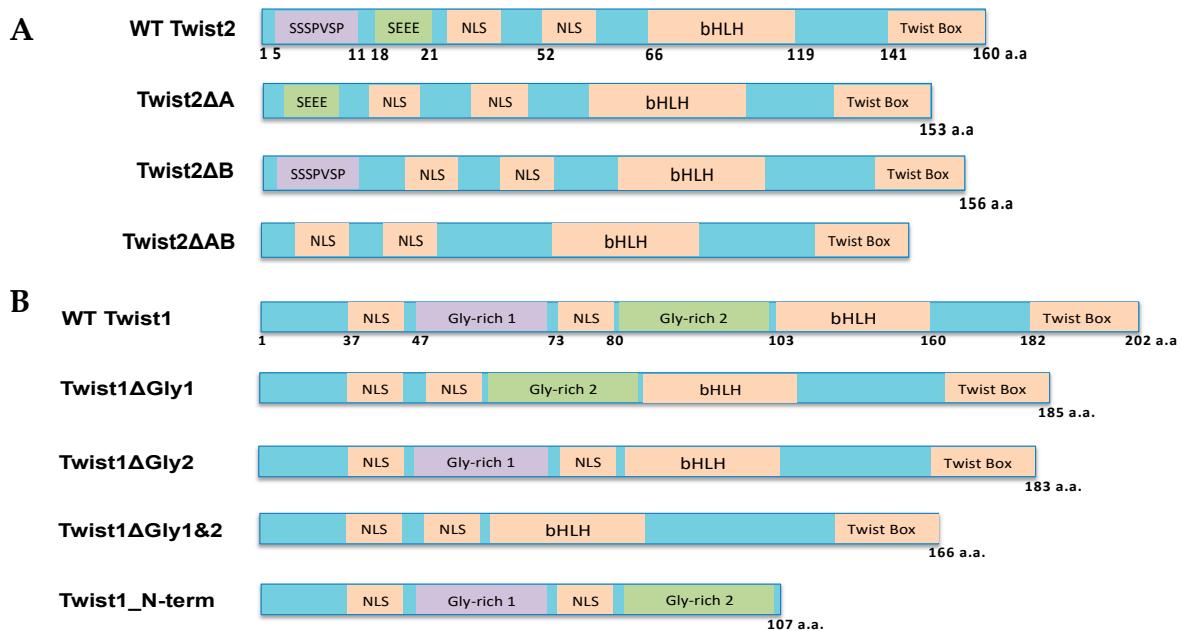


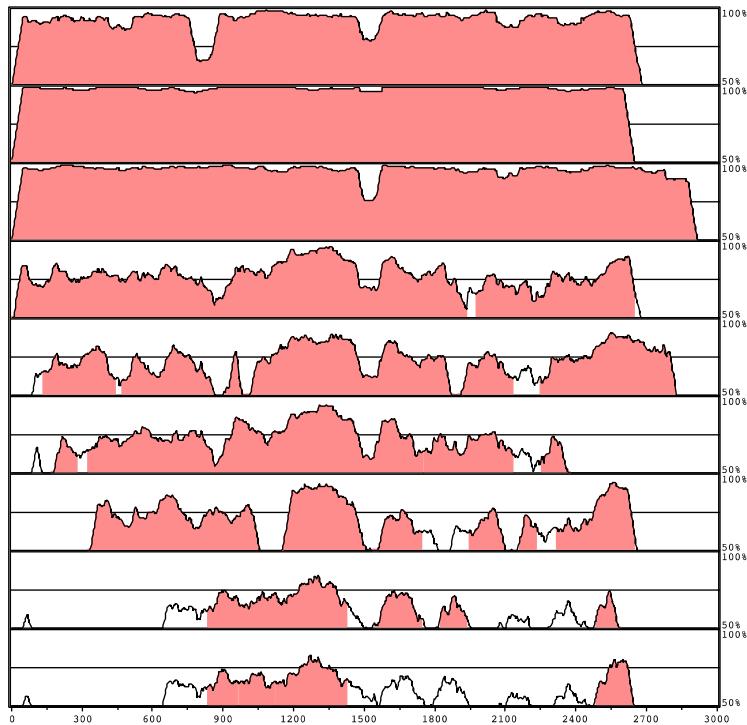
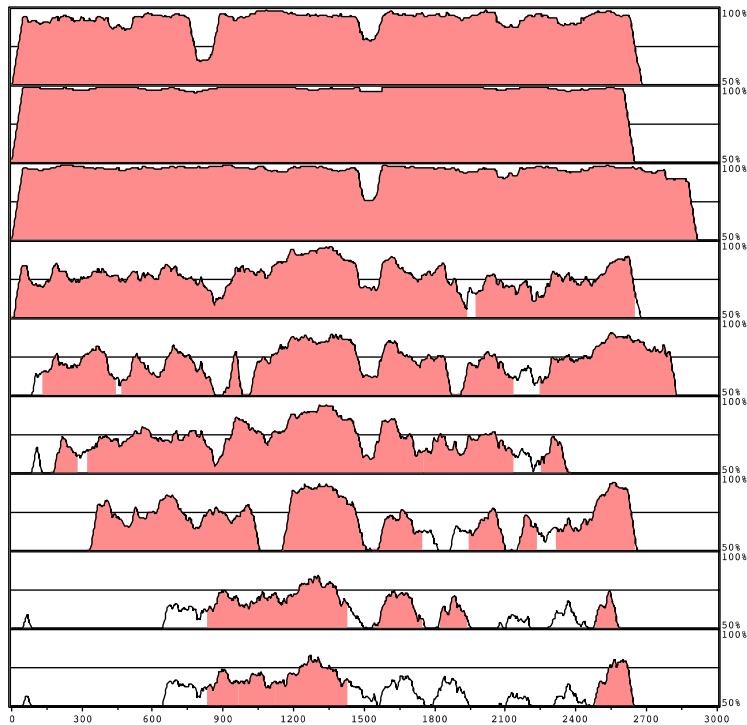
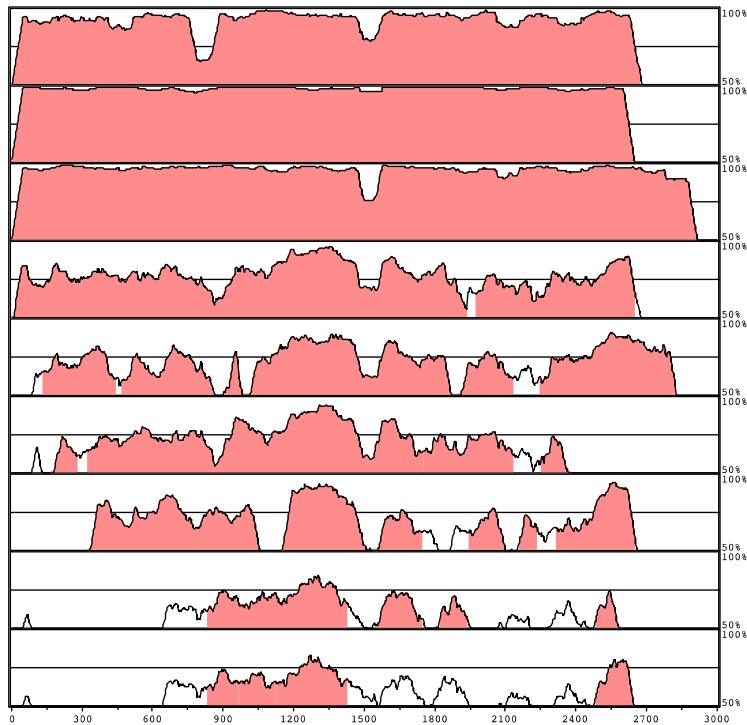
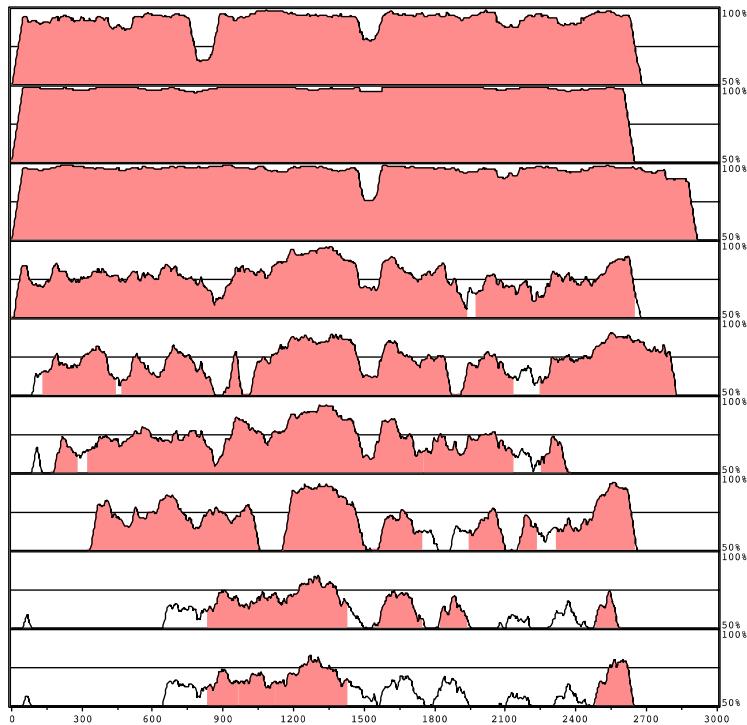
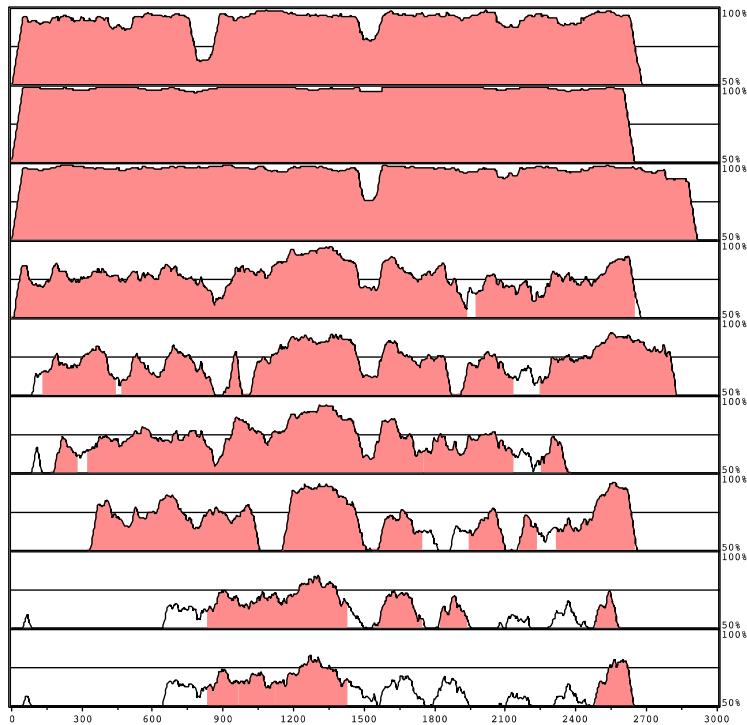
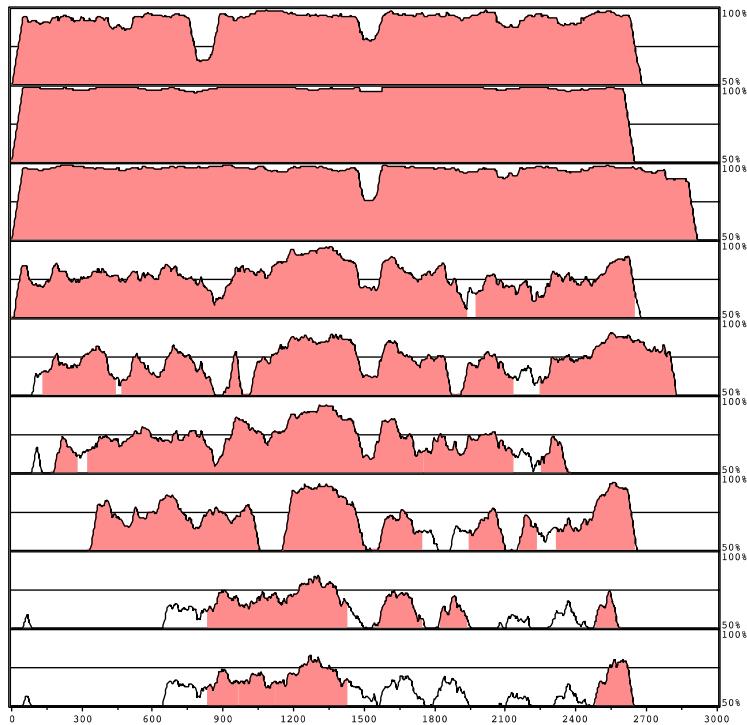
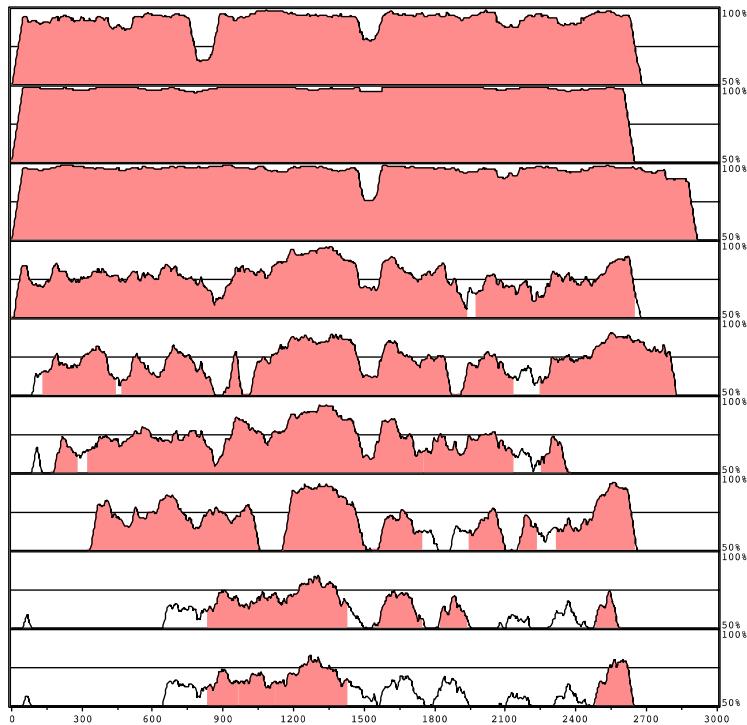
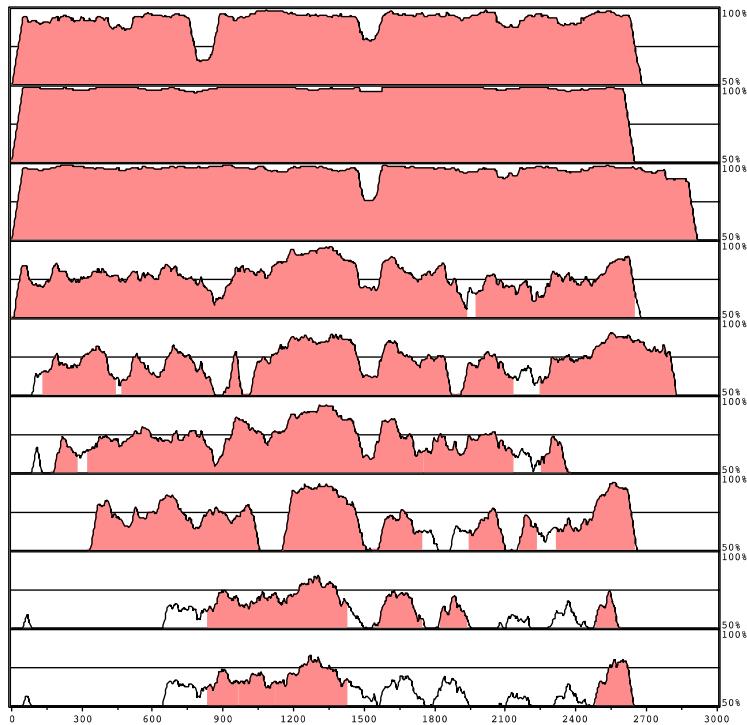
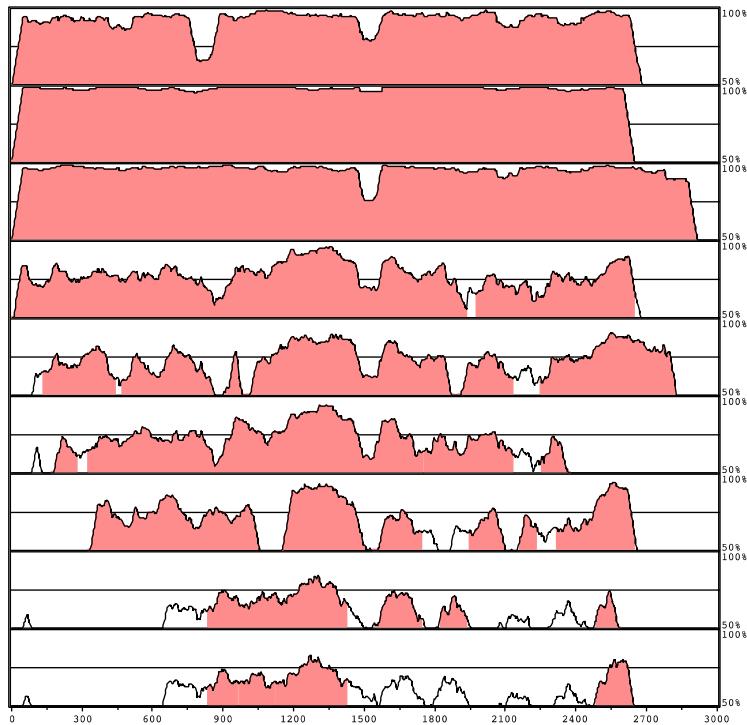
**Supplementary Table S1.** Sequences of Primers used in this study for deletion of conserved regions in the TWIST2 coding sequence, amplification of the human *CHRD1* gene upstream region and of the *TCF3* gene coding region for production of an expression construct. The primers used to amplify the human *CHRD1* gene sequences for reporter gene assays contained additional nucleotides to add KpnI and XhoI sites at the ends of the PCR product for directional cloning into the pGL4.14 vector. Sequences of the oligonucleotides used for competition reactions in electrophoretic mobility shift assays are also included, these oligonucleotides were used as double stranded molecules. E-box and SP1 elements (normal or altered) are shown in italics. E-box bases changed are underlined.

Name of Primers/Oligo-Nucleotide	Sequence	Size	Application
Twist2ΔA	FW: 5'-CGATGAGGAGGGCGTGGACAGCCTGG-3'  RV: 5'-CCAGGCTGTCCACGCCCTCCTCATCG-3'	26 bases	Deletion Mutagenesis of first sub-motif SSSPVSP
Twist2ΔB	FW: 5'-CAGCCTGGCACCCCTCGAGAGGCAGC-3'  RV 5'-GCTGCCTCTCGAGGGTGCCAGGCTG-3'	26 bases	Deletion Mutagenesis of second sub-motif SEEE
CH3KLuc-FW	5'- <u>ACGGTACC</u> GGAAAGGGAAAAGATGGGTGT-3'  Includes KpnI site (underlined) for cloning	28 bases	PCR forward primer used for Preparation of -3K <i>CHRD1</i> -pGL4 Luc construct
CH3KLuc-RV	5'- <u>ACTCGAGT</u> GCTCACTAACCTGGGCACT-3'  Includes XhoI site (underlined) for cloning	27 bases	PCR reverse primer used for Preparation of -3K <i>CHRD1</i> -pGL4 Luc construct
TCF3FW	5'-ATGAACCAGCCGCAGAGGATGG-3'	22 bases	PCR forward primer used for preparation of TCF3 expression construct in pCRII-TOPO vector
TCF3RV	5'-CGGAGGCATACCTTCACAT-3'	20 bases	PCR reverse primer used for preparation of TCF3 expression construct in pCRII-TOPO vector
SP1 oligo	5'- ATTCGATCGGGCGGGCGAGC-3' 3'-TAAGCTAGCCCCGCCCGCTCG-5'	22 base pairs	Non-specific competitor
CH2600TWI	5'-TATATACACAGGCAAATGAGTGCATATAAA-3' 3'-ATATATGTGTCGTTACTCACGTATATT-5'	30 base pairs	EMSA probe

CH2700EA	5'-TGGTGGTGGTCACATGAATATAACACATGCGATAAAATT-3' 3'-ACCACCACCA <u>GAGTGTACTTATATGTGACGCTATTTAA</u> -5'	38 base pairs	Specific competitor
CH2700EMAW	5'-TGGTGGTGGTC <u>CGCACGAATATAACACATGCGATAAAATT-3'</u> 3'-ACCACCACCA <u>GCGTGTACTTATATGTGACGCTATTTAA</u> -5'	38 base pairs	Competitor
CH2700EWAM	5'-TGGTGGTGGTCACATGAATATA <u>ACGCA<u>CGCGATAAAATT-3'</u></u> 3'-ACCACCACCA <u>GCGTGTACTTATATGCGTGC<u>GCTATTTAA</u>-5'</u>	38 base pairs	Competitor



**Supplementary Figure S1. Diagram of the TWIST2 and TWIST1 N-terminal mutant proteins produced by deletion mutagenesis.** A. We generated mutant TWIST2 expression constructs where we removed two conserved sub-motifs [43], which were predicted to be associated with protein binding, the first sub-motif SSSPVSP (construct TWIST2 $\Delta$ A), the second sub-motif SEEE (construct TWIST2 $\Delta$ B), or both sub-motifs (construct TWIST2 $\Delta$ AB). B. Diagram of TWIST1 glycine-rich deletion mutant proteins and the N-Terminal construct used for EMSAs, which were generated by gene synthesis. NLS = nuclear localization signal; bHLH = basic helix loop helix; TWIST box = protein domain which interacts with the Runx2 DNA binding domain to inhibit its function [9].

**A****Macaque****Chimpanzee****Orangutan****Horse****Dog****Pig****Squirrel****Mouse****Rat**

B.

<b>-1239Twist</b>						
Human	000002058	AACTC-TAACACTT-----	GAGACCTCAGGTAGTT----	CGGTTGGTT	000002095	
Macaque	000002144	AACTCTAACACTT-----	GAGACCTCAGGTAGTT----	CGGTTGGTT	000002182	
Chimpanzee	000002112	AACTCTAACACTT-----	GAGACCTCAGGTAGTT----	CGGTTGGTT	000002150	
Orangutan	000001739	AACTC-TAA <sub>CGCTT</sub> -----	GAGACCTCAGGTAGTT----	CGGTTGGTT	000001776	
Horse	000002139	AACAC-TGACACTT-----	CAGGCCACAGACAGTTCA--	CAGTTGGTT	000002178	
Dog	000002151	GGATTCTGACACCG-----	GAGGCCACAGACAGTTCAC--	AGTTGGTC	000002191	
Pig	000001808	AACTT-TAACACTT-----	GAGGCTATAACAGAGATAATT	CAGTTGGTT	000001849	
Squirrel	000001470	AACTT-TAACACTT-----	GAAGCCACTGTAGATGTT--	AGTTTCTT	000001505	
Mouse	000002160	-----GCATCTCATCAGGAAGGCTACTGATAGTTATTTAAGTTGTA	-----	000002201		
Rat	000002104	-----GCATCTTAAGAAAAGCTACTGATAACTTATTAAGTTGTA	-----	000002145		

C.

<b>-2661SREBP1 -2648SREBP1</b>						
Human	000000616	CTGTTTATATTTACTGTGGTGGTGGT	CACATGAATATAACACATGCGA	000000665		
Macaque	000000708	CTGTTCTATATTTACTGTGGTGGTGGT	CACATGAATGTACACATGCGA	000000757		
Chimpanzee	000000677	CTGTTCTATATTTACTGTGGTGGTGGT	CACATGAATGTACACATGCGA	000000726		
Orangutan	000000280	CTGTTTATATTTACTGTGGTGGTGGT	CACATGAATATAACACATGCGA	000000329		
Horse	000000722	CTGTTCACTATCTGATTGTGGCGCTGGT	CACATGAACCCAACACATGCGA	000000771		
Dog	000000535	TTG-----TTGACTGTGGTGATGCGATCACCTA	CACATGCGA	000000584		
Pig	000000377	TTG-----TTGACTGTGGTGATGCGATCACCTA	CACATGCGA	000000417		
Squirrel	000000096	CTGTTTGTACTGTGGTGAT	CACCTGAACCTACCTATGCGA	000000145		
Mouse	000000000	-----	-----	000000000		
Rat	000000000	-----	-----	000000000		

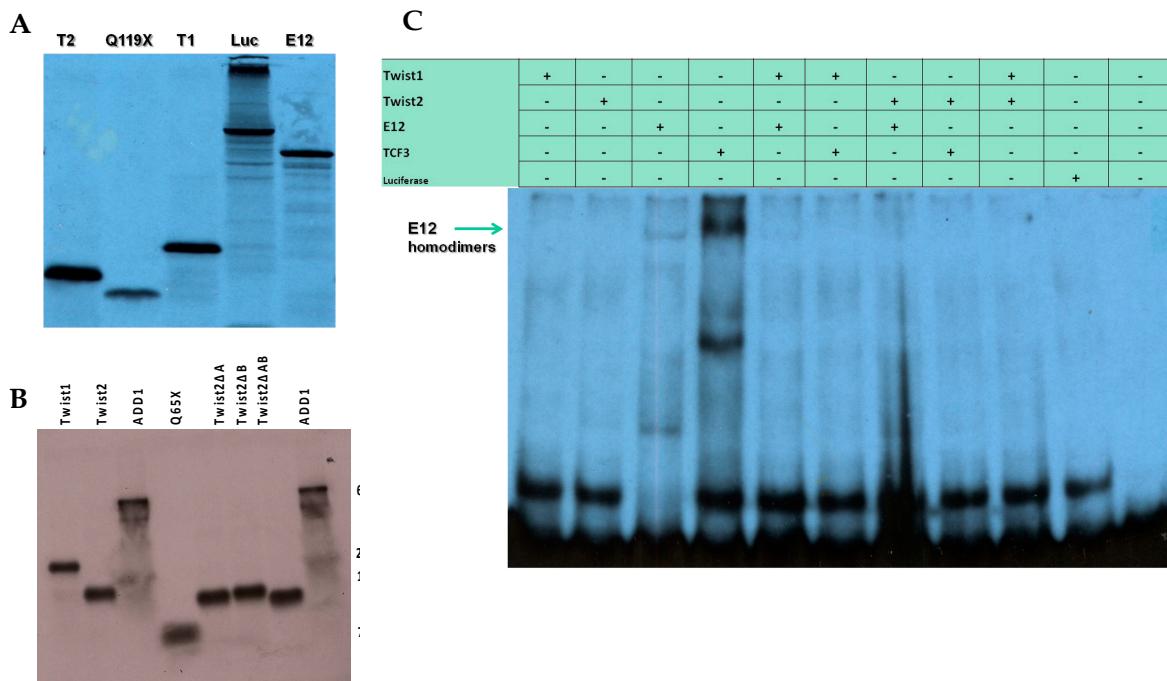
<b>-2611Twist</b>						
Human	000000666	TAAAAATTGCATAGAATTATACACAGGCAAATGACTGCATATAAAACTG	CAAATGACTGCATATAAAACTG	000000715		
Macaque	000000758	TAAAAATTGCATAGAATTATACATAGGCAAATGACTGGATATAAAACTG	CAAATGACTGGATATAAAACTG	000000807		
Chimpanzee	000000727	TAAAAATTGCATAGAATTATACATAGGCAAATGACTGGATATAAAACTG	CAAATGACTGGATATAAAACTG	000000776		
Orangutan	000000330	TAAAAATTGCATAGAATTATACACAGGCAAATGAGTACATATAAAACTG	CAAATGAGTACATATAAAACTG	000000379		
Horse	000000772	TAAAAATTGCATAGAATTATACACAGGCAAATGGTGCACGTAAAGTG	CAAATGGTGCACGTAAAGTG	000000821		
Dog	000000585	TAAAAATTGCATATAACTATACACAGGCAAATGGATGCATGCAAACACTG	CAAATGGATGCATGCAAACACTG	000000634		
Pig	000000418	TAAAAATTCTTAGAGCTATACGCATAGGCAAATGGTGCCTATAATG-A	CAAATGGTGCCTATAATG-A	000000466		
Squirrel	000000146	TAAAAATTGCCTAGAATTATACATGCAGGCAAATGAGTGCATGCAAACACTG	CAAATGAGTGCATGCAAACACTG	000000195		
Mouse	000001075	-----AGGCACA-----	-----AGGCACA-----	000001081		
Rat	000001023	-----AGGCACA-----	-----AGGCACA-----	000001029		

**Supplementary Figure S2. Bioinformatic analysis of the 5' upstream region of the human *CHRD1* gene to identify putative TF binding sites and sequence conservation**

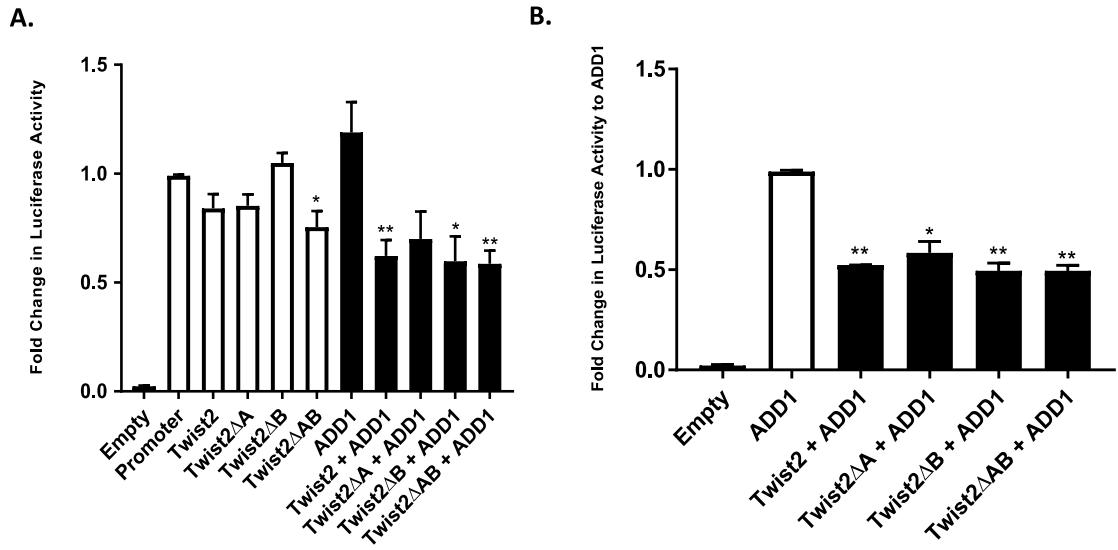
**A.** mVISTA bioinformatic analysis of the upstream region (-3000 to +1) of the *CHRD1* gene among selected mammals. Schematic representation of the global multiple sequence alignment of the different animals when compared to the human *CHRD1* gene upstream sequence. Red areas indicate conservation based on mVISTA parameters.

**B)** Multiple sequence alignment of the E-boxes found in probe #3 (from -1297 to -1148).

**C)** Multiple sequence alignment of the E-boxes found in probe #5 (from -2697 to -2548). The region contained in probe #5 is the most conserved in most of the species considered in the mVISTA analysis. The -2661 site is conserved in human, macaque, chimpanzee, orangutan, horse, dog, and squirrel. The -2648 site is conserved in human, macaque, chimpanzee, orangutan, horse, and pig. Interestingly the -2611 TWIST site is conserved in 8 out of 10 species considered in this analysis.



**Supplementary Figure S3. SDS-PAGE of *in vitro* transcribed/translated proteins and Electrophoretic Mobility Shift Assay for probe #1 of the *CHRDL1* gene.** **A.** This SDS-PAGE autoradiography represents an example of the typical results obtained for the proteins synthesized *in vitro* using the TnT Quick-Coupled Transcription/Translation kit (TnT) and labeled by incorporation of  $^{35}$ S-Methionine to assess protein production. **B.** Western Blot of SDS -PAGE of TnT reactions of TWIST proteins and ADD1, detected with an anti-Myc antibody. Luciferase was used as a positive control for protein synthesis and as a negative control (mock) for EMSA binding reactions. In both A and B, similar amounts of protein were obtained for most proteins. Abbreviations used: T2=TWIST2; Q119X and Q65X= mutant forms of TWIST2 found in Setleis Syndrome patients, T1= TWIST1; Luc=Luciferase. **B.** EMSA carried out with *in vitro* synthesized proteins using probe #1. For this probe we detected binding of E12 and in the reaction with the TCF3 construct, which codes for E12 as well. The region in probe #1 contains a putative binding site for E12 at position -167 and a TWIST site at -194.



**Supplementary Figure S4. Suppression of the transcriptional activity of ADD1/SREBP1c by TWIST2 and its deletion mutant N-terminal forms in HeLa cells.**

Luciferase reporter gene assay using HeLa cells were used to assess the effect of the different proteins (TWIST2, TWIST2 $\Delta$ A, TWIST2 $\Delta$ B, TWIST2 $\Delta$ AB and ADD1) on the *CHRD1* gene upstream region. **A.** Comparison of the basal levels of reporter expression (promoter) signal to the expression signal obtained when transfected with the constructs observed in the figure. All comparisons were made relative to the basal promoter activity. **B.** The expression levels of the co-transfected reactions were compared to ADD1-dependent activation of the *CHRD1* gene promoter (reporter expression activity was normalized against the expression of the reporter gene when transfected with ADD1). Transfection assays were performed in duplicate and repeated independently at least three times (N=3). Statistical analysis was done using one-way ANOVA followed by unpaired Student's T-test. Error bars represent standard error of the mean (SEM). \* = p<0.05, \*\* = p<0.01.