Table S1. Primer sequences used for qRT–PCR analysis of hypertrophic markers and CELF

Gene Name	Forward Primer (5′-3′)	Reverse Primer (5′-3′)
Rat-Gapdh	AGGTCGGTGTGAACGGATTTG	TGTAGACCATGTAGTTGAGGTCA
Rat-Nppa	CAACACAGATCTGATGGATTTCA	CCTCATCTTTACCGGCATC
Rat-Nppb	GTCTCAAGACAGCGCCTTCC	AACCTCAGCCCGTCAC AGC
Rat-Myh7	GCGGACATTGCCGAGTCCCAG	GCTCCAGGTCTCAGGGCTTCACA
Rat-Rcan1.4	GTGGATGGGTGTGTACTCTGG	TGGTCCACGTGTGTG AGAGT
Rat-CELF1	GTTGGCTCATTGGCAGGAATG	GATACCAGAATACGCCTGGGT

Table S2. Primer sequences used for RIP-qPCR

Gene Name	Forward Primer (5′-3′)	Reverse Primer (5′-3′)
Tead1	CAGCGCCCCGAAGTTTG	ATCCTCTCCATGTTGTCGGC
Fhod3	CAGCAGCAACTCTTTCCAATCC	CCTCCTCTGAATTGGGCTCC
R3hdm2	CACAAAATGTGGTCCAGCC	GAAGGTGACACGCCTGAGTA
Rab30	TCAAGGAGCCACAATTGGAGT	GGTCAATCTTGTTGCCCAC
Myh14	TCAGCACCATGTCTTATGGGG	ATTCGTCCTTGGCCTGTCTG
Ubn1	CATCGTCTACCTCCTTGCCA	GCCAGAAGACTGTGGCTG
Mbnl1	GGACGAGTAATCGCCTGCTT	AACTGAAAACATTGGCACGGG

Table S3. Primer sequences used for Tead1 alternative splicing RT–PCR

Gene Name	Forward Primer (5′-3′)	Reverse Primer (5′-3′)
Tead1-alternative splicing	GTGTCTAGTCACATTCAGGT	CCCCGGGCCCCCTGGGAAGG

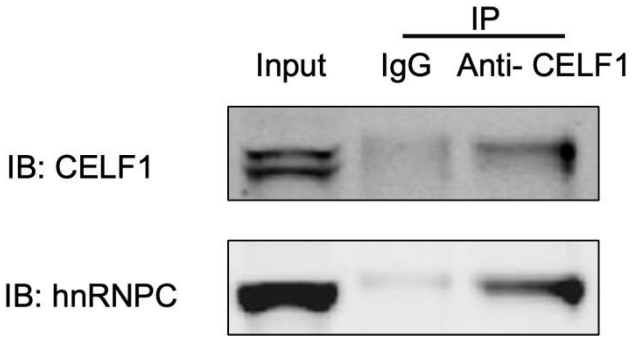


Figure S1. Endogenous interaction between CELF1 and hnRNPC in NRCMs.
Endogenous immunoprecipitation was performed in neonatal rat cardiomyocytes (NRCMs). Immunoprecipitation of endogenous CELF1 from NRCM lysates resulted in the co-precipitation of endogenous hnRNPC, supporting their association under physiological conditions.