

Supporting Information

Palladium-Catalyzed sp³ C–H Acetoxylation of α,α -Disubstituted α -Amino Acids

Atsushi Matsumura, Yoshinosuke Usuki* and Tetsuya Satoh*

Department of Chemistry, Graduate School of Science, Osaka Metropolitan University, 3-3-138 Sugimoto, Sumiyoshi-ku, Osaka 558-8585, Japan

List of Contents

Experimental Section

General: S-2

Procedure for *N*-Phthalimide Protection of Amino Acids: S-2 – S-3

Preparation of **1a-e** and **1g-k**: S-3

Preparation of **1f**: S-3

Preparation of **3a-b**: S-3 – S-4

Characterization Data of **1** and **3**: S-4 – S-8

General Procedure for Pd-Catalyzed Acetoxylation of **1** and **3**: S-8

Characterization Data of Products: S-8 – S-11

Procedure for Methanolysis of **2a**: S-11 – S-12

X-ray Crystal structure Analysis: S-12 – S-13

References: S-13

¹H and ¹³C NMR Spectra of Starting Materials: S-14 – S-23

¹H and ¹³C NMR Spectra of Products: S-24 – S-34

EXPERIMENTAL SECTION

General. ^1H and ^{13}C NMR spectra were recorded at 400 and 100 MHz, respectively, for CDCl_3 and $\text{DMSO}-d_6$ solution. NMR measurements were performed at 80 °C if necessary. HRMS data were obtained by DART using a TOF mass spectrometer. The structures of all products listed below were unambiguously determined by ^1H and ^{13}C NMR and X-ray crystal structure analysis.

Amino acid derivative **1** and **3** [35,42-45] and mono-*N*-protected amino acid ligand **L6** [46] were prepared according to published procedures. Other reagents were purchased from commercial resources and used without further purification.

The following experimental procedures may be regarded as typical in methodology and scale.

Procedure for *N*-Phthalimide Protection of Amino Acids

Procedure A [42]: Amino acid (1.0 equiv.) and phthalic anhydride (1.2 equiv.) were placed into a round-bottom flask and heated to 190 °C until visible formation of water vapor ceased (approximately 45 min.). The reaction mixture was allowed to cool slightly before adding the contents to sat. NaHCO_3 (aq.). The reaction flask was rinsed with sat. NaHCO_3 (aq.) and 1% NaOH (aq.). The combined aqueous portions were filtered, and the filtrate was acidified to a pH below 2 with concentrated HCl (aq.). The resulting precipitate was isolated by filtration and dried under vacuum to give the corresponding *N*-phthalimide protected amino acid, which was used without further purification.

Procedure B [43]: 1M toluene solution of amino acid (1.0 equiv.) and phthalic anhydride (1.0 equiv.) was prepared in a round-bottom flask. Then, NEt_3 (0.10 eq.) were added to the solution. The suspension was refluxed at 130 °C overnight equipped with a Dean-Stark system to collect the water produced during the reaction. The resulting mixture was cooled down to room temperature and solvents were removed under vacuum. The crude product was then dissolved in DCM and washed by aqueous 1 M HCl (aq., 3 times). The combined organic layers were dried over anhydrous MgSO_4 , filtered, and evaporated under reduced pressure to obtain the corresponding *N*-phthalimide protected amino acid, which was used without further purification. If the crude product was not colorless, it was dissolved in DCM and extracted with saturated NaHCO_3 (aq.). The combined aqueous portions were filtered, and the filtrate was acidified to a pH below 2 with concentrated HCl (aq.). The resulting precipitate was isolated by filtration and dried under vacuum to give the corresponding *N*-phthalimide protected amino acid, which was used without further purification.

Procedure C [44]: Amino acid (1.0 equiv.) and Na_2CO_3 (1.0 equiv.) were dissolved in water (1 M) at room temperature. *N*-Ethoxycarbonylphthalimide (1.0 equiv.) was added to the solution in small portions. The mixture was stirred for 3 hours at room temperature, and then the aqueous solution was cooled to 0 °C and slowly acidified with 6 M HCl (aq.) until pH of 1-2 was attained and white

precipitate was observed. The precipitate was collected and washed with 1 M HCl (aq.) and EtOAc/hexanes = 1/5 to give the corresponding *N*-phthalimide protected amino acid, which was used without further purification.

Preparation of **1a-e** and **1g-k** [43]

To a round-bottom flask were added the amino acid (1.0 equiv.), DCM (0.50 M) and a few drops of DMF. Oxalyl chloride (2.0 eq.) was added slowly to the mixture and then the resulting mixture was stirred for 3 h at room temperature. The excess of oxalyl chloride and DCM were removed in vacuo. The crude acid chloride was dissolved in DCM (0.5 M) and primary or secondary amine (1.5 equiv.) and *N,N*-diisopropylethylamine (1.5 equiv.) were added dropwise under cooling with an ice bath. The mixture was stirred for 2 hours at room temperature and then quenched with sat. NaHCO₃ (aq.). The aqueous layer was extracted with EtOAc (3 times). The combined organic layers were washed with 1 M HCl (aq., 3 times) and brine (3 times), dried over Na₂SO₄, filtered and concentrated. The crude residue was purified by column chromatography on silica gel using hexane–EtOAc as eluent to afford substrate **1**.

Preparation of **1f** [45]

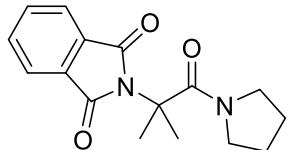
To a stirring solution of 1-(1,3-dioxoisooindolin-2-yl)cyclopropane-1-carboxylic acid (1.3 mmol) and *N*-(3-dimethylaminopropyl)-*N'*-ethylcarbodiimide hydrochloride (307 mg, 1.6 mmol) in DCM (5.2 mL) were added pyrrolidine (134 µL, 1.6 mmol) and a small amount of 4-(dimethylamino)pyridine (15.9 mg, 0.13 mmol). The reaction mixture was stirred for 24 h at room temperature. Upon completion, the organic layer was washed with 0.2 M HCl (aq.) and sat. Na₂CO₃ (aq.). The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The crude residue was purified by column chromatography on silica gel using hexane–EtOAc as eluent to afford substrate **1f** (238 mg, 64%).

Preparation of **3a-b** [35]

To a solution of amino acid methyl ester hydrochloride (3.6 mmol) and NEt₃ (1.7 mmol) in DMF (10 mL) were added 2-(1,3-dioxoisooindolin-2-yl)-2-methylpropanoic acid (3 mmol) and 1-hydroxy-7-azabenzotriazole (HOAt, 3.03 mmol). The mixture was cooled in an ice bath and subsequently 1-ethyl-3-(3-(dimethylamino)propyl)carbodiimide hydrochloride (EDCI, 3.6 mmol) was added in one portion. After 1.5 h at 0 °C and 6 h (**3a**) or 3 d (**3b**) at room temperature, the mixture was partitioned between H₂O and EtOAc. The aqueous phase was extracted with EtOAc (3 times). The organic phase was washed successively with H₂O, 0.5 M HCl (aq.), sat. NaHCO₃ (aq.) and brine. Then it was dried

over anhydrous Na₂SO₄, filtered, and concentrated. The crude residue was purified by column chromatography on silica gel using hexane–EtOAc as eluent to afford substrate **3**.

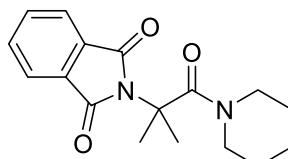
Characterization Data of **1** and **3**



2-(2-Methyl-1-oxo-1-(pyrrolidin-1-yl)propan-2-yl)isoindoline-1,3-dione (1a)

¹H NMR (400 MHz, CDCl₃) δ 7.83 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.73 (dd, *J* = 5.5, 3.0 Hz, 2H), 3.56 (t, *J* = 6.8 Hz, 2H), 3.20 (t, *J* = 6.2 Hz, 2H), 1.84 (s, 6H), 1.82–1.73 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 170.4, 168.3, 134.4, 131.6, 123.4, 61.3, 47.9, 46.8, 27.2, 24.8, 23.1; HRMS: *m/z* calcd for C₁₆H₁₉N₂O₃ [M+H]⁺ 287.13957, found: 287.13826.

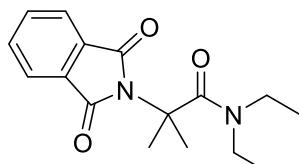
N-Phthalimide protection was performed by Procedure A.



2-(2-Methyl-1-oxo-1-(piperidin-1-yl)propan-2-yl)isoindoline-1,3-dione (1b)

¹H NMR (400 MHz, DMSO-*d*₆) δ 7.85 (d, *J* = 0.8 Hz, 4H), 3.40–3.37 (m, 4H), 1.72 (d, *J* = 1.0 Hz, 6H), 1.52–1.46 (m, 2H), 1.38–1.32 (m, 4H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 168.4, 167.2, 134.3, 130.9, 122.5, 59.9, 44.5, 25.0, 24.9, 24.7, 23.5; HRMS: *m/z* calcd for C₁₇H₂₁N₂O₃ [M+H]⁺ 301.15522, found: 301.15555.

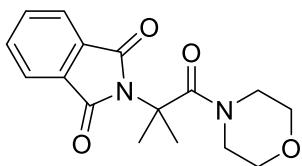
N-Phthalimide protection was performed by Procedure A.



2-(1,3-Dioxoisooindolin-2-yl)-N,N-diethyl-2-methylpropanamide (1c)

¹H NMR (400 MHz, CDCl₃) δ 7.83 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.73 (dd, *J* = 5.5, 3.0 Hz, 2H), 3.35 (br s, 4H), 1.84 (s, 6H), 1.15 (br s, 3H), 0.95 (br s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.6, 168.2, 134.3, 131.8, 123.3, 61.1, 41.5, 41.1, 25.8, 13.5, 12.5; HRMS: *m/z* calcd for C₁₆H₂₁N₂O₃ [M+H]⁺ 289.15522, found: 289.15469.

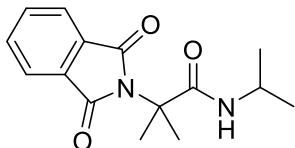
N-Phthalimide protection was performed by Procedure A.



2-(2-Methyl-1-morpholino-1-oxopropan-2-yl)isoindoline-1,3-dione (1d)

¹H NMR (400 MHz, DMSO-*d*₆) δ 7.85 (d, *J* = 1.4 Hz, 4H), 3.48–3.40 (m, 8H), 1.72 (s, 6H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 169.0, 167.2, 134.3, 130.9, 122.6, 65.4, 59.7, 44.2, 24.7; HRMS: *m/z* calcd for C₁₆H₁₉N₂O₄ [M+H]⁺ 303.13448, found: 303.13354.

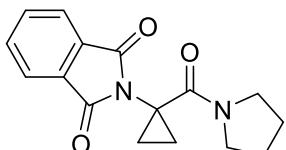
N-Phthalimide protection was performed by Procedure A.



2-(1,3-Dioxoisodolin-2-yl)-N-isopropyl-2-methylpropanamide (1e)

¹H NMR (400 MHz, DMSO-*d*₆) δ 7.82 (s, 4H), 7.68 (d, *J* = 7.9 Hz, 1H), 3.85 (dp, *J* = 7.9, 6.6 Hz, 1H), 1.64 (s, 6H), 0.99 (d, *J* = 6.6 Hz, 6H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 171.2, 168.3, 134.2, 131.9, 122.6, 60.6, 40.7, 24.8, 22.1; HRMS: *m/z* calcd for C₁₅H₁₉N₂O₃ [M+H]⁺ 275.13957, found: 275.13969.

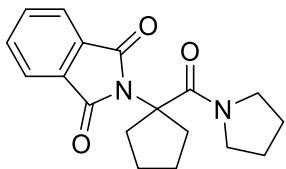
N-Phthalimide protection was performed by Procedure A.



2-(1-(Pyrrolidine-1-carbonyl)cyclopropyl)isoindoline-1,3-dione (1f)

¹H NMR (400 MHz, DMSO-*d*₆) δ 7.87 (d, *J* = 1.1 Hz, 4H), 3.32–3.28 (m, 4H), 1.74–1.70 (m, 4H), 1.63–1.60 (m, 2H), 1.42–1.39 (m, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 167.5, 166.9, 134.3, 130.8, 122.8, 46.3, 33.2, 24.3, 14.5; HRMS: *m/z* calcd for C₁₆H₁₇N₂O₃ [M+H]⁺ 285.12392, found: 285.12352.

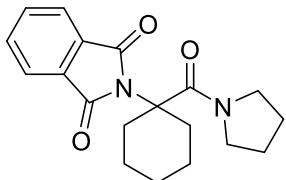
N-Phthalimide protection was performed by Procedure C.



2-(1-(Pyrrolidine-1-carbonyl)cyclopentyl)isoindoline-1,3-dione (1g)

¹H NMR (400 MHz, CDCl₃) δ 7.82 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.73 (dd, *J* = 5.5, 3.1 Hz, 2H), 3.53 (t, *J* = 6.8 Hz, 2H), 3.20 (t, *J* = 6.4 Hz, 2H), 2.66–2.59 (m, 2H), 2.50–2.43 (m, 2H), 1.84–1.72 (m, 8H); ¹³C NMR (100 MHz, CDCl₃) δ 170.0, 168.4, 134.3, 131.6, 123.4, 70.9, 47.6, 46.6, 35.5, 27.1, 24.9, 23.3; HRMS: *m/z* calcd for C₁₈H₂₁N₂O₃ [M+H]⁺ 313.15522, found: 313.15321.

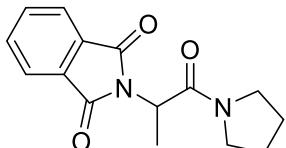
N-Phthalimide protection was performed by Procedure B.



2-(1-(Pyrrolidine-1-carbonyl)cyclohexyl)isoindoline-1,3-dione (1h)

¹H NMR (400 MHz, CDCl₃) δ 7.82 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.73 (dd, *J* = 5.5, 3.1 Hz, 2H), 3.54 (br s, 2H), 3.20 (br s, 2H), 2.58 (ddd, *J* = 12.7, 8.1, 3.7 Hz, 2H), 2.24 (ddd, *J* = 13.4, 8.7, 3.8 Hz, 2H), 1.83–1.77 (m, 6H), 1.65–1.57 (m, 2H), 1.52–1.48 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 170.1, 168.7, 134.3, 131.5, 123.3, 65.7, 48.1, 46.6, 31.9, 27.3, 25.1, 23.0, 22.9; HRMS: *m/z* calcd for C₁₉H₂₃N₂O₃ [M+H]⁺ 327.17087, found: 327.17166.

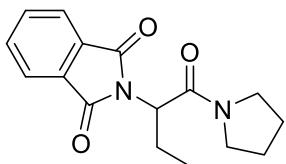
N-Phthalimide protection was performed by Procedure B.



2-(1-Oxo-1-(pyrrolidin-1-yl)propan-2-yl)isoindoline-1,3-dione (1i) [S1]

¹H NMR (400 MHz, CDCl₃) δ 7.85 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.72 (dd, *J* = 5.5, 3.1 Hz, 2H), 4.99 (q, *J* = 7.2 Hz, 1H), 3.52–3.41 (m, 3H), 3.27–3.21 (m, 1H), 1.94–1.85 (m, 3H), 1.83–1.74 (m, 1H), 1.71 (d, *J* = 7.2 Hz, 3H); HRMS: *m/z* calcd for C₁₅H₁₇N₂O₃ [M+H]⁺ 273.12392, found: 273.12338.

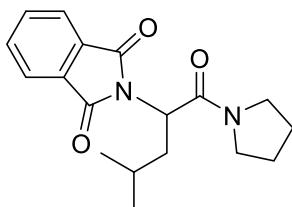
N-Phthalimide protection was performed by Procedure C.



2-(1-Oxo-1-(pyrrolidin-1-yl)butan-2-yl)isoindoline-1,3-dione (1j)

¹H NMR (400 MHz, CDCl₃) δ 7.85 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.72 (dd, *J* = 5.5, 3.0 Hz, 2H), 4.79 (dd, *J* = 10.4, 5.2 Hz, 1H), 3.50–3.43 (m, 3H), 3.29 (dt, *J* = 10.1, 7.1 Hz, 1H), 2.41 (ddq, *J* = 14.8, 10.4, 7.4 Hz, 1H), 2.15 (dqd, *J* = 14.8, 7.4, 5.2 Hz, 1H), 1.93–1.71 (m, 4H), 0.96 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.0, 167.3, 134.2, 131.7, 123.5, 54.8, 46.5, 46.5, 26.5, 23.9, 22.0, 11.1; HRMS: *m/z* calcd for C₁₆H₁₉N₂O₃ [M+H]⁺ 287.13957, found: 287.13883.

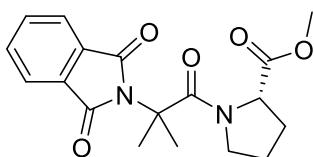
N-Phthalimide protection was performed by Procedure A.



2-(4-Methyl-1-oxo-1-(pyrrolidin-1-yl)pentan-2-yl)isoindoline-1,3-dione (1k) [S2]

¹H NMR (400 MHz, CDCl₃) δ 7.84 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.71 (dd, *J* = 5.5, 3.0 Hz, 2H), 5.00 (dd, *J* = 11.4, 4.3 Hz, 1H), 3.51–3.45 (m, 3H), 3.40–3.34 (m, 1H), 2.61–2.53 (m, 1H), 1.98–1.90 (m, 2H), 1.89–1.70 (m, 3H), 1.59–1.51 (m, 1H), 0.95 (dd, *J* = 6.6, 5.4 Hz, 6H); HRMS: *m/z* calcd for C₁₈H₂₃N₂O₃ [M+H]⁺ 315.17087, found: 315.17149.

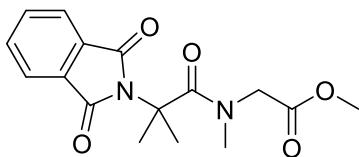
N-Phthalimide protection was performed by Procedure B.



Methyl (2-(1,3-dioxoisodolin-2-yl)-2-methylpropanoyl)-L-proline (3a) [S3]

¹H NMR (400 MHz, CDCl₃) δ 7.84 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.74 (dd, *J* = 5.5, 3.0 Hz, 2H), 4.56 (dd, *J* = 8.5, 3.7 Hz, 1H), 3.74 (s, 3H), 3.62–3.57 (m, 1H), 3.16–3.10 (m, 1H), 2.08–1.95 (m, 5H), 1.90–1.85 (m, 1H), 1.82–1.77 (m, 1H), 1.73 (s, 3H); HRMS: *m/z* calcd for C₁₈H₂₁N₂O₅ [M+H]⁺ 345.14505, found: 345.14554.

N-Phthalimide protection was performed by Procedure A.



Methyl N-(2-(1,3-dioxoisindolin-2-yl)-2-methylpropanoyl)-N-methylglycinate (3b)

¹H NMR (400 MHz, CDCl₃) δ 7.84 (dd, *J* = 5.4, 3.0 Hz, 2H), 7.74 (dd, *J* = 5.4, 3.1 Hz, 2H), 4.08 (s, 2H), 3.74 (s, 3H), 3.00 (s, 3H), 1.87 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 172.1, 169.8, 168.0, 134.4, 131.7, 123.4, 60.8, 52.1, 51.4, 37.0, 25.1; HRMS: *m/z* calcd for C₁₆H₁₉N₂O₅ [M+H]⁺ 319.12940, found: 319.13002.

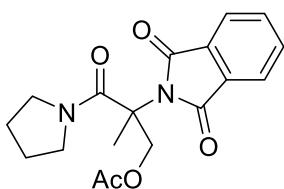
N-Phthalimide protection was performed by Procedure A.

Amino acid methyl ester hydrochloride was prepared according to literature report [S4].

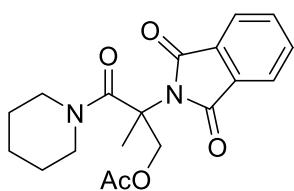
General Procedure for Pd-Catalyzed Acetoxylation of 1 and 3

To a flame dried 5 mL vial or 15 mL pressure resistant tube were added **1** or **3** (0.3 mmol), PhI(OAc)₂ (0.9 mmol, 290 mg), Pd(CH₃CN)₄(BF₄)₂ (0.015 mmol, 7 mg), *N*-acetyl-L-valine (**L1**, 0.03 mmol, 5 mg) and HFIP/DME/Ac₂O (5/4/1, 0.7 mL). The mixture was stirred under Ar (1 atm) at 80 °C (hot plate temperature or oil bath) for 20 h. The mixture was diluted with EtOAc (1 mL) and then passed through a short column with activated alumina to remove insoluble solids. After removal of the solvents under vacuum, the crude residue was purified by column chromatography on silica gel using hexane–EtOAc as eluent to afford acetoxylated products **2** or **4**. Further purification by GPC (gel permeation chromatography) was performed, if needed.

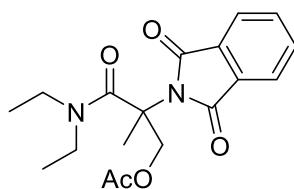
Characterization Data of Products



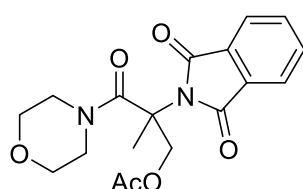
2-(1,3-Dioxoisindolin-2-yl)-2-methyl-3-oxo-3-(pyrrolidin-1-yl)propyl Acetate (2a): white solid; 67 mg (65%); m.p. 95–97 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.84 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.75 (dd, *J* = 5.5, 3.1 Hz, 2H), 4.87 (d, *J* = 11.7 Hz, 1H), 4.64 (d, *J* = 11.7 Hz, 1H), 3.61–3.44 (m, 3H), 2.97 (q, *J* = 7.6 Hz, 1H), 2.00 (s, 3H), 1.94 (s, 3H), 1.84–1.65 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 170.7, 168.4, 168.3, 134.6, 131.4, 123.6, 66.8, 62.9, 47.7, 46.4, 27.2, 23.0, 20.9, 19.4; HRMS: *m/z* calcd for C₁₈H₂₁N₂O₅ [M+H]⁺ 345.14505, found: 345.14492.



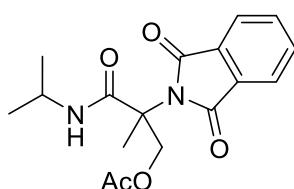
2-(1,3-Dioxoisindolin-2-yl)-2-methyl-3-oxo-3-(piperidin-1-yl)propyl Acetate (2b): white solid; 58 mg (54%); m.p. 142-144 °C; ¹H NMR (400 MHz, DMSO-*d*₆, 80 °C) δ 7.88 (d, *J* = 1.1 Hz, 4H), 4.73 (d, *J* = 11.5 Hz, 1H), 4.38 (d, *J* = 11.5 Hz, 1H), 3.46 – 3.34 (m, 4H), 1.93 (s, 3H), 1.86 (d, *J* = 1.1 Hz, 3H), 1.54 – 1.46 (m, 2H), 1.36 (td, *J* = 12.0, 6.1 Hz, 4H); ¹³C NMR (100 MHz, DMSO-*d*₆, 80 °C) δ 169.5, 167.3, 166.7, 134.5, 130.5, 122.8, 66.3, 61.5, 44.1, 24.8, 23.4, 19.9, 19.2; HRMS: *m/z* calcd for C₁₉H₂₃N₂O₅ [M+H]⁺ 359.16070, found: 359.16146.



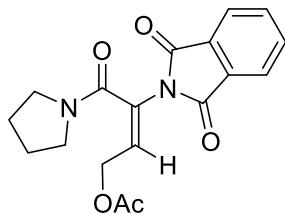
3-(Diethylamino)-2-(1,3-dioxoisindolin-2-yl)-2-methyl-3-oxopropyl Acetate (2c): white solid; 53 mg (51%); m.p. 91-93 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.83 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.74 (dd, *J* = 5.5, 3.1 Hz, 2H), 4.87 (d, *J* = 11.7 Hz, 1H), 4.54 (d, *J* = 11.7 Hz, 1H), 3.42–3.27 (m, 4H), 2.01 (s, 3H), 1.90 (s, 3H), 1.13 (t, *J* = 7.1 Hz, 3H), 0.92 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.7, 168.8, 168.3, 134.5, 131.5, 123.4, 67.3, 62.7, 41.1, 40.8, 20.8, 20.1, 13.4, 12.4; HRMS: *m/z* calcd for C₁₈H₂₃N₂O₅ [M+H]⁺ 347.16070, found: 347.16035.



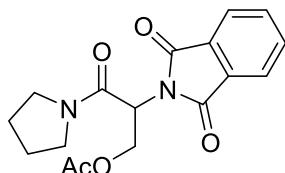
2-(1,3-Dioxoisindolin-2-yl)-2-methyl-3-morpholino-3-oxopropyl Acetate (2d): white solid; 28 mg (26%); m.p. 138-140 °C; ¹H NMR (400 MHz, DMSO-*d*₆, 80 °C) δ 7.88 (d, *J* = 0.7 Hz, 4H), 4.70 (d, *J* = 11.5 Hz, 1H), 4.41 (d, *J* = 11.5 Hz, 1H), 3.53 – 3.41 (m, 8H), 1.93 (s, 3H), 1.86 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆, 80 °C) δ 169.5, 167.3, 167.2, 134.5, 130.5, 122.8, 66.1, 65.3, 61.4, 43.9, 19.89, 19.3; HRMS: *m/z* calcd for C₁₈H₂₁N₂O₆ [M+H]⁺ 361.13996, found: 361.14043.



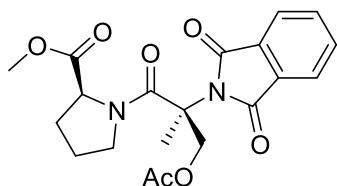
2-(1,3-Dioxoisooindolin-2-yl)-3-(isopropylamino)-2-methyl-3-oxopropyl Acetate (2e): white solid; 22 mg (22%); m.p. 191–193 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.82 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.73 (dd, *J* = 5.5, 3.1 Hz, 2H), 5.66 (br d, *J* = 8.0 Hz, 1H), 4.77 (d, *J* = 11.6 Hz, 1H), 4.69 (d, *J* = 11.6 Hz, 1H), 4.11 (dp, *J* = 8.0, 6.6 Hz, 1H), 2.00 (s, 3H), 1.86 (s, 3H), 1.15 (dd, *J* = 6.6, 4.1 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 170.4, 169.2, 168.5, 134.4, 131.8, 123.4, 65.9, 63.3, 42.0, 22.6, 20.9, 20.8; HRMS: *m/z* calcd for C₁₇H₂₁N₂O₅ [M+H]⁺ 333.14505, found: 333.14550.



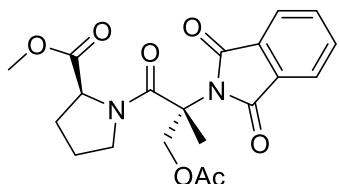
(E)-3-(1,3-Dioxoisooindolin-2-yl)-4-oxo-4-(pyrrolidin-1-yl)but-2-en-1-yl Acetate (2f'): white solid; 47 mg (46%); m.p. 148–150 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.86 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.75 (dd, *J* = 5.5, 3.1 Hz, 2H), 6.26 (t, *J* = 7.3 Hz, 1H), 4.83 (d, *J* = 7.3 Hz, 2H), 3.57–3.48 (m, 4H), 2.08 (s, 3H), 1.93 (dq, *J* = 6.8, 4.0 Hz, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 170.7, 166.3, 162.6, 134.8, 131.5, 129.0, 124.0, 123.7, 60.5, 48.2, 45.9, 26.0, 24.4, 20.9; HRMS: *m/z* calcd for C₁₈H₁₉N₂O₅ [M+H]⁺ 343.12940, found: 343.12997.



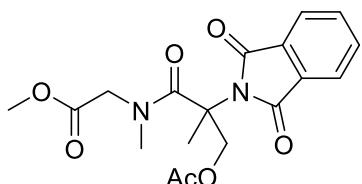
2-(1,3-Dioxoisooindolin-2-yl)-3-oxo-3-(pyrrolidin-1-yl)propyl Acetate (2i): pale yellow oil; 21 mg (21%); ¹H NMR (400 MHz, CDCl₃) δ 7.88 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.76 (dd, *J* = 5.5, 3.1 Hz, 2H), 5.18 (dd, *J* = 10.0, 4.5 Hz, 1H), 4.89 (dd, *J* = 11.9, 4.5 Hz, 1H), 4.74 (dd, *J* = 11.9, 10.0 Hz, 1H), 3.51–3.44 (m, 3H), 3.16 (dt, *J* = 10.4, 7.4 Hz, 1H), 1.97 (s, 3H), 1.93–1.85 (m, 3H), 1.79–1.72 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 170.8, 167.5, 164.5, 134.5, 131.6, 123.8, 61.3, 51.4, 46.6, 46.5, 26.5, 23.9, 20.8; HRMS: *m/z* calcd for C₁₇H₁₉N₂O₅ [M+H]⁺ 331.12940, found: 331.12932.



Methyl ((S)-3-Acetoxy-2-(1,3-dioxoisoindolin-2-yl)-2-methylpropanoyl)-L-proline ((S)-4a): white solid; 30 mg (24%); m.p. 174–175 °C; $[\alpha]_D^{20} = -89.4$ ($c = 1.5$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.84 (dd, $J = 5.5$, 3.1 Hz, 2H), 7.75 (dd, $J = 5.5$, 3.1 Hz, 2H), 4.85 (d, $J = 11.8$ Hz, 1H), 4.57 – 4.52 (m, 2H), 3.72 (s, 3H), 3.67 (ddd, $J = 9.8$, 7.2, 3.1 Hz, 1H), 3.02 (td, $J = 8.9$, 6.3 Hz, 1H), 2.06 (s, 3H), 2.04–1.88 (m, 6H), 1.80 (dt, $J = 8.6$, 3.1 Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 172.7, 170.7, 168.9, 168.2, 134.6, 131.3, 123.6, 66.6, 62.6, 60.3, 52.3, 47.0, 27.9, 25.6, 20.8, 18.7; HRMS: m/z calcd for $\text{C}_{20}\text{H}_{23}\text{N}_2\text{O}_7$ [$\text{M}+\text{H}]^+$ 403.15053, found: 403.14996.



Methyl ((R)-3-Acetoxy-2-(1,3-dioxoisoindolin-2-yl)-2-methylpropanoyl)-L-proline ((R)-4a): colorless oil; 10 mg (8%); $[\alpha]_D^{20} = 17.1$ ($c = 0.48$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.88 (dd, $J = 5.5$, 3.0 Hz, 2H), 7.76 (dd, $J = 5.5$, 3.0 Hz, 2H), 4.88 (d, $J = 11.8$ Hz, 1H), 4.77 (d, $J = 11.8$ Hz, 1H), 4.52 (dd, $J = 8.7$, 5.2 Hz, 1H), 3.76 (s, 3H), 3.44 – 3.41 (m, 2H), 2.15–2.07 (m, 1H), 2.00–1.80 (m, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 172.4, 170.5, 168.7, 167.9, 134.6, 131.5, 123.7, 66.2, 63.0, 61.0, 52.3, 47.1, 28.1, 26.0, 20.9, 20.4; HRMS: m/z calcd for $\text{C}_{20}\text{H}_{23}\text{N}_2\text{O}_7$ [$\text{M}+\text{H}]^+$ 403.15053, found: 403.14996.

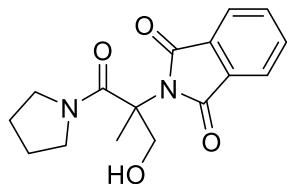


Methyl N-(3-Acetoxy-2-(1,3-dioxoisoindolin-2-yl)-2-methylpropanoyl)-N-methylglycinate (4b): pale yellow oil; 24 mg (21%); ^1H NMR (400 MHz, CDCl_3) δ 7.85 (dd, $J = 5.5$, 3.0 Hz, 2H), 7.76 (dd, $J = 5.5$, 3.0 Hz, 2H), 4.85 (d, $J = 11.8$ Hz, 1H), 4.57 (d, $J = 11.8$ Hz, 1H), 4.49 (d, $J = 17.2$ Hz, 1H), 3.74 (s, 3H), 3.66 (d, $J = 17.2$ Hz, 1H), 3.00 (s, 3H), 2.09 (s, 3H), 1.93 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 170.7, 170.3, 169.6, 168.1, 134.6, 131.4, 123.6, 66.9, 62.4, 52.3, 51.0, 36.6, 20.8, 19.4; HRMS: m/z calcd for $\text{C}_{18}\text{H}_{21}\text{N}_2\text{O}_7$ [$\text{M}+\text{H}]^+$ 377.13488, found: 377.13509.

Procedure for Methanolysis of **2a** [47]

To a 50 mL flask were added **2a** (0.26 mmol, 89 mg), HCl-MeOH (5-10%, 10 mL) and MeOH (5 mL). The mixture was stirred under air (1 atm) at 60 °C for 0.5 h. After removal of the solvents under

vacuum, product was purified by column chromatography on silica gel using hexane–EtOAc as eluent. Further purification by GPC (gel permeation chromatography) was performed to afford **5** (63 mg, 80%).



2-(3-Hydroxy-2-methyl-1-oxo-1-(pyrrolidin-1-yl)propan-2-yl)isoindoline-1,3-dione (5):
colorless oil; 63 mg (80%); ^1H NMR (400 MHz, CDCl_3) δ 7.83 (dd, $J = 5.5, 3.1$ Hz, 2H), 7.74 (dd, $J = 5.5, 3.1$ Hz, 2H), 4.01–3.92 (m, 2H), 3.76 (dd, $J = 8.0, 6.6$ Hz, 1H), 3.61–3.52 (m, 2H), 3.34 (dd, $J = 10.7, 5.8$ Hz, 1H), 3.18–3.15 (m, 1H), 1.91 (s, 3H), 1.83–1.76 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 170.2, 168.6, 134.6, 131.5, 123.6, 67.4, 64.2, 47.7, 46.8, 27.0, 23.1, 18.8; HRMS: m/z calcd for $\text{C}_{16}\text{H}_{19}\text{N}_2\text{O}_4$ [$\text{M}+\text{H}]^+$ 303.13448, found: 303.13263.

X-ray Crystal structure Analysis

The structure of **2f'** has been determined for white needle-like crystals obtained from hexane/DCM (**Figure S1**). CCDC 2240420 contains the crystallographic data of **2f'**.

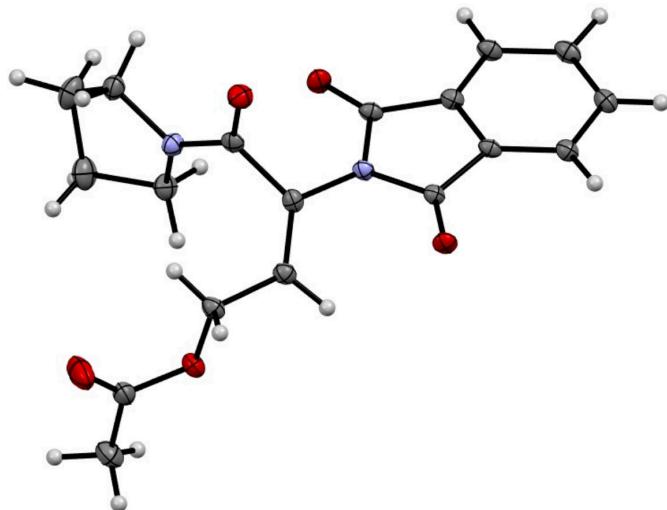


Figure S1. ORTEP drawing of compound **2f'**. Crystal data : $\text{C}_{18}\text{H}_{18}\text{N}_2\text{O}_5$, Mw = 342.34, monoclinic, space group $C\ 2/c$, $T = 110$ K, $a = 31.8056(9)$, $b = 7.9378(2)$, $c = 13.0969(4)$, $\beta = 94.205(3)$, $V = 3297.63(16)$, $Z = 8$, 14902 reflections measured, $R = 0.0527$, $wR2 = 0.1424$.

The structure of **4a** has been determined for white needle-like crystals obtained from hexane/DCM (**Figure S2**). CCDC 2240422 contains the crystallographic data of **4a**.

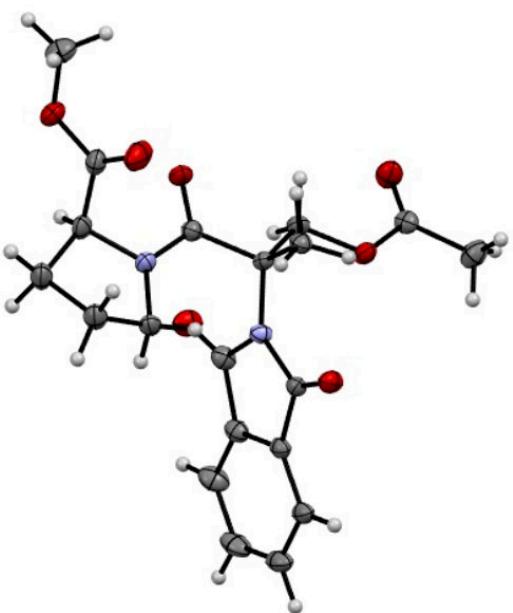
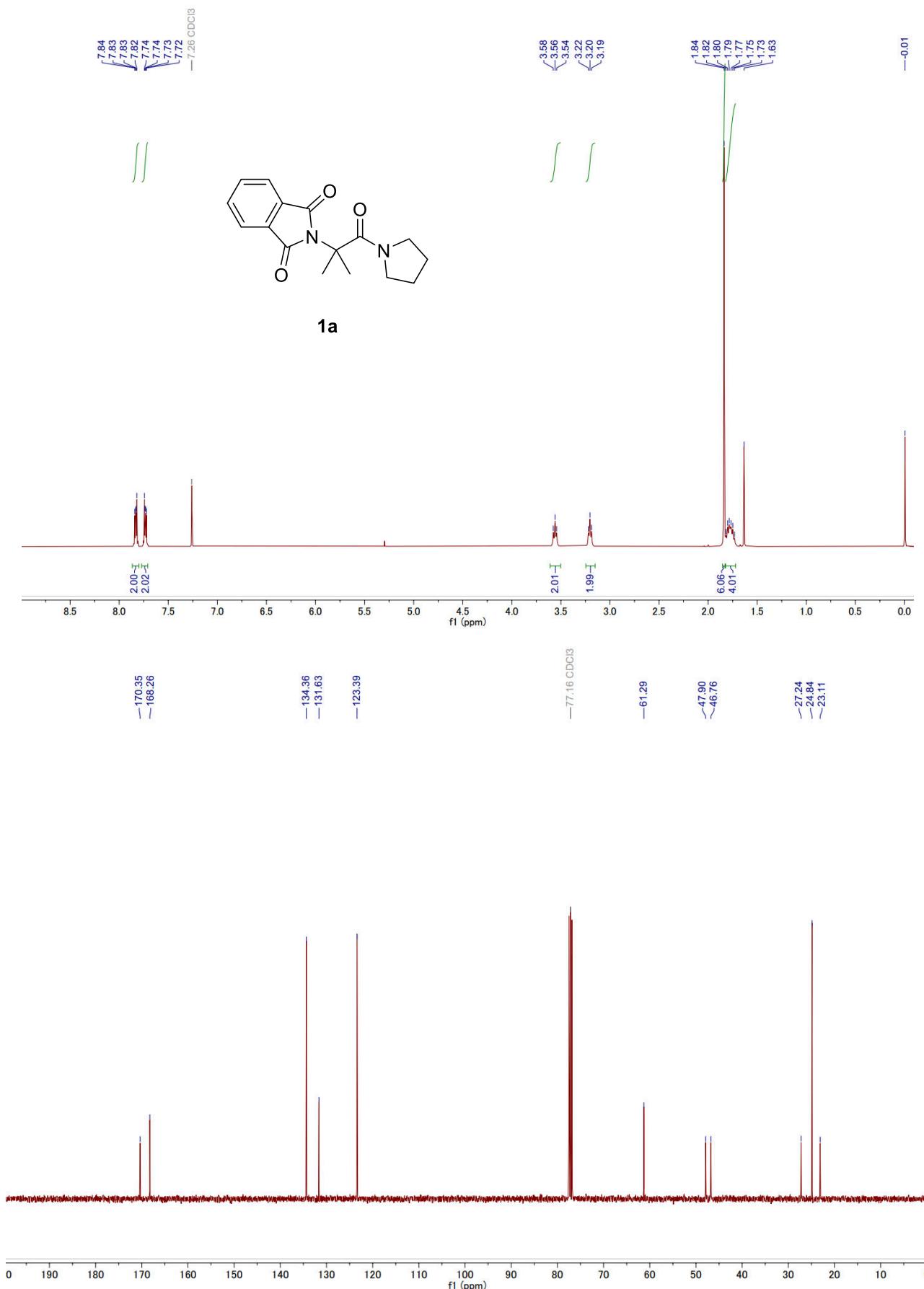


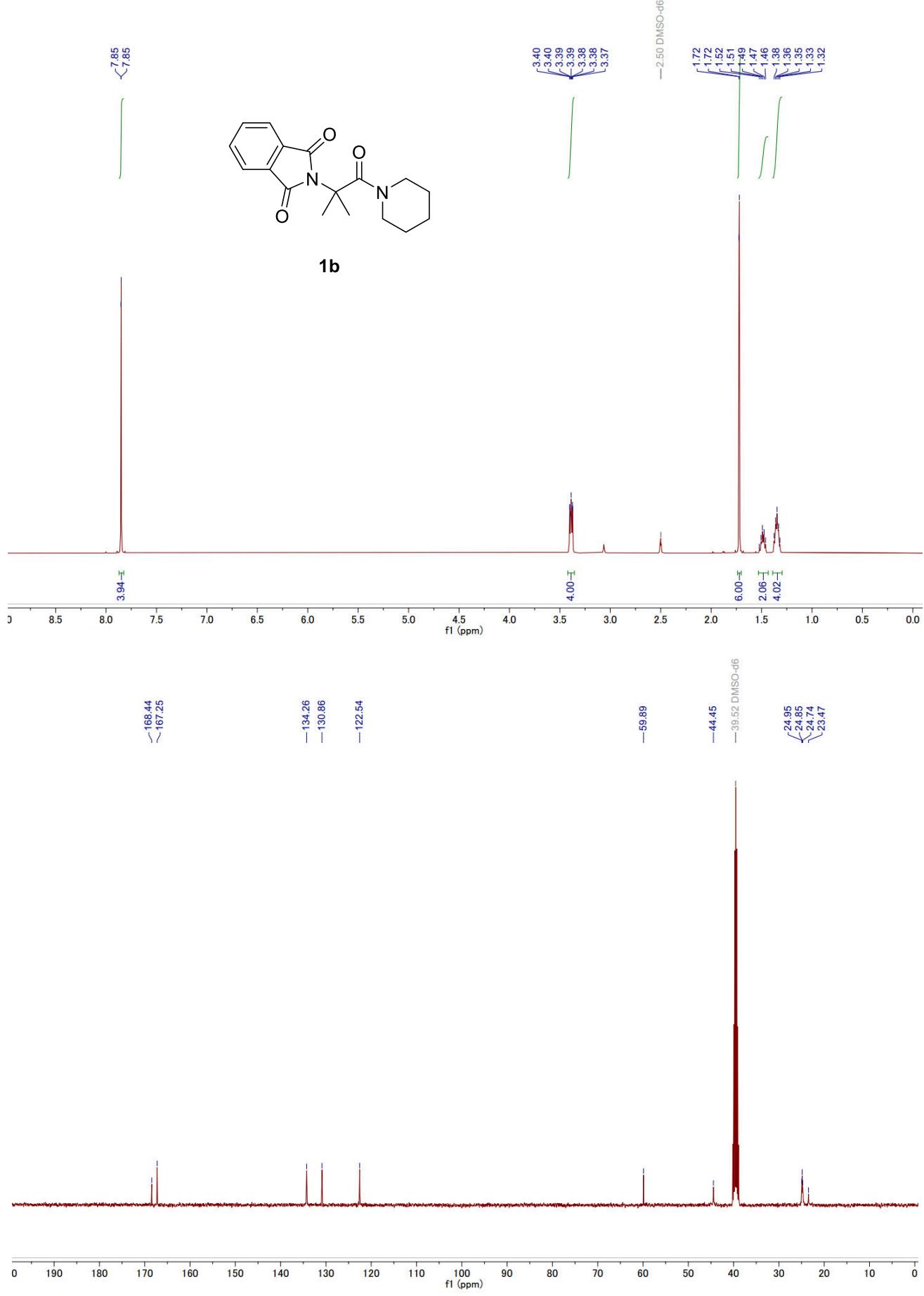
Figure S2. ORTEP drawing of compound **(S)-4a**. Crystal data: $C_{20}H_{22}N_2O_7$, Mw = 402.39, orthorhombic, space group $P\bar{1}\bar{1}\bar{1}\bar{1}$, $T = 110\text{ K}$, $a = 9.863(4)$, $b = 10.830(4)$, $c = 18.514(7)$, $\beta = 90^\circ$, $V = 1977.5(13)$, $Z = 4$, 20287 reflections measured, $R = 0.0326$, $wR2 = 0.0696$.

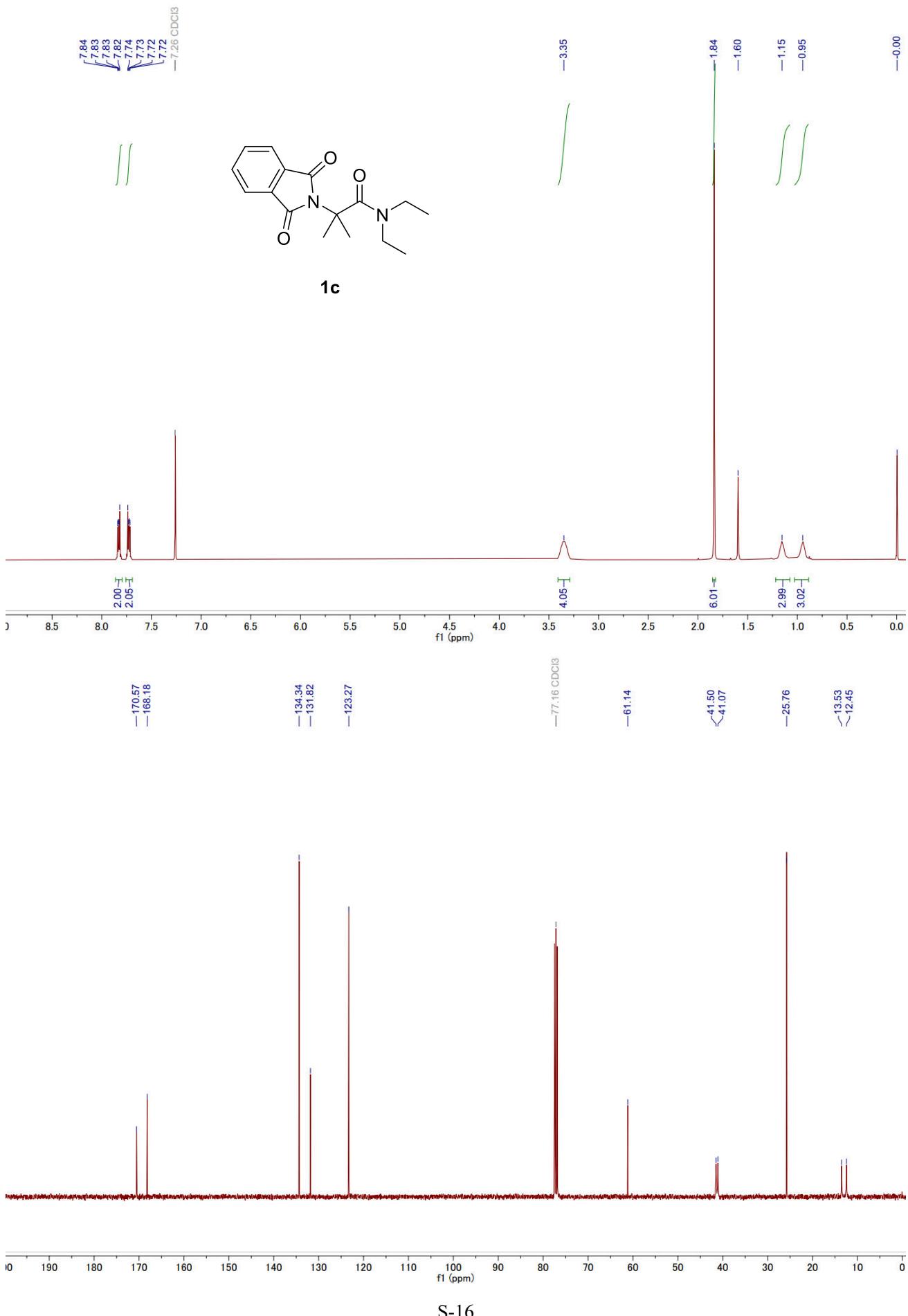
References

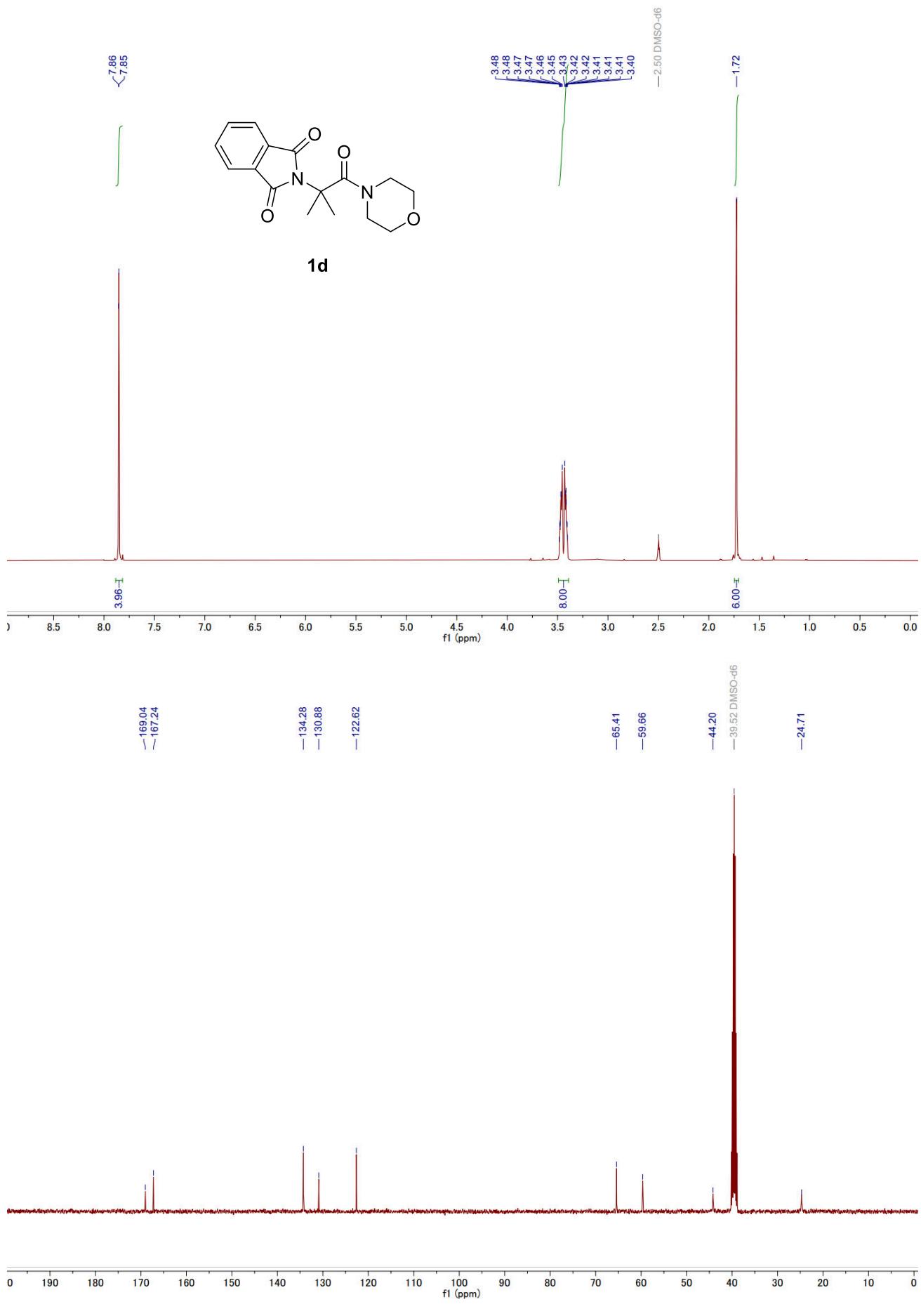
- 35 W. Gong, G. Zhang, T. Liu, R. Giri and J.-Q. Yu, *J. Am. Chem. Soc.*, 2014, **136**, 16940.
- 42 M. A. DeNardo, M. R. Mills, A. D. Ryabov and T J. Collins, *J. Am. Chem. Soc.*, 2016, **138**, 2933.
- 43 L. Vicens, M. Bietti and M. Costas, *Angew. Chem. Int. Ed.*, 2021, **60**, 4740.
- 44 G. Chen, T. Shigenari, P. Jain, Z. Zhang, Z. Jin, J. He, S. Li, C. Mapelli, M. M. Miller, M. A. Poss, P. M. Scola, K.-S. Yeung and J.-Q. Yu, *J. Am. Chem. Soc.*, 2015, **137**, 3338.
- 45 H. Park, Y. Li and J.-Q. Yu, *Angew. Chem. Int. Ed.*, 2019, **58**, 11424.
- 46 M. Erdélyi, V. Langer, A. Karlén and A. Gogoll, *New J. Chem.*, 2002, **26**, 834.
- 47 M. Caillard, A. Emm, A. Jones, T Matthews and M. Williams, *J. Chem. Research*, 1998, **12**, 806.
- S1 M. A. El-Zahabi, L. M. Gad, F. H. Bamanie and Z. Al-Marzooki, *Med. Chem. Res.* 2012, **21**, 75.
- S2 A. K. Singh, R. Kishan, N. Vijayan, V. Balachandran, T. Singh, H. K. Tiwari, B. K. Singha and B. Rathi, *RSC Adv.*, 2013, **3**, 14750.
- S3 A. Matsumoto, Z. Wang and K. Maruoka, *J. Org. Chem.*, 2021, **86**, 5401.
- S4 J. Zhang, Z.-X. Chen, T. Du, B. Li, Y. Gu and S.-K. Tian, *Org. Lett.*, 2016, **18**, 4872.

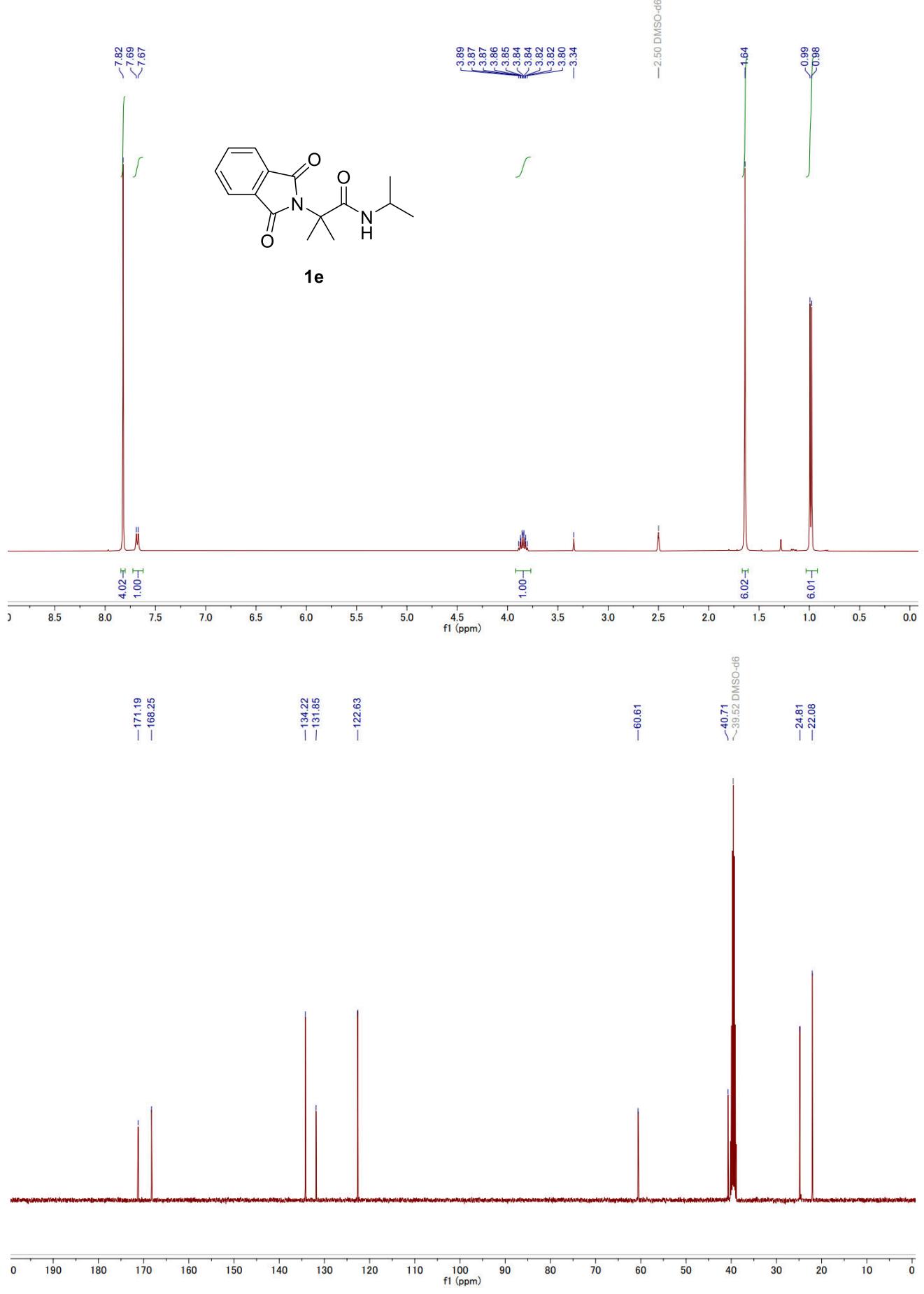
¹H and ¹³C NMR Spectra of Starting Materials

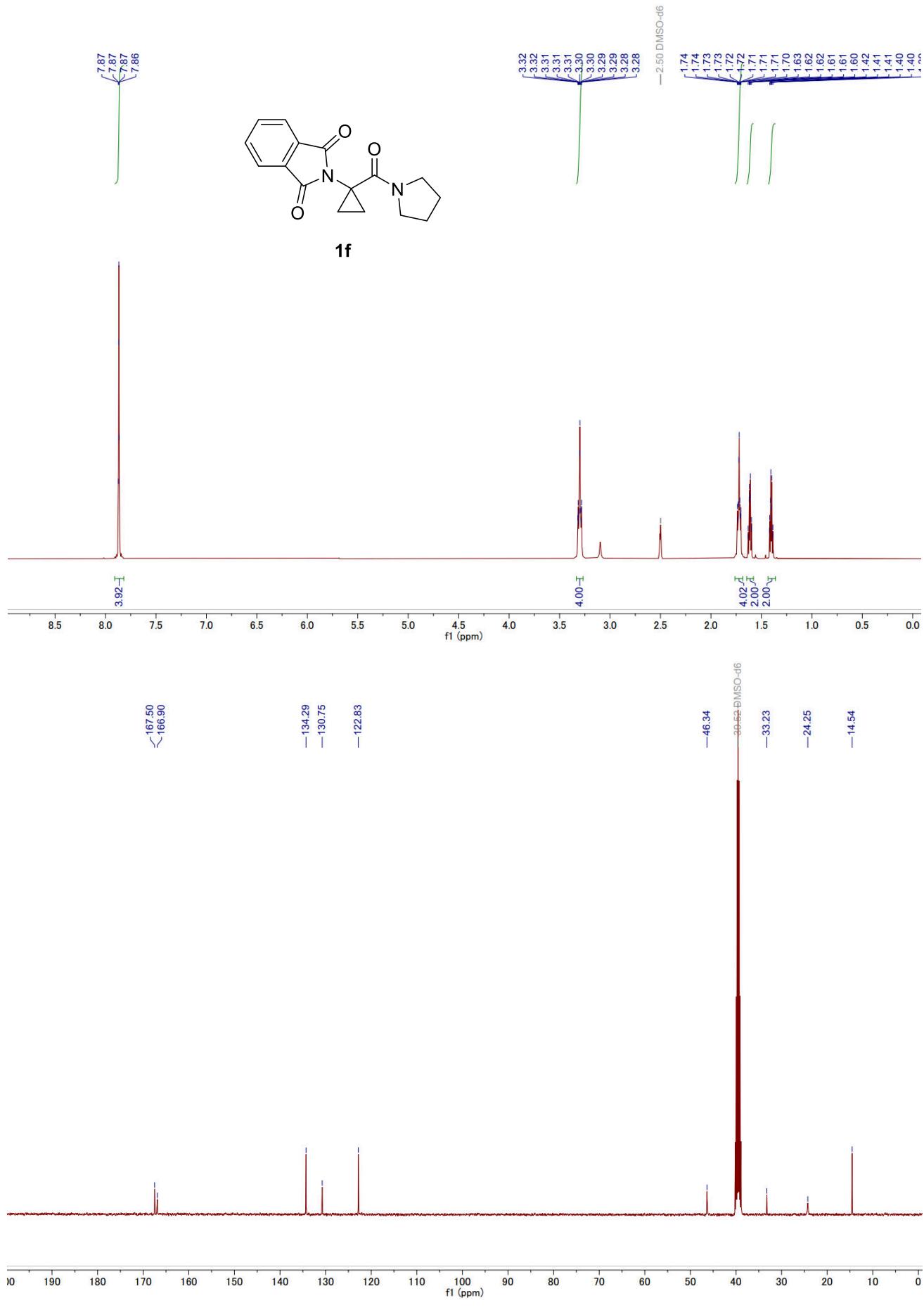


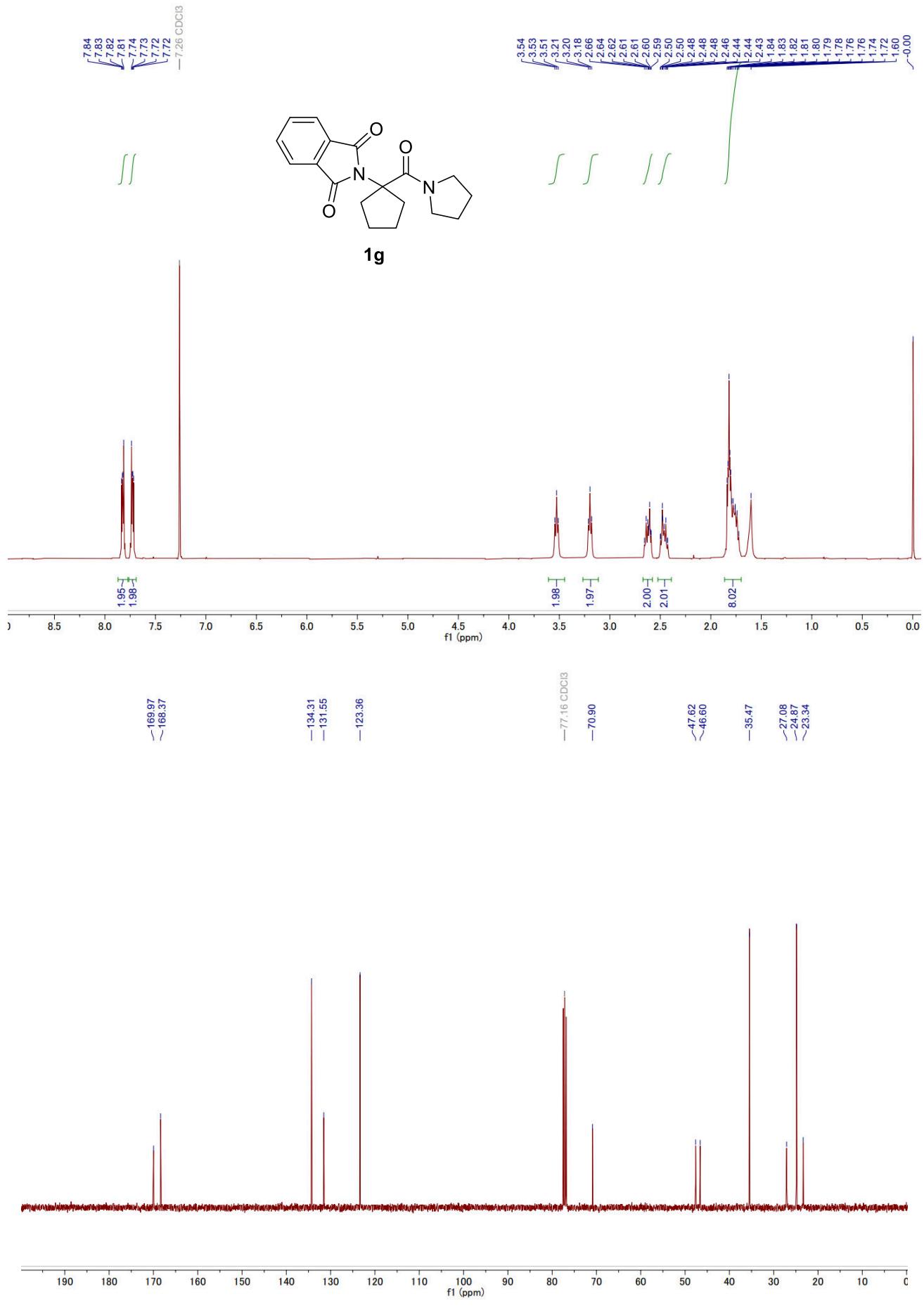


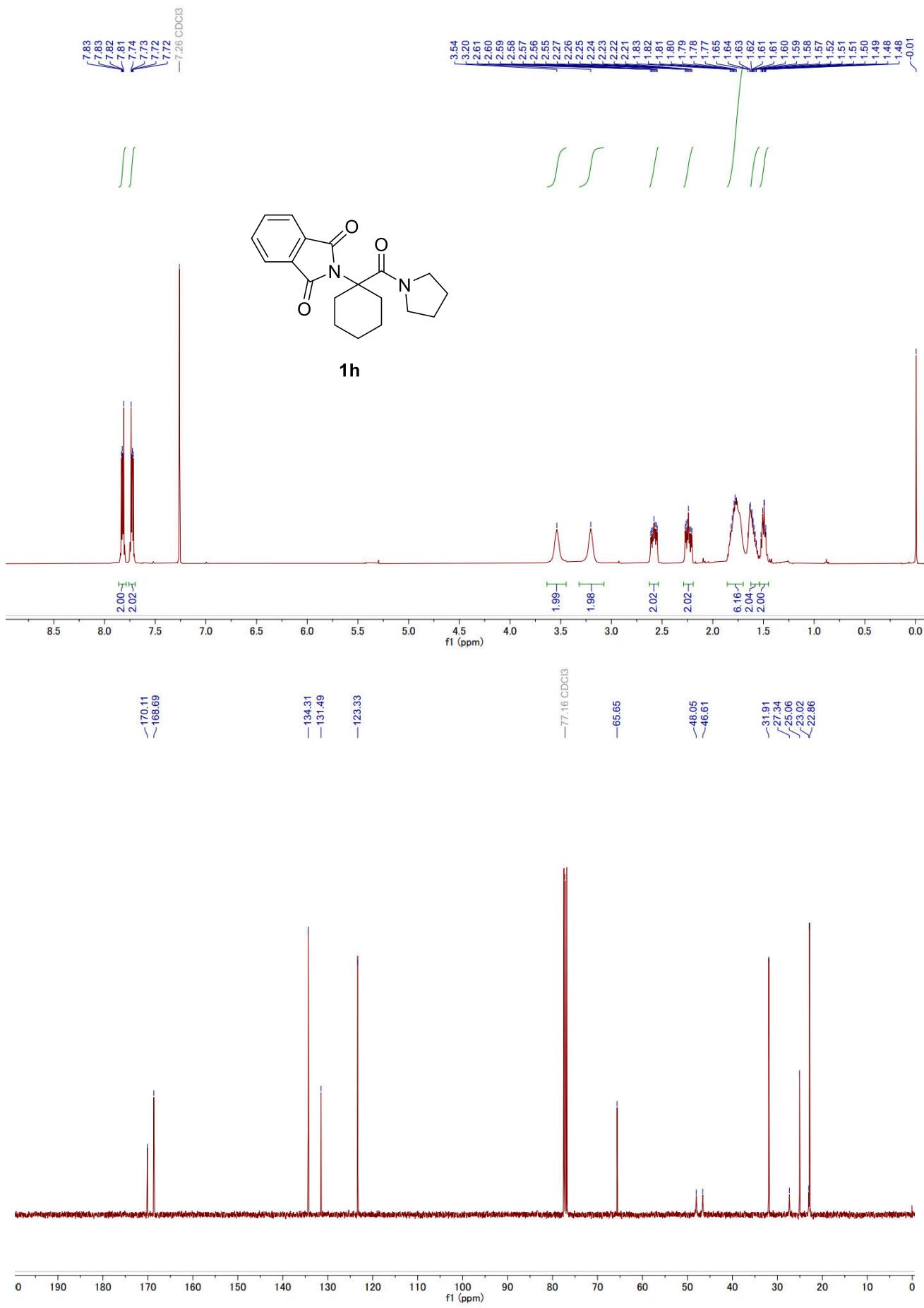


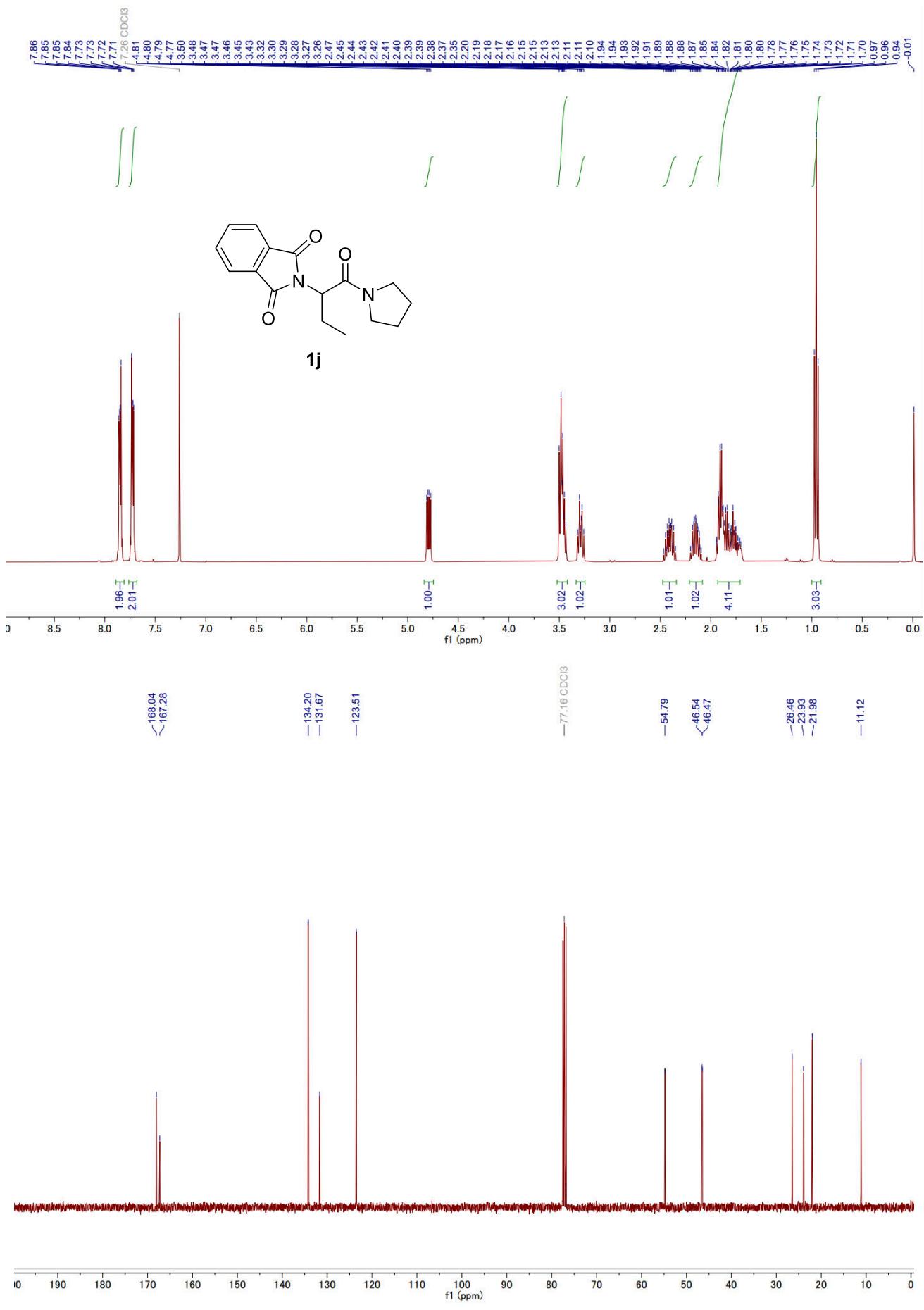


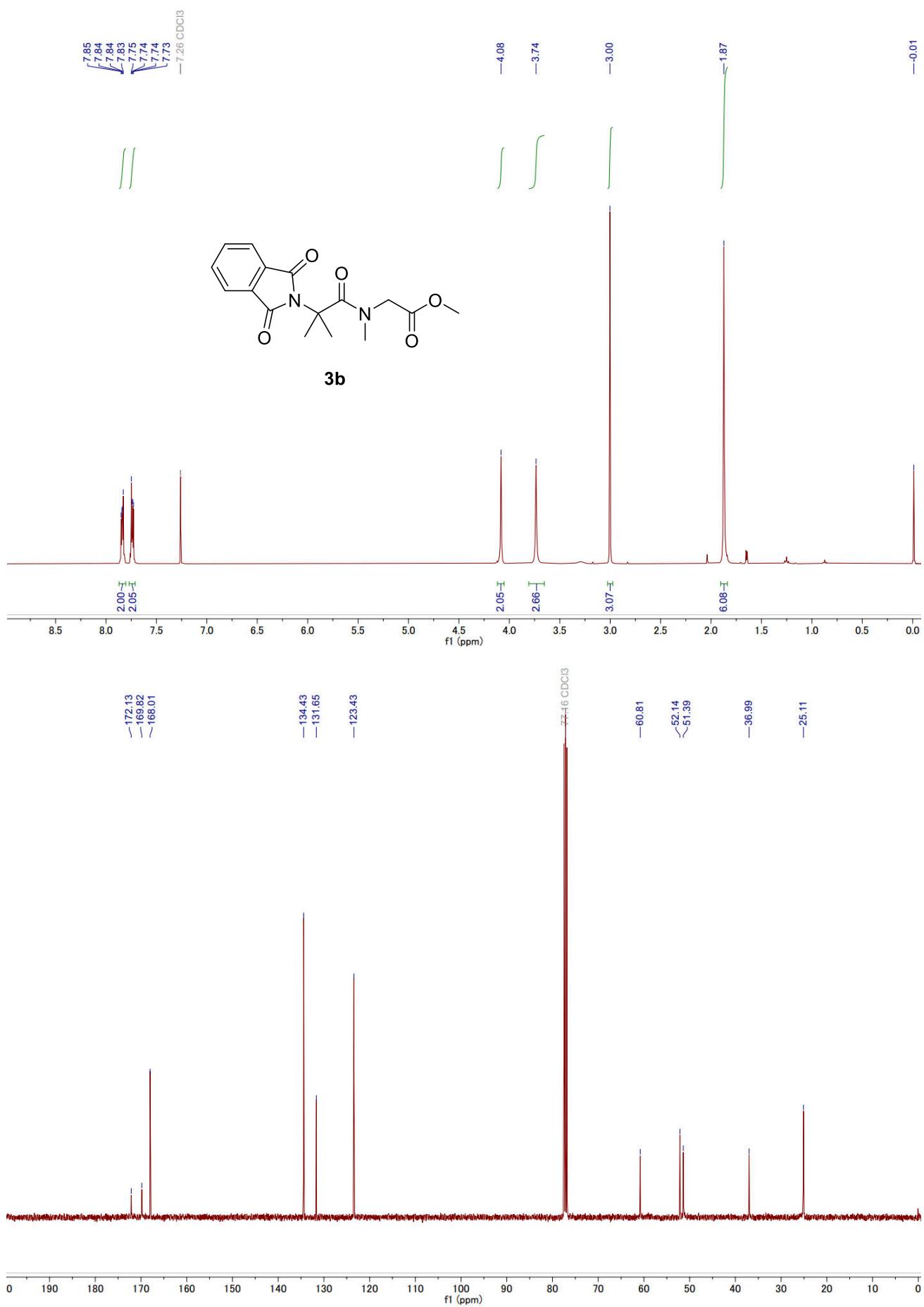












¹H and ¹³C NMR Spectra of Products

