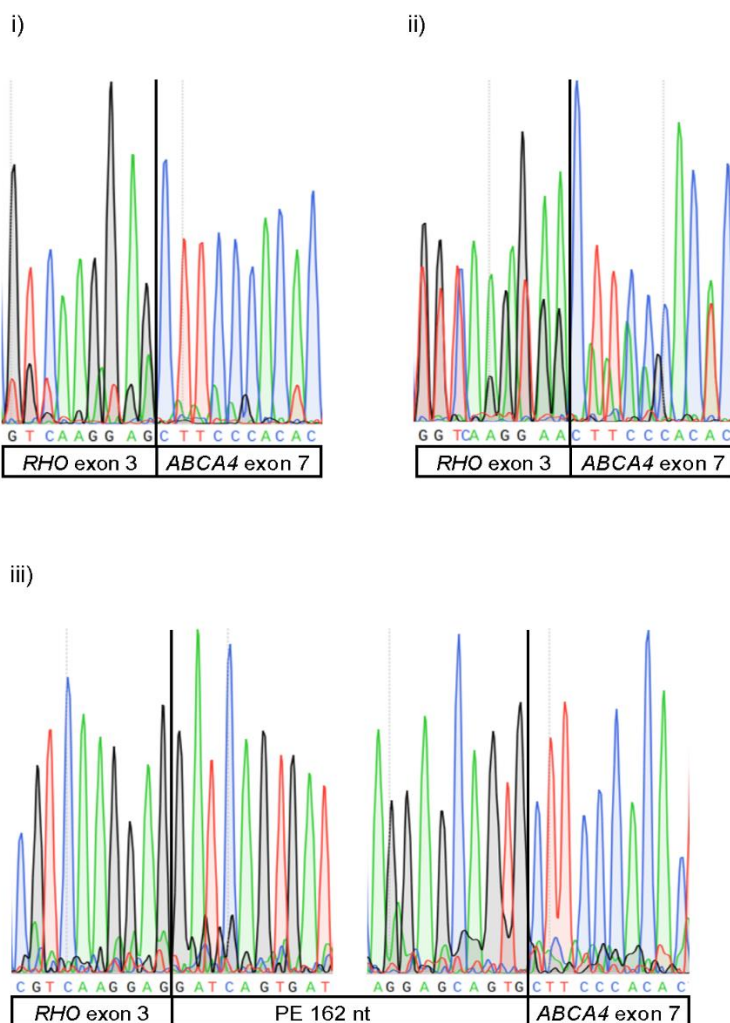
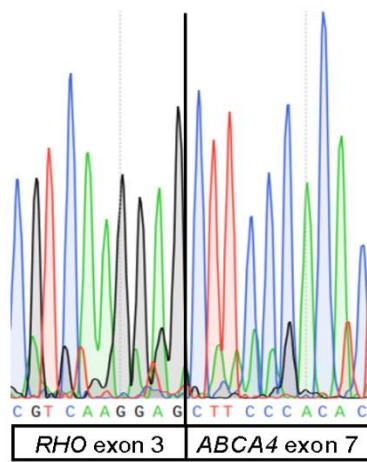


Supplementary Figure S1. Sequence analysis of the resulting RT-PCR products from rescue experiments of the deep-intronic variants c.769-784C>T causing insertion of pseudoexon. Sanger sequencing chromatograms from splicing correction experiments targeting c.769-784C>T. **A)** Sequencing results in HEK293T-midigene system: sequences of the correct transcript from wild-type *ABCA4* midigene condition (i), sequences of the correct transcript from mutant *ABCA4* midigene condition (ii), sequences of the PE transcript from mutant *ABCA4* midigene condition (iii), AON-corrected transcript in mutant *ABCA4* midigene condition (iv) and mutant transcript in AON-treated *ABCA4* midigene (v) are shown. **B)** Sequencing results in fibroblasts: sequences of the correct *ABCA4* transcript from control fibroblasts condition (i), sequences of the PE *ABCA4* transcript from patient-derived fibroblasts condition (ii), AON-corrected transcript from patient-derived fibroblasts condition (iii), mutant transcript from AON-treated patient-derived fibroblasts condition (iv), AON artifact (v), cycloheximide artifact (vi). **C)** Sequencing results in photoreceptor precursor cells: sequences of the correct *ABCA4* transcript from control PPC condition (i), sequences of the PE *ABCA4* transcript from patient-derived PPCs condition (ii), AON-corrected transcript in patient-derived PPCs condition (iii), mutant transcript in AON-treated in patient-derived PPCs condition (iv).

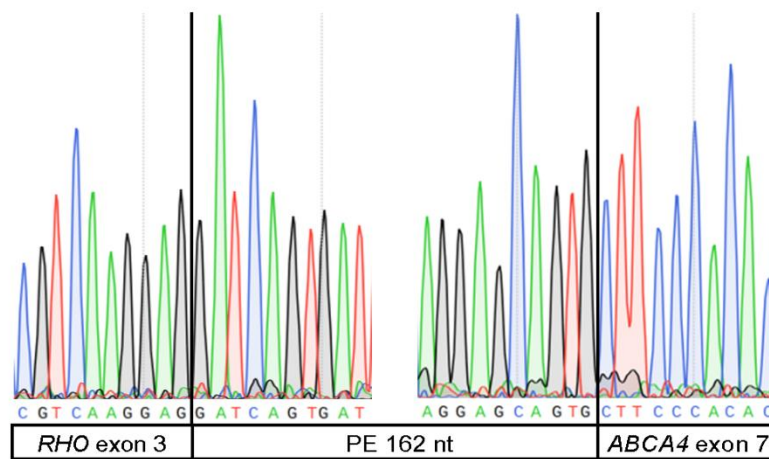
A. Sequencing results HEK-midigene system



iv)

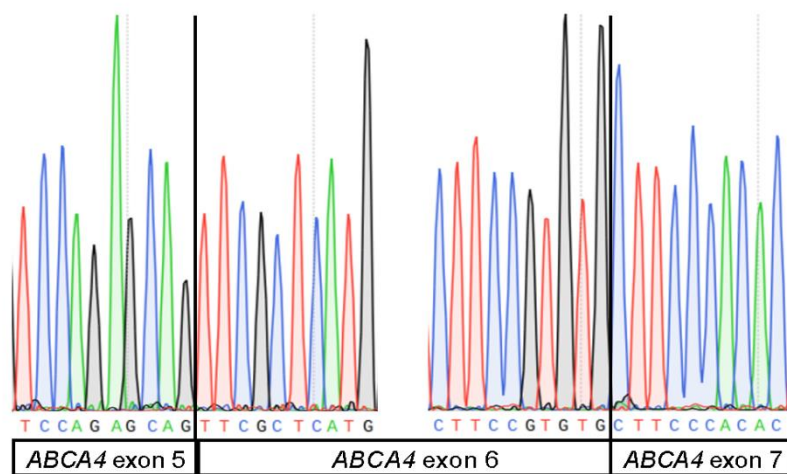


v)

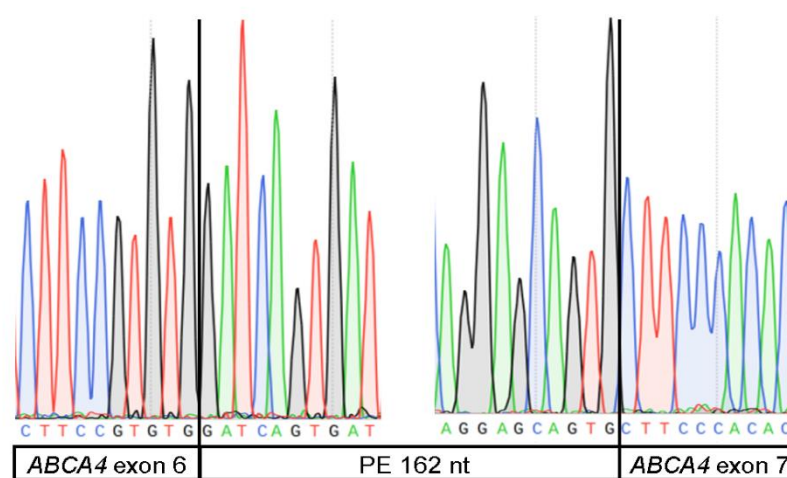


B. Sequencing results fibroblasts

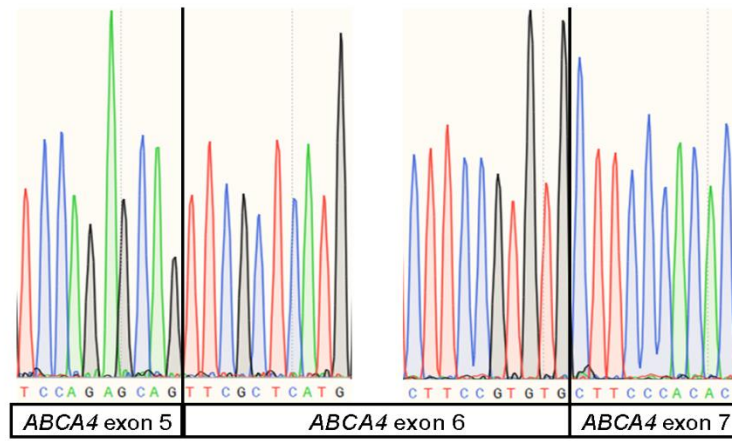
i)



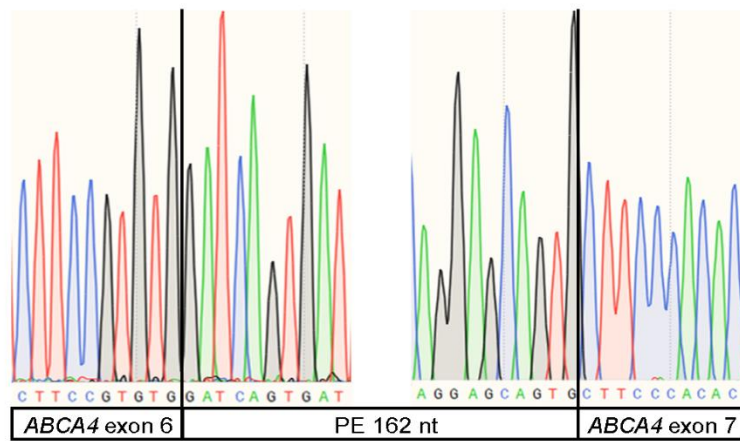
ii)



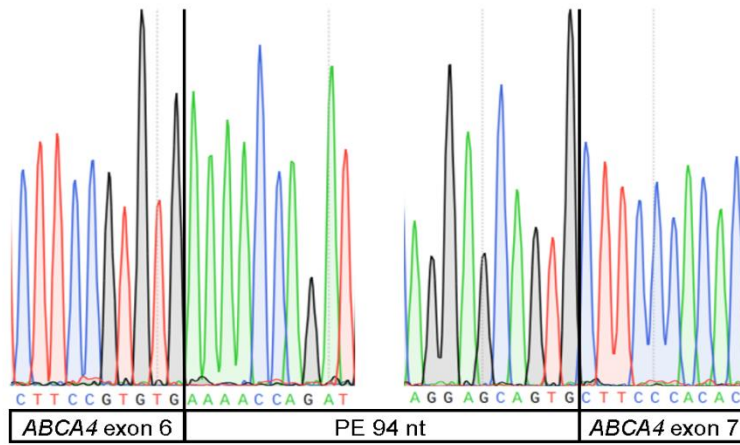
iii)



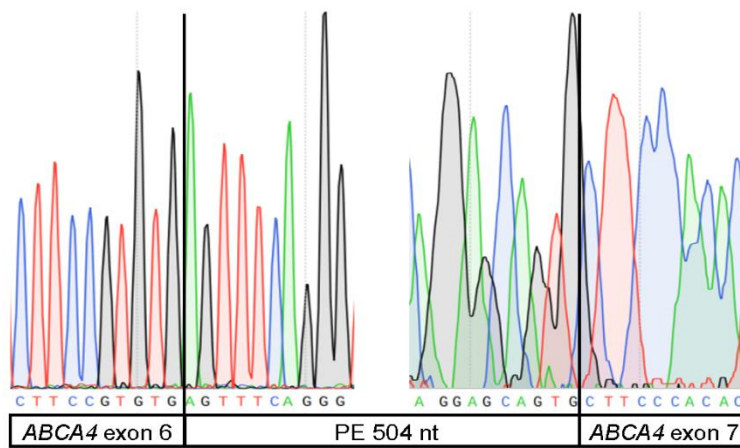
iv)



v)

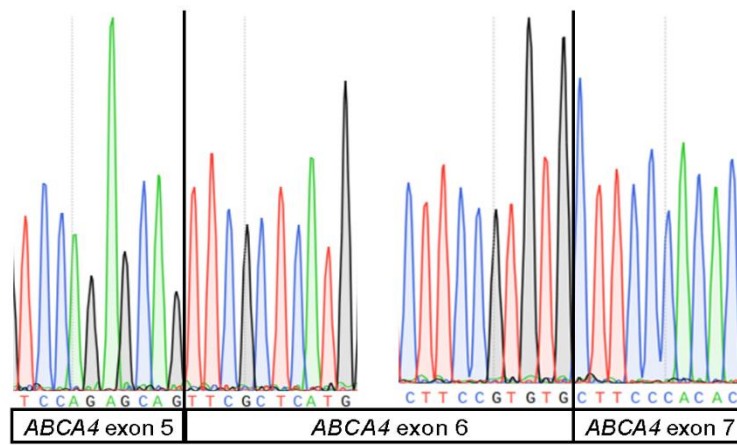


vi)

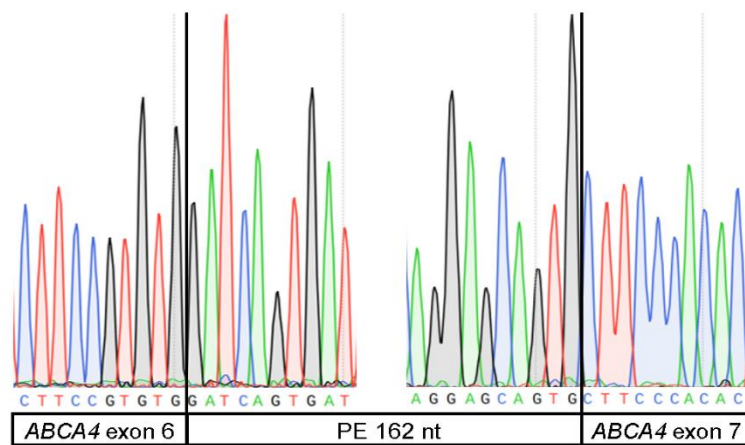


C. Sequencing results photoreceptor precursor cells

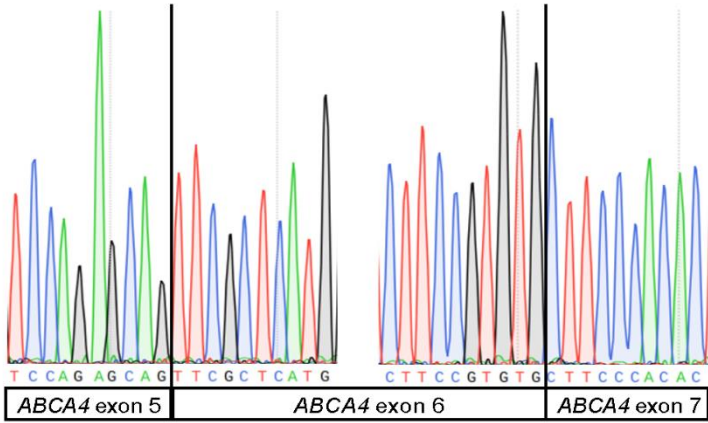
i)



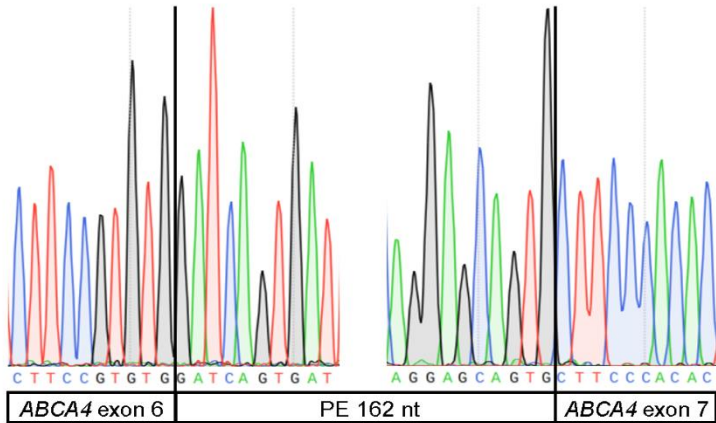
ii)



iii)

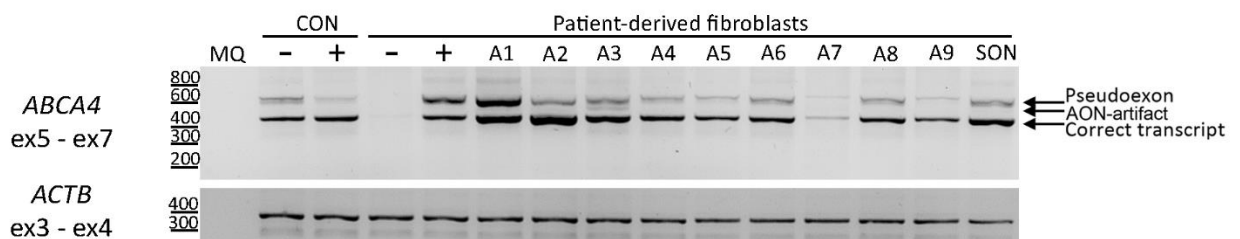


iv)

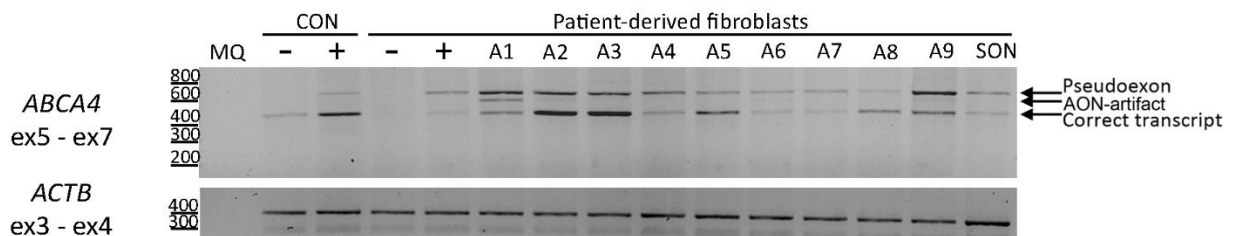


Supplementary Figure S2. AON-based rescue for the deep-intronic c.769-784C>T variant in patient-derived fibroblasts. The nine AONs and SON were tested in patient-derived fibroblasts. AONs and SON were transfected to patient-derived fibroblasts carrying the c.769-784C>T variant and were labelled from A1 to SON. Cycloheximide was used for cells labelled '+' and in all cells to which AONs and SON were transfected. The '-' indicates lack of cycloheximide treatment. The AON-not transfected control (CON) and patient-derived fibroblasts were used as a reference point for splicing-correction. MQ shows the negative control of the PCR reaction and amplification of exon 3 – exon 4 of *ACTB* gene was used as a loading control. **A)** In replicate 1 the PE band is stronger in the cycloheximide not treated control sample than in cycloheximide treated control. The PE band is weaker than expected in SON treated sample. AON-artifact is observed in sample treated with AON3. **B)** AON-artifact is observed in sample treated with AON1.

A. Antisense oligonucleotides screen in patient-derived fibroblasts, replicate 1

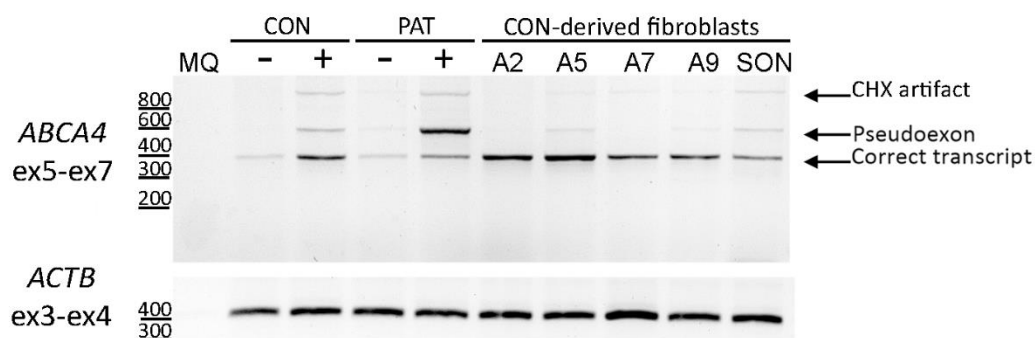


B. Antisense oligonucleotides screen in patient-derived fibroblasts, replicate 3

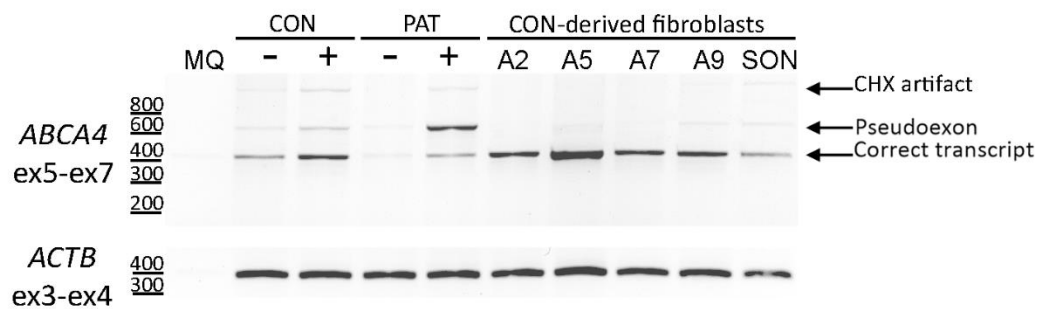


Supplementary Figure S3. AON-based rescue for the deep-intronic c.769-784C>T variant control-derived fibroblasts. A&B) Four AONs and SON were tested in control-derived fibroblasts. AONs and SON were transfected to control-derived fibroblasts and were labelled from A2, A5, A7, A9 and SON. Cycloheximide was used for cells labelled '+' and in all cells to which AONs and SON were transfected. The '-' indicates lack of cycloheximide treatment. The AON-not transfected control (CON) and patient-derived fibroblasts (PAT) were used as a reference point for splicing-correction. **C)** The graph represents the semi-quantification of the resulting RT-PCR products, showing the % of correct and pseudoexon (PE) inclusion transcript. Statistically significant PE exclusion was achieved for all four AONs. Milli-Q water (MQ) shows the negative control of the PCR reaction and amplification of exon 3 – exon 4 of *ACTB* gene was used as a loading control. Data (n=2) are presented as mean±SD and compared to the AON-untreated condition (*p<0.05, **p<0.01, ***p<0.001, ****p<0.0001).

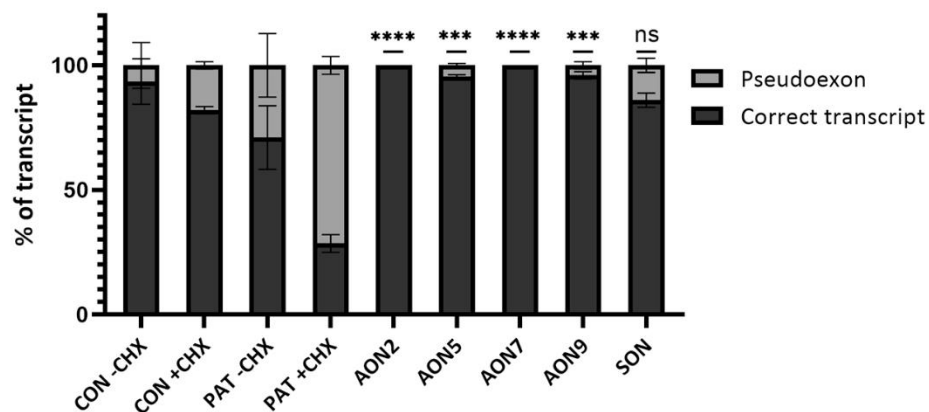
A. Replicate 1 of AONs screen in control fibroblasts



B. Replicate 2 of AONs screen in control fibroblasts

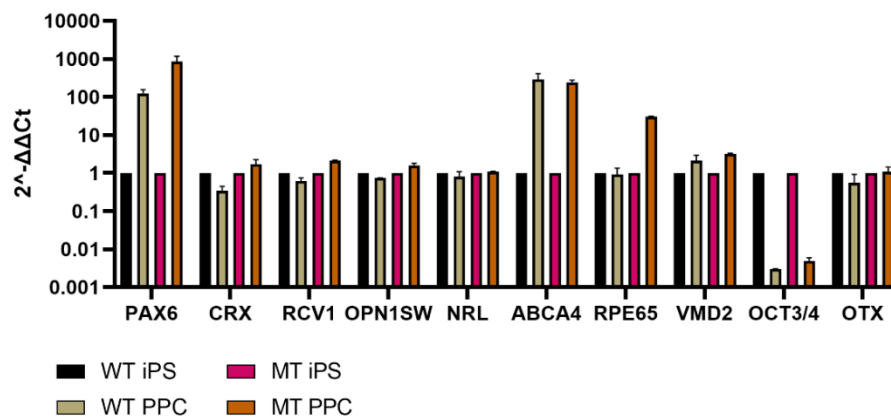


C. Average semi-quantification

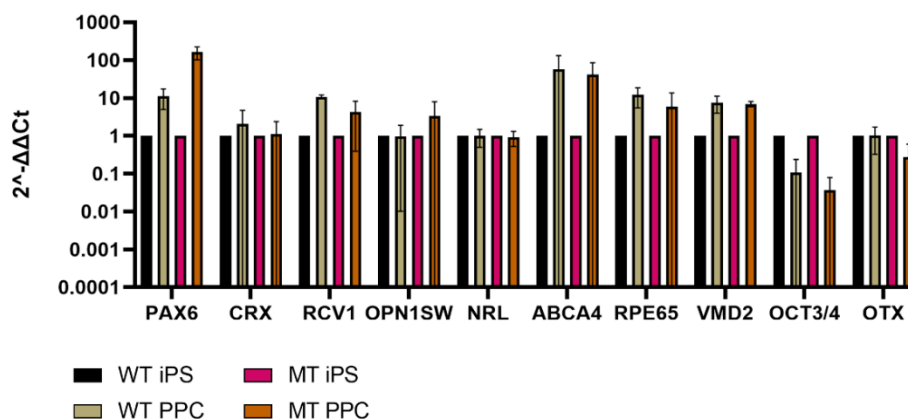


Supplementary Figure S4. Relative pluripotency and retinal gene expression profiles of photoreceptor precursor cells. Average gene expression profile of control iPSC differentiation for replicate 1 and 2. The abbreviations under each graph represents; wild-type induced pluripotent stem cells in black (WT iPS), wild-type photoreceptor precursor cells in grey (WT PPC), mutant carrying *ABCA4* c.769-784C>T variant in induced pluripotent stem cells in magenta (MT iPS) and mutant carrying *ABCA4* c.769-784C>T variant in photoreceptor precursor cells in orange (MT PPC) **(A)**. Strong increase in expression was observed in early retina marker such as *PAX6* and with retina specific genes such as *ABCA4*. The expression of pluripotency markers *OCT3/4* and *OTX* were decreased as the cells acquired more retina like phenotype. The remaining retinal gene markers had variable expression. **B)** Average gene expression profile of patient-derived differentiation for replicate 1 and 2 showing the relative gene expression of differentiated PPCs relative to iPSC. An increase in expression was observed in early retina marker such as *PAX6* and with retina specific genes such as *ABCA4*. Other retinal markers were also increased such as *RCV1*, *RPE65* and *VMD2*. The expression of pluripotency markers *OCT3/4* and *OTX* were decreased as the cells acquired more retina like phenotype. The remaining retinal gene markers had variable expression.

A. Pluripotency and retinal gene expression in *ABCA4* wild-type rep1&2



B. Pluripotency and retinal gene expression in *ABCA4* c.769-784C>T rep1&2



Supplementary Table S1. Primers for reverse transcription-PCR analysis and qPCR.

List of primers				
Forward Primer		Reverse Primer		Use
Gene	Sequence 5' -> 3'	Gene	Sequence 5' -> 3'	
<i>RHO</i> _exon3	CGGAGGTCAACAACGAGTCT	<i>ABCA4</i> exon7	CTTGAATTCTTGGTGACATATCAG	RT-PCR
<i>RHO</i> _exon5	ATCTGCTGCGGCAAGAAC	<i>RHO</i> _exon5	AGGTGTAGGGGATGGGAGAC	
<i>ABCA4</i> exon5	GGAATACGAATAAGGGATATCTTG	<i>ABCA4</i> exon7	CTTGAATTCTTGGTGACATATCAG	
<i>ACTB</i> _exon3	ACTGGGACGACATGGAGAAG	<i>ACTB</i> _exon4	TCTCAGCTGTGGTGGTGAAG	
<i>GUSB</i>	AGAGTGGTGCTGAGGATTGG	<i>GUSB</i>	CCCTCATGCTCTAGCGTGTC	qPCR
<i>ABCA4</i>	CATCCTGTTCCACCACCTCA	<i>ABCA4</i>	CTGTGTCCTCCAACATGGCT	
<i>CRX</i>	CCCCAGTGTGGATCTGATG	<i>CRX</i>	CAACAGTGCCTCCAGCTC	
<i>NRL</i>	GGCTCCACACCTTACAGCTC	<i>NRL</i>	AGCCAGTACAGCTCCTCCAG	
<i>OCT3/4</i>	GTTCTTCATTCACTAAGGAAGG	<i>OCT3/4</i>	CAAGAGCATCATTGAACCTCAC	
<i>OPN1SW</i>	TGGTATTGGCGTCTCCATC	<i>OPN1SW</i>	ACTCGCTGCGGTATTTGG	
<i>OTX2</i>	TATCTTAAGCAACCGCCTTACG	<i>OTX2</i>	GGAGGGGTGCAGCAAGTC	
<i>PAX6</i>	GCTGCAAAGAAATAGAACATCC	<i>PAX6</i>	TTGGCTGCTAGTCTTTCTCG	
<i>RCV1</i>	ACACCAAGTTCTCGGAGGAG	<i>RCV1</i>	ACTTGGCGTAGATGCTCTGG	
<i>RPE65</i>	TTACTACGCTTGACAGAGACC	<i>RPE65</i>	GCCCCATTGACAGAGACATAG	
<i>VMD2</i>	TCAGTGTGGACACCTGTATGC	<i>VMD2</i>	AAGCTGTACACCGCCACAG	

Supplementary Table S2. Averaged semi-quantification analysis of transcripts from RT-PCR products from HEK293T-midigene screen, control and patient-derived fibroblasts screen and control and patient-derived PPC screen. The quantity of the different transcripts is represented as an averaged percent (%) of the total transcript for each condition or lane. SD: standard deviation.

	HEK293T-midigene MT screen																							
Condition	WT		MT		AON1		AON2		AON3		AON4		AON5		AON6		AON7		AON8		AON9		SON	
	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD
Correct transcript	100%	0	79%	0.069	100%	0	100%	0	91%	0.053	90%	0.074	100%	0	100%	0	100%	0	93%	0.054	100%	0	73%	0.016
PE inclusion	0%	0	21%	0.069	0%	0	0%	0	9%	0.053	10%	0.074	0%	0	0%	0	0%	0	7%	0.054	0%	0	27%	0.016

	Patient-derived fibroblasts MT screen																												
Condition	CONT -CHX		CONT +CHX		PAT -CHX		PAT +CHX		AON1		AON2		AON3		AON4		AON5		AON6		AON7		AON8		AON9		SON		
	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	
Correct transcript	88	0.203335	87	0.023401	100	0	35	0.114734	40	0.132073	78	0.125521	73	0.05148	51	0.228015	80	0.065079	51	0.167247	64	0.303136	77	0.053773	69	0.290877	48	0.212768	
AON-artifact	0	0	0	0	0	0	0	0	9	0.107319	0	0	1	0.015389	1	0.014779	0	0	0	0	0	0	0	0	0	0	0	0	0
PE inclusion	12	0.203335	13	0.023401	0	0	65	0.114734	51	0.025342	22	0.125521	27	0.057593	48	0.223532	20	0.065079	49	0.167247	36	0.303136	23	0.053773	31	0.290877	52	0.212768	

Patient-derived fibroblasts WT screen																		
Condition	CONT -CHX		CONT +CHX		PAT -CHX		PAT +CHX		AON2		AON5		AON7		AON9		SON	
	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD
Correct transcript	93	0.093	81	0.004	43	0.298	27	0.038	100	0	95	0.001	100	0	96	0.023	87	0.016
PE inclusion	7	0.093	19	0.004	57	0.298	73	0.038	0	0	5	0.001	0	0	4	0.023	13	0.016

Patient-derived PPC WT screen																						
Condition	CONT -CHX		CONT +CHX		PAT -CHX		PAT +CHX		AON2@0.5 μM		AON2@1 μM		AON5@0.5 μM		AON5@1 μM		AON7@0.5 μM		AON7@1 μM		SON 1@μM	
	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD
Correct transcript	100%	0	95%	0.013	85%	0.02	70%	0.003	97%	0.002	97%	0.015	97%	0.003	95%	0.006	97%	0.002	97%	0.004	96%	0.008
PE inclusion	0%	0	5%	0.013	15%	0.02	30%	0.003	3%	0.002	3%	0.015	3%	0.003	5%	0.006	3%	0.002	3%	0.004	4%	0.008

Patient-derived PPC MT screen																						
Condition	CONT -CHX		CONT +CHX		PAT -CHX		PAT +CHX		AON2@0.5 μM		AON2@1 μM		AON5@0.5 μM		AON5@1 μM		AON7@0.5 μM		AON7@1 μM		SON 1@μM	
	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD	%	SD
Correct transcript	100%	0	97%	0.015	84%	0.056	66%	0.058	95%	0.024	92%	0.034	79%	0.009	81%	0.077	95%	0.017	97%	0.005	74%	0.025
PE inclusion	0%	0	3%	0.015	16%	0.056	34%	0.058	5%	0.024	8%	0.034	21%	0.009	19%	0.077	5%	0.17	3%	0.005	26%	0.025