

Supp. Table S1: classification of the detected variants in *ABCA12*

<b>Mutation</b>		<b>gnomAD</b>	<b>ClinVar</b>	<b>Splice prediction tools</b>	<b>ACMG classification</b>	<b>literature</b>
c.130C>T	p.(Arg44Trp)	0x homo 8x het MAF 0.003%	2x likely pathogenic	-	pathogenic (PM2, PM5, PP3, PP5)	Scott <i>et al.</i> (2013)
c.596G>A	p.(Trp199*)	n/a	1x pathogenic	-	pathogenic (PVS1, PM2, PM3, PP5)	Thomas <i>et al.</i> (2006)
<b>c.1270G&gt;T</b>	<b>p.(Glu424*)</b>	n/a	n/a	-	pathogenic (PVS1, PM2)	<b>this study</b>
<b>c.2864-6T&gt;A, p.?</b>	<b>p.?</b>	0x homo 16x het MAF 0.006%	n/a	reduction of score of wt acceptor splice site from 0.90 to 0.42 (NN) and from 0.63 to 0.23 (NG2)	uncertain significance (PM2, PM3)	<b>this study</b>
<b>c.2864-2A&gt;T, p.?</b>	<b>p.?</b>	n/a	n/a	NN, NG2: loss of wt acceptor splice site	pathogenic (PVS1, PM2)	<b>this study</b>
c.3809A>G	p.(Tyr1270Cys)	n/a	n/a	-	likely pathogenic (PM1, PM2, PM3, PP3)	Ennouri <i>et al.</i> (2022)
c.4139A>G	p.(Asn1380Ser)	0x homo 5x het MAF 0.002%	2x likely pathogenic, 5x pathogenic	-	pathogenic (PM1, PM2, PP3, PP5)	Lefèvre <i>et al.</i> (2003)
c.4544G>A	p.(Arg1515Gln)	0x homo 3x het MAF 0.001%	n/a	-	pathogenic (PM1, PM2, PP3, PP5)	Israeli <i>et al.</i> (2013)
<b>c.6611G&gt;A</b>	<b>p.(Arg2204Gln)</b>	0x homo 89x het MAF 0.03%	1x likely benign, 1x uncertain significance	-	likely pathogenic (PM1, PM2, PM3, PP3)	<b>this study</b>
c.6852G>C	p.(Glu2284Asp)	0x homo 30x het MAF 0.01%	n/a	-	pathogenic (PM1, PM2, PM3, PP3, PP5)	Bastaki <i>et al.</i> (2017), Ennouri <i>et al.</i> (2022)
c.6962+1G>A	p.?	0x homo 10x het MAF 0.004%	n/a	NN, NG2: loss of wt donor splice site	pathogenic (PVS1, PM2, PP5)	Hou <i>et al.</i> (2020)

n/a: not listed in the database. Homozygous. Het: heterozygous. MAF: minor allele frequency, NN: NNSplice. NG2: NetGene2. wt: wild-type. -: not evaluated.

gnomAD: The Genome Aggregation Database version v2.1.1 (<http://gnomad.broadinstitute.org/>). The data set provided on this website spans 125,748 exome sequences and 15,708 whole-genome sequences from unrelated individuals sequenced as part of various disease-specific and population genetic studies.ClinVar: Version september 2023 (<https://www.ncbi.nlm.nih.gov/clinvar/>).

NNSplice: Splice site predictions for 1 sequence with donor score cutoff 0.40, acceptor score cutoff 0.40. Score ranging from 0 to 1, higher score implies a more potential splice site.

NetGene2: Cutoff values used for confidence: highly confident donor sites (H): 95.0 %, nearly all true donor sites: 50.0 %, highly confident acceptor sites (H): 95.0 %, nearly all true acceptor sites: 20.0 %. Confidence score ranging from 0 to 1, higher score implies a higher confidence of true site.

ACMG: American College of Medical Genetics and Genomics, Richards *et al.* (2015).

Scott *et al.* Targeted sequence capture and high-throughput sequencing in the molecular diagnosis of ichthyosis and other skin diseases. *J Invest Dermatol.* 2013 Feb;133(2):573-6.

Thomas *et al.* ABCA12 is the major harlequin ichthyosis gene. *J Invest Dermatol.* 2006 Nov;126(11):2408-13.

Ennouri *et al.* Clinical and genetic investigation of ichthyosis in familial and sporadic cases in south of Tunisia: genotype-phenotype correlation. *BMC Med Genomics.* 2022 Jan 5;15(1):4.

Lefévre *et al.* Mutations in the transporter ABCA12 are associated with lamellar ichthyosis type 2. *Hum Mol Genet.* 2003 Sep 15;12(18):2369-78.

Israeli *et al.* Non-syndromic autosomal recessive congenital ichthyosis in the Israeli population.

Bastaki *et al.* Summary of mutations underlying autosomal recessive congenital ichthyoses (ARCI) in Arabs with four novel mutations in ARCI-related genes from the United Arab Emirates. *Int J Dermatol.* 2017 May;56(5):514-523.

Hou *et al.* Precision medicine integrating whole-genome sequencing, comprehensive metabolomics, and advanced imaging. *Proc Natl Acad Sci U S A.* 2020 Feb 11;117(6):3053-3062.