Table S1. Primers used in this study.

Primers	Sequence (5'→3')	Product of PCR amplicon (bp)	Description
For PCR			
WFB1	GACCAGCCACACTGGGAC	360	Specific-primers of *P. citrulli* (Walcott et al. 2007)
WFB2	CTGCCGCACTCCAGCGA		
Km^r-F	TGCTCTGATGCCGCCGTGTT	634	Amplification of Km resistance gene
Km^r-R	CGAGGAAGCGGTCAGCCCA		
18F	CAGGAAACAGCTATGAC	-	pK18*mobsacB* sequencing primers
18R	GTAAAACGACGGCCAGT		
F-*hrpG*-S	CGCTCTAGAACTAGTGGATCCCGATGCGTTGGGAGGAGA	1336	Amplification of *hrpG*
F-*hrpG*-A	GGTAAGCTTGATATCGAATTCACTGCCCAGGGGCGGCAT		
D46A-S	CCGCGGTCATCCTGGAAGACT	-	The aspartic acid 46 of HrpG was replaced with alanine
D46A-A	CGGGCGGATGCCGGCGGGCC		
D52A-S	CCTGCGGCGGTACGCTGGCCG	-	The aspartic acid 52 of HrpG was replaced with alanine
D52A-A	CTTCCAGGATGACCGCGTCG		
D60A-S	CTCTGGGCCTGATCCGAAGCT	-	The aspartic acid 60 of HrpG was replaced with alanine
D60A-A	CCTCGGCCAGCGTACCGCCG		
dHrpG-S	CTATGACATGATTACGAATTCGAACCGAAGCGAGCACCA	1195	Amplification point mutant sequence of *hrpG*
dHrpG-A	ACGACGGCCAGTGCCAAGCTTGGAATGCATGGCGGGAAG		
For qPCR*			
hrpG-F	GGGAGGCATTCAAGCCATCT	104	Amplification of *Aave_0445* fragment
hrpG-R	GCGTACCGCCGCAGTCT		
hrpX-F	CTGTGCGAATCGGGGGA	113	Amplification of *Aave_0444* fragment
hrpX-R	AAGGCGTGGTCCGACAGTT		
rpoB-1	GCGACAGCGTGCTCAAAGTG	134	Amplification of control gene *rpoB* fragment
rpoB-2	GCCTTCGTTGGTGCGTTTCT		

* Primer dry powder for qPCR is dissolved in enzyme-free water.