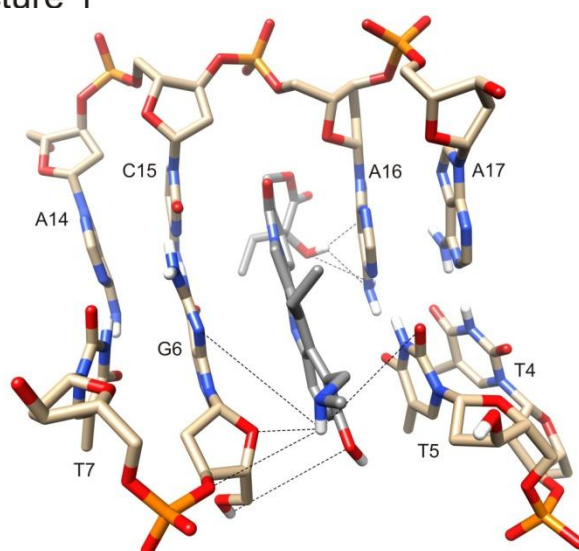
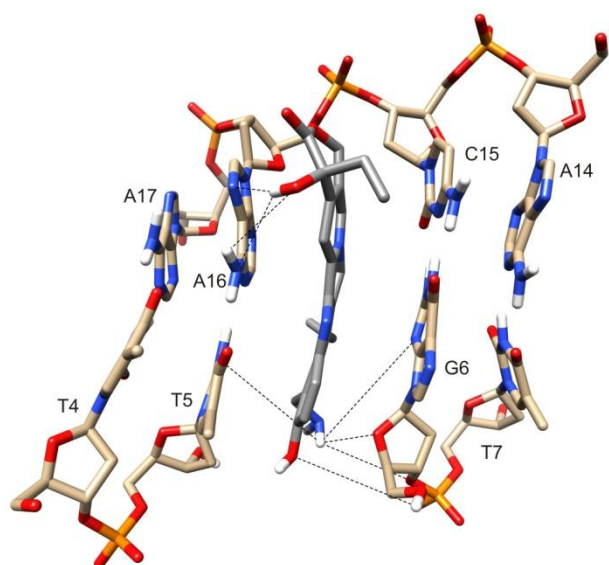


Structure 3

**Fig. S12** The second best structure from modeling showing potential sites of hybrid formation in a molecular complex **1+2**

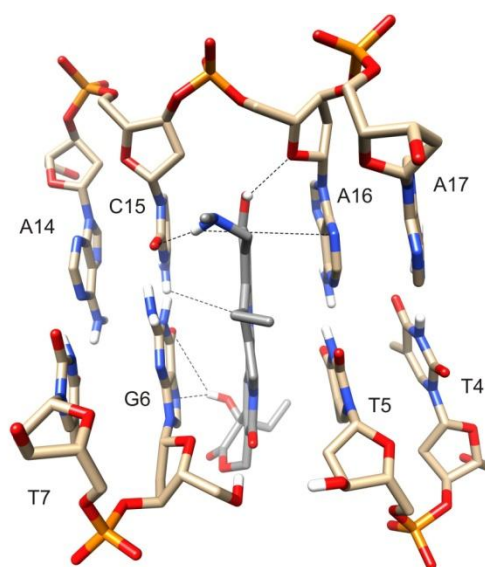
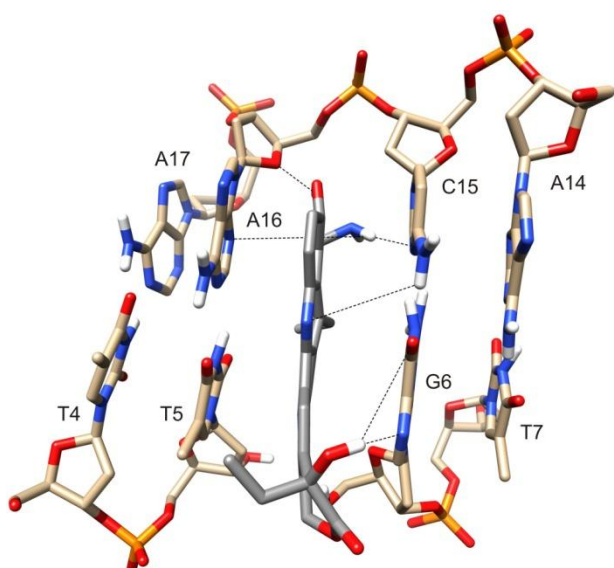


Structure 1

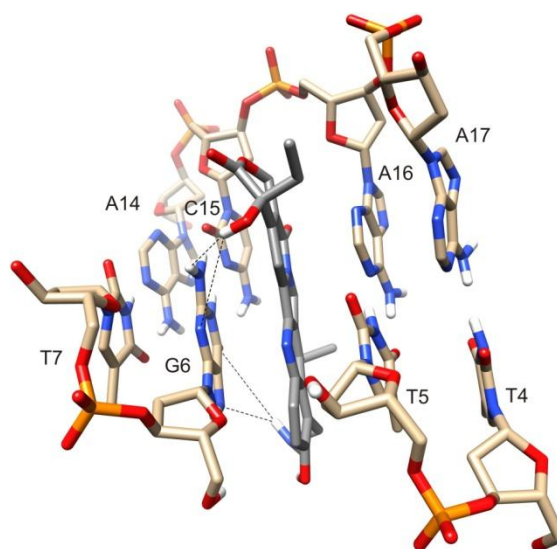
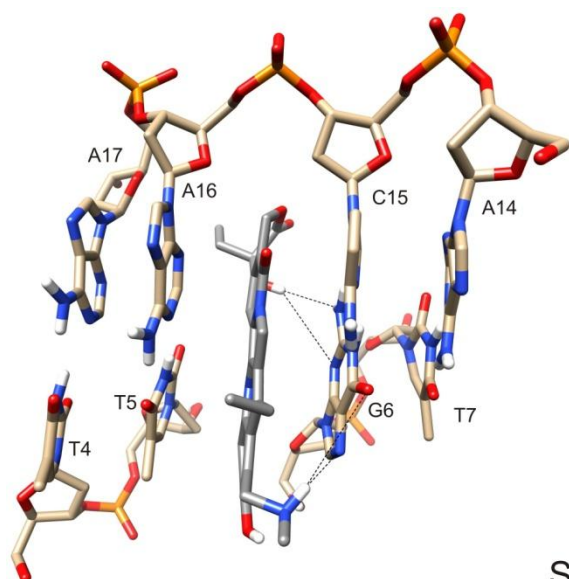


Structure 2





Structure 3

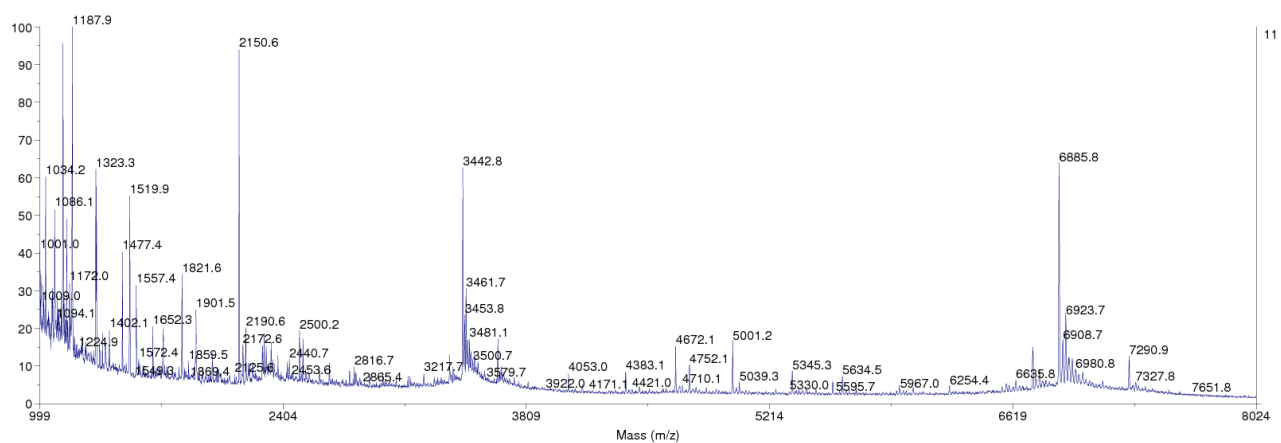


Structure 4

**Fig. S13** The hydrogen bonding in structures best representing the most populated cluster obtained by cluster analysis in a molecular complex **1+2**.

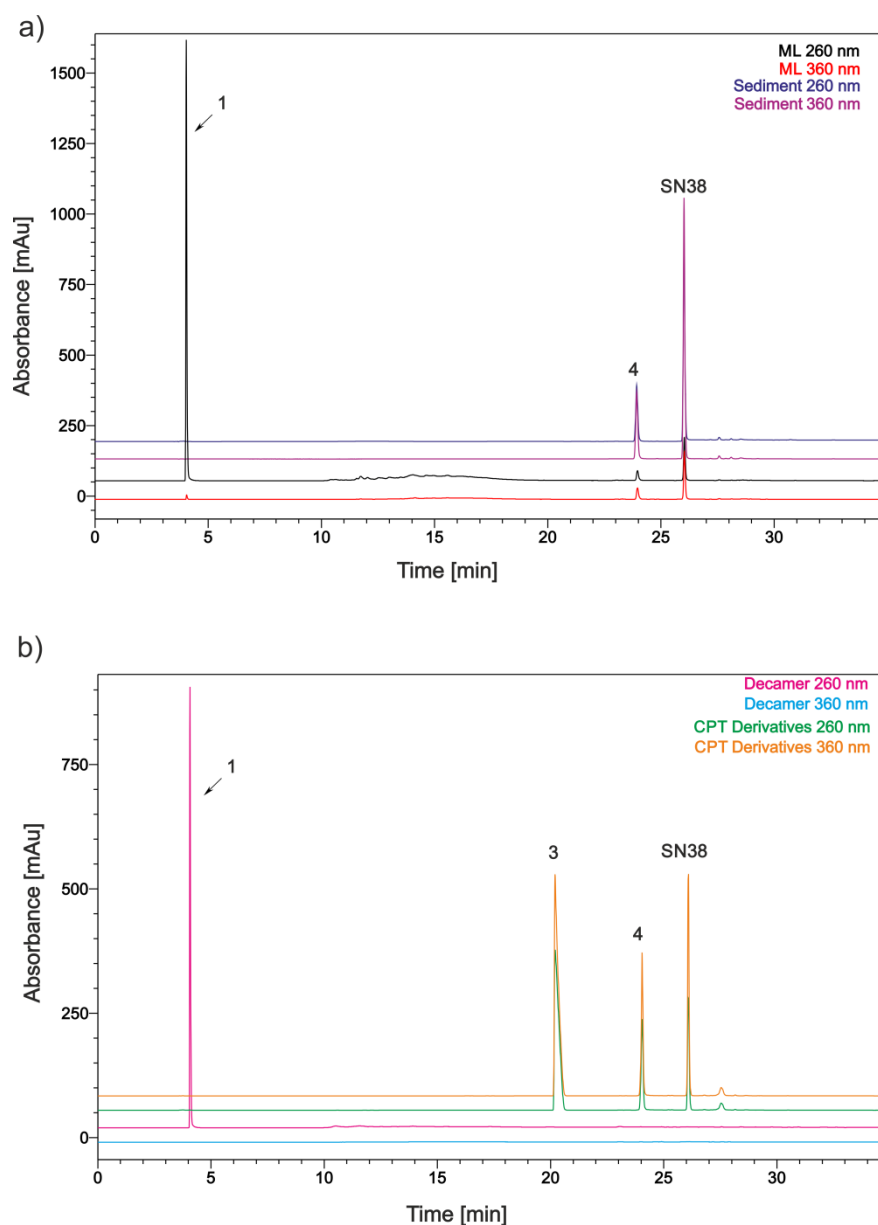
**Table S4 .** The real hydrogen bonds in **1+2** complex from PM7 calculations.

Atom names		Structure 1	
DNA	CMP	HB length [Å]	population
T5- <u>O</u> 2	20 - <u>O</u> <u>H</u>	2.08 +/- 0.29	75.5 %
G6- <u>H</u> O5'	16 >C= <u>O</u>	1.93 +/- 0.27	44.4 %
A16- <u>N</u> 7	9-CH <sub>2</sub> N <u>H</u> Me	2.26 +/- 0.10	31.5 %
T5- <u>H</u> O3'	21 >C= <u>O</u>	2.48 +/- 0.42	19.1 %
T5- <u>H</u> O3'	21 > <u>O</u>	2.85 +/- 0.28	14.0 %
A16- <u>O</u> P2	10 - <u>O</u> <u>H</u>	2.49 +/- 0.34	13.6 %
C15- <u>O</u> P2	10 - <u>O</u> <u>H</u>	2.32 +/- 0.39	7.5 %
A16- <u>O</u> 5'	10 - <u>O</u> <u>H</u>	2.90 +/- 0.27	6.8 %
G6- <u>O</u> 6	9-CH <sub>2</sub> N <u>H</u> Me	2.49 +/- 0.28	4.1 %
		Structure 2	
A16- <u>N</u> 7	20 - <u>O</u> <u>H</u>	2.32 +/- 0.29	63.5 %
T5- <u>O</u> 2	9-CH <sub>2</sub> N <u>H</u> Me	2.23 +/- 0.11	44.6 %
A16- <u>H</u> 62	20 - <u>O</u> <u>H</u>	2.93 +/- 0.35	30.0 %
G6- <u>N</u> 3	9-CH <sub>2</sub> N <u>H</u> Me	2.27 +/- 0.12	20.6 %
G6- <u>O</u> 4'	9-CH <sub>2</sub> N <u>H</u> Me	2.18 +/- 0.19	18.8 %
G6- <u>H</u> O5'	10 - <u>O</u> <u>H</u>	1.83 +/- 0.22	16.8 %
G6- <u>O</u> 3'	9-CH <sub>2</sub> N <u>H</u> Me	2.65 +/- 0.31	10.2 %
G6- <u>O</u> 4'	10 - <u>O</u> <u>H</u>	1.81 +/- 0.25	6.8 %
G6- <u>O</u> 5'	10 - <u>O</u> <u>H</u>	1.78 +/- 0.29	6.2 %
		Structure 3	
C15- <u>O</u> 2	9-CH <sub>2</sub> N <u>H</u> Me	2.19 +/- 0.11	88.1 %
A16- <u>O</u> 4'	10 - <u>O</u> <u>H</u>	2.08 +/- 0.33	78.2 %
G6- <u>N</u> 7	20 - <u>O</u> <u>H</u>	2.21 +/- 0.14	75.3 %
G6- <u>H</u> O5'	16 >C= <u>O</u>	1.74 +/- 0.17	71.9 %
G6- <u>O</u> 6	20 - <u>O</u> <u>H</u>	3.08 +/- 0.38	24.4 %
A16- <u>N</u> 3	9-CH <sub>2</sub> N <u>H</u> Me	2.37 +/- 0.24	3.2 %
A16- <u>O</u> 5'	10 - <u>O</u> <u>H</u>	3.03 +/- 0.34	3.1 %
G6- <u>H</u> O5'	21 > <u>O</u>	2.27 +/- 0.36	2.9 %
		Structure 4	
G6- <u>N</u> 7	9-CH <sub>2</sub> N <u>H</u> Me	2.38 +/- 0.23	47.7 %
G6- <u>O</u> 6	9-CH <sub>2</sub> N <u>H</u> Me	2.51 +/- 0.33	29.1 %
G6- <u>H</u> 21	20 - <u>O</u> <u>H</u>	2.54 +/- 0.46	15.1 %
G6- <u>N</u> 3	20 - <u>O</u> <u>H</u>	2.41 +/- 0.34	13.9 %
T5- <u>O</u> 4	9-CH <sub>2</sub> N <u>H</u> Me	2.16 +/- 0.13	13.3 %
G6- <u>H</u> O5'	10 - <u>O</u> <u>H</u>	2.71 +/- 0.50	5.2 %

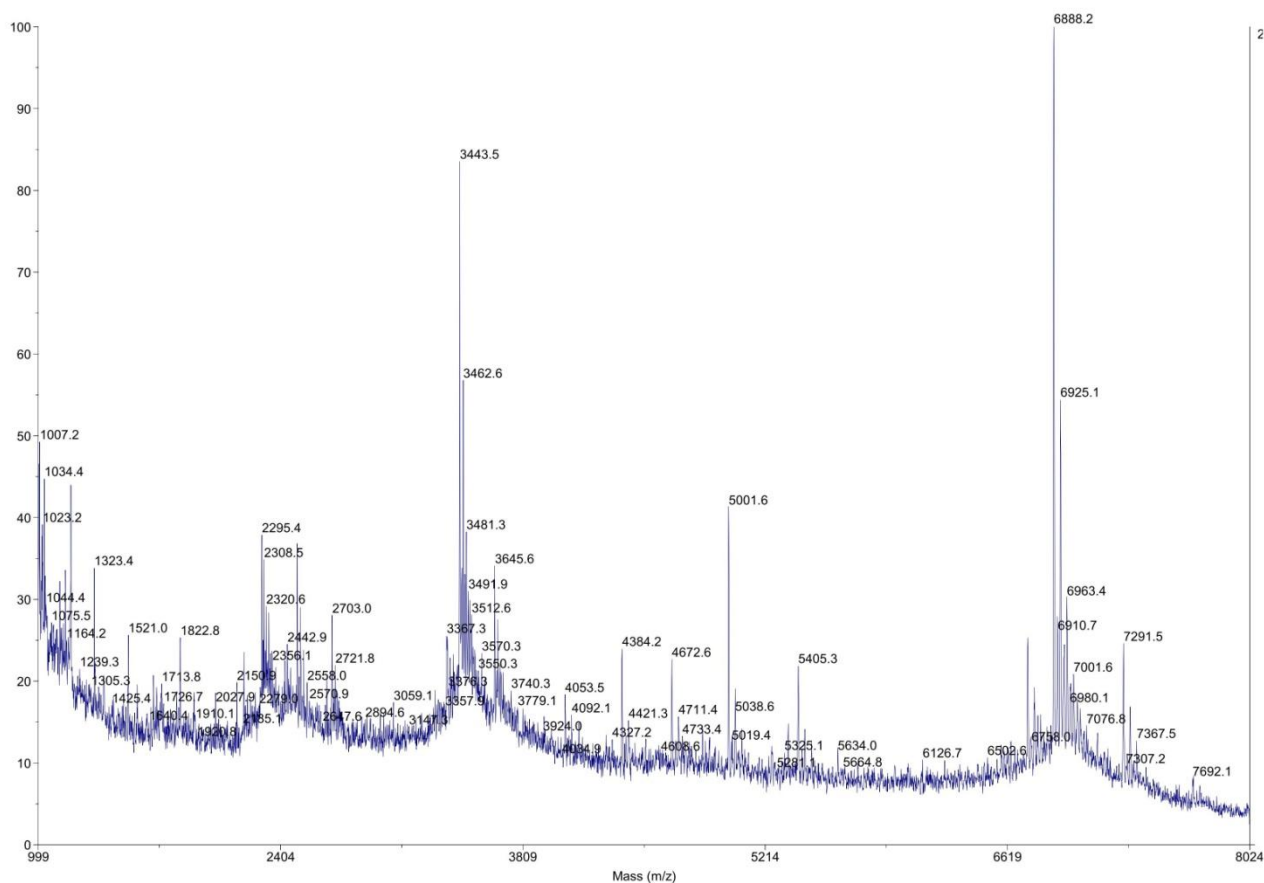


**Fig. S14** The MALDI MS spectrum of lyophilized ML showing neat DNA decamer ( $m/z=6885.8$ ,  $[M-H]^-$ ) and alkylated biohybrid ( $m/z= 7290.9$ ,  $[M-H]^-$ ) with compound **2**.

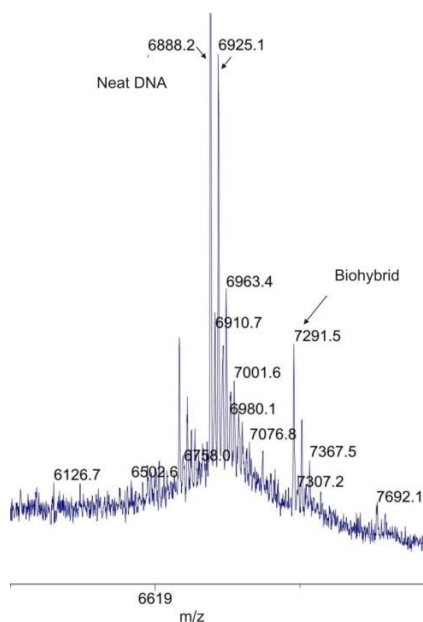




**Fig. S15** Upper panel: The overlay of two HPLC runs of reaction **1** + **3**; ML in black and red and sediment in navy blue and purple. The ML run evidences compounds strongly bound to nicked DNA which partly precipitate into sediment. Lower panel: The HPLC run of reference compounds.



**Fig. S16a** The MALDI MS spectrum of lyophilized ML showing neat DNA decamer ( $m/z=6888.2$ ) and alkylated biohybrid with compound **3** ( $m/z=7291.5$ ).



**Fig. S16b** The part of MALDI MS spectrum of lyophilized ML showing neat DNA decamer ( $m/z=6888.2$ , peak cut for clarity) and alkylated biohybrid with compound **3**.