

Table S1 Root traits associated QTLs in direct-seeded rice (DSR) conditions

Trait	Donor	Population	QTLs	Chromosome	Flanking marker	References
Root length	-	HKR47 × MAS26	<i>qRL</i> _{8.1}	8	RM8243	[1]
	-	HKR47 × MAS26	<i>qRL</i> _{9.1}	9	RM524-RM257 (13.2-66.1 cM)	
	IR94225-B-82-B, IR94226-B-177-B	MASARB25 × PB1460	<i>qRL</i> _{8.2}	8	RM547-RM331 (58.1-65.0 cM), RM310-RM547 (57.0-58.1 cM)	[1,2]
	IR94225-B-82-B, IR94226-B-177-B	HKR47 × MAS26	<i>qRHL</i> _{1.1}	1	-	[3]
Root hair length	IR94225-B-82-B, IR94226-B-177-B	-	<i>qRHD</i> _{1.1}	1	-	[3]
	IR94225-B-82-B, IR94226-B-177-B	-	<i>qRHD</i> _{5.1}	5	-	
Nodal root number	IR94225-B-82-B, IR94226-B-177-B	-	<i>qNR</i> _{5.1}	5	-	[3]
			<i>qNR</i> _{4.1}	4	-	
Root volume	-	-	<i>qRV</i> _{2.1}	2	RM485-RM7033 (0.0-7.9 cM)	[4,5]
Root thickness	-	HKR47 × MAS26	<i>qRT</i> _{1.1}	1	RM488-RM237 (101.4-112.9 cM)	[4-6]
Fresh root weight	-	MASARB25 × PB1460	<i>qFRW</i> _{2.1}	2	RM485-RM7033 (0.0-7.9 cM)	[4,5]
Anaerobic germination	IR93312-30- 101-20-13- 66-6	Donor: Khao Hlan On, Recipient: IR64	<i>qAG</i> _{9.1}	9	RM8303-RM5526	[7]
Early uniform emergence	IR91648-B- 32-B	Donor: Moroberekan	<i>qEUE</i> _{11.1}	11	id1100085-id11001535	[8]
Early vigor	IR94226-B- 177-B	Recepient: Swarna	<i>qEMM</i> _{1.1}	1		[8]
		Aus276 × MTU1010	<i>qEVV</i> _{9.1} (Donor: Aus276)	9	Flanking markers: id5000759- id5001182, ud9000737- id9002704,id1005271- id1006691; peak markers: id5001182, ud9000737, id1005271	[3]
Drought tolerance	IR74371-46- 1-1	IR74371-46-1-1/2*Sabitri	<i>qDTY</i> _{12.1}	12	<i>qDTY</i> _{1.2.1} RM28166-RM28199	[4,5,9,10]
			<i>qDTY</i> _{2.3}	2	<i>qDTY</i> _{2.3} RM3212-RM250	
			<i>qDTY</i> _{3.2}	3	<i>qDTY</i> _{3.2} RM22-RM545	

IR6322-34- 223	Donors: N22, Apo N22/Swarna, Apo/2*Swarna Ldg Rm8213	$qDTY_{3,1}$	3	$qDTY_{3,1};$ RM416	[6,8,10–12]
Lodging resistance	IR91648-B- 289-B	$qDTY_{1,1}$	1	$qDTY_{1,1};$ RM11943-RM12091	
Lodging resistance	IR91648-B- 289-B	Donor: Swarna	$qLDG_{3,1}$	2	$qDTY;$ RM324 id3000090-id300057
		Donor Moroberekan	$qLDG_{4,1}$	3	[8] id4004461-id4009390
				4	[8]

Table S2 Chromosome wise QTLs/genes related to root developmental traits in rice along with their locus ID and encoded protein

Root Phenotype	Gene	Chromosome	Locus ID		Encoded protein	References
			RAP	MSU		
Crown root	<i>OsRAA1</i>	1	Os01g0257300	LOC_Os01g15340	GTP-binding protein	[13]
	<i>OsYUC1</i>	1	Os01g0645400	LOC_Os01g45760	Flavin monooxygenase	[14]
	<i>OsPIN3t (OsPIN10a)</i>	1	Os01g0643300	LOC_Os01g45550	Auxin efflux carrier	[15]
	<i>OsmiR393a</i>	1	-	-	MicroRNA	[16]
	<i>FIB/OsTAA1</i>	1	Os01g0169800	LOC_Os01g07500	Aminotransferase	[17]
	<i>ERF3</i>	1	Os01g0797600	LOC_Os01g58420	AP2/ERF transcription factor	[18]
	<i>REH1/ OsPIN1b</i>	2	Os02g0743400	LOC_Os02g50960	Auxin efflux carrier	[19]
	<i>OsRR2</i>	2	Os02g0557800	LOC_Os02g35180	Type-A response regulator	[20]
	<i>OsCAND1</i>	2	Os02g0167700	LOC_Os02g07120	SCF ^{TIR1} ubiquitin ligase	[21]
	<i>CRL1/ARLI</i>	3	Os03g0149100	LOC_Os03g05510	LOB/ASL transcription factor	[22,23]
	<i>OsCOW1 (OsYUC8)</i>	3	Os03g0162000	LOC_Os03g06654	Flavin monooxygenase	[24]
	<i>CRL4/ OsGNOM1</i>	3	Os03g0666100	LOC_Os03g46330	Arf-GEF	[25,26]
	<i>OsRRI</i>	4	Os04g0442300	LOC_Os04g36070	Type-A response regulator	[27]
	<i>OsmiR393b</i>	4	-	-	MicroRNA	[16]
	<i>OsMT2b</i>	5	Os05g0111300	LOC_Os05g02070	Metallothionein	[28]
	<i>OsPIN2</i>	6	Os06g0660200	LOC_Os06g44970	Auxin efflux carrier	[29]
	<i>WOX11</i>	7	Os07g0684900	LOC_Os07g48560	WUS-related homeobox protein	[20]
	<i>CRL5</i>	7	Os07g0124700	LOC_Os07g03250	AP2/ERF transcription factor	[27]
	<i>CRL6</i>	7	Os07g0497100	LOC_Os07g31450	CHD family protein	[30]
Lateral root	<i>OsIAA3 (OsIAA31)</i>	12	Os12g0601400	LOC_Os12g40900	Aux/IAA protein	[31]
	<i>OsPID</i>	12	Os12g0614600	LOC_Os12g42020	Serine/threonine kinase	[32]
	<i>OsAUX1</i>	1	Os01g0856500	LOC_Os01g63770	Auxin influx carrier	[33]
	<i>LRL2/ OsCYP2</i>	2	Os02g0121300	LOC_Os02g02890	Cyclophilin	[34]
	<i>OsIAA11</i>	3	Os03g0633500	LOC_Os03g43400	Aux/IAA protein	[35]
	<i>OsIAA13</i>	3	Os03g0742900	LOC_Os03g53150	Aux/IAA protein	[36]
	<i>OsHO1</i>	6	Os06g0603000	LOC_Os06g40080	Heme oxygenase	[29]
	<i>OsORC3</i>	10	Os10g0402200	LOC_Os10g26280	Origin recognition complex protein	[37]

	<i>OsWOX3A/ OsNS (NAL2)</i>	11	Os11g0102100	LOC_Os11g01130	WUS-related homeobox protein	[38]
	<i>OsWOX3A/ OsNS (NAL3)</i>	12	Os12g0101600	LOC_Os12g01120	WUS-related homeobox protein	[38]
	<i>SOR1</i>	4	Os04g0101800	LOC_Os04g01160	E3 ubiquitin ligase	[39]
Root hair number	<i>OsFH1</i>	1	Os01g0897700	LOC_Os01g67240	Multidomain protein	[40]
	<i>SRH2</i>	3	Os03g0300000	LOC_Os03g18820	Xyloglucan 6-xylosyltransferase	[41]
	<i>OsRHL1</i>	6	Os06g0184000	LOC_Os06g08500	bHLH transcription factor	[42]
	<i>OsEXPA17</i>	6	Os06g0108600	LOC_Os06g01920	Expansin	[43]
	<i>RTH1/ OsAPY1</i>	7	Os07g0682800	LOC_Os07g48430	ATP diphosphohydrolase	[44]
	<i>OsSNDP1</i>	10	Os10g0122600	LOC_Os10g03400	Phosphatidylinositol transfer protein	[45]
	<i>OsEXPA30</i>	10	Os10g0535900	LOC_Os10g39110	Expansin	[43]
	<i>RTH2/ OsCSLD1</i>	10	Os10g0578200	LOC_Os10g42750	Cellulose synthase-like protein	[46,47])
	<i>QHB</i>	1	Os01g0854500	LOC_Os01g63510	WUSCHEL-type homeobox protein	[48]
	<i>DOCS1</i>	2	Os02g0236100	LOC_Os02g14120	LRR-RLK	[49]
Root meristem formation	<i>OsIAA23</i>	6	Os06g0597000	LOC_Os06g39590	Aux/IAA protein	[50]
	<i>OsSHR1</i>	7	Os07g0586900	LOC_Os07g39820	GRAS transcription factor	[48,51]
	<i>OsSCRI</i>	11	Os11g0124300	LOC_Os11g03110	GRAS transcription factor	[51,52]
	<i>OsSPRI</i>	1	Os01g0898300	LOC_Os01g67290	Armadillo-like repeat domain protein	[53]
	<i>OsEXPA8</i>	1	Os01g0248900	LOC_Os01g14650	Expansin	[54]
	<i>OsMOGS</i>	1	Os01g0921200	LOC_Os01g69210	Mannosyl-oligosaccharide glucosidase	[55]
	<i>OsCKII</i>	2	Os02g0622100	LOC_Os02g40860	Putative casein kinase I	[45]
	<i>OsCYT-INV1</i>	2	Os02g0550600	LOC_Os02g34560	Alkaline/neutral invertase	[56]
	<i>OsARF12</i>	4	Os04g0671900	LOC_Os04g57610	Auxin response factor	[57]
	<i>RT/OsGLU3</i>	4	Os04g0497200	LOC_Os04g41970	Endo-1,4- β -D-glucanase	[58,59]
Root length	<i>OsRPK1</i>	5	Os05g0486100	LOC_Os05g40770	LRR-RLK	[60]
	<i>OsDGL1</i>	7	Os07g0209000	LOC_Os07g10830	Dolichyl-diphosphooligosaccharide-protein glycosyltransferase	[61]

	<i>OsGNA1</i>	9	Os09g0488000	LOC_Os09g31310	Glucosamine-6-P acetyltransferase	[62]
Nutrient uptake related genes	<i>DRO1</i>	9	Os09g0439800	LOC_Os09g26840		[63]
	<i>PSTOL1</i>	12	Os12g0448900	LOC_Os12g26290	receptor-like cytoplasmic kinase	
	<i>OsPT1</i>	3	Os03g0150600	LOC_Os03g05620	inorganic phosphate transporter	[64]
	<i>OsPT8</i>	10	Os10g0444700	LOC_Os10g30790	inorganic phosphate transporter	[53]
	<i>OsNAR2.2</i>	4	Os04g0480200	LOC_Os04g40410	high affinity nitrate transporter	[65]
	<i>OsEXP3</i>	5	Os05g0276500	LOC_Os05g19570	Alpha-expansin-3	[66]
	<i>OsZIP4</i>	8	Os08g0207500	LOC_Os08g10630	Zrt-Irt-like protein 4, metal cation transporter	[67]
	<i>TOND1</i>	12	Os12g0630100	LOC_Os12g43440	thaumatin	[68]

References

- [1] Y. Qu, P. Mu, H. Zhang, C.Y. Chen, Y. Gao, Y. Tian, F. Wen, Z. Li, Mapping QTLs of root morphological traits at different growth stages in rice, *Genetica*. 133 (2008) 187–200. <https://doi.org/10.1007/s10709-007-9199-5>.
- [2] N. Sandhu, A. Singh, S. Dixit, M.T. Sta Cruz, P.C. Maturan, R.K. Jain, A. Kumar, Identification and mapping of stable QTL with main and epistasis effect on rice grain yield under upland drought stress, *BMC Genet.* (2014). <https://doi.org/10.1186/1471-2156-15-63>.
- [3] N. Sandhu, R.O. Torres, M.T. Sta Cruz, P.C. Maturan, R. Jain, A. Kumar, A. Henry, Traits and QTLs for development of dry direct-seeded rainfed rice varieties, *J. Exp. Bot.* (2015). <https://doi.org/10.1093/jxb/eru413>.
- [4] J. Bernier, A. Kumar, V. Ramaiah, D. Spaner, G. Atlin, A large-effect QTL for grain yield under reproductive-stage drought stress in upland rice, *Crop Sci.* 47 (2007) 507–516.
- [5] R. Venuprasad, M.E. Bool, C.O. Dalid, J. Bernier, A. Kumar, G.N. Atlin, Genetic loci responding to two cycles of divergent selection for grain yield under drought stress in a rice breeding population, *Euphytica*. 167 (2009) 261–269. <https://doi.org/10.1007/s10681-009-9898-3>.
- [6] K.H. Ghimire, L.A. Quiatchon, P. Vikram, B.P.M. Swamy, S. Dixit, H. Ahmed, J.E. Hernandez, T.H. Borromeo, A. Kumar, Identification and mapping of a QTL (qDTY1. 1) with a consistent effect on grain yield under drought, *F. Crop. Res.* 131 (2012) 88–96.
- [7] S.A. Angaji, E.M. Septiningsih, D.J. Mackill, A.M. Ismail, QTLs associated with tolerance of flooding during germination in rice (*Oryza sativa* L.), *Euphytica*. 172 (2010) 159–168. <https://doi.org/10.1007/s10681-009-0014-5>.
- [8] S. Dixit, A. Grondin, C.R. Lee, A. Henry, T.M. Olds, A. Kumar, Understanding rice adaptation to varying agro-ecosystems: Trait interactions and quantitative trait loci, *BMC Genet.* (2015). <https://doi.org/10.1186/s12863-015-0249-1>.

- [9] K.K. Mishra, P. Vikram, R.B. Yadaw, B.P.M. Swamy, S. Dixit, M.T.S. Cruz, P. Maturan, S. Marker, A. Kumar, QDTY12.1: A locus with a consistent effect on grain yield under drought in rice, *BMC Genet.* 14 (2013) 1–10. <https://doi.org/10.1186/1471-2156-14-12>.
- [10] P. Vikram, B.P.M. Swamy, S. Dixit, R. Singh, B.P. Singh, B. Miro, A. Kohli, A. Henry, N.K. Singh, A. Kumar, Drought susceptibility of modern rice varieties: An effect of linkage of drought tolerance with undesirable traits, *Sci. Rep.* (2015). <https://doi.org/10.1038/srep14799>.
- [11] A. Kumar, N. Sandhu, S. Dixit, S. Yadav, B.P.M. Swamy, N.A.A. Shamsudin, Marker-assisted selection strategy to pyramid two or more QTLs for quantitative trait-grain yield under drought, *Rice.* (2018). <https://doi.org/10.1186/s12284-018-0227-0>.
- [12] M.S. BP, H.U. Ahmed, A. Henry, R. Mauleon, S. Dixit, P. Vikram, R. Tilatto, S.B. Verulkar, P. Perraju, N.P. Mandal, Genetic, physiological, and gene expression analyses reveal that multiple QTL enhance yield of rice mega-variety IR64 under drought, *PLoS One.* 8 (2013) e62795.
- [13] L. Ge, H. Chen, J.F. Jiang, Y. Zhao, M.L. Xu, Y.Y. Xu, K.H. Tan, Z.H. Xu, K. Chong, Overexpression of OsRAA1 causes pleiotropic phenotypes in transgenic rice plants, including altered leaf, flower, and root development and root response to gravity, *Plant Physiol.* (2004). <https://doi.org/10.1104/pp.104.041996>.
- [14] Y. Yamamoto, N. Kamiya, Y. Morinaka, M. Matsuoka, T. Sazuka, Auxin biosynthesis by the YUCCA genes in rice, *Plant Physiol.* (2007). <https://doi.org/10.1104/pp.106.091561>.
- [15] Q. Zhang, J. Li, W. Zhang, S. Yan, R. Wang, J. Zhao, Y. Li, Z. Qi, Z. Sun, Z. Zhu, The putative auxin efflux carrier OsPIN3t is involved in the drought stress response and drought tolerance, *Plant J.* 72 (2012) 805–816.
- [16] H. Bian, Y. Xie, F. Guo, N. Han, S. Ma, Z. Zeng, J. Wang, Y. Yang, M. Zhu, Distinctive expression patterns and roles of the miRNA393/TIR1 homolog module in regulating flag leaf inclination and primary and crown root growth in rice (*Oryza sativa*), *New Phytol.* 196 (2012) 149–161. <https://doi.org/10.1111/j.1469-8137.2012.04248.x>.
- [17] T. Yoshikawa, M. Ito, T. Sumikura, A. Nakayama, T. Nishimura, H. Kitano, I. Yamaguchi, T. Koshiba, K. Hibara, Y. Nagato, The rice FISH BONE gene encodes a tryptophan aminotransferase, which affects pleiotropic auxin-related processes, *Plant J.* 78 (2014) 927–936.
- [18] Y. Zhao, S. Cheng, Y. Song, Y. Huang, S. Zhou, X. Liu, D.-X. Zhou, The interaction between rice ERF3 and WOX11 promotes crown root development by regulating gene expression involved in cytokinin signaling, *Plant Cell.* 27 (2015) 2469–2483.
- [19] M. Xu, L. Zhu, H. Shou, P. Wu, A PIN1 family gene, OsPIN1, involved in auxin-dependent adventitious root emergence and tillering in rice, *Plant Cell Physiol.* (2005). <https://doi.org/10.1093/pcp/pci183>.
- [20] Y. Zhao, Y. Hu, M. Dai, L. Huang, D.X. Zhou, The WUSCHEL-Related homeobox gene WOX11 is required to activate shoot-borne crown root development in rice, *Plant Cell.* (2009). <https://doi.org/10.1105/tpc.108.061655>.
- [21] X.F. Wang, F.F. He, X.X. Ma, C.Z. Mao, C. Hodgman, C.G. Lu, P. Wu, OsCAND1 is required for crown root emergence in rice, *Mol. Plant.* (2011). <https://doi.org/10.1093/mp/ssq068>.
- [22] Y. Inukai, T. Sakamoto, M. Ueguchi-Tanaka, Y. Shibata, K. Gomi, I. Umemura, Y. Hasegawa, M. Ashikari, H. Kitano, M. Matsuoka, Crown rootless1, which is essential for crown root formation in rice, is a target of an Auxin Response Factor in auxin signaling, *Plant Cell.* (2005). <https://doi.org/10.1105/tpc.105.030981>.

- [23] H. Liu, S. Wang, X. Yu, J. Yu, X. He, S. Zhang, H. Shou, P. Wu, ARL1, a LOB-domain protein required for adventitious root formation in rice, *Plant J.* (2005). <https://doi.org/10.1111/j.1365-313X.2005.02434.x>.
- [24] Y.M. Woo, H.J. Park, M. Su'udi, J. Il Yang, J.J. Park, K. Back, Y.M. Park, G. An, Constitutively wilted 1, a member of the rice YUCCA gene family, is required for maintaining water homeostasis and an appropriate root to shoot ratio, *Plant Mol. Biol.* (2007). <https://doi.org/10.1007/s11103-007-9203-6>.
- [25] Y. Kitomi, A. Ogawa, H. Kitano, Y. Inukai, CRL4 regulates crown root formation through auxin transport in rice, *Plant Root.* 2 (2008) 19–28. <https://doi.org/10.3117/plantroot.2.19>.
- [26] S. Liu, J. Wang, L. Wang, X. Wang, Y. Xue, P. Wu, H. Shou, Adventitious root formation in rice requires OsGNOM1 and is mediated by the OsPINs family, *Cell Res.* 19 (2009) 1110–1119.
- [27] Y. Kitomi, H. Ito, T. Hobo, K. Aya, H. Kitano, Y. Inukai, The auxin responsive AP2/ERF transcription factor CROWN ROOTLESS5 is involved in crown root initiation in rice through the induction of OsRR1, a type-A response regulator of cytokinin signaling, *Plant J.* 67 (2011) 472–484. <https://doi.org/10.1111/j.1365-313X.2011.04610.x>.
- [28] J. Yuan, D. Chen, Y. Ren, X. Zhang, J. Zhao, Characteristic and expression analysis of a metallothionein gene, OsMT2b, down-regulated by cytokinin suggests functions in root development and seed embryo germination of rice, *Plant Physiol.* (2008). <https://doi.org/10.1104/pp.107.110304>.
- [29] Y.H. Chen, Y.Y. Chao, Y.Y. Hsu, C.Y. Hong, C.H. Kao, Heme oxygenase is involved in nitric oxide- and auxin-induced lateral root formation in rice, *Plant Cell Rep.* (2012). <https://doi.org/10.1007/s00299-012-1228-x>.
- [30] Y. Wang, D. Wang, T. Gan, L. Liu, W. Long, Y. Wang, M. Niu, X. Li, M. Zheng, L. Jiang, J. Wan, CRL6, a member of the CHD protein family, is required for crown root development in rice, *Plant Physiol. Biochem.* 105 (2016) 185–194. <https://doi.org/10.1016/j.plaphy.2016.04.022>.
- [31] A. Nakamura, I. Umemura, K. Gomi, Y. Hasegawa, H. Kitano, T. Sazuka, M. Matsuoka, Production and characterization of auxin-insensitive rice by overexpression of a mutagenized rice IAA protein, *Plant J.* 46 (2006) 297–306. <https://doi.org/10.1111/j.1365-313X.2006.02693.x>.
- [32] Y. Morita, J. Kyozuka, Characterization of OsPID, the rice ortholog of PINOID, and its possible involvement in the control of polar auxin transport, *Plant Cell Physiol.* 48 (2007) 540–549. <https://doi.org/10.1093/pcp/pcm024>.
- [33] H. Zhao, T. Ma, X. Wang, Y. Deng, H. Ma, R. Zhang, J. Zhao, OsAUX1 controls lateral root initiation in rice (*Oryza sativa*L.), *Plant Cell Environ.* (2015). <https://doi.org/10.1111/pce.12467>.
- [34] B. Kang, Z. Zhang, L. Wang, L. Zheng, W. Mao, M. Li, Y. Wu, P. Wu, X. Mo, OsCYP2, a chaperone involved in degradation of auxin-responsive proteins, plays crucial roles in rice lateral root initiation, *Plant J.* 74 (2013) 86–97. <https://doi.org/10.1111/tpj.12106>.
- [35] Z.X. Zhu, Y. Liu, S.J. Liu, C.Z. Mao, Y.R. Wu, P. Wu, A gain-of-function mutation in OsIAA11 affects lateral root development in rice, *Mol. Plant.* 5 (2012) 154–161. <https://doi.org/10.1093/mp/ssr074>.
- [36] Y. Kitomi, H. Inahashi, H. Takehisa, Y. Sato, Y. Inukai, OsIAA13-mediated auxin signaling is involved in lateral root initiation in rice, *Plant Sci.* 190 (2012) 116–122. <https://doi.org/10.1016/j.plantsci.2012.04.005>.
- [37] X. Chen, J. Shi, X. Hao, H. Liu, J. Shi, Y. Wu, Z. Wu, M. Chen, P. Wu, C. Mao, OsORC3 is required for lateral root development in rice, *Plant J.* 74 (2013)

- 339–350. <https://doi.org/10.1111/tpj.12126>.
- [38] S.H. Cho, S.C. Yoo, H. Zhang, D. Pandeya, H.J. Koh, J.Y. Hwang, G.T. Kim, N.C. Paek, The rice narrow leaf2 and narrow leaf3 loci encode WUSCHEL-related homeobox 3A (OsWOX3A) and function in leaf, spikelet, tiller and lateral root development, *New Phytol.* (2013). <https://doi.org/10.1111/nph.12231>.
- [39] Y. Uga, E. Hanzawa, S. Nagai, K. Sasaki, M. Yano, T. Sato, Identification of qSOR1, a major rice QTL involved in soil-surface rooting in paddy fields, *Theor. Appl. Genet.* (2012). <https://doi.org/10.1007/s00122-011-1688-3>.
- [40] J. Huang, C.M. Kim, Y.H. Xuan, J. Liu, T.H. Kim, B.K. Kim, C.D. Han, Formin homology 1 (OsFH1) regulates root-hair elongation in rice (*Oryza sativa*), *Planta.* 237 (2013) 1227–1239. <https://doi.org/10.1007/s00425-013-1838-8>.
- [41] C. Wang, S. Li, S. Ng, B. Zhang, Y. Zhou, J. Whelan, P. Wu, H. Shou, Mutation in xyloglucan 6-xylosyltransferase results in abnormal root hair development in *Oryza sativa*, *J. Exp. Bot.* (2014). <https://doi.org/10.1093/jxb/eru189>.
- [42] W. Ding, Z. Yu, Y. Tong, W. Huang, H. Chen, P. Wu, A transcription factor with a bHLH domain regulates root hair development in rice, *Cell Res.* (2009). <https://doi.org/10.1038/cr.2009.109>.
- [43] Z. Yu, B. Kang, X. He, S. Lv, Y. Bai, W. Ding, M. Chen, H.T. Cho, P. Wu, Root hair-specific expansins modulate root hair elongation in rice, *Plant J.* (2011). <https://doi.org/10.1111/j.1365-313X.2011.04533.x>.
- [44] T. Yuo, M. Toyota, M. Ichii, S. Taketa, Molecular cloning of a root hairless gene rth1 in rice, *Breed. Sci.* 59 (2009) 13–20. <https://doi.org/10.1270/jsbbs.59.13>.
- [45] J. Huang, C.M. Kim, Y.H. Xuan, S.J. Park, H.L. Piao, B. Il Je, J. Liu, T.H. Kim, B.K. Kim, C.D. Han, OsSNDP1, a Sec14-nodulin domain-containing protein, plays a critical role in root hair elongation in rice, *Plant Mol. Biol.* 82 (2013) 39–50. <https://doi.org/10.1007/s11103-013-0033-4>.
- [46] M.K. Chul, H.P. Sung, I.J. Byoung, H.P. Su, J.P. Soon, L.P. Hai, Y.E. Moo, L. Dolan, C.D. Han, OsCSLD1, a cellulose synthase-like D1 gene, is required for root hair morphogenesis in rice, *Plant Physiol.* (2007). <https://doi.org/10.1104/pp.106.091546>.
- [47] T. Yuo, K. Shiotani, N. Shitsukawa, A. Miyao, H. Hirochika, M. Ichii, S. Taketa, Root hairless 2 (RTH2) mutant represents a loss-of-function allele of the cellulose synthase-like gene OSCSLD1 in rice (*Oryza sativa* L.), *Breed. Sci.* (2011). <https://doi.org/10.1270/jsbbs.61.225>.
- [48] N. Kamiya, H. Nagasaki, A. Morikami, Y. Sato, M. Matsuoka, Isolation and characterization of a rice WUSCHEL-type homeobox gene that is specifically expressed in the central cells of a quiescent center in the root apical meristem, *Plant J.* 35 (2003) 429–441. <https://doi.org/10.1046/j.1365-313X.2003.01816.x>.
- [49] S. Sun, M. Gu, Y. Cao, X. Huang, X. Zhang, P. Ai, J. Zhao, X. Fan, G. Xu, A constitutive expressed phosphate transporter, OsPht1;1, modulates phosphate uptake and translocation in phosphate-replete rice, *Plant Physiol.* (2012). <https://doi.org/10.1104/pp.112.196345>.
- [50] J. Ni, G. Wang, Z. Zhu, H. Zhang, Y. Wu, P. Wu, OsIAA23-mediated auxin signaling defines postembryonic maintenance of QC in rice, *Plant J.* (2011). <https://doi.org/10.1111/j.1365-313X.2011.04698.x>.
- [51] H. Cui, M.P. Levesque, T. Vernoux, J.W. Jung, A.J. Paquette, K.L. Gallagher, J.Y. Wang, I. Blilou, B. Scheres, P.N. Benfey, An evolutionarily conserved

- mechanism delimiting SHR movement defines a single layer of endodermis in plants, *Science* (80-.). 316 (2007) 421–425. <https://doi.org/10.1126/science.1139531>.
- [52] N. Kamiya, J. Itoh, A. Morikami, Y. Nagato, M. Matsuoka, The SCARECROW gene's role in asymmetric cell divisions in rice plants, *Plant J.* 36 (2003) 45–54.
- [53] H. Jia, H. Ren, M. Gu, J. Zhao, S. Sun, X. Zhang, J. Chen, P. Wu, G. Xu, The phosphate transporter gene *ospt1;8* is involved in phosphate homeostasis in rice, *Plant Physiol.* (2011). <https://doi.org/10.1104/pp.111.175240>.
- [54] S. Ray, J. Vijayan, R.K. Sarkar, Germination stage oxygen deficiency (GSOD): An emerging stress in the era of changing trends in climate and rice cultivation practice, *Front. Plant Sci.* 7 (2016) 1–4. <https://doi.org/10.3389/fpls.2016.00671>.
- [55] Jaiswal, 基因的改变NIH Public Access, *Bone*. 23 (2014) 1–7. <https://doi.org/10.1111/tpj.12497.OsMOGS>.
- [56] L. Jia, B. Zhang, C. Mao, J. Li, Y. Wu, P. Wu, Z. Wu, OsCYT-INV1 for alkaline/neutral invertase is involved in root cell development and reproductivity in rice (*Oryza sativa* L.), *Planta*. 228 (2008) 51–59. <https://doi.org/10.1007/s00425-008-0718-0>.
- [57] Y. Qi, S. Wang, C. Shen, S. Zhang, Y. Chen, Y. Xu, Y. Liu, Y. Wu, D. Jiang, OsARF12, a transcription activator on auxin response gene, regulates root elongation and affects iron accumulation in rice (*Oryza sativa*), *New Phytol.* 193 (2012) 109–120.
- [58] Y. Inukai, T. Sakamoto, Y. Morinaka, M. Miwa, M. Kojima, E. Tanimoto, H. Yamamoto, K. Sato, Y. Katayama, M. Matsuoka, H. Kitano, ROOT GROWTH INHIBITING, a Rice Endo-1,4- β -d-Glucanase, Regulates Cell Wall Loosening and is Essential for Root Elongation, *J. Plant Growth Regul.* (2012). <https://doi.org/10.1007/s00344-011-9247-3>.
- [59] J.W. Zhang, L. Xu, Y.R. Wu, X.A. Chen, Y. Liu, S.H. Zhu, W.N. Ding, P. Wu, K.K. Yi, OsGLU3, a putative membrane-bound endo-1,4-beta-glucanase, is required for root cell elongation and division in rice (*Oryza sativa* L.), *Mol. Plant.* (2012). <https://doi.org/10.1093/mp/ssr084>.
- [60] Y. Zou, X. Liu, Q. Wang, Y. Chen, C. Liu, Y. Qiu, W. Zhang, OsRPK1, a novel leucine-rich repeat receptor-like kinase, negatively regulates polar auxin transport and root development in rice, *Biochim. Biophys. Acta - Gen. Subj.* 1840 (2014) 1676–1685. <https://doi.org/10.1016/j.bbagen.2014.01.003>.
- [61] C. Qin, Y. Li, J. Gan, W. Wang, H. Zhang, Y. Liu, P. Wu, OsDGL1, a homolog of an oligosaccharyltransferase complex subunit, is involved in N-glycosylation and root development in rice, *Plant Cell Physiol.* 54 (2013) 129–137. <https://doi.org/10.1093/pcp/pcs159>.
- [62] H. Jiang, S. Wang, L. Dang, S. Wang, H. Chen, Y. Wu, X. Jiang, P. Wu, A novel short-root gene encodes a glucosamine-6-phosphate acetyltransferase required for maintaining normal root cell shape in rice, *Plant Physiol.* 138 (2005) 232–242. <https://doi.org/10.1104/pp.104.058248>.
- [63] Y. Uga, K. Sugimoto, S. Ogawa, J. Rane, M. Ishitani, N. Hara, Y. Kitomi, Y. Inukai, K. Ono, N. Kanno, H. Inoue, H. Takehisa, R. Motoyama, Y. Nagamura, J. Wu, T. Matsumoto, T. Takai, K. Okuno, M. Yano, Control of root system architecture by DEEPER ROOTING 1 increases rice yield under drought conditions, *Nat. Genet.* (2013). <https://doi.org/10.1038/ng.2725>.
- [64] H.M. Seo, Y. Jung, S. Song, Y. Kim, T. Kwon, D.H. Kim, S.J. Jeung, Y.B. Yi, G. Yi, M.H. Nam, J. Nam, Increased expression of OsPT1, a high-affinity phosphate transporter, enhances phosphate acquisition in rice, *Biotechnol. Lett.* (2008). <https://doi.org/10.1007/s10529-008-9757-7>.
- [65] M. Yan, X. Fan, H. Feng, A.J. Miller, Q. Shen, G. Xu, Rice OsNAR2.1 interacts with OsNRT2.1, OsNRT2.2 and OsNRT2.3a nitrate transporters to provide

- uptake over high and low concentration ranges, *Plant, Cell Environ.* 34 (2011) 1360–1372. <https://doi.org/10.1111/j.1365-3040.2011.02335.x>.
- [66] S. Qiu, N. Ma, S. Che, Y. Wang, X. Peng, G. Zhang, G. Wang, J. Huang, Repression of OsEXPA3 expression leads to root system growth suppression in rice, *Crop Sci.* (2014). <https://doi.org/10.2135/cropsci2013.11.0746>.
- [67] S.A. Ramesh, R. Shin, D.J. Eide, D.P. Schachtman, Differential metal selectivity and gene expression of two zinc transporters from rice, *Plant Physiol.* (2003). <https://doi.org/10.1104/pp.103.026815>.
- [68] Y. Zhang, L. Tan, Z. Zhu, L. Yuan, D. Xie, C. Sun, TOND1 confers tolerance to nitrogen deficiency in rice, *Plant J.* 81 (2015) 367–376. <https://doi.org/10.1111/tpj.12736>.