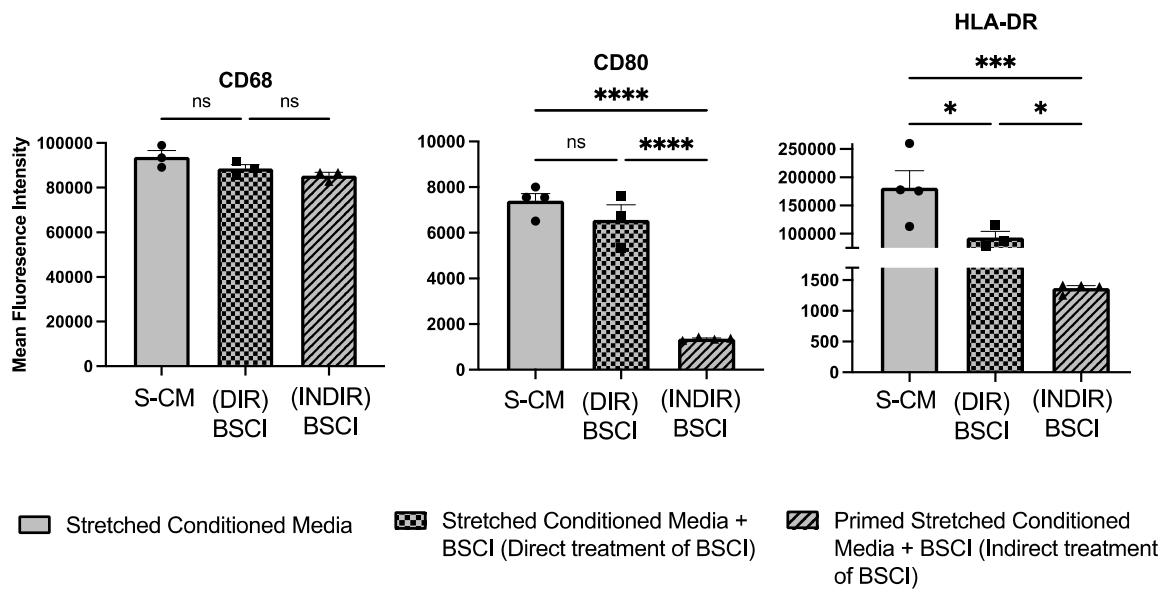
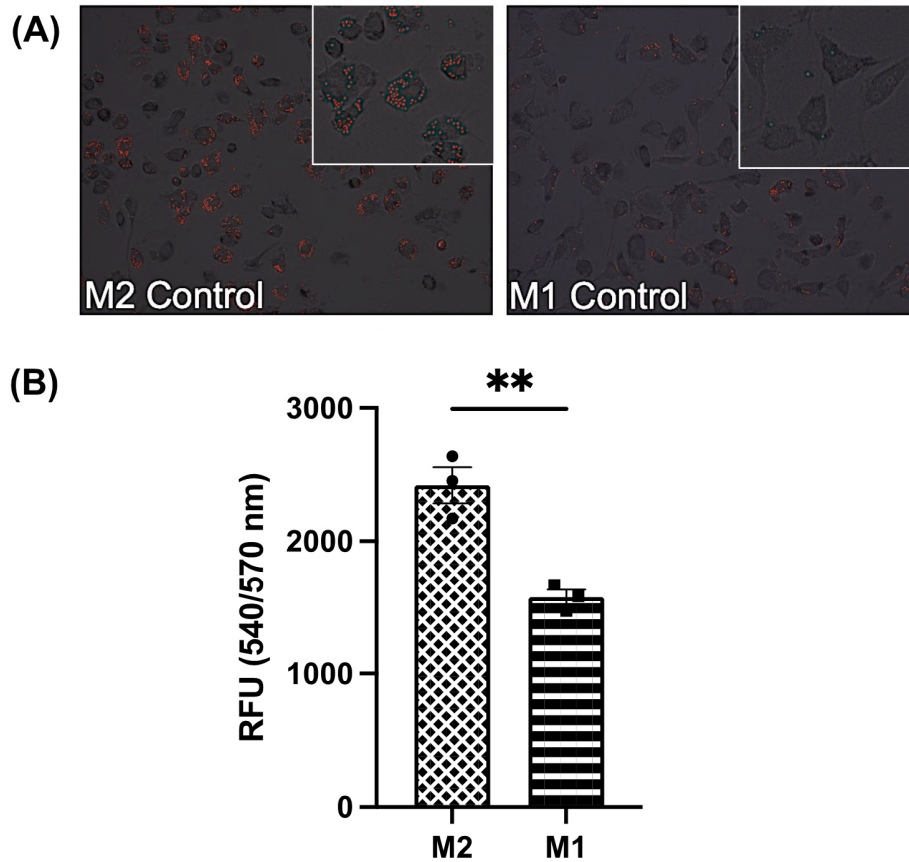
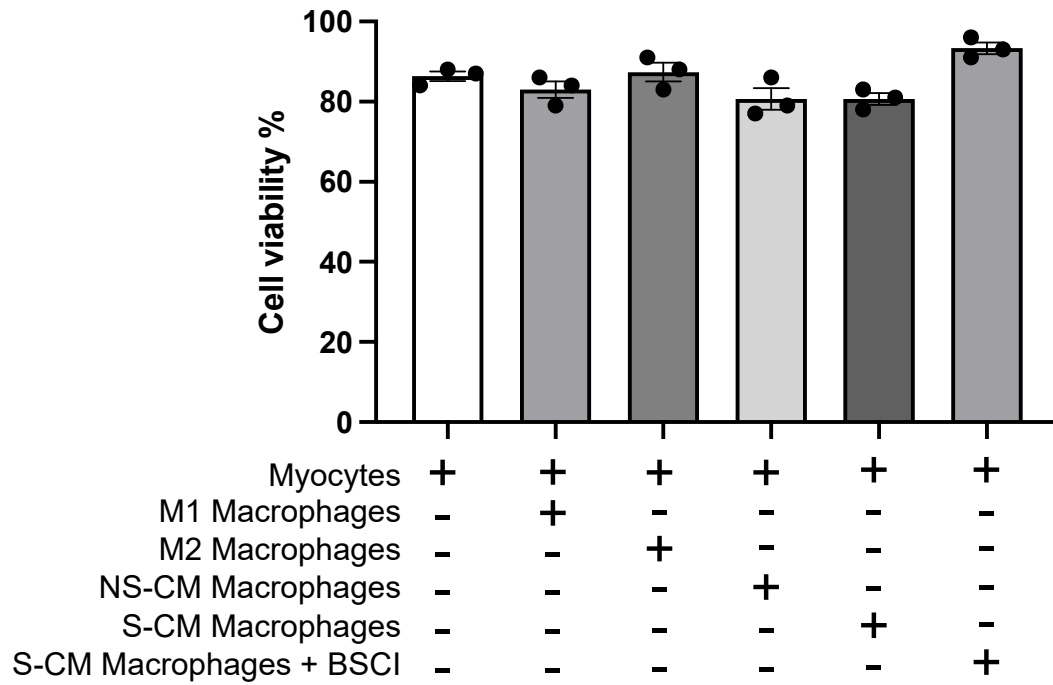






Supplemental Figure S3: Control M1 vs. M2 macrophage phagocytosis zymosan assay. Maternal peripheral blood was collected from pregnant term non-laboring women (N=3/group). For M1 polarization cells were differentiated for 6 days in media containing GM-CSF (100 ng/mL) and polarized for 2 days of LPS (100 ng/mL) and IFN γ (25 ng/mL); for M2 polarization cells were differentiated for 6 days in M-CSF (100 ng/mL), and 2 days in IL-4 (25 ng/mL), and IL-3 (25 ng/mL). After treatment, macrophages (5×10^4) were incubated at 37°C with 5 μ l of zymosan particles for 1 hour. Plates were then evaluated by real-time confocal analysis using a Leica DM IL LED-Inverted fluorescence microscope. **(A)** Representative immunofluorescent images of zymosan particles phagocytized by macrophages. **(B)** Plates were quantified in duplicates for relative fluorescence units (RFU), and absorbance readings were conducted using μ QuantTM software. The average of three macrophage lines (N=3/group) is shown. Original magnification, $\times 20$, inset magnification $\times 40$. Data are presented as mean $\pm$ SD. Statistical significance was determined by one-way ANOVA followed by Dunnett's multiple comparisons test. "***" denotes statistical significance at $p < 0.01$.

Co-Culture Cell Viability



Supplemental Figure S4: Viability of collagen-embedded human myocytes-macrophage co-cultures. After experimental completion (48 hours), myocytes and macrophages from collagen gels were released via 0.1% collagenase digestion. Cells were collected by centrifugation, and viability was measured using a hemocytometer with Trypan blue viability dye. Data are presented as Mean $\pm$ SD. Statistical significance was determined through One-Way ANOVA followed by Dunnette's multiple comparisons test.